TRIAZOLE-LINKED GLYCOAMINO ACIDS AND GLYCOPEPTIDES

A~N~N~N~B

(1)

B~N~N~N~A

(1')

Abstract: The invention concerns staff analogues of glycoamino acids and glycopeptides in which the non-sugar acetal heteroatom is replaced by a triazole moiety e.g. compounds of formula (I) or (I') wherein A represents a carbohydrate which is coupled via its anomeric center to the triazole ring and B represents an amino acid residue.
Triazole-linked glycoamino acids and glycopeptides

FIELD OF THE INVENTION

The present invention concerns stable analogues of glyco-amino acids and glycopeptides. The compounds of the invention are of use in for example medical applications, nutritional applications as well as research tools.

BACKGROUND OF THE INVENTION

Glycoproteins are widely distributed in all forms of life, with the possible exception of the eubacteria. They occur in cells, both in soluble and membrane-bound forms, as well as in the extracellular matrix and in extracellular fluids. The most common glycoproteins are those in which the carbohydrate is linked to the protein by glycosyl linkages. Glycosylation represents one of the important co-translational and post-translational modifications of proteins.

In naturally occurring glycoproteins and in glycopeptide hormones, carbohydrate moieties play key roles in intercellular and intracellular transport of gene products (exit passport hypothesis), as well as extending the biological half-life of the active peptides in vivo (proteolytic protection). Additional roles supported by experimental evidence include the alteration of peptide backbone conformation (protein folding), control of membrane permeability, and molecular recognition (the concept of carbohydrate "antennae").

Thus it becomes increasingly more apparent that glycoproteins and -peptides play important roles in organisms. For example abnormalities in O-linked glycopeptides are implicated in numerous disease states. Abnormal post-translational modification of the tau protein has been implicated in the formation of neurofibrillary tangles of Alzheimer's disease. The antigenic T-epitopes and TN-epitopes of cell-surface glycopeptides have long been associated with cancer and used as tumor cell markers.

Therefore there is a need for glycopeptides that can be of use as research tools, but also for glycopeptides that can be of use in medicaments in case of glycopeptide related disorders. The chemical synthesis of glycopeptides provides an important tool for the
study of glycopeptide hormones, glycoproteins and other complex carbohydrate structures found at the cell surface and in the glycocalyx. However, the synthesis and potential applications of glycopeptides are complicated due to the sensitivity toward chemical and enzymatic hydrolysis of the glycosidic bond linking the carbohydrate to the peptide. In this respect in a review in Trends in Glycoscience and Glycotechnology, vol 13, no. 69, 2001, pp11-30, Mizuno concludes that most methods for glycopeptide synthesis are cumbersome and give unsatisfactory yields. The same picture arises from a review by Dondoni and Marra in Chem. Rev. 2000, vol 100, pp4395-4421 in which in view of the key role of oligosaccharides in glycoprotein biological activity such as intercellular trafficking and receptor binding and signalling on one side and the enzymatic degradation via the hydrolysis of the O- or N-glycosidic bonds on the other side, the focus for preparing chemically and metabolically resistant analogues is on the incorporation of C-glycosyl amino acids in glycopeptides.

What is needed in the art are stable analogues of glycopeptides that are readily accessible preferably via chemical synthesis. This can be translated in a need for stable analogues of amino acid glycosides, hereinbelow also referred to as glyco-amino acid, which can be used to make glycopeptides.

Calvo-Flores et al in Organic Letters (2000), vol 2, pp2499-2502 describe the synthesis of carbohydrate conjugates by means of the 1,3-dipolar cycloaddition reaction. Carbohydrate moieties having a terminal alkyne attached to the anomic centre through an ether bond are reacted with azide derivatives. Bröder et al. in Carbohydrate Research (1993), vol 249, pp221-241 describe glycosyl azides as reactants for glycosylation reactions. In particular the azide group is converted to a triazole moiety by means of a cycloaddition reaction with an alkyne moiety. Then, in the resulting conjugate the triazole group functions as a leaving group for the introduction of a fluoride, thereby demonstrating the implicit instability of the triazole conjugate.

In Tornøe et al., J.Org.Chem. (2002) 67, 3057-3064 the Cu-catalyzed cycloaddition reaction between terminal alkynes and azides for the preparation of peptide derivatives is disclosed. Peptides and amino acids are disclosed which are conjugated through a carbohydrate moiety through a triazole linker. In particular peptidotriazoles linked to 2-
deoxygalactose have been described. No glyco-peptide conjugates linked through a triazole via the anomic center of the carbohydrate are disclosed.

US 6,664,399 discloses oligosaccharides having a triazole linkage between the saccharide units as well as methods for their production involving glycosyl azides as reactants. In view of the statements by the inventors that the compounds as disclosed are expected to be synthesized by more general methods than ether-oxygen linked carbohydrates, are expected to be more stable to enzymatic and chemical hydrolysis and are expected to be amenable to automated synthesis methods, the supposed advantages in terms of ease of preparation and improved stability compared to oligosaccharides having standard glycosidic bonds are merely speculative, let alone that they can only be read in connection with the preparation of oligosaccharides and not in relation to the preparation of glycopeptides. Actual data on the stability of the triazole-linked oligosaccharides is absent and there is no suggestion to apply the process of the preparation of triazole-linked oligosaccharides to any other compound than carbohydrates.

Thus altogether, triazoles, that are substituted with amino acids or peptides, and that are linked to the anomic carbon in carbohydrates are strikingly absent in the art.

