Genetic Variants in Toll-Like Receptors Are Not Associated with Rheumatoid Arthritis Susceptibility or Anti-Tumour Necrosis Factor Treatment Outcome

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6 Arthritis Research Campaign Epidemiology Unit, The University of Manchester, Manchester, United Kingdom
7 Department of Rheumatology, Ziekenhuis Gelderse Vallei, Ede, The Netherlands
8 Department of Rheumatology, Medical Centre Leeuwarden, Leeuwarden, The Netherlands
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Abstract

Background: Several studies point to a role of Toll-like receptors (TLRs) in the development of rheumatoid arthritis (RA). We investigated if genetic variants in TLR genes are associated with RA and response to tumour necrosis factor blocking (anti-TNF) medication.

Methodology and Principal Findings: 22 single nucleotide polymorphisms (SNPs) in seven TLR genes were genotyped in a Dutch cohort consisting of 378 RA patients and 294 controls. Significantly associated variants were investigated in replication cohorts from The Netherlands, United Kingdom and Sweden (2877 RA patients and 2025 controls). 182 of the Dutch patients were treated with anti-TNF medication. Using these patients and a replication cohort (269 Swedish patients) we analysed if genetic variants in TLR genes were associated with anti-TNF outcome. In the discovery phase of the study we found a significant association of SNPs rs2072493 in TLR5 and rs3853839 in TLR7 with RA disease susceptibility. Meta-analysis of discovery and replication cohorts did not confirm these findings. SNP rs2072493 in TLR5 was associated with anti-TNF outcome in the Dutch but not in the Swedish population.

Conclusion: We conclude that genetic variants in TLRs do not play a major role in susceptibility for developing RA nor in anti-TNF treatment outcome in a Caucasian population.

Introduction

Rheumatoid arthritis (RA) is a severe chronic inflammatory disorder leading to joint damage. The causes of RA are largely unknown, however, the role of genetic factors is evident, with the MHC region as the major contributor [1]. To date, more than 20 non-MHC regions explaining approximately one third of the genetic contribution to RA have been identified [2]. The role of Toll-like receptors (TLRs) in the development of RA is supported by several studies. Synovial fibroblasts of RA patients constitutively express TLR 1-6, and it has been demonstrated that TLR 2, 3, 4 and 7 are up-regulated in RA synovial tissue compared to that of osteoarthritis patients and healthy controls [3]. Furthermore, the ligands for TLRs have been detected in synovium of RA patients [4],[5]. Since TLRs are potent activators of pro-inflammatory cytokines, including tumour necrosis factor (TNF)
alpha, this also makes them interesting candidates for treatment outcome prediction, especially for TNF-neutralizing therapy [6].

Several groups explored the contribution of genetic variants in TLR genes to the development of RA. A Korean study showed an association of a dinucleotide repeat in TLR2 with RA [7]. One other study focusing on two SNPs in TLR2 resulting in amino acid substitutions (Arg677Trp and Arg753Gln) could not detect an association between these rare variants and RA [8]. Although conflicting results have been reported on the functional variant Asp299Gly (rs4986790) in TLR4 [8]. Although most studies performed to date point to a lack of association of a dinucleotide repeat in TLR2 with RA [8].

Table 1. Results of the case control association analysis in the Dutch discovery set.

