Manipulating Heat Shock Factor-1 in *Xenopus* Tadpoles: Neuronal Tissues Are Refractory to Exogenous Expression

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**Abstract**

**Background:** The aging related decline of heat shock factor-1 (HSF1) signaling may be causally related to protein aggregation diseases. To model such disease, we tried to cripple HSF1 signaling in the *Xenopus* tadpole.

**Results:** Over-expression of heat shock factor binding protein-1 did not inhibit the heat shock response in *Xenopus*. RNAi against HSF1 mRNA inhibited the heat shock response by 70% in *Xenopus* A6 cells, but failed in transgenic tadpoles. Expression of XHSF380, a dominant-negative HSF1 mutant, was embryonic lethal, which could be circumvented by delaying expression via a tetracycline inducible promoter. HSF1 signaling is thus essential for embryonic *Xenopus* development. Surprisingly, transgenic expression of the XHSF380 or of full length HSF1, whether driven by a ubiquitous or a neural specific promoter, was not detectable in the larval brain.

**Conclusions:** Our finding that the majority of neurons, which have little endogenous HSF1, refused to accept transgene-driven expression of HSF1 or its mutant suggests that HSF1 levels are strictly controlled in neuronal tissue.

**Introduction**

In healthy cells, accumulation of abnormally folded proteins in the cytoplasm results in the activation of a stress response system, the heat shock response (HSR). The HSR is essential for maintaining proteostasis. Mouse knockout models have shown that heat shock transcription factor-1 (HSF1) is the key regulator of the HSR [1–3]. Under normal physiological conditions, HSF1 is thought to exist as part of an inactive hetero-complex that also includes Hsp90, p23 and immunophilin. Exposure of cells to toxic protein aggregates ([7,8]; see also [9,10]). A number of *in vivo* model systems have been described in which the role of a failing HSR in the etiology of neurological diseases can be studied. Mice carrying null mutant HSF1 genes or expressing a dominant-negative HSF1 are attractive model systems, because the mouse brain closely resembles the human brain; however, small offspring and high costs make rodents less attractive for large scale studies. Invertebrates, such as *C. elegans* and *Drosophila*, are particularly suitable for high throughput experiments, because they have a large offspring and can be easily manipulated. Unfortunately, their nervous system differs considerably from the human brain. Simple vertebrates, such as *Xenopus* and *Danio*, are attractive *in vivo* model systems to study many aspects of human neurological diseases. In addition to the large offspring and low maintenance costs, the basic anatomy of the fish and amphibian brain is highly similar to that of the mammalian brain. Furthermore, tadpoles and zebrafish are translucent, allowing live image analysis of fluorescently labeled proteins. Microinjection of antisense oligonucleotides has already been used to transiently inhibit the expression of HSF1 in zebrafish. This resulted in increased heat shock-induced apoptosis [12], reduced basal expression levels of Hsp70 and abnormal eye development [13].

To be able to study the long term effects of decreased HSF1 activity in neurons and perform high throughput experiments, it
would be desirable to develop a model in which the expression of HSF1 is stably inhibited. In this study, we examined if we could mimic the aging-associated decline of the HSR in a stable manner by crippling endogenous HSF1 in Xenopus tadpoles. Since a technique for targeted mutagenesis of endogenous genes in *Xenopus laevis* is not yet available, we tried to manipulate *Xenopus* HSF1 via three alternative strategies: over expression of heat shock factor binding protein-1 (HSBP1), reported to be a natural inhibitor of HSF1 [14], over expression of a dominant-negative mutant of HSF1, and stable transgene-driven RNA interference (RNAi) directed against HSF1 mRNA. Our surprising finding is that XHSBP1 tadpole brain is largely refractory to exogenous expression of HSF1.

