Oral versus Intravenous Flucytosine in Patients with Human Immunodeficiency Virus-Associated Cryptococcal Meningitis

Annemarie E. Brouwer, Hendrikus J. M. van Kan, Elizabeth Johnson, Adul Rajanuwong, Prapit Teparrukkul, Vannaporn Wuthiekanun, Wirongrong Chierakul, Nick Day and Thomas S. Harrison

Updated information and services can be found at:
http://aac.asm.org/content/51/3/1038

These include:

REFERENCES

This article cites 18 articles, 8 of which can be accessed free at:
http://aac.asm.org/content/51/3/1038#ref-list-1

CONTENT ALERTS

Receive: RSS Feeds, eTOCs, free email alerts (when new
articles cite this article), more»

Information about commercial reprint orders: **http://journals.asm.org/site/misc/reprints.xhtml**
To subscribe to to another ASM Journal go to: **http://journals.asm.org/site/subscriptions/**

Oral versus Intravenous Flucytosine in Patients with Human Immunodeficiency Virus-Associated Cryptococcal Meningitis[▽]

Annemarie E. Brouwer,[1,2,3] Hendrikus J. M. van Kan,[4] Elizabeth Johnson,[5] Adul Rajanuwong,[6]
Prapit Teparrukkul,[6] Vannaporn Wuthiekanun,[2] Wirongrong Chierakul,[2]
Nick Day,[2,7] and Thomas S. Harrison[1]*

*Centre for Infection, St. George's University of London, United Kingdom[1]; Faculty of Tropical Medicine, Mahidol University, Bangkok,
Thailand[2]; Department of Internal Medicine and Nijmegen University Centre for Infectious Diseases, University Medical Centre Nijmegen,
The Netherlands[3]; Department of Clinical Pharmacy, Academic Medical Centre, Amsterdam, The Netherlands[4];
Mycology Reference Laboratory, Health Protection Agency South-West Regional Laboratory, Bristol,
United Kingdom[5]; Department of Medicine, Sappasithiprasong Hospital, Ubon Ratchathani, Thailand[6];
and Centre for Tropical Medicine, Nuffield Department of Clinical Medicine, University of
Oxford, John Radcliffe Hospital, Oxford, United Kingdom[7]*

In a randomized controlled trial of amphotericin B-based therapy for human immunodeficiency virus (HIV)-associated cryptococcal meningitis in Thailand, we also compared the mycological efficacy, toxicity, and pharmacokinetics of oral versus intravenous flucytosine at 100 mg/kg of body weight/day for the initial 2 weeks. Half of 32 patients assigned to the two arms containing flucytosine were randomized to oral and half to intravenous flucytosine. Early fungicidal activity was determined from serial quantitative cultures of cerebrospinal fluid (CSF), and toxicity was assessed by clinical and laboratory monitoring. Flucytosine and fluorouracil concentrations in plasma and CSF were measured by high-performance liquid chromatography. No significant bone marrow or hepatotoxicity was seen, there was no detectable difference in bone marrow toxicity between patients on intravenous and those on oral formulation, and no patients discontinued treatment. In patients receiving intravenous flucytosine, the median 24-h area under the concentration-time curve was significantly higher than in the oral group. Despite this difference, there was no difference in early fungicidal activity between patients on intravenous compared with patients on oral flucytosine. The results suggest that either formulation can be used safely at this dosage in a developing country setting, without drug concentration monitoring. The bioavailability of the oral formulation may be reduced in late-stage HIV-infected patients in Thailand. Concentrations of flucytosine with intravenous formulation at 100 mg/kg/day may be in excess of those required for maximal fungicidal activity.

Flucytosine (5FC) in combination with amphotericin B (AMB) is standard therapy for cryptococcal meningitis in the United States and Europe. 5FC is taken up by fungal cells by cytosine permease and converted into fluorouracil (5FU) by fungal cytosine deaminase. Further metabolism of 5FU leads to the formation of 5-fluorouridine triphosphate, which is incorporated into fungal RNA, and 5-fluorodeoxyuridine monophosphate, an inhibitor of thymidylate synthetase. This results in inhibition of protein and DNA synthesis in the fungal cell (19).