<table>
<thead>
<tr>
<th>gene</th>
<th>chromosome</th>
<th>SNP</th>
<th>BP</th>
<th>functional effect</th>
<th>major allele</th>
<th>minor allele</th>
<th>MAF cases</th>
<th>MAF controls</th>
<th>chi-square</th>
<th>p-value</th>
<th>OR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>TLR2</td>
<td>4</td>
<td>rs1898830</td>
<td>154827903</td>
<td>Promoter (−15607)</td>
<td>A</td>
<td>G</td>
<td>0.34</td>
<td>0.33</td>
<td>0.05</td>
<td>0.83</td>
<td>1.03 (0.81–1.30)</td>
</tr>
<tr>
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<td>154845401</td>
<td>Pro631His</td>
<td>T</td>
<td>C</td>
<td>A</td>
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<td>0.05</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
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<td></td>
<td>rs743708</td>
<td>154845767</td>
<td>Arg753Gln</td>
<td>G</td>
<td>A</td>
<td>C</td>
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<td>0.04</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
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<td>rs3775291</td>
<td>187241068</td>
<td>Leu412Phe</td>
<td>C</td>
<td>T</td>
<td>0.30</td>
<td>0.30</td>
<td>0.01</td>
<td>0.92</td>
<td>1.01 (0.79–1.29)</td>
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<td>19515123</td>
<td>Asp299Gly</td>
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<td>G</td>
<td>0.05</td>
<td>0.05</td>
<td>0.003</td>
<td>0.96</td>
<td>0.99 (0.60–1.63)</td>
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<td></td>
<td>rs4986791</td>
<td>19515423</td>
<td>Thr399Ile</td>
<td>C</td>
<td>T</td>
<td>G</td>
<td>0.05</td>
<td>0.05</td>
<td>0.003</td>
<td>0.96</td>
<td>0.99 (0.60–1.63)</td>
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<tr>
<td></td>
<td>rs7873784</td>
<td>19518757</td>
<td>3' UTR</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>0.18</td>
<td>0.16</td>
<td>0.54</td>
<td>0.46</td>
<td>1.12 (0.83–1.51)</td>
</tr>
<tr>
<td>TLR5</td>
<td>1</td>
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<td>221350916</td>
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<td>C</td>
<td>0.47</td>
<td>0.46</td>
<td>0.12</td>
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<td>1.04 (0.83–1.30)</td>
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<tr>
<td></td>
<td>rs2072493</td>
<td>221351222</td>
<td>Asn592Ser</td>
<td>G</td>
<td>A</td>
<td>T</td>
<td>0.10</td>
<td>0.15</td>
<td>0.63</td>
<td>0.35</td>
<td>0.64 (0.46–0.91)</td>
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<td>TLR6</td>
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<td>rs744168</td>
<td>221351823</td>
<td>Arg392STOP</td>
<td>C</td>
<td>T</td>
<td>0.08</td>
<td>0.07</td>
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<td>0.39</td>
<td>0.80 (0.57–1.28)</td>
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<td>221352752</td>
<td>Thr82Ile</td>
<td>G</td>
<td>A</td>
<td>C</td>
<td>0.02</td>
<td>0.01</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
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<tr>
<td>TLR7</td>
<td>X</td>
<td>rs2302267</td>
<td>12795499</td>
<td>Exon/intron boundary</td>
<td>G</td>
<td>T</td>
<td>0.05</td>
<td>0.04</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>TLR8</td>
<td>X</td>
<td>rs719008</td>
<td>12813580</td>
<td>Gln11Leu</td>
<td>A</td>
<td>T</td>
<td>0.21</td>
<td>0.24</td>
<td>0.94</td>
<td>0.33</td>
<td>0.86 (0.63–1.17)</td>
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<tr>
<td></td>
<td>rs5743781</td>
<td>12814891</td>
<td>Ala448Val</td>
<td>C</td>
<td>G</td>
<td>T</td>
<td>0.0003</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
<td>n.a.</td>
</tr>
<tr>
<td>r38553839</td>
<td>12817579</td>
<td>3' UTR</td>
<td>C</td>
<td>G</td>
<td>0.13</td>
<td>0.21</td>
<td>8.95</td>
<td>0.003</td>
<td>0.59</td>
<td>0.42 (0.42–0.84)</td>
<td></td>
</tr>
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<td>3</td>
<td>rs7543836</td>
<td>52235822</td>
<td>Promoter (−1237)</td>
<td>A</td>
<td>G</td>
<td>0.16</td>
<td>0.18</td>
<td>1.21</td>
<td>0.27</td>
<td>0.85 (0.63–1.14)</td>
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<tr>
<td></td>
<td>rs187084</td>
<td>52236071</td>
<td>Promoter (−1468)</td>
<td>A</td>
<td>G</td>
<td>0.41</td>
<td>0.43</td>
<td>0.49</td>
<td>0.48</td>
<td>0.92 (0.73–1.16)</td>
<td></td>
</tr>
</tbody>
</table>

MAF: minor allele frequency.
OR: odds ratio.
CI: confidence interval.
n.a.: not analysed.
doi:10.1371/journal.pone.0014326.t001

alpha, this also makes them interesting candidates for treatment outcome prediction, especially for TNF-neutralizing therapy [6].

