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Apolipoprotein E Genotype Regulates Amyloid- β Cytotoxicity

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The $\epsilon 4$ allele of apolipoprotein E (ApoE) is a risk factor for Alzheimer's disease (AD), whereas the $\epsilon 2$ allele may be relatively protective. Both alleles are risk factors for cerebral amyloid angiopathy (CAA)-related hemorrhages. CAA is associated with degeneration of smooth muscle cells and pericytes. Previously, we described that synthetic amyloid- β_{1-40} peptide ($A\beta_{1-40}$) with the ²²Glu→Gln "Dutch" mutation caused pericyte death *in vitro* by a mechanism that involves $A\beta$ fibril-like assembly at the cell surface. It is known that ApoE binds to $A\beta$ and may modify its biological activities. In the present study, we evaluated the effect of ApoE on $A\beta$ -mediated toxicity of cerebrovascular cells. We observed that cultured cells with an $\epsilon 4/\epsilon 4$ genotype were more vulnerable to $A\beta$ than cultures with an $\epsilon 3/\epsilon 3$ or $\epsilon 3/\epsilon 4$ genotype. The one cell culture with the $\epsilon 2/\epsilon 3$ genotype was relatively resistant to $A\beta$ compared with other cultures. Furthermore, we observed a dose-dependent protective effect of native ApoE against $A\beta$ -mediated toxicity of cerebrovascular cells and, in addition, ApoE $\epsilon 2/\epsilon 3$ cells secreted more ApoE protein compared with cells with other ApoE genotypes, in particular, compared with $\epsilon 4/\epsilon 4$ cells. Thus, the disparity between ApoE genotype and $A\beta$ -mediated toxicity might be related to differences in the cellular capacity to secrete ApoE. The present data suggest that one mechanism by which ApoE may alter the risk for AD is a genotype-dependent regulation of $A\beta$ cytotoxicity, possibly via variations in its secretion levels, whereby extracellular ApoE may bind to $A\beta$ and thereby modify $A\beta$ -mediated cell death.

Key words: apolipoprotein E; Alzheimer's disease; amyloid- β protein; cerebrovascular cells; CAA; cytotoxicity

Introduction

Senile plaques (SPs) and cerebral amyloid angiopathy (CAA) are two of the neuropathological hallmarks of Alzheimer's disease (AD). The amyloid- β protein ($A\beta$), which consists of 40–42 amino acids, is the major constituent of both SPs and CAA. Apolipoprotein E (ApoE) has been identified as a major risk factor for the sporadic late-onset form of AD. The *ApoE* gene has three alleles ($\epsilon 2$, $\epsilon 3$, and $\epsilon 4$), the most frequent of which is the $\epsilon 3$ allele. Inheritance of one or two copies of the $\epsilon 4$ allele is associated with a dose-dependent increased risk for AD and an earlier age of onset of AD (Corder et al., 1993; Saunders et al., 1993), although individuals homozygous for $\epsilon 4$ may reach a high age without cognitive impairment (Henderson et al., 1995). On the contrary, the $\epsilon 2$ allele seems to have a protective effect against AD (Corder et al., 1994). Surprisingly, however, both the $\epsilon 2$ and the $\epsilon 4$ alleles

are associated with an increased risk for CAA or CAA-related hemorrhages (Greenberg et al., 1996, 1998; Premkumar et al., 1996; Nicoll et al., 1997).

Inheritance of one or two copies of the ApoE $\epsilon 4$ allele is associated with a higher load of SPs and CAA (Schmechel et al., 1993; Premkumar et al., 1996). *In vitro* studies have shown that ApoE is able to bind to $A\beta$ (Strittmatter et al., 1993; Pillot et al., 1999); native ApoE3 binds $A\beta$ more efficiently than ApoE4 (LaDu et al., 1994). ApoE may also accelerate the formation of $A\beta$ fibrils (Strittmatter et al., 1993; Sanan et al., 1994; Wisniewski et al., 1994). In mice transgenic for human amyloid precursor protein (APP) and deficient in ApoE, the amount of amyloid, but not that of $A\beta$ immunoreactivity, was strongly reduced compared with transgenic APP/ApoE^{+/+} mice (Bales et al., 1999), suggesting that ApoE plays a critical role in facilitating $A\beta$ deposition. In contrast to these studies, early $A\beta$ deposition is suppressed in mice transgenic for both APP and human ApoE on an ApoE knock-out background (Holtzman et al., 1999). These results show that the role of ApoE and its specific isoforms in $A\beta$ fibrillogenesis, deposition, or clearance still remains unclear.

