PTHrP Potentiating Estradiol-induced Vitellogenesis in Sea Bream

(Sparus auratus, L.)

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Abstract

In fish, vitellogenin is an important nutritional precursor protein produced solely in the liver and released into the blood where it binds calcium. In the gilthead sea bream (Sparus auratus) 17β-Estradiol (E2) plays an important role in the synthesis of vitellogenin, but also the pituitary hormones prolactin (PRL) and growth hormone (GH) can stimulate vitellogenin induction in fish. Considering the emerging involvement of PTHrP in fish calcium metabolism and the importance of calcium regulation in reproduction, we investigated the possible role of PTHrP in vitellogenesis. E2-naive and E2-primed sea bream hepatocytes were used in an in-vitro primary hepatocyte culture and stimulated with a recombinant sea bream PTHrP (sbPTHrP) to establish the contribution of sbPTHrP alone or in combination with E2 to the regulation of hepatic vitellogenin synthesis. Hepatocytes stimulated solely with sbPTHrP were not affected in their vitellogenesis. However, in hepatocytes stimulated with E2 in combination with sbPTHrP a higher vitellogenin production was seen than with E2 alone. It is concluded that sbPTHrP has a potentiating effect on estradiol stimulation of vitellogenin production by sea bream hepatocytes. The sea bream provides a unique model where vitellogenesis regulation can be studied on E2-naive liver cells, both in vivo and in vitro.


Introduction

In fish, and other oviparous species, vitellogenin is an important nutritional precursor protein produced solely in the liver and released into the blood where it binds calcium. Circulating vitellogenin is transported to the ovary and taken up by developing oocytes to give rise to two major yolk proteins: lipovitellin and phosvitin. Lipovitellin supplies the necessary amino acids and lipids for the developing embryo, while phosvitin delivers the minerals important for skeletal development (Polzonetti-Magni et al., 2004). In the gilthead sea bream (*Sparus auratus*) 17β-Estradiol (E₂) plays an important role in the synthesis of vitellogenin (Guerreiro et al., 2002; Mosconi et al., 2002). Several studies have demonstrated that not only E₂ but also the pituitary hormones prolactin (PRL; Kwon and Mugiya, 1994) and growth hormone (GH; Mosconi et al., 2002; Peyon et al., 1996) stimulate vitellogenesis in fish. In Japanese eel, *Anguilla japonica*, a multihormonal control by GH and/or PRL as well as E₂ is essential for proper vitellogenin synthesis (Kwon and Mugiya, 1994). In female European silver eel (*Anguilla anguilla*) GH potentiates the effect of E₂ on cultured hepatocytes (Mosconi et al., 2002; Peyon et al., 1996). In sea bream the liver responds directly to stimulation with recombinant sea bream GH and a homologous pituitary homogenate as well as E₂ stimulation (Mosconi et al., 2002; Peyon et al., 1996).

When E₂ induces vitellogenesis also plasma total calcium levels rise as vitellogenin binds calcium and the vitellogenin concentration in plasma becomes very significant. Importantly, in primary rainbow trout hepatocyte cultures E₂ induced vitellogenesis, and this induction is dependent on extracellular calcium (Yeo and Mugiya, 1997). The hypercalcemic effects of E₂ result from calcium mobilization from bone (scales), enhanced intestinal absorption and kidney reabsorption (Persson, 1997). In sea bream
however, E₂ does not induce bone resorption in scales and the high plasma calcium levels cannot be explained by renal and intestinal calcium reabsorption.

A 13-fold increase of plasma hypercalcemic parathyroid hormone related protein (PTHrP) levels was shown in sea bream treated with E₂-implants; the rise in PTHrP precedes the rise in plasma calcium levels, and this establishes that PTHrP probably mediates the hypercalcemic effects of E₂; PTHrP is thought to stimulate calcium uptake from the water via the gills in this species (Guerreiro, 2002).

