The following full text is a publisher's version.

For additional information about this publication click this link.
http://hdl.handle.net/2066/96386

Please be advised that this information was generated on 2019-11-14 and may be subject to change.
TLR1/TLR2 Heterodimers Play an Important Role in the Recognition of Borrelia Spirochetes

Marije Oosting1,2, Hadewych ter Hofstede1, Patrick Sturm3, Gosse J. Adema4, Bart-Jan Kullberg1,2, Jos W. M. van der Meer1,2, Mihai G. Netea1,2, Leo A. B. Joosten1,2*

1 Department of Medicine, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, 2 Nijmegen Institute of Infection, Inflammation and Immunity (N4i), Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, 3 Department of Microbiology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, 4 Department of Tumor Immunology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands

Abstract

After infection with Borrelia species, the risk for developing Lyme disease varies significantly between individuals. Recognition of Borrelia by the immune system is mediated by pattern recognition receptors (PRRs), such as TLRs. While TLR2 is the main recognition receptor for Borrelia spp., little is known about the role of TLR1 and TLR6, which both can form functionally active heterodimers with TLR2. Here we investigated the recognition of Borrelia by both murine and human TLR1 and TLR6. Peritoneal macrophages from TLR1- and TLR6-gene deficient mice were isolated and exposed to Borrelia. Human PBMCs were stimulated with Borrelia with or without specific TLR1 and TLR6 blocking using specific antibodies. Finally, the functional consequences of TLR polymorphisms on Borrelia-induced cytokine production were assessed. Splenocytes isolated from both TLR1−/− and TLR6−/− mice displayed a distorted Th1/Th2 cytokine balance after stimulation with B.burgdorferi, while no differences in pro-inflammatory cytokine production were observed. In contrast, blockade of TLR1 with specific neutralizing antibodies led to decreased cytokine production by human PBMCs after exposure to B.burgdorferi. Blockade of human TLR6 did not lead to suppression of cytokine production. When PBMCs from healthy individuals bearing polymorphisms in TLR1 were exposed to B.burgdorferi, a remarkably decreased in vitro cytokine production was observed in comparison to wild-type controls. TLR6 polymorphisms lead to a minor modified cytokine production. This study indicates a dominant role for TLR1/TLR2 heterodimers in the induction of the early inflammatory response by Borrelia spirochetes in humans.

Editor: Jean-Pierre Gorvel, Universite de la Mediterranee, France
Received May 23, 2011; Accepted September 15, 2011; Published October 5, 2011

Copyright: © 2011 Oosting et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was supported by the Dutch Arthritis Association (NR 10-1-303). M.G.N. was supported by a Vici grant of the Netherlands Organization for Scientific Research. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: ljoosten@kig.umcn.nl

Introduction

Ticks of the Ixodes family are able to transmit bacteria of the Borrelia burgdorferi sensu stricto family, which causes Lyme Disease [1]. Within this family, three species are described to be pathogenic, namely B.burgdorferi sensu stricto, B.afzelii, and B.garvani, which are differentially distributed between the United States and Europe. Clinical signs that develop after infection with Borrelia spirochetes are diverse, ranging from skin abnormalities (erythema migrans) to arthritis or carditis [2]. Infection with Borrelia results in release of inflammatory mediators and recruitment of inflammatory cells to the site of infection [3,4]. To induce inflammation, recognition of the bacteria by pattern recognition receptors (PRRs) is necessary.

Distinct classes of PRRs have been described, including C-type lectins (CLR), NOD-like receptors (NLRs), and Toll-like receptors (TLRs). TLR4, the main receptor for bacterial lipopolysaccharides [5], has been shown not to be involved in the recognition of Borrelia [6]. On the other hand, TLR2 plays an important role in the recognition of Borrelia components. It has been demonstrated that TLR2-knockout mice produced significantly lower concentrations of antibodies against Borrelia after immunization with OspA [7]. TLR2 knockout mice harbor higher amounts of Borrelia spirochetes at the site of inflammation than wild-type animals and hence exhibit more cell influx in infected joints [8,9]. In humans, TLR2 can recognize several components of Borrelia [10,11], and TLR2 blockade in human peripheral blood mononuclear cells (PBMCs) resulted in decreased cytokine production after exposure to intact Borrelia spirochetes [6]. In order to recognize its ligands, TLR2 forms heterodimers with other members of the TLR family (TLR1 or TLR6) [12]. TLR1/TLR2 heterodimers recognize mainly triacylated lipopeptides, whereas TLR2/TLR6 heterodimers recognize diacylated lipopeptides [13,14]. TLR1/2 heterodimers induce also a different immune response against pathogens as compared to TLR2/6 heterodimers [15,16]. When TLR1/2 molecules are absent, less induction of early cytokines were observed, whereas TLR2/6 seems able to modulate the balance between a Th1/Th2 immune response.