Side effects of 5FC include nausea, vomiting, diarrhea, bone marrow depression, and hepatotoxicity. The latter two are thought to be due to effects of 5FU. Human cells lack the enzyme cytosine deaminase and are unable to convert 5FC into 5FU. However, the human intestinal microflora has been shown to be capable of converting 5FC into 5FU in vitro (8, 10, 17), and 5FU, at concentrations known to be associated with bone marrow depression, has been measured in the plasma of patients treated with oral 5FC (6). If intestinal bacteria do play

a role in conversion of 5FC to 5FU in patients, then oral administration of 5FC might be associated with increased 5FU concentrations and more side effects than intravenous (i.v.) administration of the drug. On the other hand, i.v. 5FC is more costly to administer in resource-poor settings and carries the added inconvenience of strict storage temperature requirements. Therefore, in the context of a trial of combination antifungal therapy for human immunodeficiency virus (HIV)-associated cryptococcal meningitis, we compared the efficacy, toxicity, and pharmacokinetics of oral versus i.v. 5FC.

MATERIALS AND METHODS

Participants and procedures. The study was approved by the ethical and scientific review subcommittee of the Thai Ministry of Public Health and by the research ethics committee of St. George's University of London and was carried out at Sappasithiprasong Hospital, Thailand, as described previously (4). With written informed consent, we enrolled 64 adults with a first episode of cryptococcal meningitis, diagnosed by cerebrospinal fluid (CSF) India ink and cryptococcal antigen tests. Exclusion criteria were an alanine aminotransferase (ALT) concentration of more than five times the upper limit of normal, a neutrophil count of less than 0.5×10^9/liter, a platelet count of less than 50×10^9/liter, pregnancy, and previous serious reaction to study drugs. The participants were randomized to give equal numbers in each of four treatment arms: amphotericin B deoxycholate alone (0.7 mg/kg of body weight daily, Fungizone; Bristol-Myers Squibb, New York, NY); amphotericin B plus fluconazole (400 mg daily, Diflucan; Pfizer, New York, NY); amphotericin B plus flucytosine (100 mg/kg daily, Ancotil [i.v. formulation; Valeant, Zoetermeer, The Netherlands]

---

and Ancobon [oral tablets; Basingstoke, United Kingdom]); and triple therapy with amphotericin B, flucytosine, and fluconazole. Half of the patients in each of the two arms containing 5FC were randomized to oral and half to i.v. 5FC. Treatment was not blinded. Unless contraindicated, patients received 1 liter of 0.9% (normal) saline daily to keep AMB nephrotoxicity to a minimum. After 2 weeks, we treated all four arms with fluconazole, 400 mg daily for 8 weeks, and 200 mg daily thereafter (Fluzoral; Government Pharmaceutical Organization, Thailand).

Efficacy. The rate of clearance of infection, or early fungicidal activity, of oral versus i.v. flucytosine was determined from quantitative cultures of CSF at baseline and days 3, 7, and 13 or 14 of treatment, using the slope of the linear regression of log CFU against time for each patient, as previously described (4). Negative CSF cultures were assigned a value of 1 CFU per ml. All data points were used, with the exception of sterile cultures at 14 days if this value reduced the slope. In these cases, CSF sterility would likely have been achieved at some time between 7 and 14 days, and use of the 14-day result would therefore lead to an underestimation of the true slope (4).

Clinical and laboratory toxicity. Patients were monitored daily during the initial 2 weeks of therapy for the development of clinical side effects. Peripheral blood samples were taken at admission and on days 3, 5, 7, 9, 11, and 13 for creatinine and electrolytes and at admission and on days 5, 9, and 13 for full blood count and liver function tests. Absolute values at day 13 and percent change at day 13 compared to baseline were taken as laboratory outcomes. If there was no day 13 value, the latest available follow-up value was used.

