Identification of BSPRY as a Novel Auxiliary Protein Inhibiting TRPV5 Activity

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Transient receptor potential vallinoid 5 (TRPV5) and TRPV6 are the most Ca\(^{2+}\)-selective members of the TRP superfamily and are essential for active Ca\(^{2+}\) reabsorption in epithelia. However, little is known about intracellular proteins that regulate the activity of these channels. This study identified BSPRY (B-box and SPRY-domain containing protein) as a novel factor involved in the control of TRPV5. The interaction between BSPRY and TRPV5 by GST pull-down and co-immunoprecipitation assays was demonstrated. BSPRY showed co-localization with TRPV5 in mouse kidney. Expression of BSPRY resulted in a significant reduction of the Ca\(^{2+}\) influx in Madin-Darby Canine Kidney cells that stably express TRPV5 without affecting channel cell-surface abundance. Finally, BSPRY expression in kidney was increased in 25-hydroxyvitamin D\(_3\) knockout mice, suggesting an inverse regulation by vitamin D\(_3\). Together, these results demonstrate the physiologic role of the novel protein BSPRY in the regulation of epithelial Ca\(^{2+}\) transport via negative modulation of TRPV5 activity.


Materials and Methods

DNA Constructs and cRNA Synthesis

Full-length BSPRY was obtained from mouse expressed sequence tag (EST) L.M.A.G.E. 5116439, cloned into the pTLN oocyte expression vector (5) in frame with an amino-terminal VSV tag and subcloned into pGEX6p2 (Amersham Biosciences, Uppsala, Sweden) and pCB7 (6). Full-length and carboxyl-termini of TRPV5 and TRPV6 were obtained as described previously (7).

Yeast Two-Hybrid System

Y153 yeast strain transformation and subsequently screening of a mouse kidney cDNA library (Clontech, Palo Alto, CA) was performed as described previously (7).

Protein–Protein Interaction Assays

Glutathione S-transferase (GST) fusion proteins were expressed in Escherichia coli BL21 and purified according to the manufacturer’s protocol (Amersham Biosciences). BSPRY cRNA was synthesized in vitro using SP\(_6\) RNA polymerase as described previously (7) and injected into Xenopus laevis oocytes. After 2 d, oocytes were lysed in PBS that contained 0.4% (vol/vol) Triton X-100. \(^{35}\)S]Methionine-labeled full-length TRPV5/TRPV6 protein was prepared using a reticulocyte lysate system. Xenopus oocyte lysates or in vitro–translated proteins were incubated with GST or GST-fusion proteins that were immobilized on glutathione-Sepharose 4B beads (Amersham Biosciences) in PBS that contained 0.4% (vol/vol) Triton X-100 for 2 h at room temperature, and bound proteins were visualized by autoradiography or immunoblotting. MDCK cells that were stably transfected with green fluorescent protein (GFP)-fused TRPV5 and VSV-BSPRY were lysed in sucrose buffer that fused contained 20 mM Tris (pH 7.4), 5 mM EDTA, 135 mM NaCl, 0.5% (vol/vol) NP-40, 0.2% (vol/vol) Triton X-100, and 10% (wt/vol) sucrose, incubated on ice for 60 min, and centrifuged for 30 min at 16,000 × g. Supernatants were incubated with guinea pig anti-TRPV5 or monoclonal anti-VSV antibodies (Sigma, St. Louis, MO).
showed complete co-localization with TRPV5 in Ca2+ down and co-immunoprecipitation assays. Second, BSPRY observations. First, the specific interaction between this channel TRPV5. This conclusion is based on four independent ob-

Reverse Transcription–PCR Analysis
Total RNA was isolated using TRIzol (Life Technologies/BRL, Life Technologies, Breda, The Netherlands). Total RNA (2 μg) was sub-

Preparation of Antibodies and Immunohistochemistry
Rabbit anti-BSPRY antibodies were raised against the peptide H2N-LFVPVFADQVLSSV-COOH of mouse BSPRY and used for immuno-

Cell-Surface Biotinylation
Proteins that were present at the cell surface of confluent stably transfected MDCK cells were biotinylated at 4°C using NHS-LC-LC-