Limited information is available about recognition of B. burgdorferi by TLR1 and TLR6 in murine and human cell systems, and the relative contribution of these receptors as components of the heterodimers with TLR2 for the recognition of Borrelia species has not been elucidated for primary cells [7,17–20]. In addition, mutations in TLR1 and TLR6 receptors are associated with differential susceptibility to bacterial and fungal infections [15,21], and the question arises to what extent these polymorphisms may
lead to changes in production of cytokines after exposure to Borrelia species, and hence might influence the clinical outcome of Lyme disease. Thus, we investigated the role of TLR1 and TLR6 in the recognition of Borrelia species by mouse cells and primary human cells, and assessed whether polymorphisms in either the TLR1 or the TLR6 gene influence the cytokine responses. We observed an important role for TLR1/2 heterodimers for the recognition of Borrelia species and for the induction of an early immune response against Borrelia spirochetes in humans.

Results

TLR1 activation enhanced the induction of IFN-γ by murine splenocytes after exposure to B. burgdorferi

Peritoneal macrophages from TLR1 knockout animals produced the same amounts of pro-inflammatory cytokines IL-1β, IL-6, and TNF-α, as compared to cells isolated from wild-type C57Bl/6 mice (Figure 1A–C). However, a significant decrease in IL-6 production could be observed when TLR1-deficient splenocytes were stimulated with B. burgdorferi for 5 days (Figure 1D). Moreover, IFN-γ production induced by B. burgdorferi in TLR1-deficient splenocytes was significantly higher than in controls (Figure 1E). Finally, IL-17 production was somewhat higher after stimulation of TLR1 knockout cells, but not found to be statistically significant (Figure 1F).

Borrelia-induced IFN-γ production by murine cells is dependent on TLR6

To assess the role of TLR6, 1×10⁶ live Borrelia burgdorferi spirochetes were added to freshly isolated peritoneal macrophages of either wild-type or TLR6 gene-deficient mice. After 24 hours of stimulation, no differences in the production of pro-inflammatory cytokines IL-1β, IL-6, and TNF-α could be detected between cells isolated from wild-type or TLR6 knockout mice (Figure 2A, 2B, and 2C, respectively). Other cytokines, such as IL-10, IL-17 and IFN-γ are known to be involved in the immune response against Borrelia [23,24], splenocytes of wild-type or TLR6 knock-out mice (Figure 2A, 2B, and 2C, respectively). Other cytokines, such as IL-10, IL-17 and IFN-γ are known to be involved in the immune response against Borrelia [23,24]. Splenocytes of wild-type or TLR6 knock-out mice were incubated for 5 days with Borrelia. Thereafter, the production of IL-10, IFN-γ, and IL-17 was measured by ELISA (Figure 2D–F). A significant decrease in IFN-γ production was detected in mice lacking functional TLR6. IL-10 and IL-17 production tended to be lower in these mice, but these differences did not reach statistically significant (Figure 2D and 2F).

Differential role of human TLR1 and TLR6 for the recognition of Borrelia

To investigate the relative roles of TLR1 and TLR6 signalling for the recognition of Borrelia by human primary cells, neutralizing antibodies were used to inhibit the function of these specific TLRs. When PBMCs from 5 healthy individuals were incubated for 24 hours with blocking anti-TLR1, anti-TLR6, anti-TLR2 antibodies, or control IgG1 antibody alone, no production of pro-inflammatory cytokines and chemokines could be detected (data not shown). Inhibition of cytokines produced upon Pam3Cys stimulation could only be observed when using the anti-TLR1 antibody and could not be inhibited by TLR6 antibodies (not shown). When human PBMCs were incubated with B. burgdorferi in the presence of a specific anti-TLR1 antibody, a significant reduction in IL-1β, IL-6 (but not TNFα), and chemokine IL-8 production was observed (Figure 3A–D). In contrast to TLR1, human TLR6 seems to play a minor role in the induction of pro-inflammatory cytokines after Borrelia stimulation of PBMCs. Production of IL-6, IL-1β, or IL-8 was not significantly inhibited after a neutralizing TLR6 antibody was added to the cultures (Figure 3A–D). As demonstrated previously, TLR2 is important for the induction of cytokine responses by Borrelia in human PBMCs (Figure 3A–D). No significant differences in cytokine production could be observed when Borrelia was co-incubated with IgG antibody alone (Figure 3A–D).