Previously, we described the toxicity of various $A\beta$ peptides toward cultures of human brain cerebrovascular smooth muscle cells (SMCs) and human brain pericytes (HBPs) (Davis-Salinas et al., 1995; Davis and Van Nostrand, 1996; Verbeek et al., 1997). We demonstrated that nonaggregated wild-type $A\beta_{1-42}$ and $A\beta_{1-40}$ carrying the Glu→Gln mutation at position 22 of $A\beta$, as

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Table 1. Overview of the source of HBP and SMC cell cultures

Number	Diagnosis	Grade (Braak)	Grade (CERAD)	CAA grade	Cell type	PMI (h)	ApoE	Age	Gender	Cause of death	Vascular risk factors
1	Control	II	++	—	HBP/SMC	5	2/3	84	F	Cardiac failure	Atherosclerosis
2	AD	VI	++++	—	HBP	4	3/3	94	F	Respiratory insufficiency	Atherosclerosis
3	AD	VI	++++	+++	HBP	4	3/3	81	F	Cardiac failure	None
4	AD	VI	++++	+++	HBP/SMC	3	3/3	75	M	Pneumonia	Atherosclerosis
5	Control	0	0	—	HBP/SMC	4	3/3	84	F	Pneumonia	Atherosclerosis
6	Control	0	0	—	SMC	6	3/3	71	F	Ruptured aortic aneurysm	Atherosclerosis
7	AD	V	+++	+	SMC	6	3/3	87	F	Bronchopneumonia	Atherosclerosis
8	AD/PD	V	+++	+++	SMC	4	3/4	91	F	Pneumonia	Atherosclerosis
9	AD	V	+++	—	HBP	5	3/4	84	M	Cardiac failure	Atherosclerosis
10	Control	III	++	—	HBP/SMC	4	3/4	83	F	Ruptured aortic aneurysm	Atherosclerosis
11	AD	VI	+++	++	HBP/SMC	3	3/4	89	F	Pneumonia	Atherosclerosis
12	AD	VI	++	+++	HBP	5	4/4	83	F	Sepsis	Atherosclerosis
13	AD	VI	+++	+++	HBP/SMC	3	4/4	69	M	Unknown	Atherosclerosis

F, Female; M, male; PD, Parkinson's disease; PMI, postmortem interval in hours. Grading of AD (Braak and CERAD scores) and CAA was performed as described in Materials and Methods.

in hereditary cerebral hemorrhage with amyloidosis-Dutch type, caused degeneration of cultured SMCs and HBPs. Either cell type secreted significant amounts of ApoE (Verbeek et al., 1999). Given the well described interactions between ApoE and A β , we investigated whether endogenously produced ApoE could interfere in an isoform-dependent manner with the A β -mediated cytotoxicity toward cultured human cerebrovascular cells. Therefore, in this study, we performed a comparative analysis of the cytotoxic effect of A β on cultures of HBPs and SMCs with different ApoE genotypes.

Materials and Methods

Reagents. A β with the "Dutch" mutation (22 Glu→Gln; D-A β ₁₋₄₀; 89% pure) was purchased from Biosource (Etten-Leur, The Netherlands). Human recombinant ApoE produced in baculovirus in Sf insect cells was obtained from PanVera (Madison, WI). ApoE obtained from the conditioned medium of transfected human embryonic kidney 293 (HEK293) cells was generously provided by Dr. G. W. Rebeck (Georgetown University, Washington, DC). Affinity-purified polyclonal anti-A β antibodies were prepared as described previously (Davis-Salinas et al., 1994). Monoclonal anti-ApoE antibody was purchased from Innogenetics (Antwerpen, Belgium). Anti-low-density lipoprotein receptor-related protein (LRP) (β -chain) antibody was obtained from American Diagnostica (Stamford, CT). Anti-A β antibody used for CAA grading was purchased from Dako (Glostrup, Denmark).

Cells. Pericytes were isolated from human brain tissue, and SMCs were isolated from leptomeningeal vessels from a number of AD patients and neurologically unaffected individuals, obtained after autopsy as described previously (Verbeek et al., 1994, 1995, 2002) (Table 1). Diagnosis and grading of AD patients was performed according to the criteria established by Braak and Braak (1991), Mirra et al. (1991), and The National Institute on Aging and Reagan Institute Working Group on Diagnostic Criteria for the Neuropathological Assessment of Alzheimer's Disease (1997). CAA grading was established by quantification of the number of A β -positive vessels in one microscopic field (magnification, 2.5 \times). At least four microscopic fields of both the occipital cortex and the gyrus temporalis medius were analyzed and categorized as follows: 0 (—, no CAA), 0–10 (+, sparse CAA), 10–20 (++, moderate CAA), and >20 (+++, severe CAA).

Both HBPs and SMCs were maintained in Eagle's modification of essential medium (EMEM) supplemented with 10% human serum (Sanquin Blood Bank, Nijmegen, The Netherlands), 20% newborn calf serum (Invitrogen, Paisley, UK), recombinant basic fibroblast growth factor (1 ng/ml), heparin (5 U/ml; Organon, Boxtel, The Netherlands), and antibiotics. Immunoelectron microscopical detection of A β accumulating at the surface of cultured cells was performed as described previously (Rensink et al., 2002, 2004). With respect to the various types of experiments, we did not observe differences between cerebrovascular cell cultures with

the same ApoE genotype but derived from different types of patients (i.e., AD or control patients).

Degeneration experiments. Triplicate wells with cultured cells were preincubated with serum-free medium (EMEM with 0.1% BSA and antibiotics) for 4 h. Subsequently, cells were incubated with fresh serum-free medium supplemented with synthetic A β peptides at 25 μ M for 6 d. Cells were routinely inspected and photographed using an Olympus (Tokyo, Japan) phase-contrast microscope. Cell viability was quantitated using a fluorescence live/dead cell assay according to the description of the manufacturer (Molecular Probes, Leiden, The Netherlands). The cultures were examined using an Olympus fluorescence microscope, and the percentage of dead cells was determined from counting in at least five microscopic fields per well. Based on previous studies (Davis and Van Nostrand, 1996; Verbeek et al., 1997), D-A β ₁₋₄₀ was used, because this peptide induced a robust degeneration of both pericytes and SMCs.

