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The Calcium-Sensing Receptor Promotes Urinary Acidification to Prevent Nephrolithiasis

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ABSTRACT

Hypercalciuria increases the risk for urolithiasis, but renal adaptive mechanisms reduce this risk. For example, transient receptor potential vanilloid 5 knockout (TPRV5−/−) mice lack kidney stones despite urinary calcium (Ca2+) wasting and hyperphosphaturia, perhaps as a result of their significant polyuria and urinary acidification. Here, we investigated the mechanisms linking hypercalciuria with these adaptive mechanisms. Exposure of dissected mouse outer medullary collecting ducts to high (5.0 mM) extracellular Ca2+ stimulated H+ ATPase activity. In TPRV5−/− mice, activation of the renal Ca2+-sensing receptor promoted H+-ATPase–mediated H+ excretion and downregulation of aquaporin 2, leading to urinary acidification and polyuria, respectively. Gene ablation of the collecting duct-specific B1 subunit of H+-ATPase in TPRV5−/− mice abolished the enhanced urinary acidification, which resulted in severe tubular precipitations of Ca2+-phosphate in the renal medulla. In conclusion, activation of Ca2+-sensing receptor by increased luminal Ca2+ leads to urinary acidification and polyuria. These beneficial adaptations facilitate the excretion of large amounts of soluble Ca2+, which is crucial to prevent the formation of kidney stones.


Hypercalciuria constitutes the main risk factor in urolithiasis. Patients with renal stone disease experience severe pain, a high rate of recurrence, slowly progressing tissue damage, and eventually renal insufficiency, and recurrent clinical treatment is needed.1,2 Different types of hypercalciuria are known. In this respect, absorptive, renal, and reabsorptive hypercalciuria are distinguished: Disturbed intestinal Ca2+ hyperabsorption, impaired renal Ca2+ reabsorption, and increased bone reabsorption, respectively, are primarily involved.3–6 Current treatment strategies of renal stones consist of (pain) medication, dietary adjustments, hydration, extracorporeal shock wave treatment, or surgery.

The transient receptor potential vanilloid member 5 (TRPV5), the epithelial Ca2+ channel, which is expressed in the renal distal convoluted tubule (DCT) and the connecting tubule (CNT), constitutes the major apical entry step of Ca2+ from the pro-urine into the renal cells.7 TRPV5 knockout (TRPV5−/−) mice demonstrate a robust hypercalciuria with hyperphosphaturia, predisposing these mice to an increased risk for renal Ca2+-phosphate stone formation.8 Concomitantly, TRPV5−/− mice display profound polyuria and increased urinary
acidiﬁcation.8 Urinary acidification occurs along different segments of the nephron and is of crucial importance in acid/base homeostasis.9,10

In combination with urinary buffers, the activity of acid/base transporters in intercalated cells (ICs) of CNT and collecting duct system (CD) accomplish the ﬁne-tuning of urinary pH. In type A ICs, the multisubunit vacuolar ATP-driven proton pump (H+-ATPase) is expressed at the apical membrane, which is mainly responsible for proton excretion.21 Inactivating mutations in the genes encoding individual subunits of the H+-ATPase proton pump are associated with distal renal tubular acidosis, characterized by a decrease in net proton secretion.12–14 The formation of less acidic urine predisposes to urolithiasis as the crystallization of Ca2+-phosphate occurs via the conversion of phosphate to its divalent form (HPO4 2−) in an alkaline rather than in an acidic environment.15,16 Previously, insufficient urinary acidification was demonstrated in recurrent stone formers.17,18 Currently, no clinical trials have documented the prevention of stone formation by stimulating urinary acidification.