**Results**

*Xenopus* heat shock factor binding protein-1 is not an efficient inhibitor of the heat shock response

HSBP1 is a ~9-kDa polypeptide that was shown to act as a negative regulator of the HSR in cultured mammalian cells and in *C. elegans* in vivo [14]. Since HSBP1 is highly conserved throughout the animal kingdom [15], we examined whether transgene-driven over expression of XHSBP1 could be used to stably inhibit the HSR in *Xenopus* tadpoles. ClustalW alignment of the open reading frames of multiple *Xenopus* laevis HSBP1 ESTs revealed slight variation in the C-terminal part of the protein. However, the central coiled coil region thought to be important for the inhibitory interaction between HSBP1 and HSF1 [15] is identical in all ESTs and nearly identical to that of the human protein (Fig. 1A). The code for XHSBP1 was PCR amplified from adult *Xenopus* brain cDNA and, to allow for quick identification of transgenic tadpoles, fused to the 3' end of the code for GFP. Tadpoles with ubiquitous expression of the GFP-HSBP1 protein developed normally (Fig. 1B,C). To test whether over expression of GFP-XHSBP1 inhibited the HSR in tadpoles, the endogenous Hsp90 level was monitored by western blot analysis. Basic Hsp90 levels are low in tadpoles (Fig. 1A); however, western blot analysis indicated that ribosomal skipping from the T2A-based bicistronic expression cassette was almost ~100% efficient, whereas the F2A peptide resulted in only ~50% skipping efficiency (Fig. 3B). A control experiment, in which GFP and myc-tagged versions of Hsp27 or dominant-negative HSF1 are expressed from a bicistronic transcription unit further demonstrated that the second part of the T2A-based plasmids is also properly expressed (Fig. 3C). The T2A-based system was then used to test the effect of native HSBP1 on co-transfected Hsp70-luciferase reporter in A6 cells. The activity of the Hsp70 promoter was inhibited, but by only ~30% in the presence of native XHSBP1 (Fig. 3D). In summary, our results indicate that over expression of XHSBP1 is not a successful strategy to inhibit the HSR of *Xenopus*.

RNAi against HSF1 inhibits HSR in A6 cells but is not effective in tadpoles

As an alternative strategy, we tried to reduce the expression of HSF1 by means of stable, transgene-driven RNAi. Earlier efforts to stably inhibit gene expression via RNAi in *Xenopus* tadpoles were only partially successful. Whereas expression of exogenous GFP could be inhibited by co-expressing long GFP dsRNA from RNA polymerase II promoters [17] or GFP shRNAs from an RNA polymerase III promoter [18], neither polymerase II nor polymerase III promoter-based inverted repeat constructs resulted in stable inhibition of endogenous target genes [17]. We already showed that the human H1 RNA promoter drives strong and ubiquitous GFP expression in transgenic *Xenopus* tadpoles [17]. Thus, we selected a 19-mer sequence optimal for HSF1 knockdown and placed the corresponding inverted repeat under the control of the human H1 RNA promoter. To allow for future identification of transgenic tadpoles, the H1-HSF1-sh1 cassette was linked in tandem with a muscle-specific GFP reporter.

To get a first impression of the inhibitory potential of the human H1-HSF1-sh1 plasmid, we tested its effect on the Hsp70-luciferase reporter in A6 cells. The humH1-HSF1-sh1 plasmid inhibited the heat shock induced activity of the Hsp70 promoter by only 27% (Fig. 4A). We reasoned that, in *Xenopus* tadpoles, the human H1 promoter may not be recognized by the proper RNA polymerase (polymerase II rather than III), which would result in untranslated RNAs with long single-stranded extensions that do not induce RNai. This would also explain why GFP expression from this promoter is so efficient [17], despite the fact that RNAs transcribed from polymerase III promoters are not usually capped or polyadenylated. Thus, we isolated an H1 (RNase P) promoter from *Xenopus tropicalis* and expressed the HSF1-sh1 from this promoter. XhH1 promoter-driven HSF1-sh1 reduced the heat shock induced activity of a co-transfected Hsp70-luciferase reporter by almost 50% (Fig. 4A). Similar analyses of HSF1-sh2 and HSF1-sh3, which are directed against other parts of the HSF1 mRNA, resulted in, respectively, 71% and 55% inhibition of the HSR (Fig. 4B).