The role of human TLR1 polymorphisms in cytokine production by B. burgdorferi

PBMCs isolated from individuals carrying different TLR1 genotypes were incubated for 24 hours with Borrelia, or the TLR2 specific ligand Pam3Cys. Cytokine production was significantly lower after stimulation of PBMCs with Pam3Cys in cells isolated from individuals homozygous for the R307T, N248S, and S602I polymorphisms in TLR1 (Figure 4). For the TLR1 R307T polymorphism, we observed a trend towards lower production of IL-1β after PBMC stimulation with 1×10⁶ B. burgdorferi microorganisms per mL (Figure 4A). These results are in line with previous reports showing that the presence of the TLR1 polymorphisms R307T, N248S and S602I led to decreased signalling and cytokine production after Pam3Cys stimulation [23–26]. A significantly decreased IL-1β production after Borrelia exposure could be detected in persons heterozygous or homozygous for the TLR1 polymorphism N248S or S602I (Figure 4B and 4C). Significant differences between the wild-type, heterozygous, and homozygous carriers of the TLR1 SNPs were found with respect to IL-6, IL-8, TNF-α, and IL-10 production, after incubation of their PBMCs with Borrelia or Pam3Cys (Supplementary tables S1, S2, and S3).

Human TLR6 polymorphisms are less involved in cytokine induction after stimulation with B. burgdorferi

The non-synonymous polymorphism S249P in the TLR6 gene, believed to result in a malfunction in the extracellular domain of the TLR6 molecule, is present with a high frequency in several different populations (>10%) [27]. However, after PBMC stimulation for 24 h with the specific TLR2/6 ligand FSL-1, we did not detect differences between individuals with or without the TLR6 polymorphism in either IL-1β, IL-6, IL-8, IL-10, or TNF-α (Figure 5A–E, respectively). On the other hand, we could observe lower IL-1β, IL-6, and IL-8 production by cells of individuals bearing the S249P SNP after stimulation with Borrelia burgdorferi.

Human TLR1 is involved in Borrelia-induced IFN-γ

Since we demonstrated that TLR1 controls the induction of IFN-γ in mice, we assessed whether TLR1 is involved in human IFN-γ and IL-17 responses upon encounter of Borrelia species. PBMCs isolated from healthy volunteers bearing either a TLR1 or TLR6 polymorphism were stimulated for 7 days with either medium, live Borrelia, or Pam3Cys. Pam3Cys-induced IFN-γ was less produced by cells isolated from individuals bearing TLR1 SNPs, but no differences could be observed for cells with the described TLR6 polymorphism (data not shown). IFN-γ levels after Borrelia stimulation were significantly decreased in individuals lacking a functional TLR1 molecule, whereas TLR6 seems not to play a major role in the induction of this pro-inflammatory cytokine (Fig. 6A–B, respectively). TLR1 might also play a role in the induction of IL-17 after Borrelia exposure, a trend towards lower production could be detected, although not found to be significant (Fig.6C). TLR6 seems not involved in Borrelia-induced IL-17 production (Fig.6D).

Discussion

The present study expands the knowledge regarding the role of both TLR1/2 and TLR2/6 heterodimers in the recognition of
Borrelia spp. We demonstrate that TLR1 in humans is an important component for the recognition and induction of an immune response against Borrelia. This was demonstrated by experiments using either TLR1- or TLR6 deficient murine cells, as well as studies performed in cells isolated from individuals bearing genetic polymorphisms in TLR1 or TLR6. Of high interest, a different pattern was found in murine cells; genetic disruption of TLR1 resulted in an upregulated IFN-γ response, whereas TLR6 deficient cells were unable to induce a sufficient IFN-γ response after Borrelia exposure. This difference in IFN-γ induction in mice between TLR1 or TLR6 could not be observed in humans. Neither TLR1 and TLR6 seem to be involved in the induction of this cytokine upon stimulation with Borrelia spirochetes.