Western blot analysis. Cells were grown in serum-free medium, and both culture supernatant and cell lysates were collected to examine secreted and cell-associated expression of proteins, respectively. Culture supernatant was diluted 1:1 with reducing sample buffer. Cells were washed twice with PBS and then solubilized in the wells with lysis buffer [50 mM Tris-HCl, 150 mM NaCl, pH 7.5, 1% SDS, 5 mM EDTA, 500 μ M 4-(2-aminoethyl)benzenesulfonyl fluoride, 10 μ g/ml leupeptin, and 10 μ g/ml chymostatin] for 15 min. The protein content of diluted samples of the cell lysates was determined using the bichinonic acid method (Pierce, Rockford, IL). Equal protein amounts were loaded, fractionated on SDS-polyacrylamide gels, and subsequently electrophoretically transferred to nitrocellulose membranes (Schleicher and Schuell, Hertogenbosch, The Netherlands) in blotting buffer (25 mM Tris-HCl, pH 8.6, 192 mM glycine, and 20% methanol). Blots were washed for 15 min in PBS containing 0.05% Tween 20 (PBST), preincubated with blocking solution (5% lowfat milk powder in PBST), washed three times with PBST, and subsequently incubated with primary antibodies and peroxidase-labeled secondary rabbit anti-mouse antibodies (Dako). Detection was performed by chemiluminescence according to the description of the manufacturer (Boehringer Mannheim, Almere, The Netherlands) and exposure to Kodak (Rochester, NY) X-OMAT-R films.

ApoE genotyping. DNA was isolated from small pieces of brain tissue, from which cell cultures were also derived, using a DNA isolation kit (Biozym, Landgraaf, The Netherlands). ApoE genotype was determined using PCR and *HhaI* restriction analysis according to methods described previously (Hixson and Vernier, 1990; Wenham et al., 1991).

ApoE quantification. ApoE secretion by cultured HBPs and SMCs was quantitated both by scanning of Western blots and by ELISA. The chemiluminescent signal of both ApoE standards (PanVera, Madison, WI), cell lysates, and culture supernatants obtained after Western blotting was scanned with a LabScan (Amersham Biosciences, Aylesbury, UK). For ELISA, 5 μ g/ml affinity-purified goat anti-ApoE (Bioscience International, Saco, ME) was coated overnight at 4°C on a 96-well plate. Subsequently, wells were incubated with 1% nonfat milk in PBST to block

specific binding, with ApoE standards (PanVera, or from transfected HEK293 cells) or (diluted) culture supernatants, with affinity-purified biotinylated goat anti-ApoE (Bioscience International), and finally with peroxidase-labeled avidin (Dako). Tetramethylbenzidine was used as substrate, and H₂SO₄ was used to terminate the peroxidase reaction. No significant differences were observed with either type of standards; the use of ApoE3 as a standard only resulted in slightly higher signals than with ApoE4.

Partial purification of ApoE from culture supernatant. Purification of extracellular ApoE protein from medium of cultured HBP was performed as described by DeMattos et al. (1999) with minor modifications. Cells were incubated with serum-free EMEM medium for 6 d. Cells remained viable during this period. Supernatant was collected and passed through a D100 weakly basic anion exchange filter (Sartorius, Goettingen, Germany) and eluted with 1 M ammonium bicarbonate. The eluted ApoE was recirculated over a HiTrap heparin column (Amersham Biosciences). Fractions containing ApoE were pooled and dialyzed against PBS. Using the ApoE sandwich ELISA described above and comparing the ApoE concentration with the total protein amount, it was demonstrated that, in the purified fractions, 25% of the total protein amount consisted of ApoE.

RNA isolation and Northern blot analysis. Total RNA (8 μ g) was isolated from cultured cells by using the RNeasy kit (Qiagen, Hilden, Germany) according to the protocol of the manufacturer. Total RNA was treated with formaldehyde (Sambrook et al., 1989), separated on a 1.5% agarose gel, and blotted onto a Hybond N⁺ membrane (Amersham Biosciences). A 1100 bp fragment containing human ApoE3 cDNA (a kind gift from Dr. G. W. Rebeck, Georgetown University) was released from a pCMV4 expression vector by *Xba*I and *Kpn*I restriction cleavage. ApoE cDNA probe was radiolabeled by ³²dCTP incorporation using a random-primed DNA labeling kit (Roche Diagnostics, Penzberg, Germany). After preincubation in a hybridization mix (0.25 M sodium phosphate buffer, pH 7.2, 7% SDS, 1% BSA, 1 mM EDTA, and 0.1 mg/ml single-strand salmon sperm DNA) at 65°C, membranes were hybridized overnight with the radiolabeled probes under the same conditions. Blots were then washed at 58°C with 0.25 M sodium phosphate buffer, pH 7.2, 1% SDS, and 1 mM EDTA and autoradiographed using Kodak X-OMAT-S films.

Results

HBPs and SMCs were isolated from 10 and 9 donor brains, respectively. The characterization of HBPs and SMCs has been described extensively in previous studies (Verbeek et al., 1994, 2002). The cultured cells were divided into four different groups according to their ApoE genotype: ϵ 2/ ϵ 3, ϵ 3/ ϵ 3, ϵ 3/ ϵ 4, and ϵ 4/ ϵ 4 (Table 1).