Since HSF1-sh2 had the strongest inhibitory activity in our in vitro system, this inverted repeat was used to generate transgenic tadpoles. Tadpoles carrying the HSF1-sh2 transgene were identified by muscle-specific expression of the GFP reporter. GFP-positive tadpoles developed normally and were equally resistant to sub lethal heat shock as non-transgenic tadpoles (data not shown). Western blot analysis indicated that heat shock mediated induction of endogenous Hsp90 expression was not inhibited in GFP positive tadpoles (Fig. 4C). Thus, despite its inhibitory activity on the HSR in A6 cells, HSF1-sh2 was not effective as an inhibitor of the HSR in tadpoles.
Figure 1. Ubiquitous transgene-driven expression of XHSBP1 in Xenopus tadpoles. (A) ClustalW alignment of human HSBP1 (top) and HSBP1 sequences deduced from five different Xenopus laevis ESTs. The central coiled coil region is highly conserved between man and Xenopus. (B) Fluorescence microscope analysis of tadpoles carrying the indicated transgenes. Tadpoles with ubiquitous, high level expression of GFP-HSBP1 look normal. (C) Western blot analysis of lysates from whole tadpoles carrying the indicated transgenes. Proteins were separated by SDS-PAGE and analyzed with an anti-GFP antibody.

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Dominant-negative HSF1 mutant is embryonic lethal and not detectable in transgenic brain

Dominant-negative mutant versions of HSF1 have been successfully used by others to inhibit the HSR in mammalian systems (reviewed by [19]). Therefore, we tried to inhibit the HSR in tadpoles via transgene-mediated expression of dominant-negative HSF1. The first 380 codons of the XHSF1 open reading frame (XHSF380), thus lacking the code for the C-terminal transactivation domain, were PCR amplified from adult Xenopus brain cDNA. The XHSF380 cDNA was cloned upstream or downstream of the GFP code and the effect of expression of the GFP-XHSF380 fusion protein on the HSR was first tested via reporter gene analysis of the Hsp70 promoter in transiently transfected A6 cells. Heat shock mediated induction of Hsp70-Luc was
Manipulating HSF1

Figure 2. The effect of exogenous expression of GFP-XHSBP1 on the HSR. (A) Heat shock induced expression of Hsp90. *Xenopus* A6 kidney epithelial cells were continuously cultured at room temperature (C) or exposed to a 1 h heat shock (33°C) and then cultured at room temperature for the indicated times. Cell lysates were subjected to SDS-PAGE and western blot analysis using anti-Hsp90 and anti-tubulin antibodies. (B) Western blot analysis of Hsp90 levels in tadpoles carrying the indicated transgenes. Lysates were made from whole tadpoles before heat shock (C) or after a 1 h heat shock at 33°C followed by 6 h recovery at room temperature (HS). The lysates were subjected to SDS-PAGE and western blot analysis using anti-GFP, anti-Hsp90 and anti-tubulin antibodies. (C) Reporter gene analysis of the effect of GFP-XHSBP1 on the HSR. A6 cells were transfected with mixtures of an Hsp70-luciferase reporter, a CMV-β-galactosidase reporter and the indicated plasmids. At 48 h after transfection, cells were exposed to a 1 h heat shock at 33°C. Control cells were left at room temperature. Cell lysates were made at 6 h after heat shock and used for reporter gene assays. Hsp70 promoter activities were determined by dividing luciferase values by the corresponding β-galactosidase values to correct for varying transfection efficiencies. The HSR is indicated as fold induction relative to the activity of the Hsp70 promoter in control cells, which was set at 1 (C). The results are the average of three independent transfections (standard deviations are indicated by error bars).

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completely inhibited by co-expression of GFP-XHSF380 or XHSF380-GFP, indicating that over expression of XHSF380 is a powerful way to inhibit the HSR (Fig. 5A).