PTHrP was first discovered in the late 80’s as a hypercalcemic factor originating from certain cancers of the head, neck, breast, lung and kidney, causing the syndrome Humoral Hypercalcemia of Malignancy (HHM). PTHrP is an important factor in various tissues in normal physiology with roles in calcium transport, cell proliferation, relaxation of vascular tissue and has specific roles during early development (Philbrick et al., 1996; Wysolmerski and Stewart, 1998).

In fish, PTHrPs and several PTH/PTHrP receptors have been characterized and cloned (Danks et al., 1993; Devlin et al., 1996; Flanagan et al., 2000; Gensure et al., 2005; Gensure et al., 2004; Ingleton, 2002; Power et al., 2000; Rotllant et al., 2003; Rubin and Juppner, 1999). High to moderate levels of immunoreactive PTHrP have been detected in various tissues such as the pituitary gland, liver, head kidney, oesophagus, gill, intestine, skin and muscle. High PTHrP levels have been measured in pituitary extracts and plasma of sea bream and flounder, which indeed supports that PTHrP in teleosts can act as an endocrine factor (Rotllant, Worthington et al. 2003).
Few physiological studies have been conducted on the effect of PTHrP in defined tissues. Piscine PTHrP (1-34) was shown to down-regulate mRNA expression of osteonectin in sea bream scales (Anjos et al., 2005) and in the same species it was found that PTHrP (1-34) stimulates cortisol production by interrenal cells (Rotllant et al., 2005). PTHrP stimulates whole body calcium influx and reduces calcium efflux in whole sea bream larvae (Guerreiro et al., 2001). Considering the emerging involvement of PTHrP in fish calcium metabolism and the pivotal importance of calcium regulation in ovarian maturation, we set out to study the possible involvement of PTHrP in hepatic vitellogenesis; recombinant, full length PTHrP (1-125) was used (Anjos et al., 2005).

Material and Methods

Fish

Juvenile sea bream of approximately 1g mass were obtained from a stock bred at a commercial fish farm (Viveiro Vilanova, Lda., V. N. Milfontes, Portugal). They were transported by air to the facilities at the Radboud University Nijmegen, where they were held in 600-litre round tanks with an aerated flow-through system; water salinity was 34% and the water temperature was 22°C. Water quality (pH, NO₂⁻, NO₃⁻, NH₄⁺) was measured once a week and the salinity was checked daily. The photoperiod was 12 h:12 h and the fish stock was fed commercial pellets (Trouvit, Trouw, Putten, The Netherlands) at a ration of 2% of the total estimated body mass per day. Fish used for experiments were juvenile sea bream of approximately 90 g mass.
Hepatocyte Isolation

Hepatocytes were isolated according to Mommsen et al. (1994), with minor modifications. After anaesthesia of the fish with 2-phenoxyethanol (0.1% v/v), the liver was perfused via the heart with Ca\(^{2+}\)-free hepatocyte buffer (HB; 175 mM NaCl, 5.4 KCl, 0.81 mM MgSO\(_4\), 0.33 mM Na\(_2\)HPO\(_4\), 0.44 mM KH\(_2\)PO\(_4\), 5 mM NaHCO\(_3\), pH 7.63) containing 1 mM EGTA for 10-15 minutes at room temperature. After clearing the blood from the liver, the liver was excised from the abdominal cavity and transferred onto a sterile petri dish, cut into smaller fragments with a razor blade and the tissue was treated with HB containing collagenase at a concentration of 0.3 mg/ml for about 30-45 minutes (depending on liver mass) to dissociate the cells. The softened liver fragments were further minced with a razor blade and the homogenate was filtered twice, through a 200 and a 50 µm sterile nylon mesh screen. The cell suspension was transferred into a sterile tube (Greiner, 50ml) and subsequently centrifuged 3 times at 80g for 4 minutes at 10°C. Cell pellet washes of HB containing 1.5 mM CaCl\(_2\) (HB-Ca\(^{2+}\)) and a 50/50 mixture of L-15 medium (L-15, Gibco) (containing antibiotics/antimycotics (Gibco), 5 mM NaHCO\(_3\) and 10 mM NaCl) and HB-Ca\(^{2+}\) were conducted in between centrifugations. After the last centrifugation, the cell pellet was resuspended in a small volume of L-15 medium; cell yield was assessed by determining the number of cells using a counting chamber and cell viability monitored by trypan blue exclusion. Only cell preparations with more than 90% viability were used for experiments. Isolated hepatocytes were plated in 24-well culture plates (Greiner) at a density of 1x10^6/ml in a volume of 1 ml. Cells were incubated at 22°C (the temperature at which the fish are kept) at saturated humidity.
Effect of E₂ and PTHrP on vitellogenin synthesis