Statistical Analyses
In all experiments, the data are expressed as mean ± SEM. Overall statistical significance was determined by ANOVA. P < 0.05 was consid-

Results and Discussion
Our study provides the first evidence of the physiologic role of BSPRY as a negative modulator of the epithelial Ca2+ channel TRPV5. This conclusion is based on four independent ob-

Identification of BSPRY as a Novel TRPV5- and TRPV6-Associated Protein
To discover novel TRPV5- and TRPV6-associated proteins, we performed a yeast two-hybrid screen on a mouse kidney cDNA library. Using the mouse TRPV6 carboxyl-terminus as bait, five clones that were identified as BSPRY were isolated. BSPRY contains a B-box and SPRY domain, whose tentative functions are protein–protein interaction modules (8,9). The association of full-

Co-Localization of BSPRY and TRPV5
The expression of BSPRY in mouse tissues was analyzed by RT-PCR. BSPRY was detected in several tissues, including kidney, small intestine, prostate, lung, and uterus. BSPRY was less abundant in heart, whereas skeletal muscle and liver were negative (Figure 2A, top). All samples expressed β-actin, which was used as a positive control to confirm cDNA integrity (Figure 2A, bottom). Using anti-BSPRY antibodies, raised in rabbits using a conserved 15-amino acid peptide of the carboxyl-terminus of mouse BSPRY, we demonstrated that expression was inversely regulated by the calciotropic hormone vitamin D₃.
the localization of BSPRY in kidney is strikingly similar to the localization of TRPV5 (Figure 2B, bottom). BSPRY was present in the apical domain of all TRPV5-immunopositive tubules, previously identified as the second part of the distal convoluted tubule and connecting tubule (11,12). Proximal tubules and glomeruli were negative, and preimmune serum did not show any staining. Furthermore, immunopositive BSPRY staining was absent upon incubation of the serum with immobilized GST-BSPRY, whereas incubation with GST alone had no effect (Figure 2B, top). Similar co-localization with TRPV5 in distal convoluted tubule and connecting tubule was observed previously for calbindin-D28K and the Na+/H+ exchanger (12,13). These proteins play an essential role in renal transcellular Ca2+ transport as they respectively facilitate the diffusion of Ca2+ from the apical to the basolateral side and the extrusion into the blood. Similarly, the robust co-localization between BSPRY and TRPV5 strongly supports a physiologic and specific function of BSPRY in the regulation of epithelial Ca2+ transport by direct association with TRPV5 in the kidney.
BSPRY-Mediated TRPV5 Inhibition of Ca\textsuperscript{2+} Influx

The effect of BSPRY co-expression on TRPV5-mediated Ca\textsuperscript{2+} influx was measured using confluent monolayers of MDCK cells. TRPV5 expression in these cells resulted in an approximately 2.5-fold increased Ca\textsuperscript{2+} influx compared with nontransfected cells. Upon co-expression with BSPRY, Ca\textsuperscript{2+} influx was inhibited by 41 ± 13% (Figure 3C), approaching levels of nontransfected cells. Total cellular TRPV5 expression levels were identical in the presence or absence of BSPRY, as was demonstrated by immunoblot analysis using anti-GFP antibodies (Figure 3, A and B). Furthermore, the correct size of the bands at approximately 95 to 115 kD in Figure 3A confirmed the integrity of the GFP-TRPV5 fusion protein. Finally, flow cytometry analysis of the GFP signal showed that >98% of the MDCK cells forming the confluent monolayer contributed to the TRPV5-dependent Ca\textsuperscript{2+} influx, both in the presence and in the absence of BSPRY (data not shown). These data provide the first evidence of a functional role of BSPRY. So far, only two studies have provided information about BSPRY. First, BSPRY was identified recently in a yeast two-hybrid screen using zyxin as bait. In epithelial cells, zyxin is involved in the formation of cell–cell contacts, which require actin cytoskeleton rearrangements (14). Schenker et al. (15) provided no data about the localization and function of BSPRY. However, the association with zyxin might hint at a role of the cytoskeleton in the BSPRY-mediated regulation of TRPV5. Second, it was shown that BSPRY interacts with 14-3-3 proteins. Although the exact role of 14-3-3 proteins remains to be elucidated fully, their role in the recognition of phosphorylated proteins is well established (16). It has been demonstrated that 14-3-3 proteins bind to specific motifs that contain a phosphorylated serine residue and have been implicated in the binding to and activation of signaling proteins (16,17). Furthermore, a role of 14-3-3 proteins in K\textsuperscript{+} channel trafficking was postulated (18). However, cell-surface biotinylation did not provide evidence for TRPV5 trafficking as an explanation for the observed inhibitory function of BSPRY (Figure 3D). Furthermore, we demonstrated that a significant fraction of BSPRY is membrane associated and present at TRPV5 trafficking as an explanation for the observed inhibitory function of BSPRY (Figure 3D). Furthermore, we demonstrated that a significant fraction of BSPRY is membrane associated and present at the plasma membrane (Figure 3, E and F, Supplemental Figure). Therefore, it is tempting to speculate that BSPRY is involved in inhibitory signaling cascades that control the activity of the epithelial Ca\textsuperscript{2+} channels at the cell surface.