We next used CMV-GFP-XHSF380 and CMV-XHSF380-GFP cassettes to generate transgenic tadpoles. GFP-positive larvae showed clear nuclear fluorescence (Fig. 5B), which is in accordance with an earlier report that *Xenopus* HSF1 is a nuclear protein even in the absence of heat stress [20]. Surprisingly, CMV promoter-driven expression of GFP-tagged XHSF380 was mainly restricted to skeletal muscle fibers and undetectable in brain and spinal cord (Fig. 5B), despite the fact that the CMV promoter is also highly active in neuronal tissue (data not shown). GFP-positive larvae (n = 40, Table 1) developed poorly, remained small, had a curved tail and died before reaching the feeding tadpole stage (Fig. 5B). Embryonic lethality could be due to the high expression level from the strong CMV promoter; however, replacing the CMV promoter with the weaker EF1α promoter, did not prevent the lethal phenotype. Again, GFP fluorescence was not detectable in neuronal tissues (data not shown), although expression of GFP-XHSBP1 from the EF1α promoter resulted in strong fluorescence in brain and spinal cord (Fig. 5B).

In a final effort to target expression of XHSF380 specifically to the brain, the CMV promoter was replaced with the neuron-specific β-tubulin (Ntub) promoter. Injections with Ntub-GFP-XHSF380 or Ntub-XHSF380-GFP cassettes resulted in ~300 normal larvae without detectable GFP fluorescence, whereas ~10% of the larvae derived from parallel injections with a Ntub-GFP cassette showed strong GFP fluorescence in brain and spinal cord (Fig. 5B, Table 1). We concluded that GFP-tagged XHSF380 cannot be stably expressed in *Xenopus* brain. The observation that both N-terminally and C-terminally GFP-tagged XHSF380 were undetectable in neuronal tissue suggested that this was due to tissue-specific degradation of the fusion protein or the corresponding mRNA, rather than inefficient translation of the transgene-derived mRNA.

To examine whether putative neuron-specific instability of XHSF380 was due to the absence of the C-terminal transcription activation domain, we also generated tadpoles expressing full length GFP-XHSF1 from the CMV promoter. Surprisingly, the expression pattern of GFP-XHSF1 was very similar to that of GFP-XHSF380. Fluorescence was pronounced in nuclei of muscle tissue, but undetectable in most parts of the brain, except for the epiphysis, the olfactory lobes and the retinal ganglion cells (Fig. 5C). In conclusion, the low neuronal expression level of transgene-derived XHSF380 is an intrinsic property of the *Xenopus* HSF1 protein and not a direct result of deletion of the transcription activation domain.

Since expressing XHSF380 was the most effective strategy to inhibit the HSR in *Xenopus* A6 cells, we tried to circumvent the
Figure 3. The effect of exogenous expression of XHSBP1 on the HSR. (A) Fluorescence microscope analysis of reporter gene expression from bicistronic plasmids. A6 cells were transfected with the indicated dual reporter gene plasmids based on viral 2A peptides. At 24 h after transfection, GFP and DsRed expression was determined by fluorescence microscope analysis. (B) Western blot analysis of gene expression from bicistronic plasmids. A6 cells were transfected with the indicated 2A peptide-based plasmids. At 24 h after transfection, cell lysates were made and subjected to SDS-PAGE and western blot analysis using anti-GFP antibodies. (C) Western blot analysis of gene expression from bicistronic plasmids. A6 cells were transfected with the indicated 2A peptide-based plasmids. At 24 h after transfection, cell lysates were made and subjected to SDS-PAGE and western blot analysis using anti-GFP and anti-myc tag antibodies. (D) Reporter gene analysis of the effect of native XHSBP1 on the HSR. A6 cells were transfected with mixtures of an Hsp70-luciferase reporter, a CMV-β-galactosidase reporter and the indicated plasmids. Relative luciferase activities and fold induction were determined as described in the legend to fig. 2C. The results are the average of three independent transfections (standard deviations are indicated by error bars).