Blood parameters after E₂ priming

Sea bream is a protandrous fish and therefore in initial experiments the (male) fish were estrogrenized to induce liver vitellogenesis. Fish received a peritoneal implant of coconut butter containing 0, 5 or 10 mg/kg body weight (BW) E₂ and were left for 8 days. After estrogrenization fish were euthanized with 2-phenoxyethanol (0.1% v/v; Sigma-Alderich, St Louis, MO, USA) and fish blood was taken from the caudal vessels with a 1 ml tuberculin syringe, rinsed with Na⁺-heparin (Leo Pharma, Weesp, The Netherlands, 5000 U ml⁻¹). Collected blood was centrifuged at 13,600 g for 10 min. Plasma was stored at –20°C for subsequent analysis of total calcium, estradiol and vitellogenin (VTG) levels (see below).

In-vivo and in-vitro priming

To establish the effect of in-vivo priming, fish received a single injection of 10 mg/kg BW E₂ and hepatocytes were isolated 8 days later. Next, the in-vitro hepatocyte culture was divided into two groups for the first 48 hrs. One group of cells was treated with E₂ (10⁻⁶ M) and one group was treated with vehicle. After the first 48 hrs of in-vitro priming the E₂ treated group was further divided into 2 groups: one was further treated with just E₂ and one group exposed to a combination of E₂ and 10⁻⁸ M sbPTHrP. The vehicle, E₂ and the E₂ plus sbPTHrP groups were cultured for 2 x 24 hrs, media was collected from each well and used to assess vitellogenin release with an enzyme-linked immunosorbent assay (ELISA; see below). As an extra control, E₂-naïve hepatocytes were cultured for 4 days, treated with vehicle and the medium analysed.
As the higher dose of 10mg/kg made the liver cells highly active and refractory to further manipulation, fish were then primed with a lower dose, 5mg/kg BW E₂. Hepatocytes were isolated 8 days later and liver material from up to 4 fish was pooled. The hepatocyte culture was continued for 4 days and the cells were treated with E₂ (10⁻⁶M) and/or sbPTHrP (10⁻⁸M). The medium was collected and changed every day from each culture plate well and analysed with the vitellogenin ELISA (see below).

In-vitro priming
The last series of experiments was run on tissues of control, i.e. E₂-naïve (no E₂ exogenously administered) fish. Hepatocytes were isolated and cultured for 6 days treated with E₂ (10⁻⁶M) and/or sbPTHrP (10⁻⁸M). The hepatocyte culture medium was sampled and changed every other day (after one day of culture VTG levels were below detection) to measure production of vitellogenin using the ELISA (see below).

Chemicals
17β-Estradiol (E₂) was purchased from Sigma (Sigma-Aldrich, St Louis, MO, USA). E₂ was dissolved in ethanol and the final concentration of ethanol in the culture medium never exceeded 0.02%. The recombinant parathyroid hormone related protein (sbPTHrP 1-125) used in this study was obtained from Manchester University where the sbPTHrP was produced and purified (Anjos et al., 2005).
**Plasma analyses**

Plasma E₂-levels were analysed by an ELISA method (IBL, Hamburg). The lower sensitivity limit of the E₂-ELISA was 10 pg/ml. Plasma total calcium was measured with a calcium kit (Roche, Mannheim, Germany). Plasma vitellogenin was measured using a vitellogenin ELISA (see below).