Several groups explored the contribution of genetic variants in TLR genes to the development of RA. A Korean study showed an association of a dinucleotide repeat in TLR2 with RA [7]. One other study focusing on two SNPs in TLR2 resulting in amino acid substitutions (Arg677Trp and Arg753Gln) could not detect an association between these rare variants and RA [8]. Although conflicting results have been reported on the functional variant Asp299Gly (rs4986790) in TLR4 [8]. Although most studies showed no association of this variant with RA susceptibility [8]–[13]. A French group investigated 10 SNPs in TLR1, 2, 4, 6 and 9 in 100 families but did not find evidence for an association of variants in these genes with RA, autoantibody production or erosions [13]. A Turkish study, investigating variants in TLR3, 9 and 10 in 100 patients showed an association between a variant in TLR9 and RA susceptibility [14]. So far, only one small pharmacogenetic study on TLRs has been published [15]. The authors did not demonstrate an association between the Asp299Gly variant in TLR4 and the response to disease modifying anti-rheumatic drugs (DMARDs).

Although most studies performed to date point to a lack of association between genetic variants in genes coding for TLRs and RA disease susceptibility or treatment response, these have either been very small or did not comprehensively test potentially functional SNPs in the genes. Therefore we performed an association study of TLRs testing involvement in RA pathogenesis and anti-TNF treatment response including seven TLR genes, three of which (TLR5, 7 and 8) have not been investigated in connection with RA before. The twenty-two SNPs investigated were selected for potential effects on gene function or regulation.

Results

Case control analysis

In the discovery phase of our study we included 378 RA patients and 294 controls and genotyped 22 SNPs in seven TLR genes. Genotyping failed for 51 samples (22 cases and 26 controls). Six SNPs were excluded from the analysis, based on a low MAF (Table 1).

Two of the investigated polymorphisms, rs2072493 in TLR5 and rs3853839 in TLR7, showed nominally significant allelic association with RA (p ≤ 0.01; Table 1). The minor allele of both SNPs conferred a protective effect for the development of RA. A stratified analysis for either the shared epitope or rheumatoid arthritis. We did not identify SNPs that were specifically associated with one of the RA subgroups. The odds ratios for SNPs rs2072493 and rs3853839 were comparable to the odds ratios observed in the complete patient population (data not shown). The frequencies of rs2072493 and rs3853839 were investigated in three additional cohorts from The Netherlands, Sweden and the UK, consisting of 2877 patients and 2025 controls. This analysis
showed no association of the SNPs in TLR5 and TLR7 with RA aetiology either in the separate cohorts or in a meta-analysis (Figure 1).

Anti-TNF treatment response

A subsample of the Dutch patients were treated with anti-TNF medication and had EULAR response information available at 3 months (n = 182, Table 2). Results of the association analysis are shown in Table 3. TLR5 rs2072493 showed a significant association with anti-TNF treatment outcome (p = 0.003). In total, 182 patients were included in the analysis. The EULAR response criteria were used to assess association of a genotype with anti-TNF response. The test statistic and p-value were from Fisher’s Exact Test for most SNPs, those indicated with * were based on Pearson Chi-square and p-value. A medication specific analysis showed that none of the investigated SNPs was associated with a specific anti-TNF (Table S1). Validation of association found with TLR5 in 269 RA patients treated with anti-TNF from Sweden did not confirm our initial findings (Fisher’s exact test p = 0.63).

The differences observed between the Dutch and Swedish cohort were not related to the anti-TNF used (Fisher’s exact test Swedish cohort for infliximab (p = 0.75) and etanercept (p = 0.49)).

Discussion

We found no evidence for association between 22 SNPs in TLR genes and RA disease susceptibility or anti-TNF treatment response, in keeping with the majority of the other candidate gene studies investigating TLRs in RA patients [7]–[13]. Furthermore, genome-wide association studies (GWAS) do not report associations between TLR genes and RA disease susceptibility (www.genome.gov/gwastudies). Accessed December 31th 2009), suggesting that genetic variants in these genes are not a major contributor to disease development [16]. However, some TLR genes were poorly covered in the initial GWAS (e.g. no SNPs in TLR5 and 9 were included in these studies) [17],[18]. The TLR genes remain strong candidates for susceptibility genes in RA given their crucial role in mediating inflammatory signalling. One
### Table 2. Patient characteristics of RA patients treated with anti-TNF medication.