TLR1/2 or TLR2/6 heterodimers do not only differ in structure, but also recognize different ligands. Whereas TLR1/2 heterodimers mainly recognize tri-acetylated structures such as Pam3Cys, di-acetylated proteins are mainly recognized by TLR2/6 heterodimers [13,14]. It is known that Borrelia spp. contains tri-acetylated proteins in the cell membrane and this fits with our observation that the TLR1 containing heterodimers are more important in recognition and signaling of Borrelia.

The genome of Borrelia burgdorferi encodes at least 105 membrane proteins, including the group of immunologically important outer surface proteins (OSPs) [28]. It has been suggested that Borrelia outer surface protein A (OspA) plays an important role in the pathogenesis of Lyme disease caused by

![Figure 1. In vitro cytokine production by TLR1−/− cells after stimulation with Borrelia.](image-url)
Borrelia species. Macrophages isolated from TLR1 deficient mice display an impaired ability to induce IL-6 after stimulation with OspA lipoprotein [7]. In contrast to these studies, we were unable to detect any differences in IL-6 production between wild-type and TLR1 knockout mice, when cells were stimulated with intact Borrelia spirochetes. This effect might be due to the presence of multiple ligands that are absent in studies that use solely purified OspA protein in supraphysiological concentrations. Using intact bacteria, the immune response is likely stimulated through several distinct PRR pathways, involving both TLRs and NODs [6].

Disruption of either the TLR1 or TLR6 gene in mice did not lead to changes in the pro-inflammatory cytokine production, with the exception of the T-cell-derived IFN-γ and IL-17. This implies that in the mouse the intact TLR1/TLR2 heterodimers dampen the interferon gamma response. This effect may be mediated through IL-10, since this cytokine was downregulated in mice lacking functional TLR1 molecules. It may be hypothesized that murine TLR1 and TLR6 exert counter-regulatory roles for the induction of Th1/Th17 cytokines by Borrelia bacteria.

Mutations in TLR1 can lead to a decreased surface protein expression of TLR1 on the cell surface and diminished activation of NF-κB [25,26]. It has been previously suggested that defects in TLR1 signaling might play a role in the early induction of the immune response in mice against Borrelia spp [7]. As genetic variants have been reported to modify the function of TLRs and influence susceptibility to infections in humans, we assessed the role of three SNPs in TLR1 and one SNP in TLR6 for their role in the induction of cytokines after recognition of Borrelia [21,26]. SNPs in TLR1

---

**Figure 2. Recognition of Borrelia species by immune cells of TLR6−/− mice.** Peritoneal macrophages (1 × 10⁵/well) of C57Bl/6 wild-type or TLR6 gene deficient mice (n = 5 per group) were stimulated for 24 hours with 1 × 10⁶ live Borrelia per mL. Using ELISA or RIA, IL-1β (A), IL-6 (B), or TNF-α (C) levels were measured in pg/mL. 5 × 10⁵ spleen cells/well were stimulated with 1 × 10⁶ live Borrelia per mL. IL-10, IFN-γ, and IL-17 levels were determined in the supernatant of 5-days spleen cell culture (D–F, respectively). White bars represent cytokine induction after stimulation of wild-type cells, black bars the TLR6 knockout cells. An asterisk indicates that the P-value is <0.05 (for comparisons between wild-type and knock-out mice), 5 animals per group, Mann-Whitney U-test. Bars represent the mean ± SEM, experiments were performed in duplicates.

doi:10.1371/journal.pone.0025998.g002
impair intracellular trafficking of the TLR1, which eventually leads to a decreased NF-κB signaling. It was described before that intracellular interactions between TLR1 and TLR2 are necessary for optimal immune signaling in human cells [29]. A different pattern of TLR involvement could be observed in peripheral blood mononuclear cells isolated from humans in comparison to the cytokines induced by mouse cells. Using cells of individuals with functional genetic polymorphisms in the TLR1 gene, we found a marked inhibition of the pro-inflammatory cytokine production when these cells were exposed to Borrelia. This was found in individuals carrying the polymorphism in both alleles (homozygous) for any of the three SNPs in TLR1 studied. In agreement with our data, Johnson et al exposed human monocytes with the S602I SNP in the TLR1 gene to TLR1/TLR2 agonists and also observed a blunted pro-inflammatory response [26].