SUMMARY OF THE INVENTION

The present invention provides stable analogues of glyco-amino acids by compounds of the following formula I and I’

\[
\begin{align*}
\text{I} & : & \text{A} & - \text{N} \equiv \text{N} - \text{N} - \text{B} \\
\text{I’} & : & \text{B} & - \text{N} \equiv \text{N} - \text{N} - \text{A}
\end{align*}
\]

wherein A represents a carbohydrate which is coupled via its anomic center (I) or parent anomic center (I’) to the triazole ring and B represents an amino acid or a mimetic of an amino acid
In a further aspect the present invention provides stable analogues of glycopeptides by compounds of the following formula II and II'

\[
\begin{align*}
\text{A} & \sim \text{N} \sim \text{N} \sim \text{N} \sim \text{C} \\
\text{II} \\
\text{C} & \sim \text{N} \sim \text{N} \sim \text{A} \\
\text{II'}
\end{align*}
\]

wherein A represents a carbohydrate which is coupled via its anomeric center (II) or parent anomeric center (II') to the triazole ring and C represents a polypeptide or a mimetic of a polypeptide.

In one embodiment the invention concerns compounds I and/or II. In one embodiment the invention concerns compounds I' and/or II'.

In terms of definition B may also be defined as one selected from the group consisting of an amino acid, a mimetic of an amino acid and C, wherein C is selected from the group consisting of a polypeptide and a mimetic of a polypeptide.

In the context of this invention the anomeric center means the central position (atom) in a (hemi)acetal, and a parent anomeric center is the carbon atom that was the central position (atom) in a (hemi)acetal in the parent carbohydrate.

**DETAILED DESCRIPTION OF THE INVENTION**

In prominent classes of naturally occurring glycoproteins and glycopeptides the hemiacetal group of a carbohydrate is linked to an amino acid in a polypeptide via an alcohol functional group or amine functional group in the amino acid resulting in an O-glycosidic bond (O,O-acetal) or N-glycosidic bond (N,O-acetal). The present invention is based on the surprising insight that such a type of linkage can be rendered stable by replacing the non-sugar acetal heteroatom by a triazole moiety. The resulting carbohydrate-triazole-amino acid or - (poly)peptide compound can be readily synthesised under mild conditions and proved to be surprisingly stable, in particular
stable compared to the otherwise readily hydrolysable glycosidic bond linking carbohydrate and peptide, and is suitable for a variety of applications.

In the context of this invention a glycopeptide is a compound containing a carbohydrate (or glycan) covalently linked via its (parent) anomic center to a peptide (or mimetic) composed of amino acids (or mimetics). The carbohydrate may be in the form of a protected or unprotected monosaccharide, disaccharide(s), oligosaccharide(s), polysaccharide(s), or their derivatives (e.g. sulfo- or phospho-substituted). One, a few, or many carbohydrate units may be present.

Generally speaking, chemical synthesis of N-glycopeptides can be approached in two general manners: 1) by incorporation of preformed glycosyl amino acids in solid phase peptide synthesis (SPPS) and 2) by modification of aspartic acid with carbohydrates after SPPS. Although excellent results have been obtained in the synthesis of N-glycopeptides with preformed glycosyl amino acids, the synthesis of the corresponding building blocks is time-consuming and cumbersome. Therefore, it is more advantageous to utilise a coupling of glycosylamine to aspartic-acid-containing peptides because they are potentially faster and offer a higher degree of convergence. Unfortunately, such an approach is plagued by a severe side reaction i.e. the formation of aspartimides, which upon hydrolysis give rise to peptides linked through the alpha- and beta-carboxygroup. This side-reaction can be bypassed by use of a backbone amide protective group but obviously additional chemistry and steps are necessary in such an approach.

Synthesis of triazole-linked glycopeptides circumvents all of these disadvantages, since the triazole can be introduced after completion of the peptide synthesis without concomittant side-reactions. An additional advantage lies in the application of protected acetylene-containing amino acids during the peptide synthesis which enables the regioselective introduction of different sugars at different locations after selective deprotection of the desired acetylene.

From literature compounds having a triazole linked to the anomic center of a carbohydrate are known, see De Las Heras et al. J.Med.Chem. (1979), 22, 496-500 and
Harmon et al. J.Chem.Soc.D (1971) 296. However, these compounds are scarce and limited in diversity, which is probably due to the harsh conditions needed for their formation. This has limited the applicability of this compound class over the years. Recently, novel mild metal-catalysed procedures for the formation of triazoles have been reported. However, these procedures employ Lewis acid catalysts, which in the light of the fact that benzotriazoles connected to acetalic carbons can readily act as a leaving group under mild (Lewis)acidic conditions (see for example Veerman et al. Eur.J.Org.Chem. (2002) 18, 3133-3139, Deniau et al. Tetrahedron Lett. (2002) 43, 8055-8058 and Katrizky et al. J.Org.Chem. (2002) 67, 8239-8242). The combination of this property and a reactive anomeric carbon atom was not expected to be viable. Yet further, in figure 7 of WO 03/101972 a compound is depicted that comprises carbohydrate units to which a substituted triazole is linked to the 2-deoxy position of the carbohydrate. As already mentioned in Tornøe et al., (supra) the triazole is also only linked to the 2-position of the carbohydrate. Similarly substituted triazoles linked to the anomic carbon in carbohydrates however are strikingly absent in the art.