Pharmacokinetics. Blood for 5FC concentrations was taken on days 3 and 9 and on an average of three other days. Times of administration of 5FC and of the vena puncture were registered at the bedside of the patient. A mean of 5 samples per patient were frozen at −20°C, and 5FC and 5FU concentrations were determined simultaneously using reversed-phase high-performance liquid chromatography with UV detection at the Amsterdam Medical Centre after completion of the study (15). 5FC concentrations in CSF were determined at day 14 with a similar method at the Mycology Reference Laboratory, Bristol, United Kingdom. Both methods had between-run variation coefficients below 6% within the quantification range. Individual pharmacokinetic parameters were calculated using a Bayesian maximum a posteriori fitting procedure using the computer program MW/Pharm version 3.60 (Mediware, The Netherlands) (12, 18). The Bayesian fitting procedure uses measured drug concentrations and population-based pharmacokinetic parameters to determine individualized pharmacokinetic parameters of a patient (12). We used a one-compartment open model. The initial pharmacokinetic parameters of 5FC were taken from a previous study (18): elimination rate constant normalized to creatinine clearance (k_{elr}), 0.0009 ± 0.0005 min/ml · h; volume of distribution normalized to weight (V_d/kg), 0.899 ± 0.413 liters/kg; and oral absorption rate constant (k_a), 2 h^{-1}.

MIC. The MICs of 5FC for the isolates were determined at the Mycology Reference Laboratory, Bristol, United Kingdom, by means of the CLSI (formerly NCCLS) method M27–A2 (11). This method was modified as suggested in the document by the use of yeast nitrogen base as the basal medium to improve the clinical relevance of the antifungal MIC. Briefly, broth microdilution was performed in 96-well round-bottom microtiter plates containing 100 μl of doubling dilutions of 5FC to which were added 100-μl volumes of an inoculum suspension of 0.5×10^3 to 2.5×10^3 cells/ml in yeast nitrogen base. Plates were incubated at 35°C and read after 72 h, with an 80% growth inhibition defined as the end point.

Statistics. Baseline characteristics of groups were compared by the chi-square test for categorical variables and the Mann-Whitney U test or Student's t test for continuous variables. The Mann-Whitney U test and Wilcoxon matched pairs test were used for comparing laboratory outcomes. Given a comparison of five laboratory variables, a P value of 0.01 was considered significant (Bonferroni adjustment for multiple comparisons). Fisher's exact test and Mann-Whitney U test were used to compare pharmacokinetic parameters, and linear regression (4, 14) was used to compare fungicidal activity.

RESULTS

Sixty-four patients were enrolled. One patient, who was HIV-seronegative, was excluded. Thirty-one patients were assigned to 5FC treatment, with 16 assigned to the oral formulation and 15 assigned to to i.v. formulation. The baseline clinical characteristics of the patients have been previously reported (4). Baseline laboratory values are shown in Table 1. Values for patients in the different groups were similar, except