Aquaporin 2 (AQP2) is the water channel localized at the apical membrane of the CNT and CD that is responsible for vasopressin-regulated urinary water reabsorption.19,20 Previous studies linked Ca2+ and water homeostasis, suggesting a functional role for the G-protein–coupled Ca2+/polyvalent cation-sensing receptor (CaSR).21–23 Immunolocalization studies of renal CaSR in rats revealed expression on the apical membrane of both proximal tubule and CD, the basolateral membrane of thick ascending limb of Henle (TAL), and a dif-

Table 1. Physiologic parameters of control and TRPV5−/− micea

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>TRPV5−/−</th>
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<tbody>
<tr>
<td>Urine</td>
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<td></td>
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<tr>
<td>Ca2+ (µmol/24 h)</td>
<td>10 ± 2</td>
<td>132 ± 31b</td>
</tr>
<tr>
<td>phosphate (µmol/24 h)</td>
<td>65 ± 8</td>
<td>142 ± 13b</td>
</tr>
<tr>
<td>citrate (mg/24 h)</td>
<td>40 ± 10</td>
<td>56 ± 13</td>
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<tr>
<td>oxalate (µmol/24 h)</td>
<td>3.4 ± 0.9</td>
<td>2.7 ± 0.5</td>
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<td>volume (ml/24 h)</td>
<td>1.5 ± 0.3</td>
<td>3.6 ± 0.5b</td>
</tr>
<tr>
<td>pH</td>
<td>7.4 ± 0.1</td>
<td>5.5 ± 0.1b</td>
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<tr>
<td>osmolality (mOsmol/kg)</td>
<td>1928 ± 201</td>
<td>1222 ± 175b</td>
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<tr>
<td>Serum</td>
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<tr>
<td>Ca2+ (mM)</td>
<td>2.36 ± 0.02</td>
<td>2.42 ± 0.02</td>
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<tr>
<td>ionized Ca2+ (mM)</td>
<td>1.27 ± 0.01</td>
<td>1.30 ± 0.01</td>
</tr>
<tr>
<td>phosphate (mM)</td>
<td>2.5 ± 0.2</td>
<td>2.3 ± 0.1</td>
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*aUrinary and serum biochemical parameters of 8-wk-old control and TRPV5−/− mice. Data are means ± SEM (n = 5).

bP < 0.05, signiﬁcant difference from control mice.

Table 2. Urinary TA in control and TRPV5−/− micea

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>TRPV5−/−</th>
</tr>
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<tbody>
<tr>
<td>NH4+/NH4+ (mmol/24 h)</td>
<td>0.033 ± 0.006</td>
<td>0.036 ± 0.007</td>
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<td>TA (mEq/24 h)</td>
<td>0.048 ± 0.007</td>
<td>0.121 ± 0.006b</td>
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<tr>
<td>HCO3− (mmol/24 h)</td>
<td>0.002 ± 0.001</td>
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<tr>
<td>NAE (mEq/24 h)</td>
<td>0.083 ± 0.012</td>
<td>0.157 ± 0.012b</td>
</tr>
<tr>
<td>pH</td>
<td>6.2 ± 0.1</td>
<td>5.6 ± 0.1b</td>
</tr>
</tbody>
</table>

*aUrinary TA in a novel experiment with 8-wk-old control and TRPV5−/− mice (n = 7). Data are means ± SEM.

bP < 0.001, signiﬁcant difference from control mice.

Figure 1. Urinary Ca2+ excretion, volume, and pH. (A and B) Correlation of urinary Ca2+ excretion with (A) urinary volume and (B) urinary pH in control and TRPV5−/− mice. Each dot represents single mouse datum. Linear regression analysis conﬁrmed signiﬁcant correlations of urinary Ca2+ excretion with urinary volume (r² = 0.83; P < 0.0001; n = 62) and urinary pH (r² = 0.70; P < 0.0001; n = 42). (C) Urinary Ca2+ excretion was determined in 24-h urine samples of 5-wk-old control, TRPV5−/−, TRPV5−/−/Atp6v1b1−/−, and Atp6v1b1−/− mice. TRPV5−/− and TRPV5−/−/Atp6v1b1−/− mice showed a signiﬁcant hypercalciuria, whereas hypocalciuria was present in Atp6v1b1−/− mice compared with control mice. (D) TRPV5−/− mice demonstrated a signiﬁcant decrease in urinary pH compared with control mice, whereas TRPV5−/−/Atp6v1b1−/− mice exhibited urinary pH levels similar to controls. Impaired urinary acidification in Atp6v1b1−/− mice resulted in an increase of urinary pH. (E) An increased urinary volume was demonstrated in TRPV5−/− compared with control mice. TRPV5−/−/Atp6v1b1−/− mice displayed a further increase in 24-h urinary volume compared with TRPV5−/− mice. In C through E, data are means ± SEM. *P < 0.05 (n = 6; signiﬁcant difference from control); #P < 0.05 (n = 6; signiﬁcant difference from TRPV5−/− mice).
fuse pattern throughout the DCT. Renal CaSR expression is highest in TAL cells and is absent from the glomerulus. The differential expression and localization of CaSR along the nephron is indicative of the variety of physiologic functions throughout the kidney. The localization of the CaSR along the CD facilitates the activation of the receptor during a hypercalciuric state. In the CD, a CaSR-mediated decrease of the urine concentrating ability could eventually lead to polyuria, assisting the excretion of large amounts of Ca\(^{2+}\).