In a recent report, the S602I SNP in TLR1 was linked to the N248S SNP in this gene [25]; also in our study these two SNPs displayed an 80% linkage. However, no linkage was observed with the third SNP in TLR1 (R80T). SNP N248S and R80T have previously been associated with invasive aspergillosis [21], and our finding of modulation of cytokines when cells with these genetic variations are stimulated with Borrelia warrants studies of these polymorphisms in patients with Lyme disease.

The presence of the TLR6 S249N SNP did also influence IL-1β, IL-6, and IL-8 production induced by Borrelia. TNF-α and IL-10 levels induced by Borrelia were not changed in the presence of the SNP. However, it did not influence cytokine responses induced by TLR6/2 ligand FSL-1. Until now, the precise function of this SNP is still unknown, although it has been negatively correlated with ulcerative colitis and the development of clinical signs of malaria, and probably offers protection against the development of asthma [27,30,31]. These genetic findings are in agreement with those we obtained in cultures in which human PBMCs were exposed to antibodies against TLR1 or TLR6. In these experiments we found that TLR6 had only a marginal effect on cytokine production (some effect on TNF-α production), whereas TLR1 clearly mediates Borrelia-induced cytokine production. TLR1 is also responsible for T-cell derived cytokines after recognition of Borrelia. We observed decreased IFN-γ and IL-17 production when TLR1 molecules were dysfunctional through the presence of polymorphisms.

In conclusion, the present study demonstrates an important role for TLR1/TLR2 heterodimers for the recognition of Borrelia in humans. Furthermore, the presence of genetic variants of TLR1 gene leads to impaired cytokine responses upon challenge of PBMCs with Borrelia. On the one hand, since initiation of host defense responses against
Borrelia is dependent on multiple pattern recognition receptors, more research is needed to elucidate the precise role of TLR1/2 in the pathogenesis of Lyme disease. On the other hand, these results give novel information regarding the mechanisms of Borrelia recognition and the role of TLR1 in this process, and warrants future studies in the role of this receptor for the susceptibility to Lyme disease.

Materials and Methods

**Borrelia burgdorferi cultures**

*B. burgdorferi*, ATCC strain 35210, was cultured at 33°C in Barbour-Stoenner-Kelley (BSK)-H medium (Sigma-Aldrich) supplemented with 6% rabbit serum. Spirochetes were grown to late-logarithmic phase and examined for motility by dark-field microscopy. Organisms were quantitated by fluorescence microscopy after mixing 10 μL aliquots of culture material with 10 μL of an acridine orange solution and counted using a Petroff-Hauser counting chamber. Bacteria were harvested by centrifugation of the culture at 7000 x g for 15 min., washed twice with sterile PBS (pH 7.4), and diluted in the specified medium to required concentrations of 1 x 10^6 spirochetes per mL. Heat-killed *B. burgdorferi* were prepared by heating cultured spirochetes at 52°C for 30 min. before dilution.

**Animals**

TLR1−/− and TLR6−/− mice were kindly provided by Dr. Shizuo Akira and are fully backcrossed to C57BL/6 background (Osaka University, Japan). C57BL/6 mice were obtained from Charles River Wiga (Sulzfeld, Germany). Female wild-type and knock-out mice between 8 and 14 weeks of age were used. The mice were fed sterilized laboratory chow (Hope Farms, Woerden, The Netherlands) and water ad libitum. The experiments were approved by the Ethics Committee on Animal Experiments of the Radboud University Nijmegen Medical Centre.

**In-vitro cytokine production**

Peritoneal macrophages were isolated by injecting 5 mL of ice-cold sterile PBS (pH 7.4) in the peritoneal cavity. After centrifugation and washing, cells were resuspended in Roswell Park Memorial Institute (RPMI) 1640 containing 1 mM pyruvate, 2 mM L-glutamine and 50 μg/mL gentamicin (culture medium). Cells were counted using a Z1 Coulter Particle Counter (Beckman Coulter, Woerden, The Netherlands) and adjusted to 1 x 10^6 cells/mL. Cells were cultured in 96-well round-bottom microtiter plates (Costar, Corning, The Netherlands) at 1 x 10^5 cells/well, in a final volume of 200 μL. After 24 hours of incubation of cells with different stimuli at 37°C in air and 5% CO2, the plates were centrifuged at 1400 x g for 8 min, and the supernatant was collected and stored at −20°C until cytokine assays were performed.

