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Antibodies against PfEMP1, RIFIN, MSP3 and GLURP Are Acquired during Controlled *Plasmodium falciparum* Malaria Infections in Naïve Volunteers

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Abstract

Antibodies to polymorphic antigens expressed during the parasites erythrocytic stages are important mediators of protective immunity against *P. falciparum* malaria. Therefore, polymorphic blood stage antigens like MSP3, EBA-175 and GLURP and variant surface antigens PfEMP1 and RIFIN are considered vaccine candidates. However, to what extent these antibodies to blood stage antigens are acquired during naïve individuals’ first infections has not been studied in depth. Using plasma samples collected from controlled experimental *P. falciparum* infections we show that antibodies against variant surface antigens, PfEMP1 and RIFIN as well as MSP3 and GLURP, are acquired during a single short low density *P. falciparum* infection in non-immune individuals including strain transcendent PfEMP1 immune responses. These data indicate that the immunogenicity of the variant surface antigens is similar to the less diverse merozoite antigens. The acquisition of a broad and strain transcendent repertoire of PfEMP1 antibodies may reflect a parasite strategy of expressing most or all PfEMP1 variants at liver release optimizing the likelihood of survival and establishment of chronic infections in the new host.

Introduction

Malaria caused by *Plasmodium falciparum* constitutes a major burden to large parts of the world despite efforts to reduce transmission and increase treatment. In malaria endemic populations, immunity to malaria is acquired slowly as a function of experienced infections. In regions with stable malaria transmission immunity to uncomplicated malaria is not acquired until adolescence [1] whereas protection against severe, non-cerebral malaria and death is obtained after only a few infections [2]. This development of immunity is thought to reflect the gradual acquisition of effective cells and antibodies directed against malaria polymorphic and variable *P. falciparum* blood stage antigens [3–6]. Identification of these effective antibodies may lead to a vaccine mimicking the natural acquired protection against malaria. Identified polymorphic blood stage vaccine candidates include merozoite surface protein 3 (MSP3), erythrocyte-binding antigen 175 (EBA-175) and glutamate-rich protein (GLURP). MSP3 and EBA-175 are located on the extracellular merozoite surface and are involved in red blood cell invasion [7,8] while GLURP is expressed in both the pre-erythrocytic and erythrocytic stage [9] but no function has yet been ascribed. Of considerable interest are also the variant surface antigens *Plasmodium falciparum* erythrocyte membrane protein 1 (PIEMP1) and RIFINs. PIEMP1 mediate adhesion to human endothelial receptors [5,10–13] probably to avoid clearance by the spleen [14] whereas the function of RIFINs is yet unknown, although it is proposed that they expose their highly polymorphic V2 region on the surface of infected erythrocytes and therefore contributing to the antigenic variation of a *P. falciparum* infection [15,16]. PIEMP1 molecules are encoded by a repertoire of around 60 different *var* genes per genome but are generally thought to be expressed one at a time [17,18], although the co-expression of two different PIEMP1 variants has been observed in the laboratory-cultured parasite clone 3D7 [19]. The extracellular variable part of PIEMP1 contains an N-terminal segment followed by segments composed of two main domain types, Duffy binding-like domains (DBL) and cysteine-rich inter-domain regions (CIDR), which can be further divided into classes and sub-classes based on sequence similarity [20]. The *var* genes are divided into four main groups (A, B, C and VAR2CSA), each group shares specific 5’ promoter regions and phylogenetically distinct DBL and CIDR domains [20,21,22]. The VAR2CSA PIEMP1 is involved in pregnancy malaria by facilitating parasite sequestration in the placenta [13]. Moreover, immunological studies imply that an antigenically conserved subset of PIEMP1
are associated with severe disease in children [23,24,25]. Recently, it was shown that children living in areas of high *P. falciparum* transmission gradually but most rapidly acquire a broad anti-PfEMP1 antibody repertoire and antibodies against DBL domains of the group A PfEMP1 variants are acquired first [26]. Group A PfEMP1 have previously been associated with severe childhood malaria by studies of both var expression and PfEMP1 antibody acquisition [27–31]. Studies of var expression in controlled experimental infections of naïve Dutch individuals infected with the NF54 parasite strain (parental strain of the 3D7 clone) have suggested that most of the different parasites released from the liver cells express different var genes and that continuous growth may favour parasite expressing PfEMP1 variants facilitating the most effective sequestration to host endothelium [32]. However, it has not been known if the short period of infection (1–5 days or 1–3 post liver release parasite life cycles) in experimentally infected volunteers and the parasite densities obtained (low maximum parasitaemia <44,000 parasites/ml) are sufficient to induce antibody responses to the above mentioned bloodstage antigens. Therefore, plasma samples collected from controlled malaria infections of naïve volunteers [33–37] were used to investigate the acquisition of antibodies to an array of 104 PfEMP1 domains, eight RIFINs, MSP3, EBA-175, and GLURP.