After 4–6 d of treatment with 25 μ M, D-A β _{1–40} cultured cells (HBPs or SMCs) lost their characteristic polygonal shape, individual cell contours became less evident, and signs of cellular atrophy were observed (data not shown). Because ApoE is codeposited with amyloid in the vessel walls in CAA, we determined whether ApoE production by perivascular cells was affected by D-A β _{1–40} treatment as well. In the absence of A β treatment, both cultured SMCs and HBPs secreted considerable amounts of ApoE into the culture medium, as shown by Western blot analysis (Fig. 1). In contrast, smaller amounts of ApoE were associated with the cells. In contrast, incubation with D-A β _{1–40} resulted in an inversion of ApoE distribution, with most of the ApoE remaining associated with the cells. This was confirmed by double-immunofluorescence staining of D-A β _{1–40}-treated cells, showing that both A β and ApoE were coassociated on the cell surface (data not shown).

These experiments suggested that an interaction occurred between the endogenously produced ApoE and the exogenously administered A β that accumulated on the cell surface (Fig. 2). Therefore, we investigated whether the ApoE genotype of the

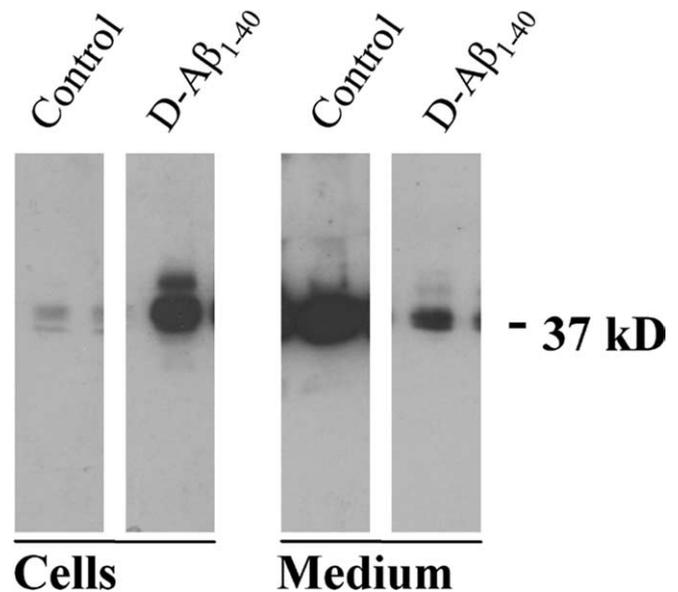


Figure 1. Western blot analysis of cell-associated (Cells) and secreted (Medium) ApoE in cultured human brain pericytes treated for 6 d with 25 μ M D-A β _{1–40}. In untreated cultures (Control), most ApoE (molecular weight, ~37 kDa) was present in the supernatant, whereas in D-A β _{1–40}-treated cells (D-A β _{1–40}), ApoE was predominantly associated with the cellular fraction.

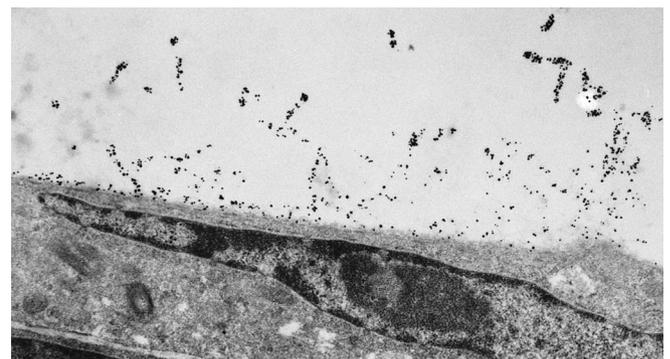


Figure 2. Immunoelectron microscopy analysis of HBP for localization of A β after 3 d of incubation with D-A β _{1–40} (25 μ M), showing decoration of A β fibrils extending from the cell surface. Magnification, 6000 \times .

cultured cells affected the toxicity of D-A β _{1–40}. The results showed that there were striking differences in the response to D-A β _{1–40} between cell cultures with a different ApoE genotype. After treatment of HBP cultures with the ϵ 4/ ϵ 4 genotype with D-A β _{1–40} for 6 d, the number of viable cells was strongly reduced (Fig. 3). In contrast, cells with either the ϵ 3/ ϵ 3 or the ϵ 3/ ϵ 4 genotype were consistently less susceptible to D-A β _{1–40} treatment. In addition, although only a single ϵ 2/ ϵ 3 HBP culture was available, we consistently observed ($n = 9$ experiments) that cell death was much lower (mean, 12 \pm 10% dead cells). In addition, similar results were obtained with SMC cultures of various ApoE genotypes (data not shown). Both HBPs and SMCs expressed the ApoE receptor LRP. The effects described above, however, could not be explained by differences in LRP expression of the cells, which was similar in the cell cultures with different ApoE genotypes (data not shown). Furthermore, the effects also could not be explained by differences in the clinicopathological state of the donors [Braak or The Consortium to Establish a Registry for Alzheimer's Disease (CERAD) grade of AD, CAA grade] (Table 1).