TABLE 1. Baseline clinical and laboratory characteristics

Parameter	Result for treatment group (n):				Result for 5FC formulation (n):		
	All (63)	No 5FC (32)	5FC (31)	P value	Oral (16)	i.v. (15)	P value
Gender, male (n [%])[a]	38 (60)	20 (63)	18 (58)	0.72	9 (56)	9 (60)	0.83
Age (yr)[a]	33 (29–36)	34 (29–38)	32 (30–34)	0.54	32 (30–34)	34 (29–36)	0.43
Weight (kg)[b]	47 (±8.4)	46 (±7.4)	49 (±9.2)	0.15	49 (±8.4)	49 (±10.3)	0.91
Altered mental status (n [%])[a]	12 (19)	7 (22)	5 (16)	0.56	3 (19)	2 (13)	0.68
CSF cryptococcal antigen titer[a]	768 (256–2,048)	512 (256–1,024)	1,024 (256–2,048)	0.53	512 (256–2,048)	512 (128–2,048)	0.41
CSF quantitative culture[a]	455,000 (53,500–1,680,000)	455,000 (54,500–1,392,500)	485,000 (47,000–2,100,000)	0.92	605,000 (129,000–1,995,000)	360,000 (43,000–2,215,000)	0.85
Hemoglobin (g/dl)[a]	10.2 (9.1–11.8)	9.8 (8.5–11.6)	10.9 (9.7–12.2)	0.14	10.6 (9.7–11.6)	11.2 (9.3–12.3)	0.75
Neutrophils (10^9/liter)[a]	4.4 (3.1–5.6)	3.6 (2.6–4.7)	4.7 (3.6–6.1)	0.008	4.5 (3.2–5.8)	5.6 (3.6–6.4)	0.14
Platelets (10^9/liter)[a]	235 (157–300)	226 (155–288)	253 (169–322)	0.30	263 (176–321)	249 (155–322)	0.48
ALT (U/liter)[a]	42 (28–54)	36 (25–53)	42 (33–57)	0.31	43 (36–52)	38 (25–63)	0.44
AST (U/liter)[a]	37 (26–59)	31 (26–59)	38 (25–63)	0.59	38 (34–61)	33 (22–75)	0.53
CrClear[c] (ml/min)[a]	72 (61–82)	69 (58–85)	74 (64–82)	0.39	75 (68–83)	74 (60–82)	0.42

[a] Median with IQR in parentheses.
[b] Mean with ±SD in parentheses.
[c] CrClear (creatinine clearance) = (140 − age in years) × (weight in kg)/(72 × serum creatinine in mg/dl); for women, multiply by 0.85.

5FC

EFA: -0.46 (SD 0.18) log CFU per day (n=27)

no 5FC

EFA: -0.34 (SD 0.17) log CFU per day (n=25)

Oral 5 FC

EFA: -0.48 (SD 0.22) log CFU per day (n=13)