**Materials and Methods**

**Ethics statement**

Informed consent form was signed by all subjects and the trials were reviewed and approved by the Ethical Committee of the Radboud University Nijmegen Medical Center (CWOM: 2004-0090, 0011-0262, 2001/203, 2002/170, and 2004/129) and the Central Committee for Research Involving Human Subjects of The Netherlands (CCMO NL24193.091.09) as previously described [33–38].

**Experimental infections of human volunteers and plasma samples**

This study benefited from the plasma collected from naïve volunteers infected with the *Plasmodium falciparum* NF54 isolate by bites of *Anopheles stephens* mosquitoes, and treated after a few rounds of asexual parasite multiplication, when parasites could be detected or the patient developed symptoms [33–37]. These controlled human malaria infection (CHMI) experiments were conducted in six series, between which the experimental set up (number of infectious mosquito bites, day of treatment relative to the initiation of the experiment) were different [37]. The microspheres were centrifuged for 1 min at 16,000 x g. One millilitre of distilled water was added to the supernatant and incubated for 2 h at room temperature, vortexed for 1 min, and transferred to Eppendorf tubes. The supernatant was removed after centrifugation for 1 min at 16,000 x g. One millilitre of distilled water was added to the microspheres, vortexed for re-suspension, followed by centrifugation for 1 min at 16,000 x g. The supernatant was removed and 1 ml of activation buffer (0.1 M NaH2PO4 [pH 6.2]) was added to the pellet and vortexed for re-suspension. In separate tubes 1-ethyl-3-[3-dimethylaminopropyl]-carbodiimide hydrochloride (EDC) and N-hydroxysulfosuccinimide (Sulfo-NHS; Pierce Biotechnology) were added to the microspheres, vortexed, and incubated at room temperature for 20 min with inversions in the dark. The microspheres were centrifuged for 1 min at 16,000 x g, re-suspended in 1 ml of 50 mM MES (pH 5.0), centrifuged for 1 min at 16,000 x g, and the supernatant was removed. The MES wash was repeated. The microspheres were re-suspended in 500 μl of MES. In separate tubes, the different protein samples (100 μg per ml of microspheres) were mixed with MES to a final volume of 500 μl and each was added to a separate microsphere population and incubated at room temperature for 2 h in the dark with inversions. The microspheres were centrifuged for 1 min at 16,000 x g and the supernatant was removed. The microspheres were washed twice in 1 ml of PBS/TBN (0.02% Tween 20, 0.1% BSA, and 0.05% sodium azide in PBS [pH 7.4]). The microspheres were re-suspended in 1 ml of PBS/TBN and stored at 4°C in the dark. To determine whether coupling was effective, aliquots of the different
microsphere sets were prepared for analysis as described below and analyzed on the Luminex instrument.

**Multiplexing and lyophilization of microspheres**

Equal volumes of the coated microspheres were pooled together and mixed by vortexing. Sucrose and Tween 20 were added to 3% and 0.05%, respectively, mixed by vortexing, and single-use aliquots were lyophilized (Advantage, Wizard 2.0; VirTis) in polypropylene vials, sealed under nitrogen gas, and stored at −80°C. Immediately before use, lyophilized microspheres were reconstituted with distilled water and used for analysis [43] as described below.