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Discussion

To mimic the aging-related decline of the HSR in a Xenopus tadpole model system, we tried to inhibit the activity of HSF1 using three different transgene-mediated strategies. XHSBP1 did not inhibit the HSR significantly, neither in A6 cells nor in transgenic tadpoles. Both untagged and GFP-tagged XHSBP1 were poor inhibitors, indicating that the absence of biological activity did not result from poor import into the nucleus due to the GFP tag. Perhaps, the expression level of XHSBP1 was insufficient to exert its inhibitory effect on HSF1, although the GFP fluorescence signal and western blot analysis suggested that the transgene was expressed at a very high level throughout the tadpole. The inhibitory effect of HSBP1 was described in a single study, wherein hemagglutinin-tagged human HSBP1 was shown to inhibit the HSR in COS7 cells and the C. elegans orthologue was shown to inhibit the HSR in nematodes in vivo [14]. The discrepancy between the published data and our results could be caused by species-specific differences in the biological activity of HSBP1, although its strong sequence conservation, especially in the hydrophobic core, suggests a conserved function. As our efforts to inhibit the HSR in human HeLa cells using human HSBP1 were also unsuccessful (unpublished results), the proposed biological function of HSBP1 may have to be reconsidered.

Stable transgene-driven RNAi directed against XHSF1 mRNA was not an effective means to inhibit the HSR in tadpoles. Earlier, it was shown that expression of exogenous GFP can be effectively inhibited by long GFP dsRNAs expressed from RNA polymerase II promoters [17] or GFP shRNAs expressed from the U6 RNA polymerase III promoter [18]. Here, we showed that XHSF1 shRNAs expressed from the Xenopus tropicalis H1 RNA promoter (RNA polymerase III promoter) inhibited the HSR by more than 70% in A6 cells. This implies that the RNAi-mediated inhibition of XHSF1 expression in A6 cells is highly efficient. Mice that are heterozygous for a null mutation in the HSF1 gene, still display a 100% HSR, whereas the HSR is completely abolished in homozygous null mutant mice [2]. There are several reasons why the XHSF1 shRNA might not be effective in transgenic tadpoles, despite its potent inhibitory activity in A6 cells. The heterogeneous genetic background of the parental frogs combined with the pseudotetraploid nature of their genome may result in mismatches between the shRNA sequence and the endogenous XHSF1 sequences, which would negatively influence the RNAi effect. Essential components of the RNAi machinery, such as the Dicer enzyme, may be present at suboptimal levels in tadpoles compared with A6 cells. In addition, activation of the RNAi machinery may require a threshold level of shRNAs that is reached in transiently transfected A6 cells, but not in stably transgenic tadpoles.

The dominant-negative XHSF380 mutant completely abolished the HSR in A6 cells. In tadpoles, its expression from a ubiquitously active promoter resulted in embryonic lethality. The Tet-on system has been successfully used to regulate the expression of GFP and thyroid hormone receptor in Xenopus tadpoles [21]. GFP-XHSF380 and XHSF380-GFP cassettes under the control of a Tet-responsive CMV promoter were mixed with a Tet-activator expression cassette and used to generate transgenic larvae. Embryos were allowed to develop in the absence of doxycyclin. Subsequently, all normal GFP-negative tadpoles (n~500) were exposed to doxycyclin to induce expression of GFP-tagged XHSF380. After 20 h doxycyclin treatment, ~1% of the tadpoles showed nuclear GFP fluorescence (Fig 5D), predominantly in muscle and again not in the nervous system, in spite of the fact that the TetO system has been shown to drive strong neuronal expression [21]. Further tadpole development in the presence of doxycyclin remained normal, indicating that the lethal effect of XHSF380 is stage-specific and that it can be circumvented by delaying the expression until after embryogenesis.
Figure 5. Transgene-driven expression of dominant-negative HSF1 in Xenopus larvae. (A) Reporter gene analysis of the effect of GFP-XHSF380 or XHSF380-GFP on the HSR. A6 cells were transfected with mixtures of an Hsp70-luciferase reporter, a CMV-β-galactosidase reporter and the indicated plasmids. Relative luciferase activities and -fold induction were determined as described in the legend to fig. 2C. The results are the average of three independent transfections (standard deviations are indicated by error bars). (B) Fluorescence microscope analysis of GFP-tagged XHSF380 constitutively expressed from either the CMV or the EF1α promoter in transgenic Xenopus larvae. Larvae with high constitutive expression of GFP-tagged XHSF380 develop poorly and never reach the feeding tadpole stage. Also shown are transgenic Xenopus larvae showing strong neuronal expression of GFP when driven by the EF1α or the Ntub promoter. (C) Fluorescence microscope analysis of transgenic Xenopus larvae expressing full
length GFP-tagged XHSF1 from the CMV promoter. Fluorescence is pronounced in kidney, epiphysis, nasal epithelium, olfactory lobes, gills, retinal ganglion cell layer (RGC) and tail muscle nuclei. (D) Fluorescence microscope analysis of doxycycline-induced expression of GFP-tagged XHSF380 expressed in transgenic Xenopus larvae. A mixture of TetO-XHSF380-GFP and CS2+rtTA2-M2 (CMV-TAM2) cassettes was used to generate transgenic Xenopus larvae. The larvae were allowed to develop in the absence of doxycycline until the feeding tadpole stage. GFP-negative larvae were exposed to doxycycline for 20 h and then monitored for GFP fluorescence.