The aim of this study was to elucidate molecular mechanisms involved in the adaptations that prevent renal stone formation during hypercalciuria. The TRPV5\(^{-/-}\) mouse model was used because polyuria and increased urinary acidification naturally occur in these hypercalciuric mice. The crucial roles of the functional H\(^+-\)ATPase proton pump, the AQP2 water channel, and the CaSR are demonstrated in these hypercalciuria-related adaptations. This study provides new insights regarding the formation and prevention of kidney stones.

**RESULTS**

**Metabolic Studies**

At 8-wk-old TRPV5\(^{-/-}\) and littermate control mice were housed in metabolic cages for 24 h and killed. Genetic ablation of TRPV5 resulted in hypercalciuria, and these mice remained normocalcemic (Table 1). In addition, TRPV5\(^{-/-}\) mice displayed hyperphosphaturia with normal serum phosphate levels and a significant polyuria with decreased urinary osmolality. Urinary citrate and oxalate excretion levels were not different between the two mouse strains. Urinary pH was significantly lower in TRPV5\(^{-/-}\) mice compared with control mice. The 24-h ammonium and HCO\(_3^-\) excretion levels were unchanged (Table 2). Furthermore, urinary titratable acidity (TA) was significantly increased in TRPV5\(^{-/-}\) mice. Consequently, renal net acid excretion (NAE), which is the sum of renal ammonium excretion and TA minus HCO\(_3^-\) excretion, was elevated in TRPV5\(^{-/-}\) mice. Ca\(^{2+}\) excretion levels correlated linearly with urinary volume as well as pH (Figure 1A and B, respectively). To address the contribution of H\(^+-\)ATPase activity to the acidified urine in TRPV5\(^{-/-}\) mice, we generated TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice that were genetically ablated for both TRPV5 and the IC-specific B1 subunit of H\(^+-\)ATPase (Atp6v1b1). Hypercalciuria remained present in 5-wk-old TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice, although less than in TRPV5\(^{-/-}\) mice (Figure 1C). Urinary pH was normalized (Figure 1D) whereas urinary volume was further increased in TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice in comparison with TRPV5\(^{-/-}\) mice (Figure 1E).

![Figure 2](https://example.com/figure2.png)

**Figure 2.** Analysis of TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice. (A) Growth curves comparing body weights of mouse genotypes, representing a litter containing one TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mouse that died at 14 d of age (dotted line) and four littermates (solid lines). (B) Growth retardation in TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice, demonstrating 5-wk-old control (left) and TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) (right) mice. (C) Mortality rate of TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) (solid line; n = 10) and littermate (dotted line; n = 55) mice. (D) Kidneys from 5-wk-old control (left) and TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice (right). (E) Renal histology of a 5-wk-old TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mouse. (F) Von Kossa staining in renal sections of a 1-wk-old TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mouse. (G) Transmission electron microscopy image of a tubular precipitate in the medullary collecting duct of a 1-wk-old TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mouse. (H) STEM image of (1) a precipitate, x-ray maps separately depicting (2) Ca\(^{2+}\), and (3) phosphorus content. (I) Energy dispersive x-ray microanalysis of the renal precipitate (arrow in H) demonstrating Ca\(^{2+}\) and phosphorus content. This spectrum represents all spot measurements performed (n = 35 spots). Magnifications: ×100 in E (left), F (left), ×250 in E (right), F (right); ×2500 in G.