**Analysis of coupled microspheres on the Luminex**

The coated microspheres were diluted 1/333 in assay buffer E (ABE buffer, 0.1% BSA, 0.05% Tween 20, 0.05% sodium azide in PBS (pH 7.4)) and 50-μl aliquots were dispensed into the wells of a 1.2-μm filter bottom 96-well microtiter plate (MSBVS 1210; Millipore) pre-wetted with ABE buffer. The microspheres in 96-well plates were washed three times with ABE using a vacuum manifold (Millipore). Frozen plasma samples were thawed at room temperature, mixed by vortexing, and spun at 16,000 x g for 5 min to remove particulates. Plasma samples were diluted 1/180 in ABE buffer and 50-μl aliquots of diluted sample were added to the microspheres and incubated in the dark on a shaking platform at 1100 rpm for 30 s followed by 300 rpm for 30 min. Excess antibody was removed using a vacuum manifold followed by three washes in ABE. 25 μl of biotinylated human IgG (Sigma-Aldrich) detection antibody diluted 1/500 in ABE was added to the microspheres, incubated in the dark with shaking at 1100 rpm for 30 s, followed by 300 rpm for 30 min and washed three times in ABE. 50 μl of streptavidin-PE (Sigma-Aldrich) diluted 1/500 in ABE was added to the microspheres and incubated in the dark with shaking at 1100 rpm for 30 s, followed by 300 rpm for 10 min. Excess streptavidin-PE was removed followed by three washes in ABE. The microspheres were then re-suspended in 125 μl of ABE and analyzed on the Luminex instrument. The reader was set to read a minimum of 100 microspheres per microsphere region and results were expressed as median fluorescent intensity.

**Statistical analyses**

The purpose of the study was to determine to which degree the volunteer’s acquired antibodies to the tested malaria antigens. To this end the reactivity (measured in MFI) in the sample collected from each individual prior to malaria exposure was used as a baseline. Individuals were defined as having acquired an IgG response if the MFI in the sample collected during the experiment/MFI in the sample collected at the beginning of the experiment was >3 and the MFI in the sample collected during the experiment was >500. This conservative cut-off definition was used to assure that responder status not reflected general increases in IgG levels or small variations in background levels. To compare acquisition of antibodies between those who were experimentally infected and those who were protected by repeated immunizations before challenge we used Wilcoxon rank-sum test for unpaired data to compare the number of PfEMP1 maximum parasitemia, number of mosquito bites and number of asexual cycles completed before treatment was done using Spearman’s rank sum test. Stata 12 (http://www stata.com/statat12/) was used for the analyses.

**Results**

**Acquisition of IgG to GLURP, MSP3, and EBA-175**

Antibody levels to N-terminal, Central and C-terminal repeat regions (R0, R1 and R2) of GLURP, C-terminal region of MSP3 and the N-terminal F2 region of EBA-175 were measured in plasma collected from 44 naïve Dutch volunteers before and after a controlled human malaria infection with Plasmodium falciparum isolate NF54 which is a relatively short infection period. Around 70% of the volunteers acquired IgG against R2 whereas IgG to R0 and R1 only were acquired by approximately 13% (Figure 1). IgG with specificity to MSP3 was acquired by ~10%. None of the volunteers acquired IgG antibodies to EBA-175 (Figure 1).