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Animal care

Female South-African claw-toed frogs (Xenopus laevis) were obtained from Xenopus Express (Cape Town, South-Africa) and kept in water tanks at 18°C at the Central Animal Facility of the Radboud University Nijmegen. Ovulation was induced by injection of 500 units hCG (Pregnyl; Organon) into the dorsal lymph sac.

DNA constructs

Oligonucleotides that were used to generate recombinant DNA constructs are listed in the on line supplementary information (Table S1). New vectors were constructed to express proteins with N- or C-terminally fused GFP in Xenopus A6 cells and tadpoles. The chimeric enhancer/promoter of the Xenopus elongation factor-1α (EF1α) gene was PCR amplified from pEF-GFP3 (kindly donated by Dr. Paul Krieg; [29]) using the EF1α primer set, cut with SstI and HindIII and used to replace the CMV promoter of the Xenopus vector pCS2+ [30], resulting in pEF2+. The code for GFPdelAUG was PCR amplified from pRES2-EGFP (Clontech) using the GFPdelAUG primer set, and cloned into the BamHI and XhoI sites of pEF2+, yielding pEF-GFPdelAUG. Similarly, the code for GFPdelSTOP was amplified from pRES2-EGFP using the GFPdelSTOP primer set, and cloned into the BamHI and XhoI sites of pEF2+, yielding pEF-GFPdelSTOP. Bicistronic expression vectors, based on viral 2A peptides, were generated as follows: pEF-GFP-T2A-delSTOP was made by annealing the T2A primer set, and cloning the double stranded oligo into the EcoRI and XhoI sites of pEF-GFPdelAUG. Similarly, pEF-GFP-T2A-delSTOP was made by annealing the T2A primer set, and cloning the double stranded oligo into the EcoRI and XhoI sites of pEF-GFPdelAUG.

Adult Xenopus laevis brain mRNA was used as a source for XHSBP1, XHSF and XHSF380 cDNA. Total RNA was isolated using the RNeasy kit (Qiagen) and copied into cDNA using the Marathon kit (Clontech). The 243-bp XHSBP1 cDNA was PCR amplified from the Marathon cDNA library using the XHSBP1 primer set, and cloned into the EcoRI and XhoI sites of pEF-GFPdelSTOP, yielding pEF-GFP-XHSBP1. To express native protein, the XHSBP1 cDNA was also cloned into the EcoRI and XhoI sites of pEF-GFP-T2A-delSTOP, resulting in pEF-GFP-

Materials and Methods

Ethics statement

Animal experiments were carried out in accordance with the European Communities Council Directive 86/609/EEC for animal welfare, and were approved by the Radboud University Animal Experimentation Committee (permit TRC 99/15072 to generate and house transgenic Xenopus).

Table 1. Expression of XHSF380-GFP, GFP-XHSF380 or GFP from ubiquitous and neuron specific promoters in transgenic Xenopus larvae.

