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Reduced Aph-1b expression causes tissue- and substrate-specific changes in γ-secretase activity in rats with a complex phenotype

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SPECIFIC AIMS

The γ-secretase enzyme is a tetrameric protein complex consisting of presenilin-1 (or -2), nicastrin, PEN-2, and Aph-1a (or -1b), and displays intramembrane catalytic activity toward a number of type I transmembrane proteins that are involved in a variety of (neuro)developmental signaling pathways. The aim of this study was to examine the functional consequences of a gene-dosage imbalance of a γ-secretase component, namely the effect of the presence of only one copy or three copies of the rat Aph-1b gene on the level of γ-secretase cleavage activity toward various endogenous substrates.

PRINCIPAL FINDINGS

1. Reduced Aph-1b expression levels are not compensated by its paralog Aph-1a

Using Northern blot analysis, we detected high levels of Aph-1b mRNA in most tissues from rats with three copies of the gene encoding the γ-secretase component Aph-1b (III/III rats), whereas in the same tissues from rats with one Aph-1b gene copy (I/I rats) the levels were not or hardly above background. In contrast, the mRNA levels of the paralogs Aph-1aS and -1aL did not differ between I/I and III/III rat tissues, also not in the tissues with large differences in Aph-1b expression. Furthermore, the ratios between the Aph-1b and Aph-1a mRNA levels greatly varied among the III/III rat tissues. Tissues such as the olfactory bulb, pons/medulla, testis and lung showed high Aph-1b/-1a ratios, while the cortex, hippocampus, spinal cord, stomach, large intestine and thymus had moderate ratios, and the cerebellum, striatum, (hypo)thalamus, eye, heart, muscle, small intestine, spleen, liver and pancreas displayed low ratios (Fig. 1). Real-time quantitative RT-PCR analysis revealed significant reductions in Aph-1b mRNA levels in cerebellum, olfactory bulb, cortex, and lung from I/I compared with III/III rats (P<0.05), with the largest difference observed for the olfactory bulb (~8-fold reduction). In contrast, the Aph-1a mRNA levels did not differ between the two rat lines in the tissues analyzed. The Aph-1b/-1a ratios determined by PCR were similar to those obtained through Northern blot analysis (high Aph-1b/-1a ratio in olfactory bulb and lung, moderate ratio in cortex and low ratio in cerebellum).

2. Tissue- and substrate-specific alterations in γ-secretase cleavage activity in rats with reduced Aph-1b levels

Since the Aph-1 protein is an essential component of the γ-secretase complex, we were interested in the effect of the differential mRNA expression of Aph-1b on the proteolytic cleavage activity of the complex in I/I and III/III rat tissues. To this end, the levels of cleavage products of various γ-secretase substrates were examined by Western blot analysis using antibodies directed against the C-terminal regions of the substrates. In general, the proteolytic processing of a γ-secretase substrate starts with shedding of its extracellular domain, leaving a C-terminal fragment (CTF) that is subsequently cleaved by γ-secretase to its intracellular domain (ICD). One of the best known substrates of γ-secretase is the Alzheimer’s disease-linked amyloid-β precursor protein (APP). APP is part of the APP superfamily that in mammals includes the two APP-like proteins APLP1 and APLP2. We compared the γ-secretase cleavage activities toward the APP superfamily members in tissues of III/III rats having different Aph-1b/-1a ratios (high: olfactory bulb and lung; moderate: spinal cord and cortex; low: (hypo)thalamus, cerebellum and spleen) with the activities in the corre-

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The levels of the direct γ-secretase substrates (CTFs) were significantly increased in the olfactory bulb (APP: 1.6-fold \( P < 0.01 \); APLP1: 2.0-fold \( P < 0.05 \); APLP2: 2.1-fold \( P < 0.05 \)), the lung (APP: 2.2-fold \( P < 0.05 \); APLP2: 1.3-fold \( P < 0.05 \)), the spinal cord (APP: 1.6-fold \( P < 0.02 \); APLP1: 1.6-fold \( P < 0.05 \); APLP2: 1.8-fold \( P < 0.05 \)) and the cortex (APP: 1.3-fold \( P < 0.05 \); APLP2: 2.2-fold \( P < 0.02 \)) of I/I rats compared with III/III rats. No significant differences in the CTF levels were observed in the (hypo)thalamus, cerebellum, and spleen of I/I and III/III rats, i.e., in tissues with a low \( \text{Aph-1b/-1a} \) ratio (Fig. 2).

We next analyzed the cleavage products of the γ-secretase substrates neurotrophin receptor p75, the neuregulin receptor ErbB4, and neuregulin-2 (NRG2). The levels of p75-CTF were similar in all I/I and III/III rat tissues tested, whereas p75-ICD showed significantly reduced levels only in the olfactory bulb of the I/I rats (1.8-fold \( P < 0.05 \)). The olfactory bulb was also the only tissue with significantly reduced ErbB4-ICD levels when comparing I/I and III/III rat tissues (1.3-fold \( P < 0.05 \)). The CTF levels of NRG2 were similar in all tissues examined.