A peptide in the context of this invention may either be an oligopeptide, comprising 2-10 amino acids, or a polypeptide, comprising more than 10 amino acids. The amino acids in the compounds of the invention and the amino acids in a peptide in the compounds of the invention may be D- or L-amino acids and include non-proteogenic amino acids. A mimic of an amino acid is a compound that mimics the structure of an amino acid and that is suitable to be incorporated into an oligomer or polymer of amino acids thereby forming a peptide comprising a single mimetics of an amino acid or multiple mimetics of amino acids or consisting substantially in total of mimetics of amino acids. The latter is referred to as a mimetic of a polypeptide. In the context of this invention mimetics of amino acids are for instance compounds selected from the group consisting of β2 amino acids, β3 amino acids, α,α-disubstituted amino acids, β-amino sulfonamides, α-amino, γ-carboxy acids, α-hydroxy acids, α-amino nitriles. In one embodiment C in structure II or II' comprises 2-100 amino acids, preferably 2-50, more preferably 2-20 or preferably 10-100, more preferably 10-50, even more preferably 10-30 amino acids.
A glyco-amino-acid is a carbohydrate attached via its (parent) anomeric center to a single amino acid by any kind of covalent bond. Commonly occurring glyco-amino-acids are (see Sharon, N. & Lis, H. (1982) in The proteins, 3rd edn (Neurath, H. & Hill, R. L., eds) vol. 5, pp. 1-144, Academic Press, New York) β-N-acetylglicosaminylasparagine ((GlcNAc-)Asn) which is widely distributed in animals, plants and micro organisms, α-N-acetylgalactosaminylerine or -threonine ((Gal-NAc)Ser, (GalNAc-)Thr) which occur in glycoproteins of animal sources, β-xylosylserine ((Xyl-)Ser) occurring in proteoglycans, human thyroglobulin, β-galactosylhydroxylysine ((Gal-)Hyl) occurring in collagens and β-L-arabinosylhydroxyproline ((L-Ara-)Hyp) which occur in plant and algal glycoproteins.

In the context of this invention a glyco-amino acid is a carbohydrate linked to an amino acid via a triazole moiety. According to the invention the (parent) anomeric carbon atom of the carbohydrate is linked to the triazole moiety.

The amino acid may be linked via its amine group, via its acid group or via its side chain. In a preferred embodiment the amino acid is linked to the triazole moiety via its side chain, leaving the amine group and the acid group intact and free to form amide bonds in a peptide chain.

In the context of this invention the generic term 'carbohydrate' includes monosaccharides, disaccharides, oligosaccharides and polysaccharides as well as substances derived from monosaccharides by reduction of the carbonyl group (alditols), by oxidation of one or more terminal groups to carboxylic acids, or by replacement of one or more hydroxy group(s) by a hydrogen atom, an amino group, a thiol group or similar heteroatomic groups. It also includes derivatives of these compounds. The term 'sugar' is frequently applied to monosaccharides and lower oligosaccharides.

Parent monosaccharides are polyhydroxy aldehydes H-[CHOH]ₙ-CHO or polyhydroxy ketones H-[CHOH]ₙ-CO-[CHOH]ₙ-H with four or more carbon atoms. The generic term 'monosaccharide' (as opposed to oligosaccharide or polysaccharide) denotes a single unit, without glycosidic connection to other such units. It includes aldoses, dialdoses, aldotoses, ketoses and diketoses, as well as deoxy sugars and amino
sugars, and their derivatives, provided that the parent compound has a (potential) carbonyl group.

The carbohydrate in the compounds of the invention may be similar to that occurring in natural glycoproteins and peptides, which are comparatively small, optionally branched, usually unsulfated carbohydrate units, often devoid of repeating units. In one embodiment the carbohydrate is in the form of oligosaccharides, linear or branched, containing up to about 20 monosaccharide residues; in a further embodiment the compounds of the invention contain linear mono-, di-, tri-, tetra-, penta-, hexa-, hepta-, octa-, nona-, or decasaccharide units, preferably mono-, di-, tri- or tetrasaccharide units, more preferably tetrasaccharide units; in yet a further embodiment the carbohydrates may contain or consist of repeating units, such as for instance repeating units of N-acetyllactosamine.


To prepare the novel glyco-amino-acids and glycopeptides the following cycloaddition reaction partners are suitable: a carbohydrate bearing an azide group and an amino acid or peptide bearing an acetylene group or alternatively a carbohydrate bearing an acetylene group and an amino acid or peptide bearing an azide group. Methods for the preparation of each of these cycloaddition partners are well known in the art and are described for instance in Shiozaki et al. Chem. Lett., 1996, 9, 735-736; Horton, D. Methods in Carbohydrate Chemistry; Whistler, R. L., BeMiller, J. N., Eds. Academic Press: New York, 1972; Vol. VI, pp 282-285; Lehnhoff et al. Angew. Chem. 1995, 107, 1208-1211.

Suitable amino acids bearing an acetylene group are for example:

\[
\begin{align*}
R^2 & R^3 & (\equiv) & n \\
R^1 & & &
\end{align*}
\]

wherein \(X\) represents \(\text{CN}, \text{CO}_2 R^4, \text{COR}^5, \text{SO}_2 R^6, \text{CH}_3 \text{OR}^7, \text{CH}_2 \text{NR}^8 R^9, \text{CR}^{10} R^{11}, \text{CONR}^{12} \text{NR}^{13} R^{14}\);

\(R^1, R^2, R^3, R^4, R^5, R^6, R^7, R^8, R^9, R^{10}, R^{11}, R^{12}, R^{13}\) and \(R^{14}\) may be the same or different, and each independently represent \(H\), linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, \(n = 1-10\).

Suitable amino acids bearing an azide group are for example:

\[
\begin{align*}
R^2 & R^3 & N & (\equiv) & n \\
R^1 & & &
\end{align*}
\]
wherein X represents CN, CO₂R⁴, COR⁵, SO₂R⁶, CH₂OR⁷, CH₂NR⁸R⁹, CR¹⁰R¹¹, CONR¹²NR¹³R¹⁴;
R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹², R¹³ and R¹⁴ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyle, optionally substituted aryl, and amine and/or acid protective groups, n = 1-10.

and

\[
\begin{array}{c}
N_3 \\
R^2 \\
N \\
R^1
\end{array}
\]

wherein X represents CN, CO₂R³, COR⁴, SO₂R⁵, CH₂OR⁶, CH₂NR⁷R⁸, CR⁹R¹⁰, CONR¹¹NR¹²R¹³;
R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹² and R¹³ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyle, optionally substituted aryl, and amine and/or acid protective groups, the ring may be saturated or (partly) unsaturated and may be 4-10 membered; the azide can be in any position on the ring.