**Acquisition of anti-PfEMP1 and anti-RIFIN IgG**

IgG reactivity to 104 recombinant PfEMP1 domains amplified from 3D7/NF54, HB3, IT/FCR3 and Dd2 genomes were measured in plasma samples from the same 44 individuals before and after P. falciparum infection. Of those 104 domains, 79 were recognized by between 3 and 12% of the volunteers (Figure 2) with no clear distinction in the recognition of domains according to domain subclass or PfEMP1 group. The pattern of antibody acquisition varied considerably among the 44 volunteers, but could be divided into six groups represented by their acquisition pattern (Figure 3). The 44 volunteers were grouped into individuals who did not acquire antibodies to any of the malaria antigens (n = 3; 7%), individuals who acquired antibodies to GLURP only (n = 11; 25%), and individuals who acquired antibodies to 1, 2, 3 or ≥3 PfEMP1 domains as well as to one or more of the merozoite antigens (n = 12, 5, 4 and 9; in total ~68%). 41% of the 44 volunteers responded to a non-NF54 PfEMP1 domains and of the 45 PfEMP1 domains amplified from P. falciparum HB3, IT/FCR3 and Dd2 genomic DNA, 31 were recognized (Figure 2). The pattern of antibody acquisition did not correlate with the parasite load, maximum parasitemia, number of mosquito bites or number of intra-erythrocytic parasite cycles. Antibody levels to the extracellular polymorphic domain [15], V2, of eight recombinant RIFIN proteins were also tested. Six volunteer acquired antibodies to in total four of the eight RIFINs (Figure 4).

**Acquisition of antibodies to blood stage antigens during an immunization study**

During an immunization study at Radboud University, Nijmegen [37], 10 volunteers were immunized by NF54 parasites by the bites of infectious mosquitoes three times with an interval of

**Table 1. Proportion of malaria naïve volunteers acquiring IgG against GLURP, MSP3 and EBA175 antigens after controlled experimental P. falciparum infections**

<table>
<thead>
<tr>
<th>Protein code</th>
<th>Gene</th>
<th>Genome</th>
<th>Domain</th>
<th>21</th>
<th>35</th>
</tr>
</thead>
<tbody>
<tr>
<td>R2 GLURP</td>
<td>na</td>
<td>F32</td>
<td>R2, C-terminal</td>
<td>73.5</td>
<td>66.7</td>
</tr>
<tr>
<td>R0 GLURP</td>
<td>na</td>
<td>F32</td>
<td>R0, N-terminal</td>
<td>14.7</td>
<td>13.3</td>
</tr>
<tr>
<td>R1 GLURP</td>
<td>na</td>
<td>F32</td>
<td>R1, Central</td>
<td>11.8</td>
<td>0.0</td>
</tr>
<tr>
<td>MSP3</td>
<td>na</td>
<td>F32</td>
<td>C-terminal</td>
<td>5.9</td>
<td>10.0</td>
</tr>
<tr>
<td>EBA-175</td>
<td>MAL7P1.176</td>
<td>3D7</td>
<td>F2, N-terminal</td>
<td>0.0</td>
<td>0.0</td>
</tr>
</tbody>
</table>

Figure 1. A proportion of malaria naïve volunteers acquiring IgG against GLURP, MSP3 and EBA-175 antigens after controlled experimental P. falciparum infections. The percentage of malaria infected volunteers with a measurable IgG response against GLURP R0, R1, R2, MSP3 and EBA175 malaria antigens on day 21 (N = 34) and/or day 35, 42 or 90 (N = 30) post infection in descending order. In total 44 volunteers were included (Table S1) and the IgG response was measured by bead Luminex technology.

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one month and concurrently receiving chloroquine, a drug that kills blood stage parasites. One month after discontinuation of the drug, when chloroquine levels were below therapeutic concentrations, the volunteers were challenged with NF54 by mosquito bites. A progressively reduced incidence and burden of blood stage parasitemia was measured by PCR during the immunization phase while there was no evidence of blood stage infection during the challenge phase [37]. These individuals acquired anti-GLURP R2 IgG after the first immunization and the antibody levels decreased after the third immunization. The peak anti-R2 IgG levels in the immunized group were comparable to the peak levels among the 44 individuals exposed to one infection (Figure 5). By contrast the acquisition of anti-PfEMP1 IgG was lower among the immunized individuals than among the individuals exposed to one