<table>
<thead>
<tr>
<th>Cohort</th>
<th>The Netherlands</th>
<th>Sweden</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of patients (N (%))</td>
<td>182 (100)</td>
<td>269 (100)</td>
</tr>
<tr>
<td>Gender (female)</td>
<td>122 (67.0)</td>
<td>211 (78.4)</td>
</tr>
<tr>
<td>RF factor positive</td>
<td>141 (77.5)</td>
<td>169 (88.9)</td>
</tr>
<tr>
<td>MTX co-medication</td>
<td>105 (61.0)##</td>
<td>185 (68.8)</td>
</tr>
<tr>
<td>Anti-TNF medication</td>
<td>61 (33.5)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Adalimumab</td>
<td>118 (64.8)</td>
<td>156 (58.0)</td>
</tr>
<tr>
<td>Etanercept</td>
<td>1 (1.6)</td>
<td>113 (42.0)</td>
</tr>
<tr>
<td>Good response*</td>
<td>45 (24.7)</td>
<td>76 (28.3)</td>
</tr>
<tr>
<td>Moderate response*</td>
<td>92 (50.5)</td>
<td>137 (50.9)</td>
</tr>
<tr>
<td>No response*</td>
<td>45 (24.7)</td>
<td>56 (20.8)</td>
</tr>
<tr>
<td>DAS28 baseline (mean ± SD)</td>
<td>5.72±1.16</td>
<td>5.84±1.13</td>
</tr>
<tr>
<td>DAS28 3 months after treatment initiation (mean ± SD)</td>
<td>4.12±1.36</td>
<td>3.89±1.30</td>
</tr>
</tbody>
</table>

*Response based on EULAR response criteria. Data available for #180, ##190 and ###172 patients.

doi:10.1371/journal.pone.0014326.t002

Possibility is that these genes might harbour rare variants with large effect sizes, which would not be well tagged with the common SNPs selected for investigation in the current study. With the introduction of next generation sequencing technologies it should become possible to analyse the genes in a thorough manner but much larger sample sizes will be required to ensure power to detect association with rare variants, even if effect sizes are larger.

We could not identify an association of the investigated SNPs with anti-TNF response. We were unable to replicate the initial finding in the Dutch cohort. Differences in ethnicity, gender, RF and co-medication might influence the observed discrepancies. Besides the study has low power to detect an association due to the small patient population. Despite this the function of TLRs makes them likely candidate genes, since they directly modulate TNF expression [6]. In addition, De Rijke and colleagues showed that anti-TNF treatment in spondylarthropathy results in a reduced expression of TLRs [7, 8 and 9]. The TLR pathway should therefore be investigated in more detail before drawing definite conclusions about the contribution of these genes to anti-TNF treatment outcome.

In conclusion, our study does not support an association between genetic variants in TLR genes and RA disease susceptibility or anti-TNF treatment response.

### Materials and Methods

#### Ethics Statement

The study was approved by the ethics committees of the participating institutions. For the Dutch samples the “Commissie Mensgebonden Onderzoek (CMO) Region Arnhem Nijmegen” of the Radboud University Nijmegen Medical Centre approved the study (CMO number 2004/014). All Dutch patients and controls gave written informed consent prior to participation in the study. Verbal consent was received from all patients participating in the EIRA study and it was registered at clinical journals according to Swedish law. Written consent was received from healthy control individuals. The ethics committee of Karolinska Institutet approved the study. All UK patients and controls were recruited with ethical approval and provided written informed consent (North-West Multi-Centre Research Ethics Committee (MREC 99/8/04) and the University of Manchester Committee on the Ethics of Research on Human Beings).

#### Patients and controls

All patients in this study fulfilled the American College of Rheumatology (ACR) criteria. In the discovery phase 378 Dutch RA patient participating in the Nijmegen Inception cohort and 294 anonymous blood donors were included [20]. Cohorts from The Netherlands, Sweden and England were included in the replication phase. The Dutch replication cohort consisted of 313 RA patients participating in the Dutch Rheumatoid Arthritis Monitoring (DREAM) registry [21]. The Dutch controls (n = 527) were part of the Nijmegen Biomedical Study [22].

