Genomewide Association Study of Leprosy

TO THE EDITOR: Zhang and colleagues (Dec. 31 issue) report that genes in the nucleotide-binding oligomerization domain containing 2 (NOD2)–mediated signaling pathway are associated with susceptibility to infection with Mycobacterium leprae in China. India has the world’s greatest leprosy disease burden. We therefore genotyped the single-nucleotide polymorphisms (SNPs) that were implicated by Zhang and colleagues in two Indian case–control cohorts (492 patients in New Delhi and 382 in Kolkata).1,2 We also genotyped 273 cases and 221 controls from Mali, West Africa.3 We observed associations between disease and SNPs at CCDC122 (the gene encoding chromosome 13q14.11 locus containing 122) (rs9533634, P = 1.1×10−5) and C13orf31 (the gene encoding coiled-coil domain containing 31) (rs3764147, P = 6.1×10−8) (Table 1); both genes were of unknown function. We did not, however, observe associations between disease and the other four non–major histocompatibility complex (MHC) genes related to the NOD2 pathway (NOD2, RIPK2 [the gene encoding receptor-interacting serine–threonine kinase 2], TNFSF15 [the gene encoding tumor necrosis factor ligand] superfAMILY member 15), and LRRK2 (the gene encoding leucine-rich repeat kinase 2) (Table 1 in the Supplementary Appendix, available with the full text of this letter at NEJM.org), despite reasonable power to detect the effect sizes observed by Zhang et al.4 An analysis of 27 additional SNPs at NOD2 in the New Delhi cohort provided support for the absence of a consistent association at this locus (Table 2 in the Supplementary Appendix). These results indicate heterogeneity among populations and suggest that future functional studies should focus on populations in which the relevant genetic association may occur. Nevertheless, a robust association of the Crohn’s disease chromosome 13q14.11 locus containing C13orf31 and CCDC122 with leprosy in India, China, and Mali provides support for a molecular link between mycobacterial infection and Crohn’s disease.

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Table 1. Associations with Leprosy for Replicated Single-Nucleotide Polymorphisms on Chromosome 13q14.11, According to Case–Control Population.

<table>
<thead>
<tr>
<th>SNP</th>
<th>Gene</th>
<th>Original Study <em>odds ratio (95% CI)</em></th>
<th>Present Study <em>odds ratio (95% CI)</em></th>
<th>New Delhi</th>
<th>Kolkata</th>
<th>Mali</th>
<th>Combined Populations</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs3764147</td>
<td>C13orf31</td>
<td>1.68 (1.57–1.80)</td>
<td>1.59 (1.34–1.89)</td>
<td>2.7×10⁻³</td>
<td>6.4×10⁻²</td>
<td>1.1×10⁻⁵</td>
<td>6.1×10⁻⁸</td>
</tr>
<tr>
<td>rs9533634</td>
<td>CCDC122</td>
<td>0.76 (0.70–0.82)</td>
<td>0.70 (0.59–0.82)</td>
<td>1.5×10⁻³</td>
<td>3.6×10⁻¹</td>
<td>1.1×10⁻⁵</td>
<td>1.1×10⁻⁵</td>
</tr>
</tbody>
</table>

* C13orf31 denotes the gene encoding chromosome 13 open reading frame 31, CCDC122 the gene encoding coiled-coil domain containing 122, CI confidence interval, and SNP single-nucleotide polymorphism.
TO THE EDITOR: A similar proinflammatory immune response has been observed in paucibacillary leprosy and in the contained form of Crohn’s disease.1 In contrast, such an immune response has not been detected in the more aggressive, multibacillary form of leprosy and in the more aggressive form of Crohn’s disease.1 These findings are now mirrored in an intriguing commonality between human genetic defects in multibacillary leprosy and Crohn’s disease reported on by Zhang et al. The accompanying editorial2 posits that a subgroup “of Crohn’s disease may have a mycobacterial cause.” A candidate culpable organism3 causes Johne’s disease in ruminants, an affliction evocative of Crohn’s disease. Interspecies comparisons4 may be illuminating. Cattle with the NOD2 defect are three times as likely to contract Johne’s disease as cattle with wild-type NOD2 in the same herd.5

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Dr. Greenstein reports submitting patent applications based on the inhibition of mycobacteria including the M. avium subspecies paratuberculosis by medications used to treat Crohn’s disease. No other potential conflict of interest relevant to this letter was reported.