Vitamin D–Dependent Regulation of BSPRY Expression in Kidney

It was demonstrated previously that TRPV5 expression is strongly regulated by 1,25-dihydroxyvitamin D\textsubscript{3}, the biologic active form of vitamin D (4,19). Therefore, we assessed the expression of BSPRY in wild-type and 25-hydroxyvitamin D\textsubscript{3}-1\textalpha-hydroxylase (1\textalpha-OHase) knockout mice. These knockout mice are unable to synthesize vitamin D\textsubscript{3} and therefore exhibit reduced expression of TRPV5 (2,19). In wild-type and 25-hydroxyvitamin D\textsubscript{3}-1\textalpha-hydroxylase (1\textalpha-OHase) knockout mice, BSPRY expression was determined by quantitative real-time PCR in kidneys of wild-type (WT; n = 11) or 25-hydroxyvitamin D\textsubscript{3}-1\textalpha-hydroxylase (1\textalpha-OHase) knockout (KO; n = 6) mice. (H) Effect of vitamin D\textsubscript{3} on BSPRY protein abundance in mouse kidney. Representative images showing kidney sections stained for BSPRY in WT or 1\textalpha-OHase KO mice. Protein abundance was determined by computerized analysis of immunohistochemical images and is presented as mean optical density (arbitrary units) for n = 24 to 30 pictures from five mice for each condition. *P < 0.05 versus all. (G) The role of vitamin D\textsubscript{3} on BSPRY expression was determined by quantitative real-time PCR in kidneys of wild-type (WT; n = 11) or 25-hydroxyvitamin D\textsubscript{3}-1\textalpha-hydroxylase (1\textalpha-OHase) knockout (KO; n = 6) mice.
the 1,25-dihydroxyvitamin D3 and are a valuable animal model to study vitamin D deficiency rickets type I (20). Quantitative real-time PCR showed significantly enhanced BSPRY mRNA expression in the 1α-OHase knockout mice compared with wild-type mice, suggesting that vitamin D negatively regulates the BSPRY expression (Figure 3G). Because the antibodies that we generated did not recognize BSPRY on immunoblot, the protein expression of BSPRY in these animals was semiquantified by immunohistochemistry. Depicted in Figure 3H are representative images of BSPRY expression in wild-type and 1α-OHase knockout mice. Computerized analysis of the immunohistochemical staining showed significantly enhanced BSPRY protein expression in the 1α-OHase knockout mice compared with wild-type mice, confirming the inverse regulation of BSPRY expression by circulating vitamin D on the protein level (Figure 3H). Together with the Ca²⁺ uptake results, this suggests that BSPRY operates as a negative modulator for TRPV5 and that this mechanism will be downregulated when vitamin D levels increase to stimulate active Ca²⁺ transport. These findings demonstrate a novel target for vitamin D₃ in the regulation of active Ca²⁺ transport and provide new insight into the factors involved in the Ca²⁺-related (patho-)physiology.

Conclusion

We have identified BSPRY as a novel auxiliary protein of the epithelial Ca²⁺ channels. The association, co-localization, and functional analyses described in this study demonstrate the first physiologic role of the novel protein BSPRY in the direct regulation of TRPV5.

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References


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