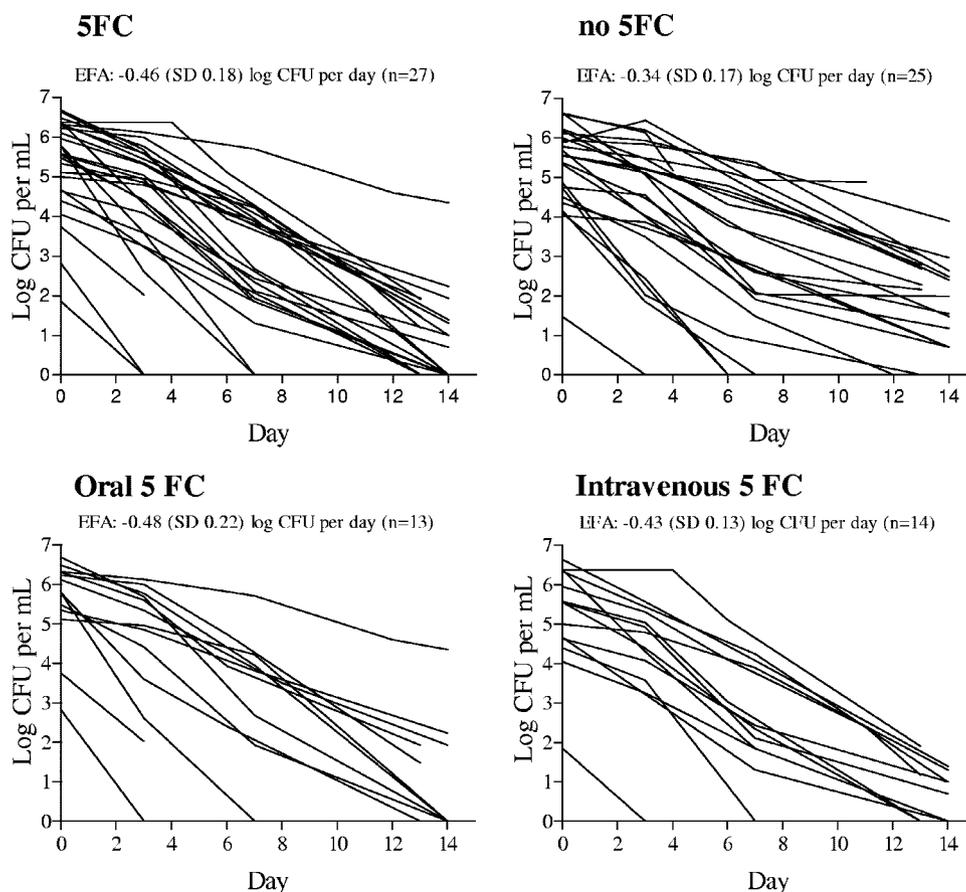**Intravenous 5 FC**

EFA: -0.43 (SD 0.13) log CFU per day (n=14)

FIG. 1. Early fungicidal activity (EFA).

for the fact that the baseline median neutrophil count was lower in patients not receiving 5FC (3.6×10^9/liter; interquartile range [IQR], 2.6 to 4.7) compared with those assigned to 5FC (4.7×10^9/liter; IQR, 3.6 to 6.1; $P = 0.008$, Mann-Whitney U test) (Table 1).

Early fungicidal activity was significantly greater in patients treated with 5FC than in patients not treated with 5FC (-0.46 [standard deviation {SD}, 0.18] versus -0.34 [SD, 0.17] log CFU/day, respectively; $P = 0.02$, linear regression) (Fig. 1). There was no difference in early fungicidal activity in patients on i.v. compared to patients on oral 5FC (-0.43 [SD, 0.13] versus -0.48 [SD, 0.22] log CFU/day, respectively; $P = 0.45$, linear regression). There was still no difference if linear regression was used to adjust for the effect of host immunity (baseline CSF gamma interferon concentrations) (14) on the rate of clearance of infection. There was no difference in mortality comparing patients on 5FC with patients not on 5FC and comparing patients on i.v. 5FC with patients on oral 5FC.

All treatment arms were well tolerated, and no drug treatment had to be withdrawn within the first 2 weeks because of clinical or laboratory side effects. Nine of 63 patients (14%) died during the first week of treatment, and one patient left the hospital on day 2, so that samples from the end of initial therapy were available from 53 (84%) patients, 25 in the non-5FC arms and 28 in the 5FC arms. There was no severe bone marrow depression (neutrophils $< 0.5 \times 10^9$/liter or plate-

lets $< 50 \times 10^9$/liter) and no clinically significant rise in liver function tests ($>$5-fold above normal).

Taking all patients, hemoglobin levels and calculated creatinine clearance decreased significantly over the initial 2 weeks of treatment ($P < 0.0001$, Wilcoxon matched pairs test). The magnitude of the reductions was not affected by treatment with 5FC or mode of administration of 5FC. There was no significant difference in any laboratory parameters, as a percentage of the baseline (Table 2) or in absolute values (data not shown), comparing patients on 5FC with patients not on 5FC and comparing patients on i.v. 5FC with patients on oral 5FC.

5FC concentrations were measured on an average of 5 occasions per patient in 28 patients on 5FC treatment. Pharmocokinetic parameters were calculated. The median oral clearance (CL/F) was 4.54 liters/h (IQR, 3.21 to 6.79 liters/h), the median volume of distribution normalized to weight (V_d/kg) was 1.16 liters/kg (IQR, 0.74 to 1.67 liters/kg), and the oral absorption rate constant (k_a) was 1.72 h^{-1} (IQR, 0.21 to 2.67 h^{-1}). The median 24-h area under the concentration-time curve (AUC$_{24}$) during steady state was significantly higher in patients on i.v. 5FC than in patients on oral 5FC (at day 7, 1,289 [IQR, 721 to 1,637] versus 576 [IQR, 455 to 847] mg · h/liter; $P = 0.002$, Mann-Whitney U test) (Table 3). There was no correlation between AUC$_{24}$ and either absolute values or percent change from the baseline in any of the laboratory variables at the end of 2 weeks of treatment (Spearman's