THE AUTHORS REPLY: Using three case–control samples from India and West Africa, Wong et al. undertook a follow-up study of our recently identified leprosy susceptibility SNPs and confirmed the strong associations of C13orf31 and CDCC122, highlighting the commonality of leprosy susceptibility loci among diverse populations. Although they did not detect the associations in the other...
non-MHC susceptibility loci, the use of combined small and mixed patient cohorts, with limited analysis of the SNPs most strongly implicated in our analysis, may have resulted in some missed associations, especially if the risk alleles were tagged by different SNPs in populations of differing ancestries, the genetic effects of risk alleles were more moderate in populations other than Chinese populations, or both.

Netea and colleagues express surprise that our study did not highlight TLR genes and further imply that our findings may be relevant only for East Asian populations. We searched our genomewide association study data for any evidence of TLR associations and found none. We agree with these correspondents that genetic variation due to differences in ancestral origin may explain this result, but we think a more likely explanation is that the NOD2 pathway is more influential than the TLR pathway in determining disease outcome. The implication of variants of C13orf31 and CCDC122 in Indian and West African populations shows that genetic risk factors for leprosy are shared by different populations.

The observation by Greenstein and Brown provides further support for the pathogenicity common to leprosy and Crohn’s disease. Although we observed an overlap of the most strongly implicated SNPs in susceptibility to both leprosy and Crohn’s disease, a joint analysis of these diseases (through a genomewide association study) should be pursued for a comprehensive investigation of common genetic risk factors. Moreover, confirmation of the association between C13orf31 and CCDC122 and susceptibility to Crohn’s disease and leprosy in diverse populations calls for an investigation of the biologic function of these genes, which will probably illuminate a pathogenic mechanism of both diseases.

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Since publication of their article, the authors report no further potential conflict of interest.


Barrett’s Esophagus

TO THE EDITOR: Sharma (Dec. 24 issue)1 says that ablation is not recommended for nondysplastic Barrett’s esophagus because of an unacceptably high number of patients who would need to be treated (the number needed to treat) to prevent one case of adenocarcinoma. The estimate of 250 as the number needed to treat is artificially high, however. The number needed to treat is 1 divided by the absolute risk reduction. In the meta-analysis by Wani et al., the incidence of adenocarcinoma in patients with untreated nondysplastic Barrett’s esophagus was 0.6% per patient-year of follow-up, as compared with 0.16% in patients treated with ablation.2 The absolute risk reduction is therefore 0.6%–0.16%, or 0.44% per patient-year of follow-up. The studies in this meta-analysis varied in the length of follow-up, but all follow-ups were multiple years in length. Assuming a follow-up of just 5 years, the number needed to treat would be 45, not 250. Other studies of ablation for nondysplastic Barrett’s esophagus also show a much lower number needed to treat — approximately 23 rather than 250.3

Sharma’s estimate of the number needed to treat limits ablation to prevention of adenocarcinoma only; however, ablation also prevents the development of high-grade dysplasia and subsequent progression to cancer.4 For the inclusion of the aggregate end points of high-grade dysplasia and cancer, the number needed to treat would be lower still.

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Dr. Ganz reports being a coinventor of the Halo system of radiofrequency ablation and having equity in and serving on the board of directors of the manufacturing company, BARRX Medical. No other potential conflict of interest relevant to this letter was reported.

2. Wani S, Puli SR, Shaheen NJ, et al. Esophageal adenocarci-