Figure 2. Proportion of malaria naïve volunteers acquiring IgG against PfEMP1 antigens after controlled experimental P. falciparum infections. The percentage of malaria infected volunteers with a measurable IgG response against 79 PfEMP1 malaria antigens on day 21 (N = 34) and/or day 35, 42 or 90 (N = 30) post infection in descending order. The PfEMP1 domain classification was according to Rask et al [19]. 25 DBL and CIDR domains were omitted from the figure as there was no IgG recognition of these domains (see Table S2). In total 44 volunteers were included (Table S1) and the IgG response was measured by bead-based Luminex technology.
doi:10.1371/journal.pone.0029025.g002
infection. The immunized individuals acquired antibodies to 0
(median and 95% confidence interval, CI) domains. Among
the 44 individuals exposed to one infection the number was 1
(0;1 9) (P = 0.02 compared to the immunized individuals). Among
the five individuals serving as controls in the immunization study
the number was 1 (0;2) (P = 0.10 compared to the immunized
individuals). Seven of the 10 immunized volunteers did not acquire
IgG against any of the PfEMP1 domains. Figure 6 shows the
reactivity pattern in one of the three volunteers who responded to
PfEMP1 during the study and one who did not. Among the
immunized individuals one acquired IgG to a single RIFIN and
none acquired IgG to the GLURP R0, GLURP R1, MSP3 or
EBA-175 domains.

Discussion

This study was conducted to assess the immune response to
PfEMP1 in experimentally infected individuals. Malaria naı ¨ ve
individuals were infected with P. falciparum after bites by
mosquitoes and the infection progressed through the liver, but
was terminated by drug treatment of blood stage parasites after a
few asexual multiplication rounds. Since the PfEMP1 repertoire of
the NF54 parasites used to infect the volunteers is known, and we
had access to recombinant PfEMP1 domains made on basis of
NF54 sequences and domains made on the basis of PfEMP1s from
other parasites, we could assess the acquisition of antibodies
recognizing homologues sequences as well as acquisition of cross
reactive antibodies. Despite the short exposure to asexual blood
stage parasites two thirds of the 44 volunteers acquired PfEMP1
antibodies. The breadth of antibody response varied considerably
between the volunteers. Some only acquired antibodies to one of
the 104 PfEMP1 domains tested while one volunteer acquired
antibodies to 38 domains. We could not establish any association
between the course of infection in the volunteers (parasite load,
maximum parasitemia, number of asexual multiplication cycles)
and the breadth of the PfEMP1 antibody response. Thus the
variation between the individuals may reflect individual differences
in the regulation and induction of immune response between the
individuals or stochastic variation. PfEMP1 can be divided into
group A-C and VAR2CSA. Antibodies were acquired to domains
present in all groups without any particular order or pattern. This
is in agreement with the hypothesis supported by var transcript
analyses in the same volunteers, that all or most variants are expressed by the parasite population at the onset of the blood stage infection to maximize the survival in a new host with unknown immune status [32,46]. The same var transcript analyses also provided the only formal evidence in support for the hypothesis that parasites that express a subset of PfEMP1, potentially due to better cytoadhesion properties, bestow higher parasite growth rates in naive individuals. The PfEMP1 antibody acquisition data presented here could not verify this hypothesis, probably because antibody measurements detect the accumulated acquired antibody repertoire and not directly reflects parasite densities. In 2007, Elliot et al [47] showed that individuals who had experienced brief infections during travels in Africa, Asia or the Western Pacific had acquired IgG recognizing variant surface antigens (VSA) on six different P. falciparum lines, indicating that IgG against VSA are broadly cross reactive. In our study, individuals were exposed to the defined repertoire of PfEMP1 present in NF54. Interestingly, 41% of the volunteers acquired IgG to non-NF54 PfEMP1 domains and two thirds of the 45 non-NF54 PfEMP1 domains were recognized by at least one volunteer. We have previously shown that there is little cross reactivity between antibodies directed against PfEMP1 domains produced on a 3D7 genetic background [48]. The present study indicates that even short exposure to a particular PfEMP1 domain can induce antibodies which react with PfEMP1 domains encoded from different genomes. Combined the studies suggest that there is little serological cross reactivity between intragenomically encoded PfEMP1 and broad cross reactivity between inter-genomically encoded PfEMP1. This cross reactive PfEMP1 immune response, which cannot in these experiments be explained by a sequential exposure to PfEMP1 epitopes, could explain why children in endemic areas relatively quickly generate a broad repertoire of anti-PfEMP1 antibodies, [26,49]. Such an antibody response may also enhance the parasites likelihood of establishing a chronic infection [50].