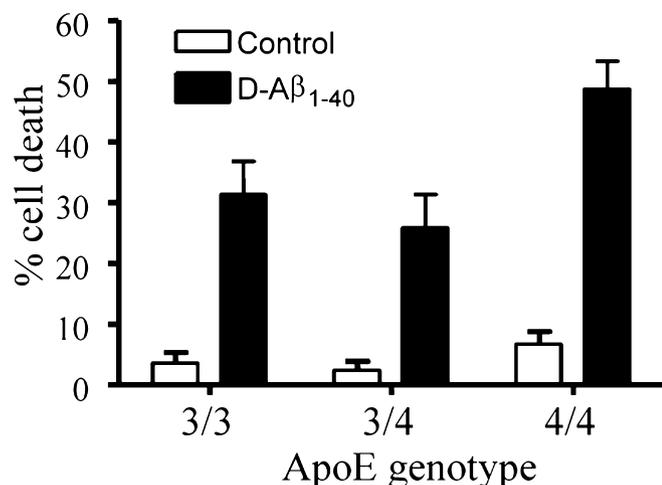


Figure 3. Comparison of the effect of treatment with 25 μ M D-A β ₁₋₄₀ for 6 d of human brain pericyte cultures with different ApoE genotypes. Cultured HBPs with the ApoE ϵ 4/ ϵ 4 genotype demonstrated more cell death than HBP cultures with the ApoE ϵ 3/ ϵ 4 or ϵ 3/ ϵ 3 genotype. Significance of the observed differences is as follows (ANOVA with Tukey's *post hoc* test for multiple comparisons): ϵ 4/ ϵ 4 versus ϵ 3/ ϵ 4 or ϵ 3/ ϵ 3, $p < 0.001$; ϵ 3/ ϵ 3 versus ϵ 3/ ϵ 4, $p < 0.05$.

These data suggested that ApoE protein modulated the association of A β with the cell surface and subsequent cellular degeneration. We therefore investigated whether simultaneous incubation of cells with D-A β ₁₋₄₀ and ApoE would lead to an ApoE isoform-dependent effect on HBP death. Addition of either ApoE3 or ApoE4 (up to 1.6 μ g/ml), obtained from supernatants of transfected HEK293 cells, resulted in a slight but nonsignificant decrease in D-A β ₁₋₄₀-induced cell death (Fig. 4, left). The addition of any of the three ApoE isoforms (between 1.5 and 12 μ g) produced by baculovirus in Sf insect cells yielded a small but significant decrease in cell death of cultured HBPs (Fig. 4, middle). Remarkably, when HBP cultures were coincubated with D-A β ₁₋₄₀ and conditioned supernatant of either ϵ 2/ ϵ 3 or ϵ 4/ ϵ 4 HBP cultures, a dramatic decrease in cell death was observed (Fig. 4, right). This effect was stronger when culture supernatant was

used from ϵ 2/ ϵ 3 cells compared with that from ϵ 4/ ϵ 4 cells. Addition of ApoE, semipurified from conditioned HBP culture supernatants (up to 20 μ l; containing 0.25 μ g of ApoE), resulted in a dose-dependent decrease in cellular toxicity (Fig. 5).

We then studied whether the differential response of HBPs with different ApoE genotypes could be explained by differences in the ApoE production capacity of the cells. ApoE production by cells with the ϵ 4/ ϵ 4, ϵ 3/ ϵ 4, or ϵ 3/ ϵ 3 genotype was not significantly different ($p = 0.55$; ANOVA), although we observed a trend toward lowest production by HBP ϵ 4/ ϵ 4 cultures (Fig. 6). In two of three experiments, HBP ϵ 4/ ϵ 4 cultures produced the smallest amounts of ApoE of all cultures. Interestingly, although based on only one HBP culture, cells with the ApoE ϵ 2/ ϵ 3 genotype produced a mean 5.0 ± 3.1 ng of ApoE per gram of protein per day ($n = 3$ experiments), which is a factor threefold to 12-fold higher than the other cell cultures ($p = 0.02$ vs ApoE ϵ 3/ ϵ 3; $p = 0.21$ vs ApoE ϵ 3/ ϵ 4; $p = 0.01$ vs ApoE ϵ 4/ ϵ 4). Similar results were obtained with SMC cultures of varying genotype, with the exception of one SMC ϵ 3/ ϵ 3 culture that consistently secreted high ApoE levels comparable with that of the ϵ 2/ ϵ 3 cell cultures. In the SMC cultures, cells with the ApoE ϵ 2/ ϵ 3 genotype produced a mean 13.9 ± 5.4 ng of ApoE per microgram of protein per day ($n = 3$ experiments), which is a factor 4–12 higher than the other cell cultures ($p = 0.01$ vs ApoE ϵ 3/ ϵ 3; $p < 0.0001$ vs ApoE ϵ 3/ ϵ 4; $p = 0.02$ vs ApoE ϵ 4/ ϵ 4; data not shown). Effects on ApoE production by the cultured cells were not correlated to differences in the clinicopathological state of the donors (Braak or CERAD grade of AD, CAA grade) (Table 1).