**CONCLUSIONS AND SIGNIFICANCE**

The γ-secretase enzyme complex requires at least four protein components to display cleavage activity: presenilin (PS-1 or -2), nicastrin, PEN-2, and Aph-1 (Aph-1aS, -1aL, or -1b). We examined the mRNA expression levels of \( \text{Aph-1aS} \) and \( \text{Aph-1b} \) and the effects of the differential \( \text{Aph-1b} \) expression on the cleavage activity of the γ-secretase complex in rats with one or three \( \text{Aph-1b} \) gene copies (I/I and III/III rats, respectively). In III/III rat tissues, ratios between the \( \text{Aph-1b} \) and \( \text{-1a} \) mRNA levels varied greatly. A reduction in \( \text{Aph-1b} \) gene copy numbers in the I/I rats was accompanied by a large drop in \( \text{Aph-1b} \) mRNA expression levels, not compensated for by its paralog \( \text{Aph-1a} \).

**Figure 1.** Northern blot analysis of \( \text{Aph-1a} \) and \( \text{-1b} \) mRNA expression in various tissues of I/I and III/III rats. The \( \text{Aph-1a} \) probe detected both the short and long isoform (\( \text{Aph-1aS} \) and \( \text{-1aL} \), respectively). The \( \text{Aph-1b/-1a} \) ratio was determined for every III/III rat tissue and categorized as high (\( \text{Aph-1b/-1a} > 2.0; \) +++) , moderate (\( \text{1.0} < \text{Aph-1b/-1a} < 2.0; \) ++) or low (\( \text{Aph-1b/-1a} < 1.0; \) +). Tissues used were cerebellum (cer), olfactory bulb (olf), cortex (ctx), hippocampus (hip), striatum (str), (hypo)thalamus (tha), pons/medulla (p/m), spinal cord (spc), eye, testis (tes), heart, (hrt), muscle (msc), stomach (sto), small intestine (s.i.), large intestine (l.i.), lung (lng), spleen (spl), liver (liv), thymus (thy), and pancreas (pan) of PND9 III/III (3) or I/I (1) rats.

**Figure 2.** Western blot analysis of the γ-secretase cleavage products derived from the APP, APLP1 and APLP2 proteins in various tissues of I/I and III/III rats. A) Levels of C-terminal fragments (CTFs) of amyloid-β precursor protein (APP), and the APP-like proteins APLP1 and APLP2 (sizes of all 3 CTFs ~10 kDa) were analyzed in neuronal tissues of PND13 I/I (1) and III/III (3) rats using specific antibodies. Tissues used were the olfactory bulb (high \( \text{Aph-1b/-1a} \) ratio), spinal cord, and cortex (moderate ratio), and (hypo)thalamus and cerebellum (low ratio). B) Levels of APP-CTF, APLP1-CTF and APLP2-CTF were analyzed in the lung (high \( \text{Aph-1b/-1a} \) ratio), spinal cord, and cortex (moderate ratio), and (hypo)thalamus and cerebellum (low ratio) of PND13 I/I and III/III rats. In each case, tubulin (~55 kDa) was used for normalization. Bars represent quantifications in arbitrary units of normalized CTF signals of 5 tissue samples with the average level in III/III rat tissues set to 1. The levels of the APP, APLP1 and APLP2 holoproteins were similar in the I/I and III/III rat tissues. *\( P < 0.05 \); **\( P < 0.02 \); ***\( P < 0.01 \); \( n = 5 \), with the 5 rats per genotype from different nests; plus sem; BD: below detection.
resulted in substrate- and tissue-specific alterations in β-secretase cleavage activity (Fig. 3). Tissues that normally have a high Apo-1b/1a ratio displayed clear differences in β-secretase cleavage activity, whereas the activity in tissues with a low ratio was either hardly affected or not at all. Within a particular tissue, substrate processing was not affected to the same extent, showing that γ-secretase complexes with different subunit compositions are not functionally redundant and that a specific complex is involved in the preferential cleavage of a subset of γ-secretase substrates.

We earlier found that the Apo-1b rat genotypes segregated with behavioral phenotypes, and rats with a natural Apo-1b knockdown (APO-SUS) display alterations in brain information processing (prepulse inhibition and latent inhibition), locomotor activity in response to novelty, fleeing, and problem-solving behavior, and HPA-axis response to stress. The APO-SUS phenotype partially overlaps with the behavioral phenotypes of a number of mouse models with altered expression of a γ-secretase substrate. For example, heterozygous ErbB4 or NRG1 knockout mice (with an overall ~50% reduction of mRNA expression) are also hyperactive in an open field and show an impaired prepulse inhibition, whereas reduced expression of Notch has no effect on the open field behavior. Conversely, overexpression of human APP751 or APP-CTF caused a general hypoactivity in mice.

In conclusion, the differential expression of Apo-1b in the I/I and III/III rats caused subtle, substrate-specific alterations in β-secretase cleavage activity toward a number of β-secretase substrates, especially in tissues with a relatively high Apo-1b level (Fig. 3). Thus, a single gene defect may affect a great variety of (neuro)developmental signaling pathways, resulting in a complex phenotype that is generally thought to have a multigenic origin. Furthermore, the γ-secretase complex, generally known because it is linked to Alzheimer’s disease (a neurodegenerative and ageing disorder), may also be associated with (neuro)developmental disorders that become apparent much earlier in life.