**Vitellogenin assay**

Vitellogenin concentration in the medium was measured by a standard ELISA protocol in 96-well plates using purified sea bream vitellogenin as standard, a polyclonal rabbit anti-sea bream vitellogenin antiserum (raised by Dr Per-Erik Olsson, Orebro University, Sweden) as primary anti-antibody and a secondary HRP-conjugated goat-anti rabbit antibody (Nordic Labs, Tilburg, The Netherlands). Briefly, the standard curve and sample dilutions were prepared in coating buffer (0.1 M Na₂CO₃, pH 9.6); 100μl of each was added to each well and incubated overnight. The wells were washed 3 times with PBS-Tween (phosphate buffered saline, 0.05% Tween-20, pH 7.4) and incubated for 1 h at RT with PBS-Tween containing 0.5% dry-milk to block non-specific binding. The wells were washed 3 times in PBS-Tween followed by incubation with primary antibody (diluted 1:3000) for 1h at RT. After washing with PBS-Tween the secondary antibody incubation (diluted 1:3000) was performed for 1 h at RT. The wells were washed for 3 times with PBS-Tween and 200μl of OPD-H₂O₂ substrate buffer (McIlvaine Buffer containing 0.05% o-phenylenediamine and 0.375% H₂O₂) was added to each well. The reaction was stopped with 4 M H₂SO₄ after 10 minutes and the plates were read using a microplate reader (Bio-Rad microplate reader model 680). The lower detection limit of the VTG-ELISA was 1 ng/ml. The intra-assay coefficient of variation was 4.6% (n=6) and inter-assay coefficient variation was 11.9% (n=6).
Statistical analysis

Data are presented as mean values ± standard error of the mean (SEM) or as mean values ± standard deviation (SD), depending on the number of experiments run.

Analysis of variance (ANOVA) was used to assess differences between groups, followed by the Bonferroni test (SPSS Windows version 13.0, Chicago, IL).

Significance was accepted when P<0.05.
Results

Blood parameters after E2 priming

In plasma of naïve juvenile sea bream E2-levels were extremely low (0.02 μM ± 0.005; Table 1). E2 implants significantly increased estradiol levels (P<0.05; Table 1) and doubling the E2 dose led to a doubling of plasma E2-levels. The significant degree of hypercalcemia (P<0.05; Table 1) essentially plateaued at the 5mg/kg dose. Both concentrations of E2 have a significant (P<0.05; Table 1) effect on plasma vitellogenin release compared to control, however VTG levels did not differ between the two treatments.

In-vivo and in-vitro priming

In-vivo primed with 10mg/kg BW E2

On day 3 of culture, addition of E2 alone or combined with sbPTHrP did not affect the VTG-production compared to vehicle treated liver cells, which resulted from priming the fish (Fig.1). Vehicle-treated liver cells produced significant amounts of VTG compared to vehicle treated liver cells of E2-naïve sea bream (P<0.05). VTG production had declined on day 4 of culture for the vehicle treated group to 100ng/ml, being significantly elevated compared to VTG levels of E2-naïve liver cells. The combined addition of E2 and sbPTHrP further stimulated VTG production compared to day 3 (P<0.05). Day 4 shows significant differences between VTG levels in control, E2 and the combination of E2 and sbPTHrP treated hepatocytes. With VTG production in E2 treated cells being significantly elevated compared to control (P<0.05) and the combination of E2 and sbPTHrP is significantly elevated compared to control and E2 treated hepatocytes (P<0.05; Fig.1).
**In-vivo primed with 5mg/kg BW E₂**