The amino acids may also be part of a peptide, in which case in the structures given above X may represent the site of attachment to one or more further amino acids, and/or R¹ and/or R² may also represent the site of attachment to one or more further amino acids.

The amino acids may be in an optically active form, and can be either in the R or S configuration at any stereocenter.

The amino group and/or the acid group in the amino acids may be suitably protected in order to facilitate further reaction steps.
Preferably the amino acid bears the azide or acetylene group in the side chain leaving for instance the amine and/or acid functionalities intact and free to form amide bonds in a peptide chain.

5 Of particular interest are the amino acids bearing azide or acetylene groups that closely resemble the amino acids in naturally occurring glycopeptides that are linked to carbohydrates. Thus in particular serine, threonine, lysine, hydroxylsine, asparagine and hydroxyproline that bear an azide or acetylene group, or in which, compared to the natural amino acid, an OH or NH₂ functional group is modified into an azide or acetylene group are of interest.

Any saccharide unit in a carbohydrate as described above may contain either an azide or acetylene group at the (parent) anomeric position. Preferably the azide or acetylene is in a terminal saccharide unit at the (parent) anomeric position of the saccharide unit.

Thus, suitable carbohydrates bearing an acetylene group are:

\[
\begin{array}{c}
\text{RO} \\
m \\
\text{OR} \\
\end{array}
\]

wherein \( R^1 \) may be \( \text{H}, \text{CH}_2\text{OR} \) or \( \text{CHORCH}_2\text{OR} \), \( R^2 \) may be \( \text{H}, \text{NHR} \) or \( \text{OR} \), \( R \) may be the same or different within one molecule and may be \( \text{H} \), a saccharide unit or a protective group, \( m = 0 \) or 1.

Suitable carbohydrates bearing an azide group are:

\[
\begin{array}{c}
\text{RO} \\
m \\
\text{OR} \\
\end{array}
\]
wherein \( R^1 \) may be H, CH\(_2\)OR or CHORCH\(_2\)OR, \( R^2 \) may be H, NHR or OR, R may be the same or different within one molecule and may be H, a saccharide unit or a protective group, \( m = 0 \) or \( 1 \).

Thus, suitable carbohydrates bearing an acetylene or azide on the anomeric carbon are derived from allo-, altro-, gluco-, manno-, gulo-, ido-, galacto-, talo-, ribo-, arabinio-, xylo- and lyxofuranoside and from allo-, altro-, gluco-, manno-, gulo-, ido-, galacto-, talo-, ribo-, arabinio-, xylo- and lyxopyranoside.

The carbohydrates can be either in the D or L configuration.

The cycloaddition of the azide and acetylene reaction partners can proceed according to any of the methods disclosed in the literature cited above. A series of advantageous conditions for the cycloaddition reaction is also disclosed in WO 03/101972. The cycloaddition results in substituted-[1,2,3]-triazoles. The cycloaddition may result in two possible regioisomers, viz. the 1,4- and 1,5-substituted triazole. The 1,4-substituted product is preferred, which also will be the most abundant, if not sole, regioisomer under standard cycloaddition conditions.

In one embodiment the invention concerns a method for the preparation of a glyco-amino acid or glycopeptide by reacting a carbohydrate having an azide group on the anomeric carbon as described above with an amino acid or peptide comprising an amino acid having an acetylene group as described above.

In one embodiment the invention concerns a method for the preparation of a glyco-amino acid or glycopeptide by reacting a carbohydrate having an acetylene group on the (parent) anomeric carbon as described above with an amino acid or peptide comprising an amino acid having an azide group as described above.

Cycloaddition of an azide or acetylene bearing amino acid with a carbohydrate reaction partner, respectively an acetylene or azide bearing carbohydrate, results in a glyco-amino-acid of the invention.
Such a glyco-amino acid may be used as a building block in the preparation of a glycopeptide according to the invention. For instance one or more of such glyco-amino acids may be used in an automated peptide synthesiser. Alternatively one or more acetylene bearing amino acids, or azide bearing amino acids may be used as building blocks for the preparation of a peptide. During the peptide synthesis, as the chain of amino acids increases, or after the synthesis of a desired peptide has been completed, the peptide having azide or acetylene functional groups may be reacted with a carbohydrate cycloaddition reaction partner in order to obtain a glycopeptide according to the invention.

Thus in an embodiment the invention relates to a method for the preparation of a glycopeptide of the invention, said method comprising reacting a glyco-amino acid of the invention as a building block in a peptide synthesis, such as a peptide synthesis on a solid support, preferably in an automated peptide synthesiser.

In another embodiment the invention relates to a method for the preparation of a glycopeptide of the invention, said method comprising reacting an azide bearing amino acid as described above and/or an acetylene bearing amino acid as described above as a building block in a peptide synthesis, such as a peptide synthesis on a solid support, preferably in an automated peptide synthesiser, and reacting azide functional groups in the resulting peptide during peptide synthesis or after peptide synthesis is completed with a carbohydrate having an acetylene group on the (parent) anomeric carbon as described above and/or reacting acetylene functional groups in the resulting peptide during peptide synthesis or after peptide synthesis is completed with a carbohydrate having an azide group on the anomeric carbon as described above.

Thus a glycopeptide of the invention may comprise one, two, three, four or more carbohydrates attached to different amino acids in the peptide chain. The glyco-amino acid can be at any position in the peptide chain.