Finally, ApoE mRNA levels in both HBP and SMC cultures were analyzed using Northern blot analysis. Although we did not observe a perfect match between protein secretion and mRNA levels, this analysis showed that the cells with the ϵ 2/ ϵ 3 genotype contained the highest levels of ApoE mRNA, whereas the cells with the ϵ 4/ ϵ 4 genotype contained the lowest levels (Fig. 7). There was some heterogeneity in the mRNA levels in the ϵ 3/ ϵ 3 and ϵ 3/ ϵ 4 cells. The same SMC culture with the ϵ 3/ ϵ 3 genotype that secreted large amounts of ApoE protein also contained a high level of ApoE mRNA, comparable with that in the ϵ 2/ ϵ 3 cell culture.

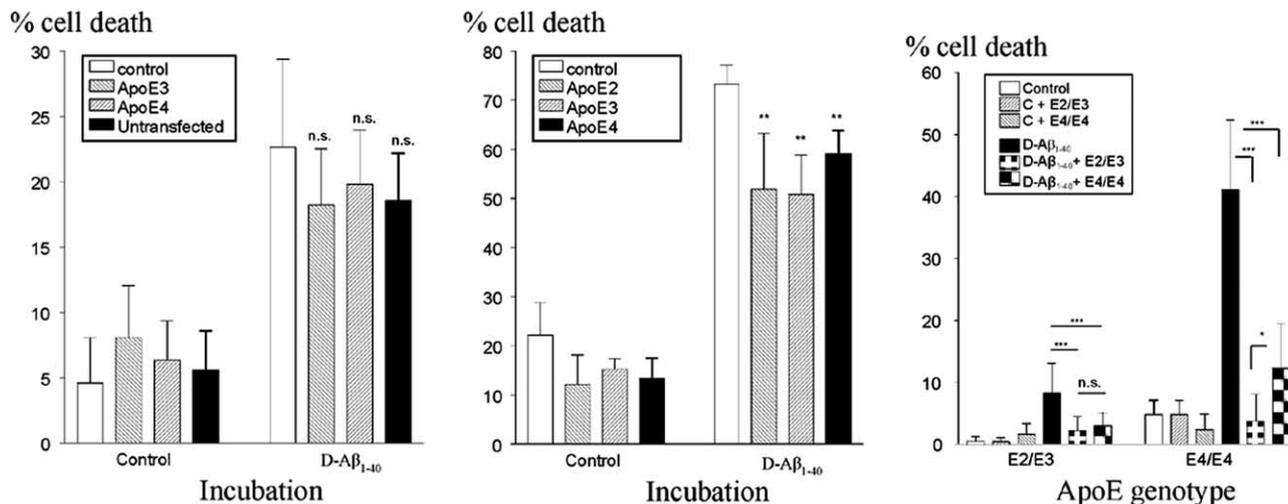


Figure 4. Effect of the presence of ApoE obtained from transfected HEK293 cells (left; 1.65 μ g of ApoE3/4) or produced by baculovirus in Sf insect cells (middle; 1.5 μ g of ApoE2/ApoE3/ApoE4) on the number of dead cells in cultures of human brain pericytes treated with 25 μ M D-A β ₁₋₄₀. Addition of HEK293 ApoE3/4 did not affect the number of dead cells induced by exposure to D-A β ₁₋₄₀ (left). Addition of any of the ApoE isoforms produced by Sf insect cells moderately, but significantly, decreased D-A β ₁₋₄₀-induced cell death (middle). Right, Incubation with conditioned medium obtained from either ϵ 2/ ϵ 3 (E2/E3) or ϵ 4/ ϵ 4 (E4/E4) cells (indicated in the inset) significantly decreased the number of dead cells in HBP cultures with an ϵ 2/ ϵ 3 or ϵ 4/ ϵ 4 genotype (indicated on the x-axis). Level of significance, * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$. n.s., Not significant.

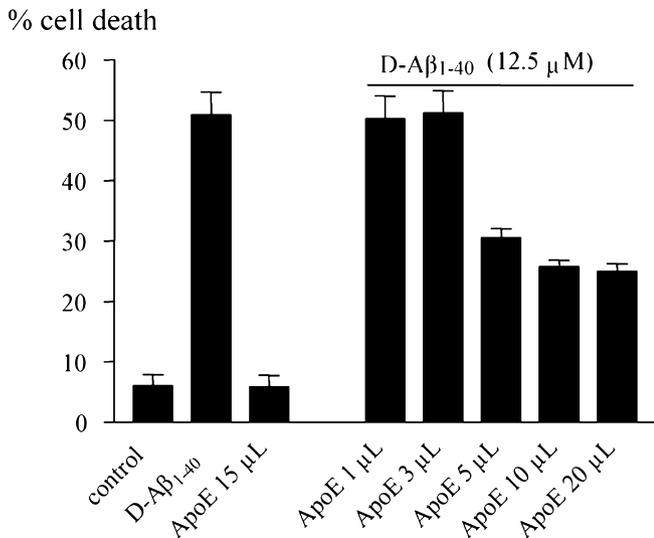


Figure 5. Addition of ApoE, semipurified from supernatants of cultured HBPs, reduced D-A β_{1-40} (12.5 μ M)-mediated HBP cell death in a concentration-dependent way.