We then set out for an experiment where fish were primed with 5mg/kg E₂. VTG production declined during the first three days of culture independently of in-vivo or in-vitro treatment (Fig.2). While control and sbPTHrP group dropped below the 50 ng/ml, the E₂ group as well as the group co-treated with E₂ and sbPTHrP stayed significantly elevated compared to control and sbPTHrP treated cells (P<0.05; Fig.2.). On day 3 a significant difference of VTG levels was detected between E₂ treated liver cells and the liver cells treated with E₂ and sbPTHrP (P<0.05). On day 4 all experimental groups show a significant rise in VTG levels compared to control (P<0.001) and significantly differ from each other (P<0.001; Fig.2). Noteworthy is the sudden significant rise of VTG levels from the sbPTHrP treated hepatocytes compared to control on day 4 of culture.

**In-vitro priming**

The effect of E₂ and/or sbPTHrP on the isolated hepatocytes of unprimed fish is shown in Fig.3. For all experimental groups almost undetectable levels of vitellogenin were measured in the first 2-4 days (< 5 ng/ml; results not shown). In control as well as sbPTHrP (10^-8M) treated liver cells the vitellogenin concentration remained very low (< 15 ng/ml) up to 6 days. Vitellogenin levels rose significantly (P<0.05) when treated with E₂ (10^-6M) after 6 days *in vitro*. When hepatocytes were co-treated with E₂ and sbPTHrP a significantly (P<0.05) higher amount of vitellogenin was produced on day 6 than with E₂ only (Fig.3)
Discussion

This study is the first to show that PTHrP is directly involved in fish vitellogenesis. Two important observations have to be taken into regard from this study. First, the recombinant sea bream PTHrP proved to have potentiating effects on E\textsubscript{2} induced VTG production and second the protandrous gilthead sea bream provides us with E\textsubscript{2}-naïve liver cells. This makes this fish a good model for in vitro hepatocyte bioassays for the study of vitellogenesis as hepatocytes can be primed \textit{in vivo} or \textit{in vitro} without prior ‘substantial experience’ to E\textsubscript{2}.

To perform this research a primary culture was set up for sea bream hepatocytes. Research has been conducted on sea bream liver before, but only \textit{in vivo} or \textit{in vitro} using liver fragments of a certain size (Carnevali et al., 2005; Mosconi et al., 2002), not on isolated cells. Isolated hepatocytes in our hands remained viable for up to at least 8 days (trypan blue exclusion) and remained E\textsubscript{2} sensitive, which is in accordance with other studies on this and other species (Flouriot et al., 1993; Islinger et al., 1999; Kim and Takemura, 2003; Kim et al., 2003; Kwon and Mugiya, 1994; Peyon et al., 1996; Riley et al., 2004; Tollefsen et al., 2003; Yeo and Mugiya, 1997).

E\textsubscript{2} is known to upregulate its own receptor, preceding the upregulation of vitellogenin mRNA (Flouriot et al., 1997) and this can be an explanation for the observed result in control (unprimed) juvenile sea bream where, after 2-4 days, no response is measured in either of the groups, only after 6 days the response can be seen clearly (Fig.3). The vitellogenic response observed after 6 days corresponds with similar kinetics in the study of Peyon et al. (Peyon et al., 1996).

In the cells of E\textsubscript{2}-naïve fish no direct effect of sbPTHrP on vitellogenesis was observed, but importantly sbPTHrP alone did stimulate VTG production in hepatocytes of fish primed with 5mg/kg BW E\textsubscript{2}; only on day 4 of subsequent culture the direct effect of sbPTHrP was observed (Fig.2). We take this as a ‘memory effect’
induced by the E₂ priming (Peyon et al., 1993). Indeed, plasma E₂ levels were strongly elevated after both priming treatments and suggest that such high E₂ levels in vivo could have lasting and dose-dependent effects on the hepatocytes in culture, as can be seen with the high dosage priming (10 mg/kg E₂; Fig. 1) where no difference was shown between vehicle and experimental treated cells. Accordingly, in controls VTG production finally decreased after 3 days of culture, whereas with a lower E₂ priming dose the decline of VTG sets in earlier (Fig.2). The potentiating effect of sbPTHrP in the ‘lower dose primed’ fish became significant as of day 4 of culture.