<table>
<thead>
<tr>
<th>Transgene</th>
<th>total number of larvae</th>
<th>number of GFP positive larvae</th>
<th>phenotype of GFP positive larvae</th>
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<tr>
<td>EF1α-GFP</td>
<td>75</td>
<td>8</td>
<td>normal</td>
</tr>
<tr>
<td>CMV-XHSF380-GFP</td>
<td>280</td>
<td>28</td>
<td>lethal</td>
</tr>
<tr>
<td>CMV-GFP-XHSF380</td>
<td>290</td>
<td>12</td>
<td>lethal</td>
</tr>
<tr>
<td>EF1α-GFP-XHSF380</td>
<td>255</td>
<td>13</td>
<td>lethal</td>
</tr>
<tr>
<td>Ntub-GFP</td>
<td>160</td>
<td>16</td>
<td>normal</td>
</tr>
<tr>
<td>Ntub-XHSF380-GFP</td>
<td>155</td>
<td>0</td>
<td>N.A.</td>
</tr>
<tr>
<td>Ntub-GFP-XHSF380</td>
<td>145</td>
<td>0</td>
<td>N.A.</td>
</tr>
<tr>
<td>CMV-GFP-XHSF1</td>
<td>225</td>
<td>6</td>
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T2A-XHSBP1. The 1143-bp XHSF380 cDNA was PCR amplified from the Xenopus brain cDNA library using the XHSF380-C primer set, corresponding to GenBank sequence BC087308 from the NIH Xenopus initiative [31]. XHSF380 was cloned into the EcoRI and XhoI sites of pH1CG2+ vector, resulting in pH1CG2+-XHSF380. In parallel, the XHSF380 cDNA was PCR amplified using the XHSF380-N primer set, cut with BglII and EcoRI, and cloned into the XhoI and EcoRI sites of pEF-GFPdelAUUG, resulting in pEF-XHSF380-GFP. To drive strong ubiquitous expression of XHSF380-GFP and GFP-XHSF380, the EF1α promoter was replaced with the CMV promoter, resulting in pCMV-GFP-XHSF380 and pCMV-XHSF380-GFP. To drive neuron-specific expression, the EF1α promoter was replaced with the neural-tubulin (Ntub) promoter, resulting in pNtub-GFP-XHSF380 and pNtub-XHSF380-GFP. Inducible expression constructs were generated by PCR amplifying the TetO promoter from pCS2+rtTA2A-M2 and reporter plasmid pCS2+[tetO]-GFP using the TetO primer set, cutting the PCR fragment with SalI and HindIII and using in a BLAST search of the X. laevis, and human H1 RNA gene. Based on the alignment of six intact copies of the gene, the Xh1 promoter set was designed and used to PCR amplify a Xenopus tropicalis H1 RNA promoter (Xh1). The human H1 RNA promoter in the pH1CG2+ vector was replaced with the Xh1 promoter using SalI and HindIII, resulting in pXh1CG2+ vector.

Target sequences for RNAi-mediated decay of XHSF1 mRNA were chosen according to http://www.promega.com/rnaiexpress (Invitrogen). The Xh1CG2+XHSF-sh1 was made by annealing of the XHSF-sh1 primer set, and cloning the double stranded oligo into the BglII and HindIII sites of pH1CG2+ vector, resulting in pXh1CG2+XHSF-sh1, -sh2, and -sh3 primer sets, respectively, and cloning the double stranded oligo into the BglII and HindIII sites of pXh1CG2+ vector.

The control plasmid pEF-GFP was made as follows: GFP cDNA was excised from pIRE2-EGFP (BD Sciences-Clontech) using MscI and NotI (filled-in), introduced into the EcoRI site of pBluescript SK-; then excised again using HindIII (filled) and EcoRI and introduced into the SfiI and EcoRI sites of pEF2+. All plasmids were sequence verified.