**Phenotypic Analysis of the TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\)Mice**

TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice were retarded in their growth directly after birth, compared with littermate mice (Figure 2A).
At 5 wk of age, TRPV5−/−/Atp6v1b1−/− mice were easily identified because of severe growth retardation (Figure 2B). Moreover, 80% of the TRPV5−/−/Atp6v1b1−/− mice died within 6 wk after birth (Figure 2C). These double-knockout mice showed bilateral hydronephrosis (Figure 2D) and abnormal dilation of the CD (Figure 2E). Von Kossa staining in kidneys of 1-wk-old TRPV5−/−/Atp6v1b1−/− mice showed massive tubular Ca2+ precipitations in the medullary CD (Figure 2F and G). Subsequently, kidney samples that included the observed precipitates were analyzed by energy-dispersive x-ray microanalysis using a Jeol 1200/STEM (Jeol Ltd., Tokyo, Japan) in combination with a Thermo Noran microanalysis SIX system (Thermo Fisher Scientific Inc., Waltham, MA) (Figure 2H). The x-ray maps and spectrum unequivocally showed the presence of Ca2+ and phosphorus (Figure 2, H and 2 and 3, and 1) in the precipitates present in the lumen of the CD suggesting Ca2+-phosphate crystals.

Renal and Intestinal Expression of Water, Acid, and Phosphate Transporters

To address renal AQP2 expression levels in control and TRPV5−/− mice, we performed immunoblot analyses. AQP2 was significantly downregulated in TRPV5−/− mice compared with control mice at 8 wk of age (Figure 3A). Moreover, a further AQP2 downregulation was detected in TRPV5−/−/Atp6v1b1−/− mice compared with TRPV5−/− mice, analyzed at 5 wk of age (Figure 3B). Membrane fractions were isolated from renal medulla tissue of 8-wk-old TRPV5−/− and control mice. Analysis of AQP2 protein expression in the membrane fractions revealed a significant downregulation in kidneys of TRPV5−/− compared with control littermate mice (Figure 3C). Immunoblotting for the B1 subunit of H+-ATPase revealed no differences in expression levels between TRPV5−/− and control mice (Figure 4A). Furthermore, a significant downregulation of the renal type IIA NaPi co-transporter (NaPi-IIa) was revealed in TRPV5−/− mice (Figure 4B). In contrast, expression of the intestinal type IIB NaPi co-transporter (NaPi-IIB) was increased in brush border membrane vesicles isolated from ileum of TRPV5−/− mice compared with control mice (Figure 4C).

Effect of Ca2+ on H+-ATPase Activity of Outer Medullary CDs

We enzymatically isolated outer medullary CDs (OMCDs) from mouse renal tissue to perform pH2 measurements. The activity of H+-ATPase was determined from the pH2 recovery rate (∆pH2/min) after intracellular acidification by NH4Cl.25 We investigated the effect on H+-ATPase activity of low (0.1 mM), normal (1.0 mM), and high (5.0 mM, mimicking the hypercalciuric state of TRPV5−/− mice) extracellular [Ca2+]. Representative recordings are depicted in Figure 5, A and C. Exposure of OMCDs to 5.0 mM [Ca2+] significantly increased H+-ATPase–mediated pH2 recovery rates in control (n = 178 cells; 15 OMCDs; five mice) and TRPV5−/− mice (n = 124 cells; nine OMCDs; four mice; Figure 5D) compared with OMCDs exposed to 0.1 mM [Ca2+] from control mice (n = 159 cells; 12 OMCDs; four mice) and TRPV5−/− mice (n = 149 cells; 13 OMCDs; five mice). Comparison of pH2 recovery rates in control and TRPV5−/− mouse OMCDs exposed to 0.1 mM [Ca2+] revealed no significant differences in H+-ATPase activity (Figure 5, B and D). To confirm the specific involvement of the H+-ATPase proton pump, we performed the following experiments. First, OMCDs were incubated with 100 nM concanamycin, a specific H+-ATPase inhibitor, which prevented the stimulatory action of 5.0 mM [Ca2+] on the pH2 recovery rate in OMCDs of control as well as