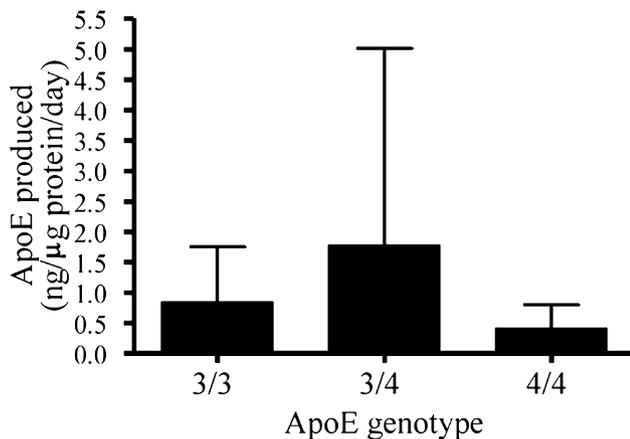


Figure 6. Quantification of ApoE secretion by cultured HBPs with the ApoE ϵ 3/ ϵ 3, ϵ 3/ ϵ 4, and ϵ 4/ ϵ 4 genotypes by ELISA. ApoE production by cells with the ϵ 4/ ϵ 4, ϵ 3/ ϵ 4, or ϵ 3/ ϵ 3 genotype was not significantly different ($p = 0.55$; ANOVA), although we observed a trend toward lowest production by HBP ϵ 4/ ϵ 4 cultures.

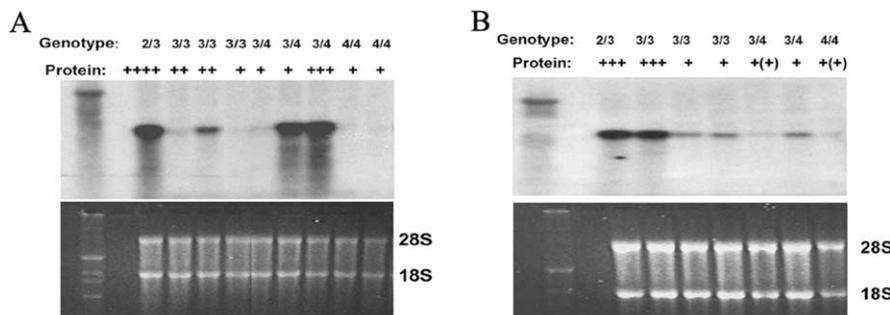


Figure 7. Northern blot analysis of ApoE mRNA expression (top) in cultured HBPs (A) and SMCs (B). Bottom, The levels of 28S and 18S RNA are shown as a control for RNA loading. ApoE mRNA expression was variable, but cells with the ApoE ϵ 4/ ϵ 4 genotype consistently contained the smallest amounts of ApoE mRNA, whereas the (single) cultures with the ϵ 2/ ϵ 3 genotype produced the highest amounts of ApoE mRNA.

Discussion

Because of the initial reports of the association of the ApoE ϵ 4 genotype with the development of AD, many research groups have tried to identify a biological explanation for this epidemiological observation. ApoE binds to A β and affects its fibrillogenesis, possibly in an isoform-specific manner (LaDu et al., 1994; Ma et al., 1994; Wisniewski et al., 1994). In other studies, isoform-specific neurotrophic or neurotoxic effects for ApoE have been described previously (Holtzman et al., 1995; DeMattos et al., 1998; Sun et al., 1998). In the present study, we provide a novel explanation that may, in part, account for the association of ApoE with AD development. The major findings of this investigation are as follows. First, the degree of A β -mediated toxicity of human cerebrovascular cells is dependent on the ApoE genotype of the cells. Cultured HBPs with an ApoE ϵ 4/ ϵ 4 genotype were more vulnerable to A β -mediated cytotoxicity than cells with other ApoE genotypes. Second, ApoE protein causes a direct, concentration-dependent inhibition of A β -mediated toxicity of human cerebrovascular cells. Third, the production level of ApoE protein may, at least in part, depend on the ApoE genotype.

It has been suggested that in the brain, a balance exists between A β deposition and A β removal. A β removal from the brain may occur by several mechanisms. For example, ApoE may play a role in internalization and subsequent degradation of A β and in A β clearance via transport to the circulation. A β may be internalized and degraded by smooth muscle cells (Urmoneit et al., 1997) and by astrocytes (Koistinaho et al., 2004), a process that is facilitated by ApoE, because astrocytes derived from ApoE $^{-/-}$ mice do not degrade A β deposits in the brain. Alternatively, A β may be cleared from the brain via transport across the blood–brain barrier (BBB). LRP-1 binds free A β and possibly is the major receptor mediating A β internalization (Urmoneit et al., 1997; Koistinaho et al., 2004) and its efflux from the brain to the circulation (Shibata et al., 2000). LRP-1 most efficiently removes wild-type A β_{1-40} , whereas A β peptides containing a higher percentage of β -sheet structures (such as wild-type A β_{42} and D-A β_{1-40}) bind less efficiently, leading to accumulation of A β in brain microvessels (Deane et al., 2004). ApoE, which is also a ligand for LRP-1, may facilitate the clearance of A β from the brain, because clearance was significantly reduced in ApoE $^{-/-}$ mice (Shibata et al., 2000). Therefore, it is possible that at least part of the cerebral A β forms complexes with ApoE and that such complexes via ApoE–LRP-1 interactions may be either degraded by astrocytes or cleared from the brain by transport over the BBB. However, although the existence of such complexes in the brain has been demonstrated previously (Russo et al., 1998), their possible interaction with LRP-1 has not yet been studied.