**Xenopus transgenesis**

All expression cassettes, consisting of promoter, cDNA and polyadenylation signal, were excised from their plasmid vector backbones using SfiI and NotI, separated by agarose gel electrophoresis, and recovered from agarose slices using the GFX gel band purification kit (Amersham). Transgenesis of Xenopus laevis was performed according to Kroll and Amaya [33], with modifications [34]. In summary: 250,000 sperm nuclei were mixed with ~200 ng DNA fragment, incubated for 15 min at room temperature and diluted in 500 μl sperm dilution buffer (250 mM sucrose, 75 mM KCl, 0.3 mM spermidine tribromohydrichloride, 0.2 mM spermidine tetrahydrochloride, 5 mM MgCl2, pH 7.4). Eggs were dejellied in 2% cysteine/1 × MMR (1 × MMR: 0.1 M NaCl, 0.02 M KCl, 0.01 M MgCl2, 0.15 M CaCl2 en 0.5 M HEPES pH 7.5), transferred to 6% Ficoll/0.1 M HEPES and incubated overnight at 17°C. At the 4-cell stage, the embryos were transferred to 6% Ficoll/0.1 × MMR and incubated overnight at 17°C. At the gastrula stage, the embryos were transferred to 0.1 × MMR and incubated at 22°C. GFP-positive tadpoles were photographed using a MZ FLIII fluorescence stereomicroscope provided with a DC200 camera (Leica Microsystems, Switzerland).

**Western blot analysis**

Tadpoles or A6 cell pellets were snap frozen in liquid nitrogen and homogenized in 100 μl SDS-PAGE sample mix. Protein samples (25 μl) were separated in 12% polyacrylamide gels and transferred to nitrocellulose transfer membrane (Portran, Schleicher and Schuell) using a Biorad Mini-PROTEAN II Electrophoresis cell according to the manufacturer’s instructions (Biorad). For western blot analysis, monoclonal ρ-GFP antibody (Cat. no. 632375; Clontech) was used at a 1:5000 dilution. Monoclonal α-tubulin antibody (kindly donated by Mr. Huib Croes, Dept. of Cell Biology, NCMLS, Radboud University Nijmegen, The Netherlands) was used at a 1:1000 dilution. Monoclonal anti-Hsp90 antibody (610418, BD Biosciences) was used at a 1:1000 dilution. Monoclonal anti-α-tubulin antibody was used at a 1:5000 dilution. Proteins were visualized using the SuperSignal West Pico Chemiluminescent Substrate kit (Pierce).

**Tissue culture, transfections, and reporter gene assays**

Xenopus A6 kidney epithelial cells (ATCC CCL-1020) were kindly donated by Ms. Stieneke van den Brink from the Hubrecht laboratory (Utrecht, the Netherlands). The cells were cultured at room temperature (~25°C) in 70% Leibovitz medium supplemented with 10% fetal calf serum (FCS) and 25 mM Heps pH 7.2. Transient transfection was performed using Eugene-6 (Roche) according to the manufacturer’s instructions. Cells were seeded on six-well plates (2.5 x 104 per well) and on the next day transfected with ~1 μg plasmid per well in serum-free medium. The following day, the transfection mix was replaced with medium supplemented with FCS. For GFP and DsRed fluorescence analysis, cells were cultured on cover slips and fixed in 4% paraformaldehyde at 24 h after transfection. Fluorescence was monitored with a Leica DM RA microscope (Leica Microsystems), coupled to a Cohu high performance CCD camera. For luciferase
assays, cells were transfected with a mixture of 100 ng pCMV-β-galactosidase, 200 ng pHλ, and 800 ng of the plasmid indicated in the figures. At 48 h after transfection, cells were either left at room temperature (control) or incubated at 33°C for 1 hour (heat shock). After 6 h recovery at room temperature, cells were lysed in 200 μl reporter lysis mix (2.5 mM, 0.05% Tween 20, 0.05% Tween 80) for 10 min. For the β-galactosidase assay, 40 μl cell lysate was mixed with 100 μl phosphate solution [100 mM Na-phosphate pH 8.2, 10 mM MgCl2, 1% Galacton-Plus (Tropix)]. After 30 min incubation at room temperature, 150 μl accelerator II (Tropix) was added and luminescence was measured with the Lumat LB 9507 tube luminometer (Berthold technologies). For the luciferase assay, 40 μl cell lysate was mixed with 50 μl luciferin solution and luminescence was again measured with the Lumat luminometer. All reporter gene assays were performed in triplicate.

References

Supporting Information

Table S1

| Found at: doi:10.1371/journal.pone.0010158.s001 (0.05 MB DOC) |

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Author Contributions

Conceived and designed the experiments: RPD NL. Performed the experiments: RPD RvG SH STvG. Analyzed the data: RFD. Wrote the paper: RPD NL.