The methods described above may require appropriate protective group manipulations on carbohydrate, amino acid and/or peptide. Such protective group manipulations are well known in the art and suitable protective groups can be found in e.g. Greene,

Glycopeptides may be used for the treatment of bacterial infections. Therefore, in another embodiment, the present invention is directed to a method for controlling a bacterial infection in a host animal, typically a warm-blooded animal, which comprises administering to the host animal an effective, antibacterial amount of a compound of the present invention. In this embodiment, the compounds can be used to control and treat infections due to various bacteria, but especially gram-positive bacteria. In a preferred embodiment, the compounds are used to control and treat infections due to bacteria resistant to existing antibacterials. For example, certain bacteria are resistant to methicillin, and yet others are resistant to vancomycin and/or teicoplanin. The present compounds provide a technique for controlling and treating infections due to such resistant bacterial species.

In carrying out this embodiment of the invention, the compounds of the present invention can be administered by any of the conventional techniques, including the oral route and parenteral routes such as intravenous and intramuscular. The amount of compound to be employed is not critical and will vary depending on the particular compound employed, the route of administration, the severity of the infection, the interval between dosings, and other factors known to those skilled in the art. In general, a dose of from about 0.5 to about 100 mg/kg will be effective; and in many situations, lesser doses of from about 0.5 to about 50 mg/kg will be effective. A compound of the present invention can be administered in a single dose, but in the known manner of antibacterial therapy, a compound of the present invention is typically administered repeatedly over a period of time, such as a matter of days or weeks, to ensure control of the bacterial infection.

Also in accordance with known antibacterial therapy, a compound of the present invention is typically formulated for convenient delivery of the requisite dose. Therefore, in another embodiment, the present invention is directed to a pharmaceutical formulation comprising a compound of the present invention, in combination with a pharmaceutically-acceptable carrier. Such carriers are well known for both oral and parenteral routes of delivery. In general, a formulation will comprise a compound of the
present invention in a concentration of from about 0.1 to about 90% by weight, and often from about 1.0 to about 3%. Thus a pharmaceutical composition comprising a compound according to the present invention and a nutritional composition comprising a compound according to the present invention are further embodiments of this invention, both in combination for that purpose with suitable carriers.

Saccharides covalently attached to proteins determine their location or destination in the cell and the metabolic routes they are supposed to follow. Glycoproteins are involved in cell-recognition and thus are involved in signalling processes, in cell-cell interactions, and immunological responses. Thus the compounds of the present invention may be used as diagnostic tools, in vitro and/or in vivo as well as research tools in vitro to study for instance signalling pathways etc. Thus in a further embodiment the present invention concerns a kit of parts that comprises as a research tool and/or diagnostic tool a compound according to the present invention and optionally comprising further reagents, diluents and/or handling instructions.

Biological evaluation of triazole-linked glycopeptides

Biosynthesis of N-glycoproteins

Glycosyltransferases in the lumen of the endoplasmatic reticulum (ER) are responsible for the transfer of a 14-mer oligosaccharide from dolichol-diphosphate to a growing protein chain. After that, the oligosaccharide part is trimmed by glycosidases and mannosidases to a 10-mer oligosaccharide, before transportation to the Golgi occurs. In the Golgi, further mannose trimming eventually leads to a core hexasaccharide which is subsequently decorated with sugars to larger N-glycans. When biosynthesis in the Golgi is complete, the entire glycoprotein is exported to the cytoplasm. In the cytoplasm, a variety of glycosidases or glycosyltransferases can further modify the glycoprotein. For example, endo-β-N-acetylglucosaminidase is able to transfer an oligosaccharide from or to the first glucosamine attached to asparagine in the protein.

Glycoamidase

An enzyme that releases intact oligosaccharides from N-glycoproteins by cleaving the β-aspartylglucosylamine amide linkage is called a glycoamidase (also N-glycanase,
peptide N-glycanase or glycopeptidase). This enzyme has been isolated from insects, plants and animal sources, and has also been found in human cytomegalovirus infected cells and human tumor cell lines. The wide occurrence of glycoamidase strongly suggests that protein N-deglycosylation mediated by the enzyme may be a universal feature in living organisms as a functionally important mechanism of regulation.

Assay for glycoamidase stability determination
Several glycoamidases are available commercially, such as glycoamidase A from almond emulsion and glycoamidase F from *Flavobacterium meningosepticum*. To examine if a particular N-glycoprotein or triazole-glycoprotein can be hydrolyzed by a glycoamidase, the substrate can be incubated with the enzyme at various temperatures or concentrations. In a typical setup, a substrate and a glycoamidase are incubated in a total volume of 50 μL of 40 mM ammonium acetate buffer (pH 5 for glycoamidase A and pH 8 for glycoamidase F). The reaction can be followed by TLC or RP-HPLC. After the conversion is complete, the reaction can be stopped by boiling in a water bath for 3 min and the products can be separated with RP-HPLC for analysis.

Application of triazolylglycopeptides
Apart from being enzymatically and chemically stable analogs of naturally occurring N-glycoproteins, the triazole technology may be conveniently applied for the glycosylation of therapeutic peptides which do NOT contain a carbohydrate naturally. In this approach the carbohydrate moiety does not fulfil an active biological role as pharmacophore or to influence protein conformation, but rather to optimize pharmacokinetic properties of the protein with retention of activity and selectivity. For example, pharmacokinetic studies of glycosylated RGD-peptides have demonstrated that uptake in the liver was significantly reduced compared to that of the non-glycosylated compound, the initial concentration in the blood was doubled and the accumulation of the tracer in tumor tissues was dramatically enhanced.
EXAMPLES

Glucopyranosyl-triazolyl-L-alanine

The following conditions are generally applicable to other cycloaddition reaction partners.

\[
\begin{align*}
\text{AcO} & \quad \text{AcO} \quad \text{N}_3 \\
\text{AcO} & \quad \text{AcO} \quad \text{OAc} \\
\end{align*}
\]

\[
\begin{align*}
\text{BocHN} & \quad \text{CO}_2\text{Me} \\
\end{align*}
\]

20 mol% CuI
40 mol% DIPEA

THF, rt, 16 h

1

2

3

To a solution of tetra-O-acetyl-β-D-glucopyranosyl azide (1) (36.3 mg, 0.10 mmol) and N-Boc-L-propargylglycine methyl ester (2) (26.3 mg, 0.12 mmol) in THF (1 mL) was added CuI (3.7 mg, 20 mol%) and DIPEA (6.4 µL, 40 mol%). The mixture was stirred for 16 hours, after which water (1 mL) was added and the product was extracted with DCM (2×). The combined organic layers were washed with a saturated aqueous solution of NaHCO₃ and brine, dried over Na₂SO₄ and concentrated in vacuo. The product was purified by flash column chromatography (EtOAc/heptane 1:2) to afford the product as a white solid (51.8 mg, 0.09 mmol, 89%).