Spleen cells were isolated by gently squeezing spleens in a sterile 200 μm filter chamber. After washing with sterile PBS and centrifugation at 4°C (1200 rpm 3 min.), cells were resuspended in 4 mL RPMI 1640 in presence of 20% FCS. Cells were counted and concentrated were adjusted to 1 x 10^6 cells/mL. Cells were cultured in 24-wells plates (Greiner, Alphen a/d Rijn, The Netherlands) at 5 x 10^5 cells/well, in a final volume of 1000 μL. After 3 days of incubation, supernatant was collected and stored at −80°C until cytokine assays were performed.

**Study populations**

Individuals in this study were foresters from the ‘Geldersch Landschap’ and ‘Kroondomein het Loo’ in the Netherlands. In this cohort, Lyme disease occurs as an occupational disease and...
hence they were invited to participate. The foresters were between 23–73 years old, and consisted of 77% males and 23% females.

Samples of venous blood of them were drawn after informed consent was obtained. Experiments were conducted according to the principles expressed in the Declaration of Helsinki.

Isolation of genomic DNA and single nucleotide polymorphism analysis

DNA was isolated using the Gentra Pure Gene Blood kit (Qiagen), in accordance with the manufacturer’s protocol for whole blood. DNA was dissolved in a final volume of 100 μL buffer. Polymerase chain reaction (PCR) amplification of TLR1 and TLR6 gene fragments bearing the polymorphisms R80T (rs5743611), N248S (rs4833095), and S249P (rs5743810) were performed using a pre-designed TaqMan® SNP genotyping assay (Applied Biosystems) in 25 μL reactions containing 2 μL of genomic DNA as well as primers, two specific probes (with either VIC or FAM label) and Universal PCR 2x Master mix (Applied Biosystems). Cycling conditions were 2 min at 50°C and 10 min at 95°C followed by 40 cycles of 95°C for 15 sec and 1 min at 60°C. Fluorescence intensities were corrected using a post-read / pre-read method for 1 min at 60°C before and after the amplification. The software automatically plotted genotypes based on a two-parameter plot with an overall success rate of more than 95%.

Figure 5. Less important role for human TLR6 in the induction of cytokines after Borrelia stimulation. Peripheral blood mononuclear cells (PBMCs) from 128 healthy volunteers carrying the S249N SNP in TLR6 were stimulated with either medium, 1 × 10^6 per mL Borrelia spirochetes, or 1 μg/mL FSL-1 for 24 h and cytokines were measured using ELISA; (interleukin 1β [IL-1β] (A); IL-6 (B); IL-8 (C); IL-10, (D); and tumor necrosis factor α [TNF-α] (E)). Bars represent individuals carrying no SNP (wild-type, wt, white bars), heterozygous SNP carriers (he, grey bars), or homozygous variation (ho, black bars). Data represent the mean ± SEM, *p<0.05; **p<0.01; Mann-Whitney U-test. The data shown are from three independent experiments each performed in duplicate.

doi:10.1371/journal.pone.0025998.g005
Intermediate samples were excluded from the analysis. Conventional PCR amplification of the TLR1 gene fragment containing polymorphism S602I (rs5743618) was performed on the Icycler (BioRad) in 50μL reactions containing 2μL of genomic DNA, 1.5 mM MgCl2, Taq DNA polymerase conc. 5 U/μL (Invitrogen), PCR buffer, 20 mM dNTP (Pharmacia), 10μM Forward primer 5′-CTA CCC GGA AAG TTA TAG AGG AAC C, and 10μM Reversed primer 5′-TTT GGC AAT AAT TCA TTC ACC. PCR consisted of one initial denaturation phase of 95°C for 10 minutes followed by 40 cycles; each cycle consisted out of one annealing step of 95°C for 30 seconds, one polymerization step of 60°C for 30 seconds, and one elongation step of 72°C for 30 seconds. Subsequently, another cycle of 72°C for 7 minutes was performed before termination. PCR products were sequenced with either 4μM of primers according to the Sanger method supported by Big Dye Terminator version 3 of Applied Biosystems. After sequencing, samples were analyzed using the 3730 Sequence analyzer and Chromas 2.33 software (Technelysium).

