

PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.

<http://hdl.handle.net/2066/182152>

Please be advised that this information was generated on 2020-11-29 and may be subject to change.

Article 25fa pilot End User Agreement

This publication is distributed under the terms of Article 25fa of the Dutch Copyright Act (Auteurswet) with explicit consent by the author. Dutch law entitles the maker of a short scientific work funded either wholly or partially by Dutch public funds to make that work publicly available for no consideration following a reasonable period of time after the work was first published, provided that clear reference is made to the source of the first publication of the work.

This publication is distributed under The Association of Universities in the Netherlands (VSNU) 'Article 25fa implementation' pilot project. In this pilot research outputs of researchers employed by Dutch Universities that comply with the legal requirements of Article 25fa of the Dutch Copyright Act are distributed online and free of cost or other barriers in institutional repositories. Research outputs are distributed six months after their first online publication in the original published version and with proper attribution to the source of the original publication.

You are permitted to download and use the publication for personal purposes. All rights remain with the author(s) and/or copyrights owner(s) of this work. Any use of the publication other than authorised under this licence or copyright law is prohibited.

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please contact the Library through email: copyright@ubn.ru.nl, or send a letter to:

University Library
Radboud University
Copyright Information Point
PO Box 9100
6500 HA Nijmegen

You will be contacted as soon as possible.



Review

Pathways towards indolent B-cell lymphoma — Etiology and therapeutic strategies



Michiel van den Brand^{a,b,*}, Blanca Scheijen^a, Corine J. Hess^c, J. Han JM van Krieken^a,
Patricia J.T.A. Groenen^a

^a Department of Pathology, Radboud university medical center, Geert Grooteplein Zuid 10, 6525GA Nijmegen, The Netherlands

^b Pathology-DNA, location Rijnstate, Wagnerlaan 55, 6815AD Arnhem, The Netherlands

^c Department of Hematology, Radboud university medical center, Geert Grooteplein Zuid 10, 6525GA Nijmegen, The Netherlands

ARTICLE INFO

Keywords:

Indolent B-cell lymphoma
Chronic lymphocytic leukemia
Marginal zone lymphoma
Follicular lymphoma
Etiology

ABSTRACT

Although patients with indolent B-cell lymphomas have a relatively good survival rate, conventional chemotherapy is not curative. Disease courses are typically characterized by multiple relapses and progressively shorter response duration with subsequent lines of therapy. There has been an explosion of innovative targeted agents in the past years. This review discusses current knowledge on the etiology of indolent B-cell lymphomas with respect to the role of micro-organisms, auto-immune diseases, and deregulated pathways caused by mutations. In particular, knowledge on the mutational landscape of indolent B-cell lymphomas has strongly increased in recent years and harbors great promise for more accurate decision making in the current wide range of therapeutic options. Despite this promise, only in chronic lymphocytic leukemia the detection of *TP53* mutations and/or del17p currently have a direct effect on treatment decisions. Nevertheless, it is expected that in the near future the role of genetic testing will increase for prediction of response to targeted treatment as well as for more accurate prediction of prognosis in indolent B-cell lymphomas.

1. Introduction

Indolent B-cell lymphomas represent approximately one third of all B-cell non-Hodgkin lymphomas. Although patients with these types of lymphoma have a relatively good survival rate, morbidity and mortality are significant, especially considering the fact the majority of these lymphomas is considered incurable. This review discusses recent developments in the etiology of indolent B-cell lymphomas which can provide a rationale for prioritizing and combining novel agents.

In form and function, cells of indolent B-cell lymphomas resemble their normal counterparts and act in close contact with and are dependent on their micro-environment. This dependence on the micro-environment is evidenced by the difficulty to culture indolent B-cell lymphoma cells which indicates that these cells cannot proliferate autonomously but require stimulatory signals. Examples of signals that can stimulate the development of B-cell lymphoma include long-standing inflammatory conditions in the context of infection or auto-immune disease. Another line of evidence for the role of antigenic stimulation of lymphomas comes from the fact that in multiple types of B-cell lymphoma, stereotyped B-cell receptors have been shown,

indicating that specific antigens are implicated in lymphomagenesis. The first part of this review is concerned with these processes driving the development of B-cell lymphomas.

A second issue is: what makes neoplastic B-cells different from normal B-cells? Which genetic events are present? It is already known for a long time that the early genetic alterations like the t(14;18) in follicular lymphoma are actually a common occurrence in healthy individuals and thus not sufficient to make a lymphocyte malignant. The genetic revolution is rapidly changing our understanding of B-cell lymphomas and in the second part of this review, the pathways and mutations involved in B-cell lymphomagenesis and the drugs that target them are discussed.

2. Micro-organisms

Multiple different species of bacteria have been implicated in the development of extranodal marginal zone lymphoma of mucosa-associated lymphoid tissues (EMZL) with *Helicobacter pylori* being the best studied and most convincing example (Table 1). *H. pylori* can be detected in up to 92% of EMZL of the stomach and eradication of the

* Corresponding author at: Department of Pathology, Radboud University Medical Center, Geert Grooteplein Zuid 10, 6525GA Nijmegen, The Netherlands.

E-mail addresses: Michiel.vandenBrand@radboudumc.nl (M. van den Brand), Blanca.Scheijen@radboudumc.nl (B. Scheijen), Corine.Hess@radboudumc.nl (C.J. Hess), Han.vanKrieken@radboudumc.nl (J.H.J. van Krieken), Patricia.Groenen@radboudumc.nl (P.J.T.A. Groenen).

<http://dx.doi.org/10.1016/j.blre.2017.08.002>

Table 1
Micro-organisms in indolent B-cell lymphomas.

Micro-organism	Related lymphoma	Summary of association
<i>Helicobacter pylori</i>	MALT lymphoma of the stomach	Strong evidence for association: <ul style="list-style-type: none"> • <i>H. pylori</i> present in up to 92% of MALT lymphomas. • Eradication results in long-term remission in three quarters of patients with early-stage disease. • Translocations in <i>BIRC3</i>, <i>MALT1</i> and/or <i>BCL10</i> predict resistance to eradication treatment.
<i>Chlamydomydia psittaci</i>	Ocular adnexal MALT lymphoma	Some evidence for association <ul style="list-style-type: none"> • <i>C. psittaci</i> present in subset of OAMZL, varying between regions and with detection methods. • Response to antibiotic treatment in a subset of patients with <i>C. psittaci</i> positive lymphoma, but also in patients with <i>C. psittaci</i> negative lymphoma.
<i>Borrelia burgdorferi</i>	Cutaneous MALT lymphoma	Some evidence for association <ul style="list-style-type: none"> • <i>B. burgdorferi</i> DNA present in a subset of cutaneous MALT lymphoma. • Anecdotal reports of lymphoma response to antibiotic treatment.
<i>Campylobacter jejuni</i>	Immunoproliferative small intestinal disease (IPSID)	Anecdotal reports of association and lymphoma response to antibiotics.
<i>Achromobacter xylosoxidans</i>	Pulmonary MALT lymphoma	<i>A. xylosoxidans</i> DNA detected more frequently in pulmonary MALT lymphoma than control tissue in a single study.
Epstein Barr virus	No specific type	Only very rarely reported in low-grade B-cell lymphoma
Hepatitis C virus	MALT lymphoma (particularly non-gastric), SMZL, NMZL, LPL, FL	Strong evidence for association <ul style="list-style-type: none"> • Co-occurrence of HCV infection and lymphoma, with strong geographical variation. • Regression of lymphoma with antiviral therapy in 62–100% of patients.

bacterium by antibiotic treatment results in complete and long-lasting remission of lymphoma in three quarters of patients with early-stage disease [1–5]. In EMZL of the stomach but also of the ocular adnexa (discussed below), the micro-organisms involved do not stimulate the neoplastic B-cells directly, but rather cause an inflammatory milieu with induction of a Th1 immune response with influx of CD4 positive T-cells which stimulate B-cell proliferation through CD40-CD40L interaction. At the same time, the cytotoxic T-cell response is impaired [6]. These factors lead to the outgrowth of autoreactive B-cells which can progress to lymphoma after the acquisition of mutations. With respect to gastric EMZL, most regress after removing the inflammatory stimulus by eradication of *H. pylori*, but some gastric EMZLs have become *H. pylori* independent. Factors associated with the lack of a response to *H. pylori* eradication include a higher disease stage and the presence of specific translocations involving the *BIRC3*, *MALT1*, or *BCL10* gene [7–9]. In both instances, it is suspected that the lymphoma cells have already become independent of the antigenic stimulus. However, even in diffuse large B-cell lymphoma (DLBCL) of the stomach, *H. pylori* eradication has been reported to result in long-term remission in up to two-thirds of patients, indicating that high-grade lymphomas may also be antigen-dependent [10,11].

A role for *Chlamydomydia psittaci* has been suggested in the pathogenesis of ocular adnexal EMZL (OAMZL), following from the observation that it can be detected in up to 80% of OAMZL [12,13] and that antibiotic treatment results in remission in a subset of patients [13]. However, the presence of *C. psittaci* in OAMZL varies greatly between studies, which can at least partially be explained by geographical differences [12–22]. In addition, differences in detection protocols for *C. psittaci* could also be part of the explanation as studies from similar regions (e.g. Italy, the Netherlands, United States) have showed strong differences in *C. psittaci* incidence in OAMZL.

With respect to treatment, most studies showed partial or complete responses in approximately 45% of patients with OAMZL treated with doxycycline antibiotics [23]. However, only two of three studies showed improved response in *C. psittaci*-positive cases. In those two studies, the response rates were 66% vs. 50% and 64% vs. 38% for *C. psittaci* positive vs. negative patients [24,25]. These results suggest that although *C. psittaci* may have a role in the development of OAMZL and is amenable to antibiotic treatment, other (Doxycycline sensitive) bacteria or direct anti-tumor effects of the antibiotics used may also

have a role in eradication of the lymphoma. Conversely, detection methods may be insufficient. In conclusion, the role of antibiotic treatment in OAMZL is speculative at best and requires additional study.

In the rare cutaneous EMZL, *Borrelia burgdorferi* has been suggested as a causative agent, with varying frequencies depending on the region where the studies were performed. In endemic areas, up to 40% of cutaneous EMZL showed evidence of *Borrelia* infection [26–28]. Anecdotal reports have shown tumor response to antibiotic therapy against *Borrelia* [23].

In immunoproliferative small intestinal disease (IPSID), *Campylobacter jejuni* has been reported as a possible causative micro-organism [29,30]. However, the evidence for this association is limited to anecdotal reports and no clinical studies have been performed. In pulmonary EMZL, *Achromobacter xylosoxidans* has been detected with increased frequency in lymphomas in comparison to controls, but these results are limited to a single study and have not been validated clinically [31]. Finally, regarding the role of bacteria in the pathogenesis of indolent lymphoma, epidemiological studies have shown an association between an increased risk of CLL and a history of pneumonia [32]. In primary intraocular B-cell lymphoma, the presence of *Toxoplasma gondii* has been reported [33].