TABLE 2. Laboratory values after 2 weeks of induction therapy, as percentage of baseline[a]

Parameter	Last value for treatment group (n):			P value	Last value for 5FC formulation (n):		P value
	All (53)	No 5FC (25)	5FC (28)		Oral (14)	i.v. (14)	
Hemoglobin	77 (70–87)	79 (71–92)	74 (67–83)	0.24	70 (61–82)	78 (72–85)	0.12
Neutrophils	78 (59–124)	80 (65–124)	66 (50–122)	0.12	73 (55–135)	59 (44–92)	0.41
Platelets	91 (67–113)	76 (67–109)	97 (71–125)	0.22	112 (80–133)	83 (71–106)	0.09
ALT	111 (68–158)	127 (106–168)	92 (67–116)	0.02	94 (42–114)	86 (67–118)	0.82
AST	84 (52–138)	97 (60–138)	74 (51–135)	0.36	69 (47–119)	100 (60–150)	0.13
CrClear[b]	80 (60–100)	84 (60–100)	77 (60–100)	0.67	83 (62–100)	76 (59–100)	0.68

[a] All values are the median percentages of baseline values with IQR indicated in parentheses.

[b] CrClear (creatinine clearance) = $(140 - \text{age in years}) \times (\text{weight in kg})/(72 \times \text{serum creatinine in mg/dl})$; for women, multiply by 0.85.

correlation, $P > 0.05$). In 5 patients, all on i.v. treatment, peak plasma 5FC concentrations above 100 mg/liter were measured on at least one occasion. No patients on oral 5FC had concentrations above 75 mg/liter at any time. High concentrations were not associated with bone marrow toxicity or hepatotoxicity (results not shown). 5FU was detected in 4 patients (0.50, 0.97, 1.25, and 1.28 mg/liter), 3 on oral 5FC and 1 on i.v. 5FC treatment. There was no association between the presence of 5FU and hemoglobin, neutrophil, or platelet count or ALT or AST (aspartate aminotransferase) at the end of 2 weeks of treatment.

The 5FC MIC ranged from 1 to 8 mg/liter. In patients treated with 5FC, there was no significant correlation of 5FC MIC, area under the concentration-time curve (AUC), AUC/MIC ratio, or time above MIC (which was 100% for all except one patient), with the rate of clearance of infection.

CSF 5FC concentrations at the end of initial therapy were 84% of corresponding plasma concentrations (median IQR, 57 to 106; $n = 19$). Again, no association was found between CSF concentrations and rate of clearance of infection from the CSF.

DISCUSSION

In this study, 5FC was safe at 100 mg/kg/day for 2 weeks in patients with cryptococcal meningitis. The results support the safety of 5FC, at this dosage and without drug concentration monitoring, seen in the last Mycoses Study Group trial (16) and extend the findings to a developing country setting. While

5FC remains a part of our most rapidly fungicidal combination for initial therapy, the results argue for more widespread access to 5FC in Southeast Asia and Africa (2), areas where increasing availability of antiretroviral drugs now offers patients with HIV-associated cryptococcal meningitis the prospect of a good long-term prognosis, provided they survive the acute cryptococcal infection.

Despite significantly lower concentrations of 5FC in patients on oral 5FC, 5FU was detected more frequently in patients on oral compared with the i.v. formulation. The data are compatible with the hypothesis that intestinal microflora plays some role in the conversion of 5FC to 5FU. However, at these doses, any conversion of 5FC to 5FU was not associated with detectable toxicity.