In a similar method the following cycloaddition reaction partners have been reacted to give the 1,4-triazole glyco amino acids and peptides:
Table 1

<table>
<thead>
<tr>
<th>entry</th>
<th>saccharide</th>
<th>glycopeptide</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td><img src="saccharide_1.png" alt="image" /></td>
<td><img src="glycopeptide_1.png" alt="image" /> 98%</td>
</tr>
<tr>
<td>2</td>
<td><img src="saccharide_2.png" alt="image" /></td>
<td><img src="glycopeptide_2.png" alt="image" /> 88%</td>
</tr>
<tr>
<td>3</td>
<td><img src="saccharide_3.png" alt="image" /></td>
<td><img src="glycopeptide_3.png" alt="image" /> 87% (α:β 1:4)</td>
</tr>
<tr>
<td>4</td>
<td><img src="saccharide_4.png" alt="image" /></td>
<td><img src="glycopeptide_4.png" alt="image" /> 88%</td>
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<tr>
<td>5</td>
<td><img src="saccharide_5.png" alt="image" /></td>
<td><img src="glycopeptide_5.png" alt="image" /> 72%</td>
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<tr>
<td>6</td>
<td><img src="saccharide_6.png" alt="image" /></td>
<td><img src="glycopeptide_6.png" alt="image" /> 30%</td>
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Table 2

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<th>Entry</th>
<th>Amino Acid</th>
<th>Glycopeptide</th>
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<tr>
<td>1</td>
<td>Fmoc</td>
<td>70%</td>
</tr>
<tr>
<td>2</td>
<td>Ts</td>
<td>64%</td>
</tr>
<tr>
<td>3</td>
<td>Boc</td>
<td>70%</td>
</tr>
<tr>
<td>4</td>
<td>Boc</td>
<td>76%</td>
</tr>
<tr>
<td>5</td>
<td>Boc</td>
<td>85% (R:S 1:1)</td>
</tr>
<tr>
<td>6</td>
<td>Boc</td>
<td>60%</td>
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Table 3

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<th>dipeptide</th>
<th>glycopeptide</th>
<th>yield</th>
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<tbody>
<tr>
<td>1</td>
<td>Boc-Ala-NH&lt;sub&gt;H&lt;/sub&gt;CO₂Me</td>
<td>AcO-O-O-N&lt;sub&gt;3&lt;/sub&gt;</td>
<td>51%</td>
</tr>
<tr>
<td>2</td>
<td>Boc-Pro-NH&lt;sub&gt;H&lt;/sub&gt;CO₂Me</td>
<td>AcO-O-O-N=NN&lt;sub&gt;MeO&lt;/sub&gt;₂C&lt;sub&gt;H&lt;/sub&gt;₂N&lt;sub&gt;H&lt;/sub&gt;</td>
<td>92%</td>
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Table 4

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<th>amino acid</th>
<th>glycopeptide</th>
<th>yield</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>N&lt;sub&gt;H&lt;/sub&gt;CO₂Me</td>
<td>AcO-O-O-N&lt;sub&gt;3&lt;/sub&gt;</td>
<td>71%</td>
</tr>
<tr>
<td>2</td>
<td>Boc-Ala-NH&lt;sub&gt;H&lt;/sub&gt;CO₂Me</td>
<td>AcO-O-O-N=NN&lt;sub&gt;MeO&lt;/sub&gt;₂C&lt;sub&gt;H&lt;/sub&gt;₂N&lt;sub&gt;H&lt;/sub&gt;</td>
<td>81%</td>
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Table 5

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<tr>
<th>entry</th>
<th>amino acid</th>
<th>n</th>
<th>R</th>
<th>glycopeptide</th>
<th>yield (%)</th>
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<td>1</td>
<td>1</td>
<td>H</td>
<td>5</td>
<td>70%</td>
</tr>
<tr>
<td>2</td>
<td>2</td>
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<td>H</td>
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<td>73%</td>
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<tr>
<td>3</td>
<td>3</td>
<td>3</td>
<td>H</td>
<td>7</td>
<td>71%</td>
</tr>
<tr>
<td>4</td>
<td>4</td>
<td>1</td>
<td>Me</td>
<td>8</td>
<td>84%</td>
</tr>
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</table>

5 Table 6

Synthesis of peptide using glyco-amino acids

First route: To a solution of N-Boc-glycoamino acid (1 equiv) in CH₂Cl₂ (0.1 M) were added H₂N-ΑΑ-ΟMe (1 equiv), DIPEA (1.2 equiv) and EDCI (1.1 equiv) at 0 °C. The reaction mixture was stirred for 16 hours, after which water was added. The product was extracted with DCM (2x). The combined organic layers were washed with a aqueous HCl (0.1 M), a saturated aqueous solution of NaHCO₃ and brine, dried over Na₂SO₄ and concentrated in vacuo. The product was purified by flash column chromatography.