Co-occurrence of Hepatitis C virus (HCV) infection and lymphoma has been reported in multiple types of non-Hodgkin lymphoma, including indolent B-cell lymphomas. Within the group of indolent B-cell lymphomas, lymphoplasmacytic lymphoma and marginal zone lymphoma (MZL, particularly splenic MZL and non-gastric MALT lymphoma) are most often associated with HCV infection. However, the association between B-cell lymphoma and HCV has varied significantly between studies, with stronger associations in those areas with a higher HCV prevalence. In systematic reviews, 13%–18% of B-cell lymphomas were HCV-associated [34–36]. In addition to epidemiological data, regression of lymphoma after eradication of HCV with antiviral therapy provides further evidence for a role of HCV in B-cell lymphoma development. Response of lymphoma to antiviral treatment has been recorded in extranodal, splenic, and nodal MZL and in smaller numbers of lymphoplasmacytic lymphoma (LPL) and follicular lymphoma (FL) [37–44]. The studies reported thus far with a total number of almost 200 patients, reported an overall response rate of 62–100% and complete response rate of 50–89% [45]. Accordingly, antiviral treatment

can be considered as first-line treatment in patients with indolent B-cell lymphoma in whom immediate antilymphoma therapy is not necessary due to limited stage of disease or low tumor burden. Novel agents against HCV (e.g. sofosbuvir, simeprevir) are also promising in the setting of lymphoma arising in patients with HCV. In two recent case reports, regression of SMZL was reported after interferon free HCV treatment [46,47], but formal studies have not been reported yet. The mechanisms by which HCV induces lymphoma are not well understood, but possibilities include chronic antigenic stimulation and/or a direct oncogenic effect of the virus [36].

3. Stereotyped B-cell receptors

Although micro-organisms have been implicated in the development of B-cell lymphoma, the neoplastic B-cells usually do not have BCRs that recognize antigens belonging to these micro-organisms. Therefore, infections appear to contribute mainly by forming a pro-inflammatory environment rather than directly stimulating the neoplastic cells. The question then arises to what extent antigen stimulation is necessary for B-cell lymphoma development and which antigens are involved. With respect to the first part of this question, several lines of evidence argue for a role of antigens in lymphomagenesis.

First, the BCR is retained in the majority of B-cell lymphomas and most lymphoma cells cannot survive autonomously *ex vivo*. This suggests a dependence on the micro-environment and on signaling through the BCR [48]. Second, studies into the structure of the B-cell receptor have revealed stereotyped B-cell receptors in different types of B-cell lymphoma.

In chronic lymphocytic leukemia (CLL), multiple subsets with specific stereotyped receptors have been recognized, with approximately one third of CLLs having a stereotypic BCR [49]. Part of these stereotypes are associated with a specific clinical presentation or prognosis. In splenic MZL, about one third of lymphomas express the same immunoglobulin heavy chain variable gene. The fact that the BCRs in different lymphomas show a similar IGH structure suggests selection of tumor cells by specific antigens or superantigens [50].

If B-cell lymphomas are dependent on antigens for their survival, then which antigens are responsible? In both SMZL and CLL, it has been shown that the BCRs of the neoplastic cells are often reactive to self-antigens, including antigens which are expressed during cell death [51–55]. In addition, reactivity to viral bacterial and fungal antigens has also been suggested, but the bacteria implicated are different from those previously discussed in the context of MALT lymphoma. In CLL, the neoplastic cells can express BCRs that react with epitopes on common bacteria [56]. Also in CLL, an association between usage of the immunoglobulin heavy chain variable 4-34 (IGH4-34) gene and activation of EBV and CMV, suggesting that viral antigens could facilitate clonal expansion and neoplastic transformation [57].

4. Auto-immune diseases

Auto-immune diseases are associated with an increased risk of lymphoma [58]. In particular Sjögren's syndrome and systemic lupus erythematosus were associated with an increased risk of lymphoma in a large pooled analysis [59]. With respect to indolent B-cell lymphomas in the context of auto-immune disease, development of MZL in Sjögren's syndrome is the most striking example. Patients with Sjögren's syndrome have a 7-fold increased risk of non-Hodgkin lymphoma and a 1000-fold increased risk of MZL of the parotid gland [59]. Clinical and laboratory features that predict lymphoma development include permanent swelling of the salivary glands, oral and ocular symptoms, vasculitis, lymphadenopathy, purpura, cryoglobulinemia, lymphopenia, low C4, M-protein and germinal center-like structures in the salivary gland.

With respect to the pathophysiology of lymphoma development in Sjögren's syndrome, it has been suggested that the B-cell clone often has

a B-cell receptor with rheumatoid factor (RF) activity, being directed against the Fc portion of IgG [60–63]. The hypothesis is then that polyclonal B-cells which are activated in the auto-immune inflammatory environment acquire additional mutations and eventually grow out to become a B-cell lymphoma [62].

5. Preneoplastic conditions

Much has been learned about the pathogenesis of indolent lymphomas from precursor lesions, which include precursors of FL and monoclonal B-cell lymphocytosis (MBL).

Deregulation of BCL2 by the t(14;18) translocation is the hallmark genetic lesion of FL. However, this translocation in itself is insufficient for neoplastic transformation. This is well illustrated by studies into FL and its precursors. With highly sensitive techniques, cells harboring the t(14;18) translocation can be detected in over half of the healthy adult population, of which only a very small minority will develop FL [64,65]. These cells carrying the t(14;18) are antigen experienced and it is hypothesized that multiple re-entries in the germinal center with aberrant somatic hypermutation induce mutations, thereby contributing to oncogenesis. *In situ* follicular neoplasia (ISFN, also termed follicular lymphoma *in situ*) is a coincidentally detected phenomenon in lymphoid tissue in which BCL2 and CD10 positive cells colonize germinal centers without architectural distortion. The cells in these follicles have a t(14;18) translocation, but also carry additional genomic alterations [66], suggesting that ISFN could represent a step in between the t(14;18) positive cells detected in healthy individuals and overt FL. Only approximately 5% of individuals with ISFN have developed FL in small studies [67].

MBL is the presence of a monoclonal or oligoclonal B-cell expansion with a B-cell count of $< 5 \times 10^9$ B-cells/L in the absence of symptoms of an overt lymphoproliferative disease. Most cases of MBL (approximately 75%) have an immunophenotype resembling CLL, but other phenotypes are also encountered. Blood samples collected before a diagnosis of CLL consistently show a monoclonal B-cell population, indicating that CLL is preceded by MBL [68]. However, the large majority of MBLs do not progress to CLL. MBL with a low number of clonal B-cells in the peripheral blood ($< 0.5 \times 10^9$ /L), termed 'low count' MBL, virtually never progress to CLL whereas 'high count' MBL progresses to CLL at a rate of 1–2% per year and requires follow-up [69,70]. With respect to the immunogenetic profile, low-count MBL is different from CLL with more frequently mutated IGHV genes and a very low prevalence of BCR stereotypy in low-count MBL, whereas high-count MBL resembles CLL with respect to IGHV mutation status and BCR stereotypy [71]. On the contrary, cytogenetic aberrations found in CLL can also be found in low-count MBL, indicating that these lesions are not necessarily associated with progression [72]. High count MBL and low-stage CLL show similar mutational patterns, further indicating that these conditions are part of the same spectrum [73]. Low-count MBL appears to be a different entity which might be related to aging [71].

6. Deregulated pathways

Recent large sequencing efforts in indolent B-cell lymphomas have substantially increased our knowledge on the molecular pathways that are deregulated in these lymphomas. With the concurrent development of drugs that target these specific pathways, the role of targeted treatment for indolent B-cell lymphoma is likely to increase (Fig. 1). Although some indolent B-cell lymphomas are characterized by highly recurrent mutations, most lymphoma subtypes show a low number of genes with a moderate mutation frequency followed by a long tail of genes that are mutated in a low number of cases. The pathways that are most frequently affected are discussed below and summarized in Table 2.

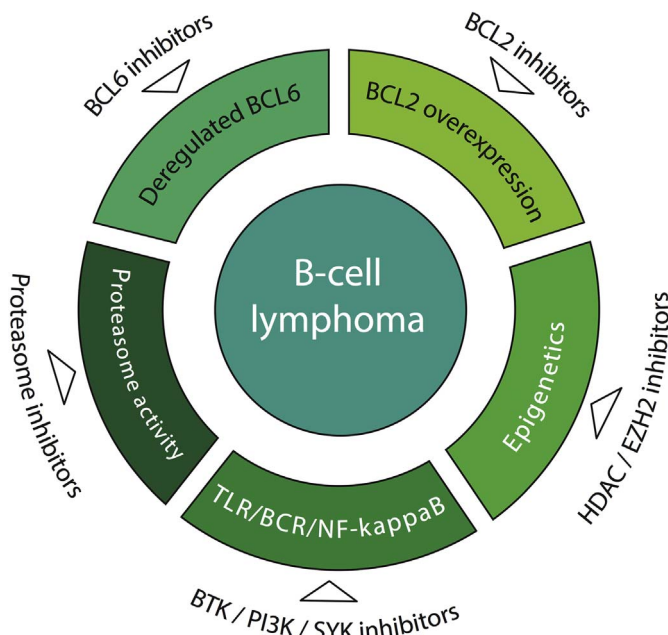


Fig. 1. Deregulated pathways in B-cell lymphoma with targeted treatments.

6.1. DNA damage response and apoptosis (TP53, BCL2, and ATM)

6.1.1. TP53

TP53 is one of the most extensively studied tumor suppressor genes and is well known for its association with a poor prognosis and refractoriness to conventional chemotherapy in CLL. In fact, TP53 mutations and del17p are the only mutations that currently have a direct impact on patient care, with a preferred treatment with novel inhibitors (ibrutinib as single agent or idelalisib combined with anti CD20 treatment) in patients with TP53 mutated CLL with a treatment indication [74,75]. The current standard method of TP53 mutation detection is Sanger sequencing and a for next-generation sequencing a cut-off of mutations present in > 10% allelic fraction is recommended by the European Research Initiative on CLL (www.ericll.org). However, small

TP53 mutated subclones are also associated with a worse prognosis and become predominant under conventional treatment, ultimately causing treatment refractoriness [76]. Therefore, low frequency TP53 mutations in CLL, which can be detected by sensitive next generation sequencing techniques, may be of clinical relevance, and should be reported with caution until validation of their prognostic and predictive impact in a larger patient group. Whether treatment with for instance BTK inhibitors will provide similar benefit as it does in patients with > 10% allelic fraction remains to be elucidated. In addition to CLL, an association between TP53 mutations and a worse prognosis has also been shown for FL and MZL [77,78].