Thus, high cerebral concentrations of ApoE may inhibit deposition of A β in the form of SPs and CAA. Our *in vitro* data suggest that, at least at the vascular level, ApoE abrogates the pathological interaction of A β with vascular cells and retains A β in a conformation that does not result in its assembly at the cell surface, possibly by a diminished interaction with LRP-1. These findings are in line with the observation that human ApoE, in an isoform-specific way, suppresses A β deposition in both young and aged APP-V717F transgenic mice (Holtzman et al., 1999; Fagan et al., 2002). Furthermore, human ApoE delays the shift in A β from soluble pools to

insoluble pools in these animals, which is in agreement with our observations of decreased accumulation of A β at the cell surface in the presence of elevated ApoE concentrations (Fagan et al., 2002).

Thus, relatively high concentrations of cerebral ApoE are beneficial. It can be anticipated that the levels of free cerebral ApoE are reflected by its concentration in CSF. It has been reported, however, that the CSF concentration of ApoE is similar in both control and AD groups with different ApoE genotypes (Lehtimäki et al., 1995; Yamauchi et al., 1999), although a trend toward elevated ApoE levels in patients carrying the ϵ 2 allele was observed (Lehtimäki et al., 1995). However, because the applied assays have all been designed to detect unbound ApoE, it remains possible that the levels of CSF A β –ApoE complexes will be different among individuals with different ApoE genotypes.

It has been suggested by Nicoll et al. (1997) and Greenberg et al. (1998) that both the ϵ 2 and ϵ 4 alleles are risk factors for CAA-related hemorrhages, the latter by increasing the vascular amyloid load and the former by increasing the susceptibility of the vessel for rupture. By incorporating these suggestions in our *in vitro* model, one would predict increased cell death in cells carrying the ϵ 2 allele, because this would lead to vessel weakening by (A β -mediated) degeneration of SMCs or HBPs. However, our *in vitro* data show less cytotoxicity associated with cells possessing an ϵ 2 allele. Perhaps the ϵ 2 allele promotes other changes leading to vessel rupture, such as increased extracellular proteolysis.

ApoE produced by cerebrovascular cells may play a prominent role in the pathogenesis of CAA. Second only to the liver, the brain is the organ that produces the largest amount of ApoE. Astrocytes are regarded as the major source of cerebral ApoE (Boyles et al., 1985), although neurons may be able to produce ApoE as well (Xu et al., 1999). Here, we demonstrated that both pericytes and SMCs secrete ApoE and, because both are located at the abluminal side of the blood–brain barrier, they may contribute to the pool of cerebral ApoE. This may be particularly important with regard to the etiology of CAA. An attractive model explaining the pathogenesis of vascular A β deposition has been proposed by Weller et al. (1998), who suggested that A β of neuronal origin might be transported from the brain parenchyma to the vasculature by drainage of interstitial fluid. Experimental support for this model was provided by the transgenic mice studies by Calhoun et al. (1999), who demonstrated extensive CAA formation in APP transgenic mice under the control of the neuronal Thy-1 promoter. This model of CAA formation requires the availability of local factors that enhance or otherwise stimulate A β deposition in the vasculature. The availability of vessel-derived ApoE, but possibly also factors such as heparan sulfate proteoglycans (van Horssen et al., 2001), may regulate A β clearance and/or deposition in the vasculature. Our data suggest that when ApoE is available in high-enough concentrations, A β may remain solubilized and be processed for clearance, whereas if ApoE is available in restricted amounts only, A β will be prone to deposition and cell-surface aggregation in the vessel wall.

The degree of lipidation of the ApoE molecule is crucial to its biological activities. Circulating ApoE is tightly bound to several classes of lipoproteins, such as high-density lipoprotein and very-low-density lipoprotein. It was demonstrated previously that lipidated and unlipidated ApoE differentially interact with A β , suggesting that the degree and type of lipidation of ApoE are of critical importance (LaDu et al., 1994). In accordance with this study, we observed differences in the reduction of A β -induced cellular toxicity of HBPs by ApoE, dependent on its source; ApoE, especially when secreted by HBPs, seemed very efficient. Previous studies by DeMattos et al. (1998) and Sun et al. (1998) showed

that ApoE is secreted by cultured cells as specific lipid-containing particles different from those in CSF (LaDu et al., 1998). Our data suggest that between different cell types (HEK293 cells vs HBPs/SMCs), the degree of ApoE lipidation may vary. The recombinant ApoE from PanVera is posttranslationally modified, but lipidation of the molecule is probably also different in Sf insect cells compared with HBPs.

Our comparative analysis of ApoE protein secretion and mRNA levels by the respective HBP and SMC cell cultures suggests that not only the ApoE genotype but also other unstudied factors may affect ApoE secretion (e.g., ApoE promoter polymorphisms) (Artiga et al., 1998) or factors involved in lipid metabolism. Furthermore, although we did not collect any evidence in this direction, we cannot completely exclude the possibility that the original cerebral environment of the cells (i.e., AD, control, or other diagnosis) contributes to the cellular phenotype *in vitro*. Finally, future studies on other cerebral cell types will have to reveal to what extent ApoE will affect other cellular responses to A β , including neuronal cell death and glial cytokine production.

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