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**Figure 3.** Immunoblotting for renal AQP2. (A) Representative immunoblots for AQP2, revealing 29- and 35- to 50-kD bands representing nonglycosylated and glycosylated AQP2, respectively, which were all included in the semiquantitative determination of renal AQP2 protein abundance from 8-wk-old control versus TRPV5−/− mice (n = 4). (B) Renal AQP2 protein abundance comparing 5-wk-old control, TRPV5−/−, TRPV5−/−/Atp6v1b1−/−, and Atp6v1b1−/− mice (n = 3). (C) AQP2 expression levels in renal medullary membrane fractions from control and TRPV5−/− mice. Broad-range protein and prestained marker bands are depicted, indicating protein size. Expression levels were corrected for β-actin expression. Data were calculated as integrated optical density (arbitrary units), were depicted as percentage of control mice, were presented as means ± SEM, and represented two separate experiments with a total of n = 4. *P < 0.05 (significant difference from control); #P < 0.05 (significant difference from TRPV5−/−).
TRPV5\(^{-/-}\) mice. Second, the pH\(_1\) recovery rates were not affected by 5.0 mM compared with 0.1 mM [Ca\(^{2+}\)] in OMCDs of Atp6v1b1\(^{-/-}\) mice, underlining an essential role of H\(^+/\)ATPase in this acidification process (Figure 5E). OMCDs from control mice were exposed to the CaSR agonist neomycin (200 \mu M) during the measurements to investigate the molecular mechanism of this Ca\(^{2+}\)-mediated increase of H\(^+/\)ATPase activity. Neomycin induced a significant increase in the pH\(_1\) recovery rate (\(n = 100\) cells; eight OMCDs; four mice) compared with nonexposed OMCDs in the presence of 1.0 mM [Ca\(^{2+}\)] (\(n = 90\) cells; nine OMCDs; four mice), indicating the involvement of CaSR activation (Figure 5F).

**DISCUSSION**

As illustrated in this study, activation of the CaSR by increased urinary Ca\(^{2+}\) levels triggers urinary acidification and polyuria that are crucial adaptations in the prevention of renal stone formation during hypercalciuria. This conclusion is based on the following observations. First, TRPV5\(^{-/-}\) mice display hypercalciuria as a result of impaired active Ca\(^{2+}\) reabsorption, concomitant hyperphosphaturia, polyuria, and increased urinary acidification, whereas renal Ca\(^{2+}\) precipitations are not detected. Second, additional gene ablation of Atp6v1b1 in TRPV5\(^{-/-}\) mice prevents the increased urinary acidification and evokes massive Ca\(^{2+}\)-phosphate precipitation. Third, activation of the renal CaSR by elevated luminal Ca\(^{2+}\) levels stimulates H\(^+/\)ATPase-mediated H\(^+\) excretion and renal AQP2 protein downregulation, responsible for the consistent increased urinary acidification and polyuria, respectively, in hypercalciuric TRPV5\(^{-/-}\) mice.

Fine-tuning of renal acid excretion is generally accomplished in the CD, where the H\(^+/\)ATPase proton pump is localized at the apical side of type A ICs. Exposure to a 5.0 mM Ca\(^{2+}\) concentration significantly enhanced H\(^+/\)ATPase activity in OMCDs, a specific stimulatory effect that was absent in OMCDs from Atp6v1b1\(^{-/-}\) mice. Also, the CaSR agonist neomycin increased H\(^+/\)ATPase activity, emphasizing that CaSR activation modulates urinary acid excretion. In TRPV5\(^{-/-}\) mice, an increased luminal Ca\(^{2+}\) concentration is consistently present in the renal DCT, CNT, and CD.\(^8\) This could activate the apical CaSR, increasing H\(^+/\)ATPase activity and thereby initiating the observed acidic urinary pH. Importantly, genetic ablation of TRPV5 increased NAE, which is attributed to a significant rise in urinary TA as a result of hyperphosphaturia. In this perspective, the hyperphosphaturia observed in TRPV5\(^{-/-}\) mice could potentially present an acid load to the animals, initially caused by an increase in NaPi-IIb-mediated phosphate absorption; however, previous investigations showed that phosphate administration in healthy volunteers evokes metabolic alkalosis with a concomitant increase of urinary pH and remarkably only transient increases of TA and NAE.\(^26\) Although a higher dietary acid load cannot be completely ruled out, it did not cause a significant change in the acid-base homeostasis of the TRPV5\(^{-/-}\) mice. A mild metabolic acidosis would also have increased urinary ammonium excretion, which was not observed; therefore, the decline of the urinary pH in TRPV5\(^{-/-}\) mice represents a significant renal acid loss in the presence of hyperphosphaturia. Previously, Nijenhuis et al.\(^27\) showed that TRPV5\(^{-/-}\) mice display normal blood pH and HCO\(_3^-\) levels under basal conditions. These mice become even more alkalemic than wild-type mice when given an NaHCO\(_3\) diet via the drinking water, which indicates that TRPV5\(^{-/-}\) mice do not have a metabolic acidosis. Together with these observations, our data demonstrate that TRPV5\(^{-/-}\) mice have increased urinary acidification without an altered systemic acid-base status.