6.1.2. BCL2

BCL2 is an anti-apoptotic protein which was initially discovered as the deregulated gene involved in the t(14;18) translocation in FL [79]. In the physiological situation, it is part of the balance between pro-apoptotic and anti-apoptotic proteins in the intrinsic/mitochondrial apoptotic pathway. In the large majority of FLs, overexpression of BCL2 can be explained by the t(14;18) translocation which puts BCL2 under the control of the IGH enhancer. More rarely, translocations between BCL2 and the immunoglobulin light chain loci are encountered. However, many small B-cell lymphomas other than FL express BCL2 without evidence of a BCL2 rearrangement. This can be explained by other mechanisms of BCL2 expression, especially physiological expression, but also BCL2 amplification, hypomethylation, or deletion of micro-RNAs that downregulate BCL2 [80–83].

BCL2 is a target for BCL2 inhibitors venetoclax and navitoclax which are currently under investigation for their effectiveness in multiple lymphoma subtypes. In Europe, the European Medicines Agency (EMA) has approved venetoclax for the treatment of CLL with 17p deletion or TP53 mutation in adult patients who are unsuitable for or have failed a B-cell receptor pathway inhibitor and also for the treatment of CLL in the absence of 17p deletion or TP53 mutation in adult patients who have failed both chemoimmunotherapy and a B-cell receptor pathway inhibitor. In the United States, venetoclax has been approved by the Food and Drug Administration (FDA) for treatment of patients with CLL with a 17p deletion who have received at least one prior therapy.

Table 2 Mutated genes and their relevance in indolent B-cell lymphomas.

Lymphoma subtype	Mutated gene	Relevance of mutation present
Chronic lymphocytic leukemia/small lymphocytic lymphoma	<u>TP53</u>	Associated with worse prognosis. Treatment with novel inhibitors (ibrutinib, idelalisib)
	<u>NOTCH1</u>	Associated with worse prognosis and rituximab resistance
	<u>BIRC3</u>	Associated with worse prognosis
	<u>SF3B1</u>	Associated with worse prognosis
	<u>ATM</u>	Associated with worse prognosis
Follicular lymphoma	<u>BCL2</u>	Translocation specific for FL. Mutations associated with disease progression
	<u>MLL2, EZH2, EP300, MEF2B</u>	Histone-modifying genes frequently affected in FL
	<u>CARD11, CD79A/B, PRKCB</u>	NF-kappaB activating mutations, present in one third of follicular lymphomas.
	<u>TP53</u>	Associated with worse prognosis
	<u>TNFRSF14</u>	Affected by loss of 1p36 or mutations
	<u>BIRC3, MALT1, BCL10</u>	Involved in translocations activating NF-kappaB and predicting resistance to H. pylori eradication in gastric MALT lymphoma
MALT lymphoma	<u>TNFAIP3</u>	Frequently mutated NF-kappaB inhibitor
	<u>NOTCH2</u>	Mutated in 10–25% of SMZLs, master regulator of marginal zone differentiation
Splenic marginal zone lymphoma	<u>KLF2</u>	Mutated in 20–40% of SMZLs, causing NF-kappaB activation. Associated with short time to first treatment.
	<u>TNFAIP3, MYD88, TRAF3</u>	NF-kappaB activating mutations
	<u>CARD11</u>	
Lymphoplasmacytic lymphoma	<u>TP53</u>	Associated with short overall survival
	<u>MYD88</u>	Mutated in > 90% of LPLs. In LPL (but not DLBCL) associated with response to ibrutinib
Nodal marginal zone lymphoma	<u>CXCR4</u>	Mutations in 30% of LPLs, associated with ibrutinib resistance
	Recurrent mutations reported in <u>MLL2, PTPRD, NOTCH2, KLF2, TNFAIP3</u> and <u>CD79B</u>	

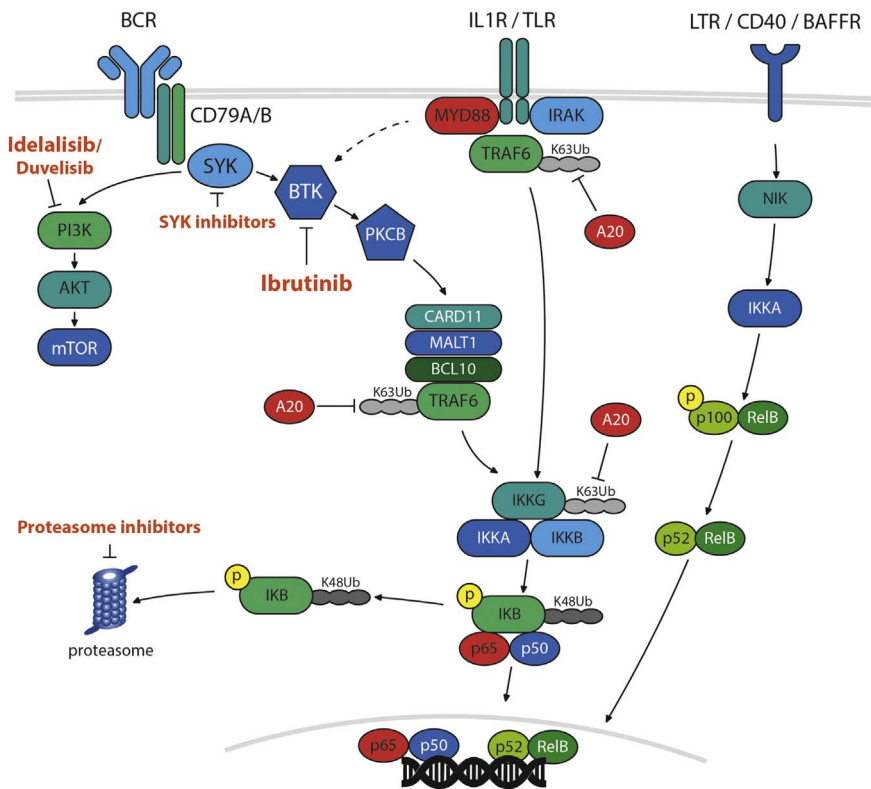


Fig. 2. Overview of BCR/TLR/NF-kappaB signaling with targeted treatments. Non-canonical signaling via LTR/CD40/BAFFR is indicated on the right. Canonical NF-kappaB signaling is indicated on the left. Activation of BCR signaling induces NF-kappaB activation via SYK, BTK, and PKCB. In addition, BCR signaling activates the PI3K pathway. IL1R and TLR signaling also induce NF-kappaB activation. In lymphoplasmacytic lymphoma, mutated MYD88 causes BTK activation. Ibrutinib, SYK inhibitors and idelalisib/duvelisib target different steps in these signaling cascades. Proteasome inhibitors decrease breakdown of inhibitors of NF-kappaB, causing decreased NF-kappaB signaling.

Abbreviations: BAFFR: B-cell activating factor receptor; BCR: B-cell receptor; IL1R: interleukin-1 receptor; LTR: lymphotoxin B receptor; NF-kappaB: nuclear factor kappa B; TLR: Toll-like receptor.

6.1.3. ATM

Inactivation of *ATM* (located at 11q22) by deletion and/or mutation is a frequent event in CLL with deletions of 11q22-23 in < 10% in newly diagnosed CLLs, rising to approximately 20% at the time of first treatment. Mutations in *ATM* are present in 10–15% of CLL at first diagnosis and approximately 15% at the time of first treatment [75]. Together, genetic lesions are present in *ATM* in CLL in approximately 20% at the time of diagnosis and approximately 35% at the time of first treatment [84]. The *ATM* gene encode for a serine/threonine kinase which prevents cell cycle progression and activated DNA repair mechanisms in the event of chromosomal double strand breaks.

6.2. B-cell receptor/toll-like receptor/NF-kappaB signaling

Signals from multiple receptors converge on the NF-kappaB pathway (Fig. 2). Canonical and non-canonical NF-kappaB signaling are distinguished. Canonical NF-kappaB signaling starts from the B-cell receptor, T-cell receptor, Toll-like receptors (TLR), interleukin-1 receptor (IL1R) or tumor necrosis factor receptor (TNFR). Through multiple steps which include the proteins CD79A/B, SYK, BTK, CARD11, BCL10, TRAF6 and MYD88, the inhibitor of kappa-B kinase (IKK) complex is activated. Activation of this complex results in the removal of inhibitory elements from NF-kappaB proteins, allowing these proteins to enter the nucleus to regulate transcription. NF-kappaB signaling is negatively regulated by deubiquitinases which include CYLD (cylindromatosis) and TNFAIP3 (also known as A20).

The non-canonical pathway is activated by signaling from the lymphotoxin B receptor, B-cell activating factor receptor (BAFFR), and CD40. This activates NF-kappaB inducible kinase (NIK, also known as MAP3K14) and CHUK, also eventually resulting in translocation of NF-kappaB proteins to the nucleus.

Deregulation of pathways that converge on NF-kappaB is frequently observed in lymphomas. In indolent B-cell lymphomas, extranodal marginal zone lymphoma is the most prominent example with frequent translocations involving *MALT1*, *BIRC3* (also known as *API2*), and *BCL10* [85–89]. In addition, extranodal MZLs harbor frequent

mutations in genes involved in NF-kappaB signaling including *TNFAIP3* [90]. Also in SMZL and NMZL, mutations in genes involved in NF-kappaB signaling are frequent and most notably include *TNFAIP3*, *IKKB*, *CARD11*, *BIRC3* and rarely *MYD88* [78,91–95]. *MYD88* mutations are present in the large majority of LPLs, which is very helpful for diagnosis [93,96–98]. Almost all mutations affect the TIR domain with NF-kappaB activation as a result. In a subset of CLL/SLL, mutations in *BIRC3* are detected with enrichment in fludarabine-refractory cases [99]. In patients with *TP53* mutated CLL, additional mutations in *BIRC3* are associated with an even worse prognosis. *MYD88* mutations have been found in a small subset of CLL/SLLs, mainly in cases with mutated IGHV [98].

In FL, mutations in genes involved in BCR/NF-kappaB signaling are relatively frequent with mutations in *CARD11*, *TNFAIP3*, *CD79A/B*, and/or *PRKCB* being present in a third of FLs [100]. Interestingly, mutations in NF-kappaB genes were gained at transformation [100]. Mutations in *TNFRSF14* are also frequently present in FL, being reported in approximately one third of cases [100–102]. TNFRSF14 is a putative tumor suppressor which can signal through the NF-kappaB pathway after binding of one of its ligands which include LIGHT, BTLA or CD160 [103]. However, as loss-of-function mutations in *TNFRSF14* are observed in FL, these mutations would cause a decrease rather than an increase of NF-kappaB activation. The tumor suppressor effect of TNFRSF14 can be explained by the fact that in the unmutated situation, triggering of TNFRSF14 by its ligand LIGHT increases the sensitivity to FAS-induced apoptosis. Disruption of TNFRSF14 would then make cells less sensitive to apoptosis, contributing to lymphomagenesis [104,105].