Second route: An amino acid immobilized on a solid support was suspended in DMF and treated with N-Fmoc-glycoamino acid (1.5 equiv), DIPEA (3 equiv) and EDCI (3
equiv) at 0 °C. The reaction mixture was agitated for 16 hours, after which the suspension was filtrated. The resin was washed several times with CH₂Cl₂ and MeOH and dried in vacuo. The resin was suspended in a mixture of CH₂Cl₂ and TFA (1:1) and agitated for 2

5 Stability
The glycoamino acid products were subjected to the following reaction conditions, to investigate the stability of the glyco-triazole linkage.

- 2.5 M HCl in EtOAc at rt
- K₂CO₃, MeOH at rt
- 1 M aq. HCl at rt
- 1 M aq. HCl at reflux
- 1 M aq. NaOH at rt
- 1 M HCl in MeOH at reflux

15 The glyco-triazole linkage appeared to be stable to all these conditions.
Claims

1. A compound of formula I or I’

\[
\begin{align*}
\text{A} & \sim \text{N} \sim \text{N} \\
\text{B} & \\
\text{I}
\end{align*}
\]

\[
\begin{align*}
\text{B} & \sim \text{N} \sim \text{N} \\
\text{A} & \\
\text{I’}
\end{align*}
\]

wherein A represents a carbohydrate which is coupled via its anomic center (I) or parent anomic center (I’) to the triazole ring and B represents an amino acid or a mimetic of an amino acid.

2. A compound of formula II or II’

\[
\begin{align*}
\text{A} & \sim \text{N} \sim \text{N} \\
\text{C} & \\
\text{II}
\end{align*}
\]

\[
\begin{align*}
\text{C} & \sim \text{N} \sim \text{N} \\
\text{A} & \\
\text{II’}
\end{align*}
\]

wherein A represents a carbohydrate which is coupled via its anomic center (II) or parent anomic center (II’) to the triazole ring and C represents a polypeptide or a mimetic of a polypeptide.

3. A compound according to claim 1 or 2 wherein the carbohydrate A comprises 1-20 saccharide units.

4. A compound according to claim 2 or 3 wherein the peptide C or mimetic thereof comprises 2-100 amino acids or mimetic(s) thereof.
5. A compound according to any of the preceding claims wherein B or C is attached to the triazole moiety via the side chain of the amino acid or mimetic thereof or via the side chain of an amino acid or mimetic thereof in the peptide or mimetic thereof.

6. A compound according to any of the preceding claims wherein the carbohydrate is attached to the triazole moiety via the (parent) anomeric carbon atom of a terminal saccharide unit.

7. Method for the preparation of a compound according to claim 1 or 2, comprising the step of the cycloaddition of

\[ R^2 R^3 \]
\[ \begin{array}{c}
R^1 \ N X \\
\end{array} \]

wherein \( X \) represents CN, CO_2R^4, COR^5, SO_2R^6, CH_2OR^7, CH_2NR^8R^9, CR^{10}R^{11}, CONR^{12}NR^{13}R^{14}, \) or \( X \) represents the site of attachment to one or more further amino acids;

\( R^1, R^2, R^3, R^4, R^5, R^6, R^7, R^8, R^9, R^{10}, R^{11}, R^{12}, R^{13} \) and \( R^{14} \) may be the same or different, and each independently represent \( \text{H} \), linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, \( R^1 \) and/or \( R^2 \) may also represent the site of attachment to one or more further amino acids,

\( n = 1-10 \)

with

\[ \begin{array}{c}
R^i \\
\end{array} O \begin{array}{c} N_3 \end{array} \]
\[ \begin{array}{c}
RO \\
\end{array} \]
\[ \begin{array}{c}
R^2 \\
\end{array} \]
\[ \begin{array}{c}
OR \\
\end{array} \]

wherein \( R^1 \) may be \( \text{H} \), CH_2OR or CHO-RCH_2OR, \( R^2 \) may be \( \text{H} \), NHR or OR, \( R \) may be the same or different within one molecule and may be \( \text{H} \), a saccharide unit or a protective group, \( m = 0 \) or 1,

or the cycloaddition of
wherein X represents CN, CO₂R⁴, COR⁵, SO₂R⁶, CH₂OR⁷, CH₂NR⁸R⁹, CR¹⁰R¹¹, CONR¹²NR¹³R¹⁴, or X represents the site of attachment to one or more further amino acids;

R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹², R¹³ and R¹⁴ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, R¹ and/or R² may also represent the site of attachment to one or more further amino acids,

n = 1-10

or

wherein X represents CN, CO₂R³, COR⁴, SO₂R⁵, CH₂OR⁶, CH₂NR⁷R⁸, CR⁹R¹⁰, CONR¹¹NR¹²R¹³ or X represents the site of attachment to one or more further amino acids;

R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹² and R¹³ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, R¹ may also represent the site of attachment to one or more further amino acids, the ring may be saturated or (partly) unsaturated and may be 4-10 membered; the azide can be in any position on the ring.

with
wherein $R^1$ may be H, CH$_2$OR or CHORCH$_2$OR, $R^2$ may be H, NHR or OR, $R$ may be the same or different within one molecule and may be H, a saccharide unit or a protective group, $m = 0$ or 1.

8. Method for the preparation of a compound according to claim 2, said method comprising reacting a compound according to claim 1 as a building block in a peptide synthesis.

9. Method according to claim 8 wherein the peptide synthesis is a peptide synthesis on a solid support, preferably in an automated peptide synthesiser.