Additional gene ablation of Atp6v1b1 in TRPV5\(^{-/-}\) mice normalized urinary pH and caused tubular precipitation of Ca\(^{2+}\)-phosphate in the medullary CD. These observations in-
action enhanced H⁺-K⁺-ATPase activity in isolated parietal cells of gastric glands, allowing maximal ionization of dietary Ca²⁺ and establishing enhanced Ca²⁺ absorption from the intestine. Furthermore, Farajov et al. showed that the activation of the CaSR at the basolateral membrane of medullary TAL cells causes increased acid secretion in a hypercalcemic state. Together with our data, this strongly suggests a key role of the CaSR in linking Ca²⁺ metabolism and acid secretion.

Polyuria can diminish the risk for renal stone formation by reducing the urinary Ca²⁺ concentration. In mice, calciuresis linearly correlated with urinary volume, because an increase of Ca²⁺ excretion leads to an enhanced urinary volume. The consistent polyuria in hypercalciuric TRPV5⁻/⁻ mice, assessed by a substantial decrease in urinary osmolality, was caused by downregulation of renal AQP2 water channels. Specifically, immunoblot analysis revealed a significant downregulation of AQP2 protein expression levels in isolated renal medullary membrane fractions from TRPV5⁻/⁻ compared with control mice. Importantly, this is in line with previous studies performed on humans and rats that suggested that activation of the apical CaSR reduces the CD water permeability when the luminal Ca²⁺ level rises. This involves the CaSR-mediated retrieval of AQP2 from the apical membrane into endocytic compartments. Recently, Bustamante et al. showed an attenuated AQP2 expression in a mouse CD cell line after exposure to extracellular Ca²⁺, neomycin, or Gd³⁺. Thus, hypercalciuria activates the apical CaSR in the CD and reduces AQP2-mediated water reabsorption, permitting the disposal of excess urinary Ca²⁺ and preventing the formation of stones.

The B1 subunit of the renal H⁺-ATPase proton pump, encoded by the Atp6v1b1 gene, is important for urinary acidification in the renal CD. Atp6v1b1⁻/⁻ mice demonstrated hypocalciuria, which is an important difference in phenotype between mice and humans with a genetic Atp6v1b1 defect, although it confirms previous observations of Finberg et al. The presence of hypocalciuria in Atp6v1b1⁻/⁻ has not been explained yet and needs further investigation. Moreover, Atp6v1b1⁻/⁻ mice displayed a higher urinary pH compared with control mice. Consequently, gene ablation of the Atp6v1b1 gene in TRPV5⁻/⁻ mice normalized urinary pH. Interestingly, TRPV5⁻/⁻/Atp6v1b1⁻/⁻ mice evoked a more severe polyuria than displayed by the TRPV5⁻/⁻ mice. To explain the increased urinary volume in TRPV5⁻/⁻/Atp6v1b1⁻/⁻ mice, the CaSR response as a function of the extracellular pH was previously investigated, showing the functional modulation of the CaSR by extracellular pH. The increased urinary

dicate that increased H⁺-ATPase-mediated urinary acidification in TRPV5⁻/⁻ mice protects against renal Ca²⁺-phosphate stone formation. Because the formation of alkaline urine adds to the risk for urinary Ca²⁺-phosphate precipitation, renal stones do occur in patients with distal renal tubular acidosis, who display a urinary acidification defect and hypercalciuria. In accordance with our findings, previous studies demonstrated that extracellular Ca²⁺ levels modulate gastric acid secretion, generally controlled by H⁺-K⁺-ATPase, CaSR