The Swedish patients (n = 2158) and controls (sex, age and residence area matched, n = 1068) were part of the Epidemiological Investigation of Rheumatoid arthritis (EIRA) study. The UK patients (n = 406) were recruited as part of the Arthritis Research Campaign National Repository of patients and families with one patient from each family selected for genotyping. UK controls without a history of inflammatory arthritis (n = 430) were recruited from General Practitioner records or from blood donors.

#### Anti-TNF treatment outcome

Patients participating in the DREAM registry were included in the study of anti-TNF treatment response. The registry includes patients with RA who start with anti-TNF treatment, according to the Dutch recommendations (Disease Activity Score (DAS28) >3.2 and previous failure on at least two disease-modifying antirheumatic drugs (DMARDs), one of which has to be methotrexate (MTX)). The DAS28 at treatment start and after 3 months of treatment were used to calculate the clinical response according to the EULAR criteria [23]. Association analysis was performed for the total patient group and a subgroup analysis for the patients treated with infliximab and adalimumab. Nominally significant findings (p<0.01) were validated in 269 anti-TNF treated RA patients from Sweden [23].

#### SNP genotyping

Twenty-two SNPs located in TLR2, 3, 4, 5, 7, 8 and 9 were genotyped in all Dutch patients and Dutch anonymous blood donors (Table 1). SNP selection criteria are described previously [24]. SNPs were genotyped using two multiplexed bead-based assays on a Luminex 100IS flow cytometer (Luminex Corporation, Austin, TX, USA). The tests were based on procedures previously described, with some modifications [24]–[26].

Replication studies were performed using TaqMan® SNP genotyping assays for SNPs rs2072493 in TLR5 (assay ID C__22273027_10) and rs3853839 in TLR7 (assay ID C__2259573_10).

#### Statistical analysis

For quality control reasons we excluded SNPs that had a minor allele frequency (MAF) <0.05. Hardy-Weinberg Equilibrium (HWE) was tested using the controls (only females were used for X-chromosomal SNPs), no deviations were observed. The whole-genome association analysis toolset (PLINK) was used for association analysis at the allele level [27]. A stratified analysis was performed for RF positive and negative as well as shared epitope positive and negative patients. SNPs with a nominal p-value ≤0.01 in the discovery sample were included in the
Table 3. Results of the association analysis for anti-TNF treatment outcome after three months of treatment initiation.

<table>
<thead>
<tr>
<th>Gene</th>
<th>chromosome</th>
<th>SNP</th>
<th>Gender</th>
<th>Test statistic</th>
<th>p-value</th>
</tr>
</thead>
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<td>rs1898830</td>
<td>males</td>
<td>11.188*</td>
<td>0.024*</td>
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<td></td>
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<td>rs5743704</td>
<td></td>
<td>n.a.</td>
<td>n.a.</td>
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<td></td>
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<td></td>
<td>n.a.</td>
<td>n.a.</td>
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<td>5.32</td>
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<td></td>
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<td>rs179008</td>
<td></td>
<td>2.97*</td>
<td>0.24*</td>
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<td>rs3853839</td>
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<td>0.089</td>
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<td></td>
<td></td>
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<td>females</td>
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<td>females</td>
<td>1.76</td>
<td>0.85</td>
</tr>
<tr>
<td>TLR9</td>
<td>3</td>
<td>rs5743836</td>
<td>males</td>
<td>0.63</td>
<td>0.80</td>
</tr>
<tr>
<td></td>
<td></td>
<td>rs187084</td>
<td>females</td>
<td>3.20</td>
<td>0.50</td>
</tr>
</tbody>
</table>

In total, 182 patients were included in the analysis. The EULAR response criteria were used to assess association of a genotype with anti-TNF response. The test statistic and p-value were from Fisher's Exact Test for most SNPs, those indicated with * were based on Pearson Chi-square and p-value. SNPs located on the X-chromosome were analyzed for males and females separately.

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Author Contributions

Conceived and designed the experiments: MJC CE PB EJT BF KB TR. Performed the experiments: CE MMS EJT. Analyzed the data: MJC CE EJT LP AK AB WK. Contributed reagents/materials/analysis tools: MJC HS LP AK LK AB WK MJR TLJ DWS PvR BF KB. Wrote the paper: MJC CE TR.

Supporting Information

Table S1 Results of the association analysis for anti-TNF treatment outcome after three months of treatment initiation.
References