10. Method for the preparation of a compound according to claim 2, said method comprising reacting

wherein $X$ represents CN, CO$_2$R$^4$, COR$^5$, SO$_2$R$^6$, CH$_2$OR$^7$, CH$_2$NR$^8$R$^9$, CR$_{10}$R$_{11}$, CONR$_{12}$NR$_{13}$R$_{14}$; $R^1$, $R^2$, $R^3$, $R^4$, $R^5$, $R^6$, $R^7$, $R^8$, $R^9$, $R^{10}$, $R^{11}$, $R^{12}$, $R^{13}$ and $R^{14}$ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, $n = 1$-10 and/or
wherein X represents CN, CO₂R⁴, COR⁵, SO₂R⁶, CH₂OR⁷, CH₂NR⁸R⁹, CR¹⁰R¹¹, CONR¹²NR¹³R¹⁴;
R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹², R¹³ and R¹⁴ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, n = 1-10
and/or

\[
\text{\begin{center}
\begin{tikzpicture}
\node (N3) at (0,0) {N_3};
\node (N) at (-0.5,0) {N};
\node (R2) at (0,-0.5) {R²};
\node (R1) at (-0.5,-0.5) {R¹};
\node (R) at (-0.5,-1) {X};
\draw (N3) -- (N);
\draw (N) -- (R2);
\draw (N) -- (R1);
\draw (N) -- (R);
\end{tikzpicture}
\end{center}
}\]

wherein X represents CN, CO₂R², COR⁴, SO₂R⁵, CH₂OR⁶, CH₂NR⁷R⁸, CR⁹R¹⁰, CONR¹¹NR¹²R¹³;
R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, R¹² and R¹³ may be the same or different, and each independently represent H, linear or branched and optionally substituted alkyl, optionally substituted acyl, optionally substituted sulfonyl, optionally substituted aryl, and amine and/or acid protective groups, the ring may be substituted and be saturated or (partly) unsaturated and may be 4-10 membered; the azide can be in any position on the ring,
as a building block in a peptide synthesis, such as a peptide synthesis on a solid support, preferably in an automated peptide synthesiser.

and reacting azide functional groups in the resulting peptide during peptide synthesis or after peptide synthesis is completed

\[
\text{\begin{center}
\begin{tikzpicture}
\node (R1) at (-0.5,0) {R¹};
\node (O) at (0,0) {O};
\node (R2) at (0.5,0) {R²};
\node (OR) at (1,0) {OR};
\node (m) at (0,0) {m};
\draw (R1) -- (O);
\draw (O) -- (R2);
\draw (O) -- (m);
\draw (m) -- (R2);
\end{tikzpicture}
\end{center}
}\]
wherein $R^1$ may be $H$, $\text{CH}_2\text{OR}$ or $\text{CHORCH}_2\text{OR}$, $R^2$ may be $H$, $\text{NHR}$ or $\text{OR}$, $R$ may be the same or different within one molecule and may be $H$, a saccharide unit or a protective group, $m = 0$ or $1$.

and/or

reacting acetylene functional groups in the resulting peptide during peptide synthesis or after peptide synthesis is completed

with

wherein $R^1$ may be $H$, $\text{CH}_2\text{OR}$ or $\text{CHORCH}_2\text{OR}$, $R^2$ may be $H$, $\text{NHR}$ or $\text{OR}$, $R$ may be the same or different within one molecule and may be $H$, a saccharide unit or a protective group, $m = 0$ or $1$.

11. Kit of parts comprising as a research tool and/or a diagnostic tool a compound according to any of claims 1-6.
## INTERNATIONAL SEARCH REPORT

**PCT/NL2005/000406**

### A. CLASSIFICATION OF SUBJECT MATTER

<table>
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According to International Patent Classification (IPC) or to both national classification and IPC.

### B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols):

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Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched.

Electronic data base consulted during the international search (name of data base and, where practical, search terms used):

- EPO-Internal, WPI Data, PAJ, CHEM ABS Data, BEILSTEIN Data

### C. DOCUMENTS CONSIDERED TO BE RELEVANT

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<th>Relevant to claim No.</th>
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<td>Y</td>
<td>TORMOE C W ET AL: &quot;Peptidotriazoles on solid phase: '1,2,3!-triazoles by regiospecific copper (I) catalyzed 1,3-dipolar cycloadditions of terminal alkynes to azides&quot; JOURNAL OF ORGANIC CHEMISTRY, AMERICAN CHEMICAL SOCIETY. EASTON, US; vol. 67, no. 9, 2002, pages 3057-3064, XP002234501</td>
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- abstract; schemes 1-3; page 3060, paragraph joining left- and right-hand columns

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### X

Further documents are listed in the continuation of box C. Patent family members are listed in annex.

**#** Special categories of cited documents:

- **"A"** document defining the general state of the art which is not considered to be of particular relevance
- **"E"** earlier document but published on or after the international filing date
- **"L"** document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- **"O"** document referring to an oral disclosure, use, exhibition or other means
- **"P"** document published prior to the international filing date but later than the priority date claimed

**"T"** later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

**"X"** document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

**"Y"** document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

**"8"** document member of the same patent family

### Date of the actual completion of the international search

18 July 2005

### Date of mailing of the international search report

27/07/2005

### Name and mailing address of the ISA

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Tel. (+31-70) 940-2540, Tx. 31 651 apo nl,
Fax. (+31-70) 340-3016

### Authorized officer

Fausti, S
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<td>BROEDER WOLFGANG ET AL: &quot;A new method of anomic protection and activation based on the conversion of glycosyl azides into glycosyl fluorides&quot; CARBOHYDRATE RESEARCH, vol. 249, no. 1, 1993, pages 221-241, XP002304299 ISSN: 0008-6215 * reaction scheme on page 224; Table I *</td>
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<td>A</td>
<td>B. G. DAVIS: &quot;Recent developments in glycoconjugates&quot; J. CHEM. SOC., PERKIN TRANS. 1, 1999, pages 3215-3237, XP002304300 * page 3222, left-hand column, last paragraph; paragraph joining pages 3222 and 3223; page 3223, right-hand column, third paragraph; schemes 12 and 13 on pages 3225 and 3226 *</td>
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