Figure 5. Acquisition of IgG to GLURP R2 during a liver stage immunization study. The anti-GLURP R2 IgG levels in 44 volunteers experimentally infected with Plasmodium falciparum (indicated by a thick arrow day one) and in 10 volunteers immunized by three exposures to P. falciparum while treated with chloroquine killing blood stage parasites (indicated by thin blue arrows day one, 33 and 61) and after challenge without a drug cover (thick blue arrow day 118). “35+” represents plasma samples taken 35, 42 or 90 days after infection. CHMI: controlled human malaria infection. doi:10.1371/journal.pone.0029025.g005

Figure 6. Acquisition of IgG Abs to 104 PfEMP1 domains, eight RIFINs, and GLURP, MSP3, and EBA-175 antigens in two immunized volunteers. The two volunteers, 1 and 2, were selected to represent volunteers with acquisition of antibodies to PfEMP1 and merozoite antigens (n = 3) and volunteers with acquisition of antibodies to the merozoite antigens only (n = 7). The letters A, B, C, D and E correspond to 32, 60, 116, 151 and 256 days after first immunization. Parasite challenges were at day 118 [37]. The results are expressed as median fluorescent intensity (MFI): Red >2000 MFI; Green > 1000 MFI; Yellow > 500 MFI (cut-off value). doi:10.1371/journal.pone.0029025.g006
We also assessed the acquisition of antibodies to selected merozoite antigens and RIFINs. Two thirds of the volunteers acquired antibodies to the C-terminal R2 repeat region of GLURP, while only between 0–15% acquired antibodies to GLURP R0, GLURP R1, MSP3, EBA-175 and the RIFINs. This is in agreement with a previous observation where no antibodies were found against a GLURPR0R1 long synthetic peptide which did not include the R2-region [37]. The high recognition of GLURP R2 could reflect that this region is particularly immunogenic [51] and/or that the antibodies are induced by liver parasites expressing GLURP [9]. It has previously been reported that antibodies to GLURP R2 and other malaria antigens can be induced by transient exposure to *P. falciparum* [52–55].

We also investigated the antibody response in individuals who were immunized by three exposures to infected mosquito bites while under chloroquine treatment. Surprisingly, these immunized individuals were protected, when they receive a fourth exposure to infected mosquitoes in the absence of chloroquine [37]. During immunization, the attenuated infections gave raise to brief very low density blood stage parasitemia. The question was whether these brief episodes were sufficient to induce anti-PfEMP1 antibodies, which could mediate protection during challenge. Our data do not support this hypothesis, since only a few of the immunized individuals acquired anti-PfEMP1 antibodies to a few of the domains. Similarly, the immunized individuals did not acquire antibodies to the most of the other blood stage antigens tested. The exception was anti-GLURP R2 antibodies, which were acquired after the first parasite exposure in the immunized individuals at levels that were similar to those measured in the non-immunized volunteers. In the immunized individuals, these antibodies could have been induced by a brief exposure to low levels of asexual parasites or induced by GLURP or other glutamine rich proteins expressed by parasites stages in the pre-erythrocytic life cycle [9].

In conclusion, PIEMP1, RIFIN, GLURP and MSP3 antibodies are acquired after short controlled *P. falciparum* infections suggesting that the immunogenicity of the variant surface antigens is similar to the less diverse merozoite antigens and broad and strain transient PIEMP1 reactivity may reflect a parasite strategy of expressing most or all PIEMP1 variants at liver release optimizing the likelihood of survival and establishment of chronic infections in the new host.

Supporting Information

Table S1 Overview of the 54 volunteers involved.

Table S2 Primers used for *Plasmodium falciparum* antigen expression.

Acknowledgments

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Author Contributions

Conceived and designed the experiments: LT CW TL RWS CCH TGT. Performed the experiments: LT. Analyzed the data: LT CW TL RWS CCH TGT. Wrote the paper: LT CW TL RWS CCH TGT.

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