6.2.1. B-cell receptor signaling inhibitors

Multiple drugs have recently become available that targets signaling to NF-kappaB (Fig. 2). Ibrutinib blocks this signaling by binding covalently to BTK at cysteine 481, thereby preventing phosphorylation and blocking of downstream signaling [106,107]. Ibrutinib has been approved by the FDA for treatment of patients with CLL, recurrent MCL, Waldenström macroglobulinemia and recently also marginal zone lymphoma. In Europe, its use has been approved for patients with CLL,

recurrent MCL, and Waldenström macroglobulinemia. In addition, efficacy of ibrutinib has been shown in DLBCL, mainly in the activated B-cell-like (ABC) subtype [108,109]. However, not all patients show a response to ibrutinib and both primary and secondary mechanisms of resistance have been identified [108]. Primary resistance mechanisms have been mainly studied in MCL and DLBCL and include mutations that cause activation the non-canonical NF-kappaB pathway (e.g. *TRAF2* and *TRAF3*), mutations that activate the canonical NF-kappaB pathway independent of *BTK* (*MYD88* mutations) [109,110]. In lymphoplasmacytic lymphoma (LPL), *MYD88* mutations predict response rather than resistance to ibrutinib, which is in line with experiments showing BTK phosphorylation after mutant MYD88 overexpression in LPL [111]. MYD88^{L265P} mutations are present in the large majority (> 90%) of lymphoplasmacytic lymphomas [96,112,113], and patients with *MYD88* mutated LPL show a major response in 91% versus only 29% in patients with *MYD88* wildtype LPL [114]. Mutations in *CXCR4* are associated with a decreased response to ibrutinib in LPL with a major response rate of 91% vs. 62% for patients with LPL without and with *CXCR4* mutations. This might be caused by activation of AKT signaling by *CXCR4* mutations in which the mechanism might be similar to that in MCL as discussed above [115,116]. CXCR4 is a G-protein coupled chemokine receptor with a role in lymphocyte homing and migration. *CXCR4* mutations of WHIM-like type are present in 30% of LPLs. WHIM-like describes the presence of nonsense and frameshift mutations that are also present in the germline of patients with Warts, Hypogammaglobulinemia, Infections, and Myelokathexis (WHIM) syndrome.

In CLL, an unmutated BCR appears to be related to a better response to ibrutinib, which could be due to a larger dependence on BCR signaling in BCR unmutated CLL [108,117]. Secondary resistance to ibrutinib occurs as well. In both CLL and MCL, C481S mutations in *BTK* have been detected after ibrutinib therapy which were not present before therapy [118,119]. This mutation is located at the active site of BTK and causes a strong decrease in the affinity for ibrutinib for BTK. In addition, mutations in *PLCG2* have been detected after treatment with ibrutinib [119]. These are gain-of-function mutations which activate BCR signaling downstream of BTK [120].

Other novel drugs in targeting B-cell receptor signaling are directed against PI3K or SYK. Idelalisib and duvelisib are inhibitors of PI3K that target specific isoforms of the p110 subunit of PI3K. Idelalisib targets the delta isoform and duvelisib targets both the delta and gamma isoform. Idelalisib has been shown to be effective in patients with relapsed CLL [121] and indolent non-Hodgkin lymphoma (NHL), including FL, MZL, and LPL [122,123]. In MCL, idelalisib was also effective but the majority of patients showed only a short response duration [124]. Resistance mechanisms for idelalisib have not been studied as well as for ibrutinib, but higher expression of other PI3K isoforms has been suggested as a mechanism from in vitro studies [125]. In addition, activation of other oncogenic pathways such as MYC has been hypothesized as a mechanism for idelalisib resistance [126]. Idelalisib has been approved by the FDA for treatment of patients with relapsed CLL in combination with rituximab and for patients with relapsed FL and SLL. In Europe, idelalisib is authorized in combination with rituximab for use in patients with refractory FL and patients with refractory CLL or CLL with 17p deletion and/or *TP53* mutation in patients who are too frail for standard chemotherapy.

6.2.2. Proteasome inhibitors

Inhibition of the proteasome in hematological as well as solid tumors induces a wide range of effects including anti-proliferative and pro-apoptotic effects in vitro [127]. Although the ubiquitin-proteasome system also has key functions in normal cells, proteasome activity has been shown to be higher in hematological malignancies, forming a basis for in vivo use of proteasome inhibitors in these tumors [128,129]. In addition to NF-kappaB pathway inhibition, proteasome inhibitors cause inhibition of ERK signaling, upregulation of pro-apoptotic proteins,

downregulation of anti-apoptotic proteins, anti-angiogenic effects, inhibition of DNA repair, disruption of tumor-microenvironment interactions, and stabilization of p21, p27, and p53 [130]. Proteasome inhibitors are currently registered for multiple myeloma (bortezomib, carfilzomib) and MCL (bortezomib). In addition, inhibitors of the immunoproteasome are under development. Immunoproteasomes are predominantly present in hematopoietic cells and immunoproteasome inhibition is therefore a potentially more targeted method of proteasome inhibition [127]. Furthermore, compounds that inhibit components of the ubiquitin-proteasome system other than the proteasome are being developed [127].

6.3. BCL6

The BCL6 gene located on chromosome 3q27 encodes a sequence specific repressor of transcription that serves as a master regulator of the germinal center phenotype. It is essential for formation of germinal centers, and mice lacking BCL6 are unable to form germinal centers [131]. In the normal germinal center, high proliferation occurs together with genomic instability due to somatic hypermutation. In this context, BCL6 functions as a repressor of normal control mechanisms of the cell cycle, DNA damage response, and cell death. Although this action is necessary to generate B-cells with high-affinity antibodies, it is also dangerous because it may lead to deregulated growth and therefore BCL6 expression is tightly regulated in the normal situation. However, deregulated BCL6 expression is a common event in B-cell lymphomas and multiple mechanisms can cause this. Translocations which put BCL6 under the control of a strong promoter such as IGH are a frequent event in DLBCL and are also present in a subset of FLs [132–134]. Alternative mechanisms include mutations in binding sites for repressive transcription factors [135], mutations in *MEF2B* which is a transcriptional activator of *BCL6* [136], hypermethylation of intron 1 preventing silencing by CTCF [137], and posttranscriptional regulation [131].

6.3.1. BCL6 inhibitors

Studies into BCL6 inhibitors are currently limited to in vitro and mice studies. These studies show a strong response of tumors to BCL6 inhibition in vitro and in vivo, but oncogene addiction to BCL2 was observed in response to BCL6 inhibition, indicating that BCL6 inhibitors could be particularly useful in combination with BCL2 inhibitors [138–140]. In addition to drugs specifically targeting BCL6, histone deacetylase (HDAC) inhibitors also counteract the effects of BCL6 [141].

6.4. NOTCH signaling

Mammals possess 4 different NOTCH genes that encode for receptors involved in self-renewal and differentiation [142]. Upon binding of their ligands, a cleavage step causes release of the intracellular part of NOTCH which then translocates to the nucleus where it forms a protein complex with other proteins including MAML and histone acetyltransferases (e.g. EP300) [143]. This complex then stimulates transcription of a number of genes, resulting in increased expression of MYC and activation of the PI3K/AKT, mTOR, and NF-kappaB pathway [142–144]. NOTCH1 is essential for normal T-cell development, but is also frequently mutated in T-cell acute lymphoblastic leukemia [145]. These mutations occur in the heterodimerization and/or PEST domain. The PEST domain is the target for the FBXW7 ubiquitin protein ligase which targets NOTCH1 for the proteasome. Mutations in the PEST domain prevent ubiquitination and downregulation of NOTCH. Mutations in the heterodimerization domain cause enhanced cleavage of membranous NOTCH [145]. In indolent B-cell lymphomas, NOTCH1 mutations have been reported in a subset of CLL, SMZL, and FL [93,98,146–148]. In CLL, NOTCH1 mutations are detected in approximately 15% with a higher frequency in CLL with unmutated IGHV and/or trisomy 12 [75]. NOTCH1 mutations in CLL

are associated with a worse prognosis and a lack of response to rituximab treatment [149]. NOTCH2 is the master regulator of marginal zone differentiation and *NOTCH2* mutations are detected in up to one quarter of SMZLs with an association with short time to first treatment [78,93,95,147].

6.5. Epigenetic deregulation

Recent large-scale sequencing efforts have identified frequent mutations in genes involved in chromatin modulation in B-cell lymphomas [100,150–152]. These have been most extensively studied in DLBCL and FL in which mutations in *MLL2* (*KMT2D*), *CREBBP*, *EP300*, and *EZH2* were most frequently observed. Most of these mutations are loss-of-function mutations, except for mutations in *EZH2* which are gain-of-function. These mutations result in epigenetic silencing of specific genetic loci involved in cell cycle progression and regulation of the germinal center reaction, contributing to lymphomagenesis [153].

6.5.1. Epigenetic therapy

In hemato-oncology, HDAC inhibitors are mainly used for the treatment of patients with T-cell lymphoma, but limited studies have also been performed in B-cell lymphomas with variable results [154,155]. In addition to HDAC inhibitors, EZH2 inhibitors are under investigation for treatment of non-Hodgkin lymphomas [156].

6.6. Mitogen activated protein kinase (MAPK) signaling

In hairy cell leukemia, the BRAF V600E mutation has been identified by next-generation sequencing as a disease-defining genetic event [157]. BRAF is downstream from KRAS and upstream from MEK and ERK in the MAPK pathway. The BRAF V600E mutation causes constitutive activation of BRAF, resulting in cell survival [143]. For patients with relapsed or refractor hairy cell leukemia, the oral BRAF inhibitor vemurafenib is an effective treatment option [158–160].

7. Prediction of effective targeted treatment

With the increase of novel targeted treatments, the pressing question is which patient should receive which treatment. To select patients for treatment, the diagnosis still forms a starting point to select patients, but refinements are already being added. It is expected that the use of molecular diagnostics for treatment prediction will increase significantly as more and more mechanisms for resistance and sensitivity become known. It can also be expected that complexity will increase as more molecular events relevant for prediction of therapy response will be detected and more drugs developed. This also means that large clinical trials will become more difficult as each patient will have a different lymphoma with a different genetic background and novel approaches to clinical studies should be introduced.

With respect to mutation detection, many laboratories are now setting up panels for next-generation sequencing in lymphomas. Recently, the European Expert Group on NGS-based Diagnostics in Lymphomas (EGNL) summarized the current state-of-the-art in NGS for lymphomas and is now performing validation of a panel of 30 recurrently mutated genes [74].

Another question that needs to be answered in the near future is what diagnostic modalities will be required for therapy prediction. Immunohistochemistry and NGS are important techniques in this respect, but a mutation or changed protein expression is no guarantee for a therapy response. Mutations in non-coding regions, epigenetic alterations, and the micro-environment could all have an impact on the response to a particular treatment. In line with this, it will be necessary in the future to integrate the features of a lymphoma and the environment on different levels (i.e. genome, epigenome, proteome, micro-environment) to provide the most accurate prediction of therapy response.