For a long time E₂ was considered to be the main trigger in the process of hepatic vitellogenesis, it is now known that this process is under a multihormonal regulation in which factors such as pituitary hormones (GH, PRL) and steroid hormones, e.g. progesterone and cortisol play a role (Polzonetti-Magni et al., 2004). In a study of Mosconi and co-workers hepatocytes were treated, (besides E₂ and GH), with a pituitary homogenate inducing VTG production (Mosconi et al., 2002), such a homogenate may well contain significant amounts of PTHrP (Ingleton, 2002; Rotllant et al., 2003). This study demonstrates PTHrP as one of the factors involved in the process of hepatic vitellogenesis with known expression of PTHrP and the PTHrP-receptor in sea bream liver (Hang et al., 2005) and the pituitary as the main source suggested (Rotllant et al., 2003). Although the mechanism for the potentiating effect of PTHrP on the E₂ induced hepatocytes has not been studied in more detail here, it is known that extracellular calcium is an important factor during vitellogenesis (Yeo and Mugiya, 1997). E₂ significantly stimulates the secretion of PTHrP into the bloodstream before the rise in plasma calcium levels and this suggests that PTHrP is a good candidate for the modulation of the hypercalcemic action of E₂ as it elevates plasma PTHrP that has hypercalcemic actions (Guerreiro, 2002).
This hypothesis of a hypercalcemic action of PTHrP and possibly even acting as the main calcium up-regulator is supported by studies performed on sea bream larvae where PTHrP was able to stimulate whole body calcium influx and reduce calcium efflux (Guerreiro et al., 2001), and elevated levels of PTHrP were measured when calcium is limited in water or in diet (Abbink et al., 2004).

However, other possible mediators of the calciotropic action of E₂ may be factors such as GH and PRL (Flik et al., 1994; Takagi et al., 1992).

For long, fish were thought to lack parathyroid hormone (PTH), but certainly fish including sea bream express PTHrP. As PTH and PTHrP signal through the same receptor (Gensure et al., 2005) it was thought that the PTHrP gene was the ancestral gene to PTH (Blair et al., 2002). Recently PTH genes and their products were demonstrated in zebrafish and the puffer fish (Canario et al., 2006; Gensure et al., 2004) and has opened new fields of research. PTH induces calcium influx in rat hepatocytes (Klin et al., 1994) and the puffer fish PTH-like ligand induces calcium whole body influx in sea bream larvae (Canario et al., 2006). In both cases the pathway, via which the PTH action is realised, involves the G protein adenylate cyclase-cAMP system. Indeed, also PTHrP can act through this pathway (Canario et al., 2006; Gensure et al., 2005) via shared receptors. Via the second messenger pathway another possible mechanism may be responsible for the potentiating action of PTHrP. Protein kinases can enhance the E₂ dependent estradiol-receptor (ER) transcription activity. E₂, cAMP and certain growth factors, for instance insulin-like growth factor-I, have been shown to mediate regulatory effects of certain genes via the ER (Cho and Katzenellenbogen, 1993).

IGF-I has been shown to stimulate vitellogenin synthesis in the frog and it was recently suggested that the GH effect on synthesis of vitellogenin is at least partly mediated through its action on IGF-I synthesis (Carnevali et al., 2005). It has been shown that
PTHrP and PTH can induce elevations of IGF-I in humans (Horwitz et al., 2005) and in cultures of 21-day fetal rat calvaria (Canalis et al., 1990). Moreover, it is speculated that during fracture healing of bone, PTHrP influences IGF-1 expression and both enhance the ossification in an autocrine/paracrine fashion (Okazaki et al., 2003). To elucidate the pathways in which PTHrP works in the process of hepatic vitellogenesis requires further study.