Isolation of human peripheral blood mononuclear cells and in-vitro cytokine production

Venous blood was drawn from the cubital vein of foresters into 10 mL ethylenediaminetetraacetic acid (EDTA) tubes (Monoject). Peripheral blood mononuclear cells (PBMCs) were isolated according to standard protocols, with minor modifications. The PBMC fraction obtained by density centrifugation of blood diluted 1:1 in phosphate-buffered saline (PBS)-buffer over Ficoll-Paque (Pharmacia Biotech). Cells were washed three times in PBS and resuspended in RPMI 1640 (Dutch modified) supplemented with 50 μg/L gentamicin, 2 mM L-glutamin, and 1 mM pyruvate. Cells were counted in a Coulter Counter Z (Beckman Coulter), and adjusted to 5×10⁶ cells/mL. Mononuclear cells (5×10⁵) in a 100 μL volume were added to round-bottom 96-wells plates (Costar, Corning, The Netherlands) and incubated with either 100 μL of medium (negative control) or B. burgdorferi (1×10⁶ spirochetes per mL). In some experiments, PBMCs were pre-incubated with neutralizing antibodies for 30 minutes (functional grade anti-human Toll-like receptor 1 (10μg/mL, eBioscience), anti-TLR6 (10μg/mL, BioLegend), anti-TLR2 (10 μg/mL, eBioscience) or control antibody (mouse IgG1κ, 10 μg/mL, eBioscience)). After pre-incubation, B. burgdorferi or specific TLR ligands were added, such as Pam3Cys or FSL-1 (10μg/mL or 1μg/mL, respectively). After 24 hours or 7 days (in the presence of 10% human pool serum) supernatants were collected and stored at −20°C until being assayed.

Cytokine measurements

Concentrations of mouse IL-1β were determined by specific radioimmunoassay (RIA; detection limit is 20 pg/mL) [22].
Mouse IL-6, IL-17, IFN-γ, and IL-10 concentrations were measured by a commercial ELISA kit (Biosource, Camarillo, CA; detection limits 16 pg/mL), according to the instructions of the manufacturer. Concentrations of human IL-1β, IL-6, IL-17, or IFN-γ were determined in duplicates using either specific or commercial ELISA kits (PeliKinE Compact, Sanquin, Amsterdam, or R&D Systems, Minneapolis), in accordance with the manufacturers’ instructions. Detection limits were 40 pg/mL, except for IFN-γ ELISA (12 pg/mL).

Ethics statement
All experiments in this study were carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health, the Dutch law on animal experiments, and FELASA regulations. The protocol was approved by the Ethics Committee on Animal Experiments of the Radboud University Nijmegen Medical Centre. All efforts were made to minimize suffering of the animals.

All human experiments were conducted according to the principles expressed in the Declaration of Helsinki. Before taking blood, informed written consent of each human subject was provided. The study was approved by the review board of the department of Medicine of the Radboud University Nijmegen Medical Centre.

Statistical Analysis
The data are expressed as mean ± SEM unless mentioned otherwise. Differences between experimental groups were tested using the two-sided Mann-Whitney U test performed on GraphPad Prism 4.0 software (GraphPad). *P* values of ≤0.05 were considered significant.

References


Supporting Information

Table S1 Cytokine production in pictograms per milliliter after stimulation of PBMCs isolated from healthy volunteers carrying R30T SNP in TLR1 molecules. All values are depicted as means plusminus the standard error of the means.

Table S2 Cytokine production in pictograms per milliliter after stimulation of PBMCs isolated from healthy volunteers carrying N248S SNP in TLR1 molecules. All values are depicted as means plusminus the standard error of the means.

Table S3 Cytokine production in pictograms per milliliter after stimulation of PBMCs isolated from healthy volunteers carrying S602I SNP in TLR1 molecules. All values are depicted as means plusminus the standard error of the means.

Acknowledgments
The authors thank Carla Bartels for culturing and counting Borrelia spirochetes.

Author Contributions
Conceived and designed the experiments: MO HH PS GJA BK JWMM. Performed the experiments: MO HH MGN LABJ. Analyzed the data: MO HH MGN LABJ. Contributed reagents/materials/analysis tools: HH PS GJA. Wrote the paper: MO HH PS GJA BK JWMN MGN LABJ.