8. Conclusion

Developments are following each other rapidly in the area of targeted treatment of indolent lymphoma and the (epi)genetic lesions involved. The challenge will be to determine which patient will benefit most from a given treatment at different time points of disease. It is expected that extensive genetic testing for prediction of treatment response at diagnosis, during follow-up and at relapse will be adopted in clinical practice soon, but for now only *TP53* mutations and del17p in CLL have a direct effect on treatment decisions. The long tail of different mutations in many types of lymphoma and the increasing number of drugs will require large studies to assess the impact of these mutations on prognosis and treatment response. This challenge is becoming even greater with the ever increasing knowledge from multiple sources (i.e. genome/epigenome, proteome, tumor micro-environment, clinical features). Tumor heterogeneity and natural and chemotherapy induced clonal evolution further increase complexity and will require sequential analyses to optimize and adapt therapy. Particularly in indolent B-cell lymphomas which are for the largest part incurable, decision making based on (epi)genetic profiling will be important.

Practice points

- Specific bacterial infections predispose to extranodal marginal zone lymphoma and are amenable to antibiotic treatment. However, the evidence is only conclusive for treatment of *Helicobacter pylori* in marginal zone lymphoma of the stomach.
- The mutational landscape of indolent B-cell lymphomas is rapidly being elucidated and it is expected that mutation detection will influence treatment decisions regarding targeted therapy.
- For now however, only demonstration of *TP53* mutations and/or del 17p in CLL/SLL have a direct impact on therapy.

Research agenda

- Correlation of mutational status with response to targeted treatment
- Development of functional assays to predict treatment responses more accurately.
- Refinement of diagnostic criteria based on subgroups that arise from large-scale sequencing efforts.

Conflict of interest

The authors declare no potential conflicts of interest.

References

- [1] Zullo A, Hassan C, Cristofari F, Andriani A, De Francesco V, Ierardi E, et al. Effects of *Helicobacter pylori* eradication on early stage gastric mucosa-associated lymphoid tissue lymphoma. *Clin Gastroenterol Hepatol* 2010;8(2):105–10.
- [2] Eidt S, Stolte M, Fischer R. *Helicobacter pylori* gastritis and primary gastric non-Hodgkin's lymphomas. *J Clin Pathol* 1994;47(5):436–9.
- [3] Nakamura S, Yao T, Aoyagi K, Iida M, Fujishima M, Tsuneyoshi M. *Helicobacter pylori* and primary gastric lymphoma. A histopathologic and immunohistochemical analysis of 237 patients. *Cancer* 1997;79(1):3–11.
- [4] Wotherspoon AC, Ortiz-Hidalgo C, Falzon MR, Isaacson PG. *Helicobacter pylori*-associated gastritis and primary B-cell gastric lymphoma. *Lancet* 1991;338(8776):1175–6.
- [5] Wotherspoon AC, Dogliani C, Diss TC, Pan L, Moschini A, de Boni M, et al. Regression of primary low-grade B-cell gastric lymphoma of mucosa-associated lymphoid tissue type after eradication of *Helicobacter pylori*. *Lancet* 1993;342(8871):575–7.
- [6] Ferreri AJ, Govi S, Ponzoni M. Marginal zone lymphomas and infectious agents. *Semin Cancer Biol* 2013;23(6):431–40.
- [7] Isaacson PG. Update on MALT lymphomas. *Best Pract Res Clin Haematol* 2005;18(1):57–68.
- [8] Hamoudi RA, Appert A, Ye H, Ruskone-Fourmesttraux A, Streubel B, Chott A, et al. Differential expression of NF-kappaB target genes in MALT lymphoma with and without chromosome translocation: insights into molecular mechanism. *Leukemia* 2010;24(8):1487–97.
- [9] Ye H, Gong L, Liu H, Ruskone-Fourmesttraux A, de Jong D, Pileri S, et al. Strong