CSF concentrations were 84% of corresponding plasma concentrations, in agreement with prior data (3). Plasma concentrations were significantly higher in patients given the i.v. formulation than in patients given the oral formulation. Absorption and bioavailability of oral 5FC has generally been high (75 to 90%) in normal volunteers and patients in North America (3, 5). The data suggest that absorption is not so complete in patients with late-stage HIV disease in Thailand, with the ratio of AUC for oral and i.v. formulations suggesting an oral bioavailability of only 45%.

Despite the difference in 5FC plasma concentrations, there was no difference in mycological efficacy, as accurately assessed by serial quantitative cultures. The results support in vitro and animal model work suggesting that 5FC has concentration-

TABLE 3. Plasma pharmacokinetic parameters

Parameter[a]	Result for 5FC formulation:		P value
	Oral (n = 14)	i.v. (n = 14)	
Calculated peak (mg/liter)[b]	30 (23–35)	63 (41–82)	<0.00001[d]
Calculated trough (mg/liter)[b]	20 (13–28)	37 (17–57)	0.004[d]
Clearance/F (liter/h)[b]	6.7 (4.8–8.3)	3.8 (2.7–4.3)	0.006[d]
k_{elr} (min/ml/h)[b]	0.0017 (0.0008–0.022)	0.0014 (0.0009–0.0024)	0.89[d]
V_d/kg (liter/kg)[b]	1.41 (0.85–2.23)	0.94 (0.60–1.18)	0.07[d]
5FC AUC_{24} (mg · h/liter)[b]	576 (455–847)	1,289 (721–1,637)	0.002[d]
5FC > 100 mg/liter[c]	0	5	0.04[e]
5FC > 75 mg/liter[c]	0	8	0.002[e]
Any 5FU[c]	3	1	0.58[e]

[a] Peak and trough concentrations were calculated on days 3, 7, and 13. The AUC_{24} was determined on day 7.

[b] Results are medians with IQR indicated in parentheses.

[c] Results are numbers.

[d] Mann-Whitney U test.

[e] Fisher exact test.

independent pharmacodynamics (1, 7, 9). Plasma concentrations between 20 and 30 mg/liter, as seen in patients on the oral formulation, were not associated with any reduction in fungicidal activity. The facts that maximal fungicidal activity, additional to the effect of AMB, was achieved, and that the time above MIC was close to 100% with this dosage of 5FC, irrespective of mode of administration, may explain why we could not find an association between any pharmacokinetic parameter and rate of clearance of infection and between the narrow range of MICs and rate of clearance of infection. With regard to the MIC, it is of interest that Schwartz and colleagues have shown additive effects with 5FC given with AMB in a murine model, even when the isolate was resistant to 5FC (MIC = 64 μg/ml) (13).

The results suggest that with i.v. formulation or in situations where absorption of oral formulation is more complete, a 5FC dose of 75 mg/kg/day, or even lower, given with AMB, may be associated with maximal additional fungicidal activity.