Figure 5. Effect of Ca²⁺ and neomycin on vacuolar H⁺-ATPase activity. (A and C) Original pH traces from control mouse single OMCD ICs exposed to low (0.1 mM) and high (5.0 mM) [Ca²⁺], respectively. Bars indicate the alternating HEPES, Na⁺-free HEPES, and NH₄Cl-containing buffers applied to the cells. (B and D) Summary of the data from the experiments performed in control and TRPV5⁻/⁻ mouse OMCD ICs, respectively. (E) ATP6v1b1⁻/⁻ OMCD ICs exposed to a high (5.0 mM) [Ca²⁺]. (F) Effect of neomycin (200 μM) on H⁺-ATPase activity in control mouse OMCD ICs. (B, D, E, and F) pH recovery rates (ΔpH/min), which are presented as means ± SEM, calculated from number of individual OMCD ICs. *P < 0.05 (significant difference from control 0.1 mM [Ca²⁺]); ²P < 0.05 (significant difference from control 5.0 mM [Ca²⁺]); §P < 0.05 (significant difference from TRPV5⁻/⁻ 0.1 mM [Ca²⁺]); †P < 0.05 (significant difference from TRPV5⁻/⁻ 5.0 mM [Ca²⁺]); #P < 0.05 (significant difference from control 1.0 mM [Ca²⁺]). Conc, concanamycin (100 nM); Neo, neomycin.
volume in TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice compared with TRPV5<sup>−/−</sup> mice was explained by a further AQP2 downregulation, demonstrated by immunoblotting. We postulate that the extended decrease of AQP2 expression in the TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice is evoked by the higher sensitivity for Ca<sup>2+</sup> of the CaSR at a neutral urinary pH. On the basis of previous studies, we conclude that, in the absence of increased urinary acidification in TRPV5<sup>−/−</sup> mice, a more severe reduction in water reabsorption is induced in an attempt to prevent renal stone formation. Stehberger et al.<sup>41</sup> demonstrated a decreased urinary osmolality in AE1-deficient (AE1<sup>−/−</sup>) mice as a result of the retrieval of AQP2 water channels to intracellular vesicles in CD cells. This effect could be the result of increased pH sensitivity of the CaSR at an alkaline environment. Despite the severe polyuria, TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice developed renal Ca<sup>2+</sup>-phosphate precipitations that obstructed the CD system at an early age. Thus, these results underscore the crucial importance of increased urinary acidification in the local prevention of renal Ca<sup>2+</sup>-phosphate stones.

TRPV5<sup>−/−</sup> mice also displayed, besides the hypercalciuria, a profound hyperphosphaturia, increasing the risk for Ca<sup>2+</sup>-phosphate precipitation. For maintaining normocalcemia, TRPV5<sup>−/−</sup> mice present with a hypervitaminosis D. Previous investigations showed that increased circulating 1,25-dihydroxyvitamin D<sub>3</sub> levels caused compensatory Ca<sup>2+</sup>-hyperabsorption by upregulation of intestinal Ca<sup>2+</sup>-transporters.<sup>8,42</sup> Moreover, 1,25-dihydroxyvitamin D<sub>3</sub> is an important key player in phosphate homeostasis,<sup>43</sup>–<sup>45</sup> increasing the abundance of intestinal NaPi-IIb transporters.<sup>46</sup> TRPV5<sup>−/−</sup> mice displayed a significant upregulation of NaPi-IIb proteins in the ileum, which might evoke phosphate hyperabsorption. Here, we postulate that downregulation of renal NaPi-IIa proteins results in hyperphosphaturia in TRPV5<sup>−/−</sup> mice, possibly as a compensatory response to the increased phosphate absorption. Whether the NaPi-IIa downregulation could also be an effect of the hypervitaminosis D needs further investigation.

TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice exhibited growth retardation and died before 6 wk of age. The lethal phenotype is likely to be affected by the hydronephrosis, which could be explained by the premature formation of renal stones that physically obstruct the urine flow. This resembles the occurrence of hydronephrosis in congenital nephropathy that is often caused by a functional obstruction in the renal pelvis.<sup>47</sup> Another explanation could be that the hydronephrosis arises in response to the pressure produced by the severe polyuria, dilating the CD and leading to kidney damage. This might also clarify the individual differences in severity of the hydronephrosis, polyuria, and early age of death observed in TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice. A similar observation was previously made in the Nkcc2 knockout mice, a mouse model of Bartter syndrome that displayed polyuria and concomitant hydronephrosis as well.<sup>48</sup> Li et al.<sup>49</sup> previously investigated the presence of uremia subsequent to urinary tract obstruction in rats, which might have occurred in the TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice as well. The less robust hypercalciuria observed in the TRPV5<sup>−/−</sup>/Atp6v1b1<sup>−/−</sup> mice in comparison with the single TRPV5<sup>−/−</sup> mice could be due to the severe increase in urinary volume resulting in a slight dehydration in these mice. This could evoke increased proximal tubular Ca<sup>2+</sup> reabsorption as a compensatory mechanism, decreasing Ca<sup>2+</sup> wasting via the urine.

In conclusion, the increased urinary acidification and polyuria in response to hypercalciuria are prerequisites to prevent the formation of renal Ca<sup>2+</sup>-phosphate stones in TRPV5<sup>−/−</sup> mice. Interestingly, these adaptations involve luminal CaSR activation by increased urinary Ca<sup>2+</sup> levels, inducing enhanced H<sup>+</sup>-ATPase activity and AQP2 downregulation (Figure 6). Renal Ca<sup>2+</sup>-phosphate stone formers could benefit from the induction of a decrease in urinary pH as well as polyuria that is accomplished by increased water intake. Knowledge of these adaptive mechanisms is relevant for the therapy and prevention of kidney stones.

![Figure 6](https://example.com/figure6.png)
CONCISE METHODS

Animal Experiments and Metabolic Data
Please see supplemental online information for detailed methods.

Renal Histology
Von Kossa staining of kidney tissue was performed on fixed 5- to 7-μm cryosections to stain Ca\(^{2+}\)-containing deposits, and further analysis of renal tubular Ca\(^{2+}\) precipitations was performed by transmission electron microscopy. Please see supplemental information for detailed methods.

Protein Isolations and Immunoblotting
To quantify the renal AQP2, the H\(^+\)-ATPase B1 subunit, and the NaPi-IIa protein expression levels in 8-wk-old control and TRPV5\(^{-/-}\) mice, we prepared renal protein lysates as described before.\(^{42}\) Membrane fractions were isolated from renal medulla tissue, according to centrifugation methods as described previously.\(^{52}\) For renal AQP2 quantification in the control, TRPV5\(^{-/-}\), Atp6v1b1\(^{-/-}\), and littermate TRPV5\(^{-/-}\)/Atp6v1b1\(^{-/-}\) mice, kidneys were dissected at 5 wk of age. Ileum brush border membrane vesicles were isolated to monitor pHi. Cells were acidified using the NH\(_4\)Cl (20 mM) pre-treatment for detailed methods.

Tubule Preparations and pH\(_i\) Measurements
Mice were transcardially perfused with a digestion buffer containing 1 mg/ml collagenase (Sigma-Aldrich Chemie, Buchs, Switzerland) to obtain renal OMCDs from control, TRPV5\(^{-/-}\), and Atp6v1b1\(^{-/-}\) mice for H\(^+\)-ATPase activity measurements.\(^{25,31,52}\) BCECF AM (1 μM: Molecular Probes, Invitrogen AG, Basel, Switzerland) was used to monitor pH\(_i\). Cells were acidified using the NH\(_4\)Cl (20 mM) pre-pulse technique, as described previously.\(^{53}\) Please see supplemental information for detailed methods.

ACKNOWLEDGMENTS

This work was supported by the European Science Foundation (EU-RYI), the Swiss National Research Foundation (31-109677/1), the EU 6th Framework project EuReGene, the ERA-EDTA, and the Dutch Kidney Foundation (C03.6017).

We thank Drs. R.P. Lifton and K.E. Finberg (Yale University School of Medicine, New Haven, CT) for providing the Atp6v1b1 knockout mice. We also acknowledge the animal facility of the Radboud University Nijmegen for technical assistance, as well as Drs. L.A. Monnens, J.J. Weening, and C.E. van der Zee for stimulating discussions and technical assistance.

DISCLOSURES

None.

REFERENCES


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