- BCL10 nuclear expression identifies gastric MALT lymphomas that do not respond to *H. pylori* eradication. *Gut* 2006;55(1):137–8.
- [10] Ferreri AJ, Govi S, Raderer M, Mule A, Andriani A, Caracciolo D, et al. *Helicobacter pylori* eradication as exclusive treatment for limited-stage gastric diffuse large B-cell lymphoma: results of a multicenter phase 2 trial. *Blood* 2012;120(18):3858–60.
- [11] Kuo SH, Yeh KH, Wu MS, Lin CW, Hsu PN, Wang HP, et al. *Helicobacter pylori* eradication therapy is effective in the treatment of early-stage *H. pylori*-positive gastric diffuse large B-cell lymphomas. *Blood* 2012;119(21):4838–44. [quiz 5057].
- [12] Yoo C, Ryu MH, Huh J, Park JH, Kang HJ, Ahn HS, et al. *Chlamydia psittaci* infection and clinicopathologic analysis of ocular adnexal lymphomas in Korea. *Am J Hematol* 2007;82(9):821–3.
- [13] Ferreri AJ, Guidoboni M, Ponzoni M, De Conciliis C, Dell'Oro S, Fleischhauer K, et al. Evidence for an association between *Chlamydia psittaci* and ocular adnexal lymphomas. *J Natl Cancer Inst* 2004;96(8):586–94.
- [14] Rosado MF, Byrne Jr. GE, Ding F, Fields KA, Ruiz P, Dubovy SR, et al. Ocular adnexal lymphoma: a clinicopathologic study of a large cohort of patients with no evidence for an association with *Chlamydia psittaci*. *Blood* 2006;107(2):467–72.
- [15] Vargas RL, Fallone E, Felgar RE, Friedberg JW, Arbin AA, Andersen AA, et al. Is there an association between ocular adnexal lymphoma and infection with *Chlamydia psittaci*? The University of Rochester experience. *Leuk Res* 2006;30(5):547–51.
- [16] Zhang GS, Winter JN, Variakojis D, Reich S, Lissner GS, Bryar P, et al. Lack of an association between *Chlamydia psittaci* and ocular adnexal lymphoma. *Leuk Lymphoma* 2007;48(3):577–83.
- [17] Mulder MM, Heddema ER, Pannekoek Y, Faridpooya K, Oud ME, Schilder-Tol E, et al. No evidence for an association of ocular adnexal lymphoma with *Chlamydia psittaci* in a cohort of patients from the Netherlands. *Leuk Res* 2006;30(10):1305–7.
- [18] Daibata M, Nemoto Y, Togitani K, Fukushima A, Ueno H, Ouchi K, et al. Absence of *Chlamydia psittaci* in ocular adnexal lymphoma from Japanese patients. *Br J Haematol* 2006;132(5):651–2.
- [19] de Cremoux P, Subtil A, Ferreri AJ, Vincent-Salomon A, Ponzoni M, Chaoui D, et al. Re: evidence for an association between *Chlamydia psittaci* and ocular adnexal lymphomas. *J Natl Cancer Inst* 2006;98(5):365–6.
- [20] Chanudet E, Zhou Y, Bacon CM, Wotherspoon AC, Muller-Hermelink HK, Adam P, et al. *Chlamydia psittaci* is variably associated with ocular adnexal MALT lymphoma in different geographical regions. *J Pathol* 2006;209(3):344–51.
- [21] Aigelsreiter A, Leitner E, Deutsch AJ, Kessler HH, Stelzl E, Beham-Schmid C, et al. *Chlamydia psittaci* in MALT lymphomas of ocular adnexals: the Austrian experience. *Leuk Res* 2008;32(8):1292–4.
- [22] Yakushijin Y, Kodama T, Takaoka I, Tanimoto K, Besho H, Sakai I, et al. Absence of chlamydial infection in Japanese patients with ocular adnexal lymphoma of mucosa-associated lymphoid tissue. *Int J Hematol* 2007;85(3):223–30.
- [23] Kiesewetter B, Raderer M. Antibiotic therapy in nongastrointestinal MALT lymphoma: a review of the literature. *Blood* 2013;122(8):1350–7.
- [24] Ferreri AJ, Ponzoni M, Guidoboni M, Resti AG, Politi LS, Cortelazzo S, et al. Bacteria-eradicating therapy with doxycycline in ocular adnexal MALT lymphoma: a multicenter prospective trial. *J Natl Cancer Inst* 2006;98(19):1375–82.
- [25] Ferreri AJ, Govi S, Pasini E, Mappa S, Bertoni F, Zaja F, et al. *Chlamydia psittaci* eradication with doxycycline as first-line targeted therapy for ocular adnexal lymphoma: final results of an international phase II trial. *J Clin Oncol* 2012;30(24):2988–94.
- [26] Cerroni L, Zochling N, Putz B, Kerl H. Infection by *Borrelia burgdorferi* and cutaneous B-cell lymphoma. *J Cutan Pathol* 1997;24(8):457–61.
- [27] Goodlad JR, Davidson MM, Hollowood K, Ling C, MacKenzie C, Christie I, et al. Primary cutaneous B-cell lymphoma and *Borrelia burgdorferi* infection in patients from the Highlands of Scotland. *Am J Surg Pathol* 2000;24(9):1279–85.
- [28] Goodlad JR, Davidson MM, Hollowood K, Batstone P, Ho-Yen DO. *Borrelia burgdorferi*-associated cutaneous marginal zone lymphoma: a clinicopathological study of two cases illustrating the temporal progression of *B. burgdorferi*-associated B-cell proliferation in the skin. *Histopathology* 2000;37(6):501–8.
- [29] Lecuit M, Abachin E, Martin A, Poyart C, Pochart P, Suarez F, et al. Immunoproliferative small intestinal disease associated with *Campylobacter jejuni*. *N Engl J Med* 2004;350(3):239–48.
- [30] Mesnard B, De Vroey B, Maunoury V, Lecuit M. Immunoproliferative small intestinal disease associated with *Campylobacter jejuni*. *Dig Liver Dis* 2012;44(9):799–800.
- [31] Adam P, Czapiewski P, Colak S, Kosmidis P, Tousseyn T, Sagaert X, et al. Prevalence of *Achromobacter xylosoxidans* in pulmonary mucosa-associated lymphoid tissue lymphoma in different regions of Europe. *Br J Haematol* 2014;164(6):804–10.
- [32] Landgren O, Rapkin JS, Caporaso NE, Mellemkjaer L, Gridley G, Goldin LR, et al. Respiratory tract infections and subsequent risk of chronic lymphocytic leukemia. *Blood* 2007;109(5):2198–201.
- [33] Shen DF, Herbort CP, Tuaille N, Buggage RR, Egwuagu CE, Chan CC. Detection of *Toxoplasma gondii* DNA in primary intraocular B-cell lymphoma. *Mod Pathol* 2001;14(10):995–9.
- [34] Negri E, Little D, Boiocchi M, La Vecchia C, Franceschi S. B-cell non-Hodgkin's lymphoma and hepatitis C virus infection: a systematic review. *Int J Cancer* 2004;111(1):1–8.
- [35] Gisbert JP, Garcia-Buey L, Pajares JM, Moreno-Otero R. Prevalence of hepatitis C virus infection in B-cell non-Hodgkin's lymphoma: systematic review and meta-analysis. *Gastroenterology* 2003;125(6):1723–32.
- [36] Vannata B, Zucca E. Hepatitis C virus-associated B-cell non-Hodgkin lymphomas. *Hematology Am Soc Hematol Educ Program* 2014;2014(1):590–8.
- [37] Hermine O, Lefrere F, Bronowicki JP, Mariette X, Jondeau K, Eclache-Saudreau V, et al. Regression of splenic lymphoma with villous lymphocytes after treatment of hepatitis C virus infection. *N Engl J Med* 2002;347(2):89–94.
- [38] Kelaidi C, Rollet F, Park S, Tulliez M, Christoforov B, Calmus Y, et al. Response to antiviral treatment in hepatitis C virus-associated marginal zone lymphomas. *Leukemia* 2004;18(10):1711–6.
- [39] Arcaini L, Vallisa D, Rattotti S, Ferretti VV, Ferreri AJ, Bernuzzi P, et al. Antiviral treatment in patients with indolent B-cell lymphomas associated with HCV infection: a study of the Fondazione Italiana Linfomi. *Ann Oncol* 2014;25(7):1404–10.
- [40] Saadoun D, Suarez F, Lefrere F, Valensi F, Mariette X, Aouba A, et al. Splenic lymphoma with villous lymphocytes, associated with type II cryoglobulinemia and HCV infection: a new entity? *Blood* 2005;105(1):74–6.
- [41] Vallisa D, Bernuzzi P, Arcaini L, Sacchi S, Callea V, Marasca R, et al. Role of anti-hepatitis C virus (HCV) treatment in HCV-related, low-grade, B-cell, non-Hodgkin's lymphoma: a multicenter Italian experience. *J Clin Oncol* 2005;23(3):468–73.
- [42] Mazzaro C, De Re V, Spina M, Dal Maso L, Festini G, Comar C, et al. Pegylated-interferon plus ribavirin for HCV-positive indolent non-Hodgkin lymphomas. *Br J Haematol* 2009;145(2):255–7.
- [43] Pellicelli AM, Marignani M, Zoli V, Romano M, Morrone A, Nosotti L, et al. Hepatitis C virus-related B cell subtypes in non Hodgkin's lymphoma. *World J Hepatol* 2011;3(11):278–84.
- [44] Michot JM, Canioni D, Driss H, Alric L, Cacoub P, Suarez F, et al. Antiviral therapy is associated with a better survival in patients with hepatitis C virus and B-cell non-Hodgkin lymphomas, ANRS HC-13 lympho-C study. *Am J Hematol* 2015;90(3):197–203.
- [45] Vannata B, Arcaini L, Zucca E. Hepatitis C virus-associated B-cell non-Hodgkin's lymphomas: what do we know? *Ther Adv Hematol* 2016;7(2):94–107.
- [46] Lim LY, La D, Cserti-Gazdewich CM, Shah H. Lymphoma remission by interferon-free HCV eradication without chemotherapy. *ACG Case Rep J* 2015;3(1):69–70.
- [47] Rossotti R, Travi G, Pazzi A, Baiguera C, Morra E, Puoti M. Rapid clearance of HCV-related splenic marginal zone lymphoma under an interferon-free, NS3/NS4A inhibitor-based treatment. A case report. *J Hepatol* 2015;62(1):234–7.
- [48] Sutton LA, Agathangelidis A, Belessi C, Darzentas N, Davi F, Ghia P, et al. Antigen selection in B-cell lymphomas—tracing the evidence. *Semin Cancer Biol* 2013;23(6):399–409.
- [49] Agathangelidis A, Darzentas N, Hadzidimitriou A, Brochet X, Murray F, Yan XJ, et al. Stereotyped B-cell receptors in one-third of chronic lymphocytic leukemia: a molecular classification with implications for targeted therapies. *Blood* 2012;119(19):4467–75.
- [50] Bikos V, Darzentas N, Hadzidimitriou A, Davis Z, Hockley S, Traverse-Glehen A, et al. Over 30% of patients with splenic marginal zone lymphoma express the same immunoglobulin heavy variable gene: ontogenetic implications. *Leukemia* 2012;26(7):1638–46.
- [51] Catera R, Silverman GJ, Hatzi K, Seiler T, Didier S, Zhang L, et al. Chronic lymphocytic leukemia cells recognize conserved epitopes associated with apoptosis and oxidation. *Mol Med* 2008;14(11–12):665–74.
- [52] Chu CK, Catera R, Hatzi K, Yan XJ, Zhang L, Wang XB, et al. Chronic lymphocytic leukemia antibodies with a common stereotypic rearrangement recognize non-muscle myosin heavy chain IIA. *Blood* 2008;112(13):5122–9.
- [53] Herve M, Xu K, Ng YS, Wardemann H, Albesiano E, Messmer BT, et al. Unmutated and mutated chronic lymphocytic leukemias derive from self-reactive B cell precursors despite expressing different antibody reactivity. *J Clin Invest* 2005;115(6):1636–43.
- [54] Lanemo Myhrinder A, Hellqvist E, Sidorova E, Soderberg A, Baxendale H, Dahle C, et al. A new perspective: molecular motifs on oxidized LDL, apoptotic cells, and bacteria are targets for chronic lymphocytic leukemia antibodies. *Blood* 2008;111(7):3838–48.
- [55] Warsame AA, Aasheim HC, Nustad K, Troen G, Tierens A, Wang V, et al. Splenic marginal zone lymphoma with VH1-02 gene rearrangement expresses poly- and self-reactive antibodies with similar reactivity. *Blood* 2011;118(12):3331–9.
- [56] Hatzi K, Catera R, Ferrarini M, Fischetti V, Herve M, Meffre E, et al. B-cell chronic lymphocytic leukemia (B-CLL) cells express antibodies reactive with antigenic epitopes expressed on the surface of common bacteria. *Blood* 2006;108:25.
- [57] Kostareli E, Hadzidimitriou A, Stavroyianni N, Darzentas N, Athanasiadou A, Gounari M, et al. Molecular evidence for EBV and CMV persistence in a subset of patients with chronic lymphocytic leukemia expressing stereotyped IGHV4-34 B-cell receptors. *Leukemia* 2009;23(5):919–24.
- [58] Zintzaras E, Voulgarelis M, Moutsopoulos HM. The risk of lymphoma development in autoimmune diseases: a meta-analysis. *Arch Intern Med* 2005;165(20):2337–44.
- [59] Ekstrom Smedby K, Vajdic CM, Falster M, Engels EA, Martinez-Maza O, Turner J, et al. Autoimmune disorders and risk of non-Hodgkin lymphoma subtypes: a pooled analysis within the InterLymph Consortium. *Blood* 2008;111(8):4029–38.
- [60] Bende RJ, Aarts WM, Riedl RG, de Jong D, Pals ST, van Noesel CJ. Among B cell non-Hodgkin's lymphomas, MALT lymphomas express a unique antibody repertoire with frequent rheumatoid factor reactivity. *J Exp Med* 2005;201(8):1229–41.
- [61] Martin T, Weber JC, Levallois H, Labouret N, Soley A, Koenig S, et al. Salivary gland lymphomas in patients with Sjogren's syndrome may frequently develop from rheumatoid factor B cells. *Arthritis Rheum* 2000;43(4):908–16.
- [62] Nocturne G, Mariette X. Sjogren Syndrome-associated lymphomas: an update on pathogenesis and management. *Br J Haematol* 2015;168(3):317–27.
- [63] Dagklis A, Ponzoni M, Govi S, Cangli MG, Pasini E, Charlotte F, et al. Immunoglobulin gene repertoire in ocular adnexal lymphomas: hints on the nature of the antigenic stimulation. *Leukemia* 2012;26(4):814–21.
- [64] Limpens J, Stad R, Vos C, de Vlaam C, de Jong D, van Ommen GJ, et al.