ACKNOWLEDGMENTS

REFERENCES

1. **Andes, D., and M. van Ogtrop.** 2000. In vivo characterization of the pharmacodynamics of flucytosine in a neutropenic murine disseminated candidiasis model. Antimicrob. Agents Chemother. **44:**938–942.
2. **Bicanic, T., and T. S. Harrison.** 2004. Cryptococcal meningitis. Br. Med. Bull. **72:**99–118.
3. **Block, E. R., and J. E. Bennett.** 1972. Pharmacological studies with 5-fluorocytosine. Antimicrob. Agents Chemother. **1:**476–482.
4. **Brouwer, A. E., A. Rajanuwong, W. Chierakul, G. E. Griffin, R. A. Larsen, N. J. White, and T. S. Harrison.** 2004. Combination antifungal therapies for HIV-associated cryptococcal meningitis: a randomised trial. Lancet **363:**1764–1767.
5. **Cutler, R. E., A. D. Blair, and M. R. Kelly.** 1978. Flucytosine kinetics in subjects with normal and impaired renal function. Clin. Pharmacol. Ther. **24:**333–342.
6. **Diasio, R. B., D. E. Lakings, and J. E. Bennett.** 1978. Evidence for conversion of 5-fluorocytosine to 5-fluorouracil in humans: possible factor in 5-fluorocytosine clinical toxicity. Antimicrob. Agents Chemother. **14:**903–908.
7. **Francis, P., and T. J. Walsh.** 1992. Evolving role of flucytosine in immunocompromised patients: new insights into safety, pharmacokinetics, and antifungal therapy. Clin. Infect. Dis. **15:**1003–1018.
8. **Harris, B. E., B. W. Manning, T. W. Federle, and R. B. Diasio.** 1986. Conversion of 5-fluorocytosine to 5-fluorouracil by human intestinal microflora. Antimicrob. Agents Chemother. **29:**44–48.
9. **Lewis, R. E., M. E. Klepser, and M. A. Pfaller.** 2000. In vitro pharmacodynamic characteristics of flucytosine determined by time-kill methods. Diagn. Microbiol. Infect. Dis. **36:**101–105.
10. **Malet-Martino, M. C., R. Martino, M. de Forni, A. Andremont, O. Hartmann, and J. P. Armand.** 1991. Flucytosine conversion to fluorouracil in humans: does a correlation with gut flora status exist? A report of two cases using fluorine-19 magnetic resonance spectroscopy. Infection **19:**178–180.
11. **National Committee for Clinical Laboratory Standards.** 2002. Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard, 2nd ed. NCCLS document M27–A2. National Committee for Clinical Laboratory Standards, Wayne, PA.
12. **Proost, J. H., and D. K. Meijer.** 1992. MW/Pharm, an integrated software package for drug dosage regimen calculation and therapeutic drug monitoring. Comput. Biol. Med. **22:**155–163.
13. **Schwarz, P., F. Dromer, O. Lortholary, and E. Dannaoui.** 2006. Efficacy of amphotericin B in combination with flucytosine against flucytosine-susceptible or flucytosine-resistant isolates of *Cryptococcus neoformans* during disseminated murine cryptococcosis. Antimicrob. Agents Chemother. **50:**113–120.
14. **Siddiqui, A. A., A. E. Brouwer, V. Wuthiekanun, S. Jaffar, R. Shattock, D. Irving, J. Sheldon, W. Chierakul, S. Peacock, N. Day, N. J. White, and T. S. Harrison.** 2005. IFN-gamma at the site of infection determines rate of clearance of infection in cryptococcal meningitis. J. Immunol. **174:**1746–1750.
15. **Toraño, J. S., A. Vermes, and H. J. Guchelaar.** 2001. Simultaneous determination of flucytosine and fluorouracil in human plasma by high-performance liquid chromatography. Biomed. Chromatogr. **15:**89–94.
16. **van der Horst, C. M., M. S. Saag, G. A. Cloud, R. J. Hamill, J. R. Graybill, J. D. Sobel, P. C. Johnson, C. U. Tuazon, T. Kerkering, B. L. Moskovitz, W. G. Powderly, and W. E. Dismukes.** 1997. Treatment of cryptococcal meningitis associated with the acquired immunodeficiency syndrome. National Institute of Allergy and Infectious Diseases Mycoses Study Group and AIDS Clinical Trials Group. N. Engl. J. Med. **337:**15–21.
17. **Vermes, A., E. J. Kuijper, H. J. Guchelaar, and J. Dankert.** 2003. An in vitro study on the active conversion of flucytosine to fluorouracil by microorganisms in the human intestinal microflora. Chemotherapy **49:**17–23.
18. **Vermes, A., R. A. Mathôt, H. van der Sijs, J. Dankert, and H. J. Guchelaar.** 2000. Population pharmacokinetics of flucytosine: comparison and validation of three models using STS, NPEM, and NONMEM. Ther. Drug Monit. **22:**676–687.
19. **Waldorf, A. R., and A. Polak.** 1983. Mechanisms of action of 5-fluorocytosine. Antimicrob. Agents Chemother. **23:**79–85.