In conclusion, we have established a new role for PTHrP showing a potentiating effect on hepatocyte vitellogenesis when induced with E2. This study with other studies on the effect of PTHrP on corticosteroidogenic activity (Rotllant et al., 2005), downregulation of osteonectin mRNA in scales (Anjos et al., 2005) and the ability to stimulate whole body calcium influx and reduce calcium efflux in whole sea bream larvae show that PTHrP in fish (Guerreiro et al., 2001), as in normal human physiology, is probably a pleiotropic hormone working on different levels in different tissues.

Acknowledgements

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Rubin, D.A., Juppner, H., 1999. Zebrafish express the common parathyroid hormone/parathyroid hormone-related peptide receptor (PTH1R) and a novel receptor (PTH3R) that is preferentially activated by mammalian and fugu fish parathyroid hormone-related peptide. J Biol Chem 274, 28185-28190.


Legends

Fig. 1. Vitellogenin release, after in-vivo priming (10mg/kg BW E₂). Hepatocytes treated with either vehicle, E₂ or a combination of E₂ and sbPTHrP. Values are expressed as means ± SEM (N= 4). On Day 3 no significant differences occurred between the groups. On Day 4 E₂ and the combination group differ significantly from control and each other. On both days VTG levels were significantly higher then VTG levels of cultured hepatocytes treated with vehicle from E₂-naive fish. Letters denote significance at P<0.05.

Fig. 2. In-vitro vitellogenin synthesis, after lower dose priming of E₂ (5mg/kg BW), during 4 days of E₂ and/or sbPTHrP treatments. After 4 days a potentiating effect can be seen of the group treated with E₂ (10⁻⁶M) and sbPTHrP (10⁻⁸M). Values are expressed as means ± SD (n= 6). Letters denote significance per day of culture. On day 2 and 3 letters show significance at P<0.05 and on day 4 letters denote significant difference at P<0.001.

Fig. 3. Effect of E₂ and/or sbPTHrP on the production of vitellogenin in control (unprimed) hepatocytes in primary culture. Comparison of the vitellogenin synthesis in control vs. sbPTHrP, E₂ and E₂ + sbPTHrP on the last day of culture. Values are expressed as means ± SEM (N=6). Bars with different letters denote significant difference (P< 0.05).
Table 1. Plasma levels of Estradiol, Total Calcium and Vitellogenin in sea bream injected with 0 (Control), 5mg or 10mg/kg BW 17β-Estradiol (E\textsubscript{2}-injected) coconut butter implants. Values are the mean ± SD, N=10 for Control or N=8 for E\textsubscript{2}-injected. Letters in the same row denote significant difference (P<0.05).

<table>
<thead>
<tr>
<th>Plasma levels</th>
<th>Control 0mg</th>
<th>E\textsubscript{2}-injected 5mg</th>
<th>E\textsubscript{2}-injected 10mg</th>
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<tr>
<td>Estradiol (µM)</td>
<td>0.02 ± 0.005\textsuperscript{a}</td>
<td>2.04 ± 1.33\textsuperscript{b}</td>
<td>4.53 ± 0.27\textsuperscript{c}</td>
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<tr>
<td>Total Calcium (mM)</td>
<td>1.85 ± 0.28\textsuperscript{a}</td>
<td>17.10 ± 2.4\textsuperscript{b}</td>
<td>20.16 ± 3.0\textsuperscript{c}</td>
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<td>VTG (ng/ml)</td>
<td>5.29 ± 2.18\textsuperscript{a}</td>
<td>770.91 ± 38.67\textsuperscript{b}</td>
<td>804.02 ± 47.78\textsuperscript{b}</td>
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