- Lymphoma-associated translocation t(14;18) in blood B cells of normal individuals. *Blood* 1995;85(9):2528–36.
- [65] Mamessier E, Broussais-Guillaumot F, Chetaille B, Bouabdallah R, Xerri L, Jaffe ES, et al. Nature and importance of follicular lymphoma precursors. *Haematologica* 2014;99(5):802–10.
- [66] Schmidt J, Salaverria I, Haake A, Bonzheim I, Adam P, Montes-Moreno S, et al. Increasing genomic and epigenomic complexity in the clonal evolution from in situ to manifest t(14;18)-positive follicular lymphoma. *Leukemia* 2014;28(5):1103–12.
- [67] Jegalian AG, Eberle FC, Pack SD, Mirvis M, Raffeld M, Pittaluga S, et al. Follicular lymphoma in situ: clinical implications and comparisons with partial involvement by follicular lymphoma. *Blood* 2011;118(11):2976–84.
- [68] Landgren O, Albitar M, Ma W, Abbasi F, Hayes RB, Ghia P, et al. B-cell clones as early markers for chronic lymphocytic leukemia. *N Engl J Med* 2009;360(7):659–67.
- [69] Karube K, Scarfo L, Campo E, Ghia P. Monoclonal B cell lymphocytosis and “in situ” lymphoma. *Semin Cancer Biol* 2014;24:3–14.
- [70] Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood* 2016;127(20):2375–90.
- [71] Scarfo L, Ghia P. What does it mean I have a monoclonal B-cell lymphocytosis?: recent insights and new challenges. *Semin Oncol* 2016;43(2):201–8.
- [72] Fazi C, Scarfo L, Pecciarini L, Cottini F, Dagklis A, Janus A, et al. General population low-count CLL-like MBL persists over time without clinical progression, although carrying the same cytogenetic abnormalities of CLL. *Blood* 2011;118(25):6618–25.
- [73] Morabito F, Mosca L, Cutrona G, Agnelli L, Tuana G, Ferracin M, et al. Clinical monoclonal B lymphocytosis versus Rai 0 chronic lymphocytic leukemia: a comparison of cellular, cytogenetic, molecular, and clinical features. *Clin Cancer Res* 2013;19(21):5890–900.
- [74] Rosenquist R, Rosenwald A, Du MQ, Gaidano G, Groenen P, Wotherspoon A, et al. Clinical impact of recurrently mutated genes on lymphoma diagnostics: state-of-the-art and beyond. *Haematologica* 2016;101(9):1002–9.
- [75] Rossi D, Gaidano G. The clinical implications of gene mutations in chronic lymphocytic leukaemia. *Br J Cancer* 2016;114(8):849–54.
- [76] Rossi D, Khiabanian H, Spina V, Ciardullo C, Bruscaggini A, Fama R, et al. Clinical impact of small TP53 mutated subclones in chronic lymphocytic leukemia. *Blood* 2014;123(14):2139–47.
- [77] O’Shea D, O’Riain C, Taylor C, Waters R, Carloti E, Macdougall F, et al. The presence of TP53 mutation at diagnosis of follicular lymphoma identifies a high-risk group of patients with shortened time to disease progression and poorer overall survival. *Blood* 2008;112(8):3126–9.
- [78] Parry M, Rose-Zerilli MJ, Ljungstrom V, Gibson J, Wang J, Walewska R, et al. Genetics and prognostication in splenic marginal zone lymphoma: revelations from deep sequencing. *Clin Cancer Res* 2015;21(18):4174–83.
- [79] Tsujimoto Y, Cossman J, Jaffe E, Croce CM. Involvement of the bcl-2 gene in human follicular lymphoma. *Science* 1985;228(4706):1440–3.
- [80] Scarfo L, Ghia P. Reprogramming cell death: BCL2 family inhibition in hematological malignancies. *Immunol Lett* 2013;155(1–2):36–9.
- [81] Monni O, Joensuu H, Franssila K, Klefstrom J, Alitalo K, Knuutila S. BCL2 overexpression associated with chromosomal amplification in diffuse large B-cell lymphoma. *Blood* 1997;90(3):1168–74.
- [82] Hanada M, Delia D, Aiello A, Stadtmayer E, Reed JC. bcl-2 gene hypomethylation and high-level expression in B-cell chronic lymphocytic leukemia. *Blood* 1993;82(6):1820–8.
- [83] Cimmino A, Calin GA, Fabbri M, Iorio MV, Ferracin M, Shimizu M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci U S A* 2005;102(39):13944–9.
- [84] Skowronska A, Parker A, Ahmed G, Oldreive C, Davis Z, Richards S, et al. Biallelic ATM inactivation significantly reduces survival in patients treated on the United Kingdom Leukemia Research Fund Chronic Lymphocytic Leukemia 4 trial. *J Clin Oncol* 2012;30(36):4524–32.
- [85] Dierlamm J, Baens M, Wlodarska I, Stefanova-Ouzounova M, Hernandez JM, Hossfeld DK, et al. The apoptosis inhibitor gene API2 and a novel 18q gene, MLT, are recurrently rearranged in the t(11;18)(q21;q21) associated with mucosa-associated lymphoid tissue lymphomas. *Blood* 1999;93(11):3601–9.
- [86] Lucas PC, Yonezumi M, Inohara N, McAllister-Lucas LM, Abazeed ME, Chen FF, et al. Bcl10 and MALT1, independent targets of chromosomal translocation in malt lymphoma, cooperate in a novel NF-kappa B signaling pathway. *J Biol Chem* 2001;276(22):19012–9.
- [87] Willis TG, Jadayel DM, Du MQ, Peng H, Perry AR, Abdul-Rauf M, et al. Bcl10 is involved in t(1;14)(p22;q32) of MALT B cell lymphoma and mutated in multiple tumor types. *Cell* 1999;96(1):35–45.
- [88] Zhang Q, Siebert R, Yan M, Hinzmann B, Cui X, Xue L, et al. Inactivating mutations and overexpression of BCL10, a caspase recruitment domain-containing gene, in MALT lymphoma with t(1;14)(p22;q32). *Nat Genet* 1999;22(1):63–8.
- [89] Thome M, Martinon F, Hofmann K, Rubio V, Steiner V, Schneider P, et al. Equine herpesvirus-2 E10 gene product, but not its cellular homologue, activates NF-kappaB transcription factor and c-Jun N-terminal kinase. *J Biol Chem* 1999;274(15):9962–8.
- [90] Chanudet E, Huang Y, Ichimura K, Dong G, Hamoudi RA, Radford J, et al. A20 is targeted by promoter methylation, deletion and inactivating mutation in MALT lymphoma. *Leukemia* 2010;24(2):483–7.
- [91] Novak U, Rinaldi A, Kwee I, Nandula SV, Rancoita PM, Compagno M, et al. The NF-(kappa)B negative regulator TNFAIP3 (A20) is inactivated by somatic mutations and genomic deletions in marginal zone lymphomas. *Blood* 2009;113(20):4918–21.
- [92] Rossi D, Deaglio S, Dominguez-Sola D, Rasi S, Vaisitti T, Agostinelli C, et al. Alteration of BIRC3 and multiple other NF-kB pathway genes in splenic marginal zone lymphoma. *Blood* 2011;118(18):4930–4.
- [93] Rossi D, Trifonov V, Fangazio M, Bruscaggini A, Rasi S, Spina V, et al. The coding genome of splenic marginal zone lymphoma: activation of NOTCH2 and other pathways regulating marginal zone development. *J Exp Med* 2012;209(9):1537–51.
- [94] Martinez N, Almaraz C, Vaque JP, Varela I, Derdak S, Beltran S, et al. Whole-exome sequencing in splenic marginal zone lymphoma reveals mutations in genes involved in marginal zone differentiation. *Leukemia* 2014;28(6):1334–40.
- [95] Clipson A, Wang M, de Leval L, Ashton-Key M, Wotherspoon A, Vassiliou G, et al. KLF2 mutation is the most frequent somatic change in splenic marginal zone lymphoma and identifies a subset with distinct genotype. *Leukemia* 2015;29(5):1177–85.
- [96] Treon SP, Xu L, Yang G, Zhou Y, Liu X, Cao Y, et al. MYD88 L265P somatic mutation in Waldenstrom’s macroglobulinemia. *N Engl J Med* 2012;367(9):826–33.
- [97] Varettoni M, Arcaini L, Zibellini S, Boveri E, Rattotti S, Riboni R, et al. Prevalence and clinical significance of the MYD88 (L265P) somatic mutation in Waldenstrom’s macroglobulinemia and related lymphoid neoplasms. *Blood* 2013;121(13):2522–8.
- [98] Puente XS, Pinyol M, Quesada V, Conde L, Ordóñez GR, Villamor N, et al. Whole-genome sequencing identifies recurrent mutations in chronic lymphocytic leukaemia. *Nature* 2011;475(7354):101–5.
- [99] Rossi D, Fangazio M, Rasi S, Vaisitti T, Monti S, Cresta S, et al. Disruption of BIRC3 associates with fludarabine chemorefractoriness in TP53 wild-type chronic lymphocytic leukemia. *Blood* 2012;119(12):2854–62.
- [100] Okosun J, Bodor C, Wang J, Araf S, Yang CY, Pan C, et al. Integrated genomic analysis identifies recurrent mutations and evolution patterns driving the initiation and progression of follicular lymphoma. *Nat Genet* 2014;46(2):176–81.
- [101] Cheung KJ, Johnson NA, Affleck JG, Severson T, Steidl C, Ben-Neriah S, et al. Acquired TNFRSF14 mutations in follicular lymphoma are associated with worse prognosis. *Cancer Res* 2010;70(22):9166–74.
- [102] Launay E, Pangault C, Bertrand P, Jardin F, Lamy T, Tilly H, et al. High rate of TNFRSF14 gene alterations related to 1p36 region in de novo follicular lymphoma and impact on prognosis. *Leukemia* 2012;26(3):559–62.
- [103] Cheung TC, Steinberg MW, Osborne LM, Macauley MG, Fukuyama S, Sanjo H, et al. Unconventional ligand activation of herpesvirus entry mediator signals cell survival. *Proc Natl Acad Sci U S A* 2009;106(15):6244–9.
- [104] Lohr JG, Stojanov P, Lawrence MS, Auclair D, Chaput B, Sougnez C, et al. Discovery and prioritization of somatic mutations in diffuse large B-cell lymphoma (DLBCL) by whole-exome sequencing. *Proc Natl Acad Sci U S A* 2012;109(10):3879–84.
- [105] Costello RT, Mallet F, Barbat B, Schiano De Colella JM, Sainy D, Sweet RW, et al. Stimulation of non-Hodgkin’s lymphoma via HVEM: an alternate and safe way to increase Fas-induced apoptosis and improve tumor immunogenicity. *Leukemia* 2003;17(12):2500–7.
- [106] Pan Z, Scheerens H, Li SJ, Schultz BE, Sprengeler PA, Burrill LC, et al. Discovery of selective irreversible inhibitors for Bruton’s tyrosine kinase. *ChemMedChem* 2007;2(1):58–61.
- [107] Honigberg LA, Smith AM, Sirisawad M, Verner E, Loury D, Chang B, et al. The Bruton tyrosine kinase inhibitor PCI-32765 blocks B-cell activation and is efficacious in models of autoimmune disease and B-cell malignancy. *Proc Natl Acad Sci U S A* 2010;107(29):13075–80.
- [108] Zhang SQ, Smith SM, Zhang SY, Lynn Wang Y. Mechanisms of ibrutinib resistance in chronic lymphocytic leukaemia and non-Hodgkin lymphoma. *Br J Haematol* 2015;170(4):445–56.
- [109] Wilson WH, Young RM, Schmitz R, Yang Y, Pittaluga S, Wright G, et al. Targeting B cell receptor signaling with ibrutinib in diffuse large B cell lymphoma. *Nat Med* 2015;21(8):922–6.
- [110] Rahal R, Frick M, Romero R, Korn JM, Kridel R, Chan FC, et al. Pharmacological and genomic profiling identifies NF-kappaB-targeted treatment strategies for mantle cell lymphoma. *Nat Med* 2014;20(1):87–92.
- [111] Yang G, Zhou Y, Liu X, Xu L, Cao Y, Manning RJ, et al. A mutation in MYD88 (L265P) supports the survival of lymphoplasmacytic cells by activation of Bruton tyrosine kinase in Waldenstrom macroglobulinemia. *Blood* 2013;122(7):1222–32.
- [112] Gachard N, Parrens M, Soubeyran I, Petit B, Marfak A, Rizzo D, et al. IGHV gene features and MYD88 L265P mutation separate the three marginal zone lymphoma entities and Waldenstrom macroglobulinemia/lymphoplasmacytic lymphomas. *Leukemia* 2013;27(1):183–9.
- [113] Hunter ZR, Xu L, Yang G, Zhou Y, Liu X, Cao Y, et al. The genomic landscape of Waldenstrom macroglobulinemia is characterized by highly recurring MYD88 and WHIM-like CXCR4 mutations, and small somatic deletions associated with B-cell lymphomagenesis. *Blood* 2014;123(11):1637–46.
- [114] Treon SP, Tripsas CK, Meid K, Warren D, Varma G, Green R, et al. Ibrutinib in previously treated Waldenstrom’s macroglobulinemia. *N Engl J Med* 2015;372(15):1430–40.
- [115] Cao Y, Hunter ZR, Liu X, Xu L, Yang G, Chen J, et al. CXCR4 WHIM-like frameshift and nonsense mutations promote ibrutinib resistance but do not supplant MYD88(L265P)-directed survival signalling in Waldenstrom macroglobulinemia cells. *Br J Haematol* 2015;168(5):701–7.
- [116] Cao Y, Hunter ZR, Liu X, Xu L, Yang G, Chen J, et al. The WHIM-like CXCR4(S338X) somatic mutation activates AKT and ERK, and promotes resistance to ibrutinib and other agents used in the treatment of Waldenstrom’s Macroglobulinemia. *Leukemia* 2015;29(1):169–76.
- [117] Byrd JC, Furman RR, Coutre SE, Flinn IW, Burger JA, Blum KA, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *N Engl J Med*

- 2013;369(1):32–42.
- [118] Furman RR, Cheng S, Lu P, Setty M, Perez AR, Guo A, et al. Ibrutinib resistance in chronic lymphocytic leukemia. *N Engl J Med* 2014;370(24):2352–4.
- [119] Woyach JA, Furman RR, Liu TM, Ozer HG, Zapatka M, Ruppert AS, et al. Resistance mechanisms for the Bruton's tyrosine kinase inhibitor ibrutinib. *N Engl J Med* 2014;370(24):2286–94.
- [120] Zhou Q, Lee GS, Brady J, Datta S, Katan M, Sheikh A, et al. A hypermorphic missense mutation in PLCG2, encoding phospholipase Cgamma2, causes a dominantly inherited autoinflammatory disease with immunodeficiency. *Am J Hum Genet* 2012;91(4):713–20.
- [121] Furman RR, Sharman JP, Coutre SE, Cheson BD, Pagel JM, Hillmen P, et al. Idelalisib and rituximab in relapsed chronic lymphocytic leukemia. *N Engl J Med* 2014;370(11):997–1007.
- [122] Gopal AK, Kahl BS, de Vos S, Wagner-Johnston ND, Schuster SJ, Jurczak WJ, et al. PI3Kdelta inhibition by idelalisib in patients with relapsed indolent lymphoma. *N Engl J Med* 2014;370(11):1008–18.
- [123] Martin P, Armas A, Gopal AK, Gyan E, Wagner-Johnston ND, Walewski J, et al. Idelalisib monotherapy and durable responses in patients with relapsed or refractory marginal zone lymphoma (MZL). *Blood* 2015;126(23):1543.
- [124] Kahl BS, Spurgeon SE, Furman RR, Flinn IW, Coutre SE, Brown JR, et al. A phase I study of the PI3Kdelta inhibitor idelalisib in patients with relapsed/refractory mantle cell lymphoma (MCL). *Blood* 2014;123(22):3398–405.
- [125] Iyengar S, Clear A, Bodor C, Maharaj L, Lee A, Calaminici M, et al. P110alpha-mediated constitutive PI3K signaling limits the efficacy of p110delta-selective inhibition in mantle cell lymphoma, particularly with multiple relapse. *Blood* 2013;121(12):2274–84.
- [126] Woyach JA, Johnson AJ. Targeted therapies in CLL: mechanisms of resistance and strategies for management. *Blood* 2015;126(4):471–7.
- [127] Crawford LJ, Irvine AE. Targeting the ubiquitin proteasome system in haematological malignancies. *Blood Rev* 2013;27(6):297–304.
- [128] Kumatori A, Tanaka K, Inamura N, Sone S, Ogura T, Matsumoto T, et al. Abnormally high expression of proteasomes in human leukemic cells. *Proc Natl Acad Sci U S A* 1990;87(18):7071–5.
- [129] Jakob C, Egerer K, Liebisch P, Turkmen S, Zavrski I, Kuckelkorn U, et al. Circulating proteasome levels are an independent prognostic factor for survival in multiple myeloma. *Blood* 2007;109(5):2100–5.
- [130] Bose P, Batalo MS, Holkova B, Grant S. Bortezomib for the treatment of non-Hodgkin's lymphoma. *Expert Opin Pharmacother* 2014;15(16):2443–59.
- [131] Hatzl K, Melnick A. Breaking bad in the germinal center: how deregulation of BCL6 contributes to lymphomagenesis. *Trends Mol Med* 2014;20(6):343–52.
- [132] Ye BH, Lista F, Lo Coco F, Knowles DM, Offit K, Chaganti RS, et al. Alterations of a zinc finger-encoding gene, BCL-6, in diffuse large-cell lymphoma. *Science* 1993;262(5134):747–50.
- [133] Katzenberger T, Ott G, Klein T, Kalla J, Muller-Hermelink HK, Ott MM. Cytogenetic alterations affecting BCL6 are predominantly found in follicular lymphomas grade 3B with a diffuse large B-cell component. *Am J Pathol* 2004;165(2):481–90.
- [134] Ott G, Katzenberger T, Lohr A, Kindelberger S, Rudiger T, Wilhelm M, et al. Cytomorphologic, immunohistochemical, and cytogenetic profiles of follicular lymphoma: 2 types of follicular lymphoma grade 3. *Blood* 2002;99(10):3806–12.
- [135] Saito M, Gao J, Basso K, Kitagawa Y, Smith PM, Bhagat G, et al. A signaling pathway mediating downregulation of BCL6 in germinal center B cells is blocked by BCL6 gene alterations in B cell lymphoma. *Cancer Cell* 2007;12(3):280–92.
- [136] Ying CY, Dominguez-Sola D, Fabi M, Lorenz IC, Hussein S, Bansal M, et al. MEF2B mutations lead to deregulated expression of the oncogene BCL6 in diffuse large B cell lymphoma. *Nat Immunol* 2013;14(10):1084–92.
- [137] Lai AY, Fatemi M, Dhasarathy A, Malone C, Sobol SE, Geigerman C, et al. DNA methylation prevents CTCF-mediated silencing of the oncogene BCL6 in B cell lymphomas. *J Exp Med* 2010;207(9):1939–50.
- [138] Cerchietti LC, Ghetu AF, Zhu X, Da Silva GF, Zhong S, Matthews M, et al. A small-molecule inhibitor of BCL6 kills DLBCL cells in vitro and in vivo. *Cancer Cell* 2010;17(4):400–11.
- [139] Dupont T, Yang SN, Patel J, Hatzl K, Malik A, Tam W, et al. Selective targeting of BCL6 induces oncogene addiction switching to BCL2 in B-cell lymphoma. *Oncotarget* 2016;7(3):3520–32.
- [140] Parekh S, Prive G, Melnick A. Therapeutic targeting of the BCL6 oncogene for diffuse large B-cell lymphomas. *Leuk Lymphoma* 2008;49(5):874–82.
- [141] Bereshchenko OR, Gu W, Dalla-Favera R. Acetylation inactivates the transcriptional repressor BCL6. *Nat Genet* 2002;32(4):606–13.
- [142] Lobry C, Oh P, Mansour MR, Look AT, Aifantis I. Notch signaling: switching an oncogene to a tumor suppressor. *Blood* 2014;123(16):2451–9.
- [143] Rossi D, Ciardullo C, Gaidano G. Genetic aberrations of signaling pathways in lymphomagenesis: revelations from next generation sequencing studies. *Semin Cancer Biol* 2013;23(6):422–30.
- [144] Aster JC, Blacklow SC, Pear WS. Notch signalling in T-cell lymphoblastic leukaemia/lymphoma and other haematological malignancies. *J Pathol* 2011;223(2):262–73.
- [145] Weng AP, Ferrando AA, Lee W, Morris JPt, Silverman LB, Sanchez-Irizarry C, et al. Activating mutations of NOTCH1 in human T cell acute lymphoblastic leukemia. *Science* 2004;306(5694):269–71.
- [146] Fabbri G, Rasi S, Rossi D, Trifonov V, Khiabanian H, Ma J, et al. Analysis of the chronic lymphocytic leukemia coding genome: role of NOTCH1 mutational activation. *J Exp Med* 2011;208(7):1389–401.
- [147] Kiel MJ, Velusamy T, Betz BL, Zhao L, Weigelin HG, Chiang MY, et al. Whole-genome sequencing identifies recurrent somatic NOTCH2 mutations in splenic marginal zone lymphoma. *J Exp Med* 2012;209(9):1553–65.
- [148] Karube K, Martinez D, Royo C, Navarro A, Pinyol M, Cazorla M, et al. Recurrent mutations of NOTCH genes in follicular lymphoma identify a distinctive subset of tumours. *J Pathol* 2014;234(3):423–30.
- [149] Stilgenbauer S, Schnaiter A, Paschka P, Zenz T, Rossi M, Dohner K, et al. Gene mutations and treatment outcome in chronic lymphocytic leukemia: results from the CLL8 trial. *Blood* 2014;123(21):3247–54.
- [150] Morin RD, Mendez-Lago M, Mungall AJ, Goya R, Mungall KL, Corbett RD, et al. Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. *Nature* 2011;476(7360):298–303.
- [151] Pasqualucci L, Dominguez-Sola D, Chiarenza A, Fabbri G, Grunn A, Trifonov V, et al. Inactivating mutations of acetyltransferase genes in B-cell lymphoma. *Nature* 2011;471(7337):189–95.
- [152] Morin RD, Johnson NA, Severson TM, Mungall AJ, An J, Goya R, et al. Somatic mutations altering EZH2 (Tyr641) in follicular and diffuse large B-cell lymphomas of germinal-center origin. *Nat Genet* 2010;42(2):181–5.
- [153] Graham C, LeBrun DP. Tumor suppressors in follicular lymphoma. *Leuk Lymphoma* 2015;56(7):1981–8.
- [154] Kirschbaum M, Frankel P, Popplewell L, Zain J, Delioukina M, Pullarkat V, et al. Phase II study of vorinostat for treatment of relapsed or refractory indolent non-Hodgkin's lymphoma and mantle cell lymphoma. *J Clin Oncol* 2011;29(9):1198–203.
- [155] Ogura M, Ando K, Suzuki T, Ishizawa K, Oh SY, Itoh K, et al. A multicentre phase II study of vorinostat in patients with relapsed or refractory indolent B-cell non-Hodgkin lymphoma and mantle cell lymphoma. *Br J Haematol* 2014;165(6):768–76.
- [156] Cheah CY, Fowler NH, Wang ML. Breakthrough therapies in B-cell non-Hodgkin lymphoma. *Ann Oncol* 2016;27(5):778–87.
- [157] Tiacci E, Trifonov V, Schiavoni G, Holmes A, Kern W, Martelli MP, et al. BRAF mutations in hairy-cell leukemia. *N Engl J Med* 2011;364(24):2305–15.
- [158] Tiacci E, Park JH, De Carolis L, Chung SS, Broccoli A, Scott S, et al. Targeting mutant BRAF in relapsed or refractory hairy-cell leukemia. *N Engl J Med* 2015;373(18):1733–47.
- [159] Dietrich S, Pircher A, Endris V, Peyrade F, Wendtner CM, Follows GA, et al. BRAF inhibition in hairy cell leukemia with low-dose vemurafenib. *Blood* 2016;127(23):2847–55.
- [160] Falini B, Martelli MP, Tiacci E. BRAF V600E mutation in hairy cell leukemia: from bench to bedside. *Blood* 2016;128(15):1918–27.