Dietary n-3 Fatty Acids Increase Spleen Size and Postendotoxin Circulating TNF in Mice; Role of Macrophages, Macrophage Precursors, and Colony-Stimulating Factor-1

Willem L. Blok,* Marella F. T. R. de Bruijn,‡ Pieter J. M. Leenen,† Wijnand M. C. Eling,‡ Nico van Rooijen,§ E. Richard Stanley,|| Wim A. Buurman,|| and Jos W. M. van der Meer‡*

In experimental studies in mice, dietary supplementation with n-3 fatty acids (FA) alleviates inflammation and increases resistance to infection. Nevertheless, TNF production capacity was found to be increased in n-3 FA-fed mice. We previously found increased relative spleen weights in n-3 FA-fed mice. In this study, the nature of this increased spleen size was further investigated. Spleen cellularity was increased significantly in mice fed n-3 FA (fish oil 15% w/w), compared with controls fed corn oil (15%) or normal lab chow (p < 0.05). Experiments with T cell-deficient nude mice and experiments using macrophage depletion through liposomal dichloromethylene-biphosphonate revealed that the increase in spleen cellularity is T cell independent and largely due to macrophage accumulation in the spleen. Accumulation of marginal zone and red pulp macrophages was histologically and immunohistochemically confirmed. n-3 FA induced peripheral blood monocytoplasia and an aspecific increase in bone marrow cellularity. Postendotoxin circulating TNF concentrations were increased significantly in n-3 FA-fed mice compared with controls. Splenectomy did not abolish this increase in circulating TNF. However, after macrophage depletion through liposomal dichloromethylene-biphosphonate, circulating TNF was not detectable after endotoxin challenge. Circulating concentrations of CSF-1 did not differ between the various experimental groups. It is suggested that the cellular changes observed relate to increased constitutive production of TNF.


Abbreviations used in this paper: FA, fatty acid; CI2 MDP, dichloromethylene-biphosphonate; CO, corn oil; FO, fish oil.

Departments of *General Internal Medicine and *Medical Microbiology, University Hospital Nijmegen, Nijmegen, The Netherlands; ‡Department of Immunology, Erasmus University, Rotterdam, The Netherlands; †Department of Cell Biology, Free University, Amsterdam, The Netherlands; ||Department of Developmental and Molecular Biology, Albert Einstein College of Medicine of Yeshiva University, Bronx, NY 10461; and Department of Surgery, University of Limburg, Maastricht, The Netherlands

Received for publication May 14, 1996. Accepted for publication September 16, 1996.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked

1 This work was supported by National Institutes of Health Grants CA 32551 and CA 26504 (to E.R.S.) and Albert Einstein Cancer Core Grant P30CA13330.

2 Address correspondence and reprint requests to Dr. Jos W. M. van der Meer, Department of Medicine, General Internal Medicine, P. O. Box 9101, 6500 HB Nijmegen, The Netherlands.

3 Abbreviations used in this paper: FA, fatty acid; CI2 MDP, dichloromethylene-biphosphonate; CO, corn oil; FO, fish oil.

Copyright © 1996 by The American Association of Immunologists

0022-1767/96/302.00
n-3 FATTY ACIDS AND MACROPHAGES

The mice were anesthetized with ether, and blood was taken from the retrobulbar vessels after eye extraction. The animals were killed by neck dislocation, and spleens were removed and weighed. Total body weight was determined immediately before bleeding, and relative spleen weight was calculated afterward. A suspension of spleen cells was made over a nylon filter, and cells were counted with a Coulter counter (Coulter Corp., Hialeah, FL). The effect of dietary n-3 FA on spleen weight was determined in normal Swiss mice, normal BALB/c mice, and T cell-deficient nude BALB/c mice (nu/nu).

Spleen cells were phenotypically characterized with flow cytometry using a set of mAbs specific for mouse cells: 59-AD 2.2 detecting Thy-1 (T lymphocytes), RA3-6B2 detecting B220 (B lymphocytes), R129.19 detecting MT4 (CD4-positive T lymphocytes), 53-6.72 detecting Ly-2 (CD8-positive T lymphocytes), KT3 (CD3-positive T lymphocytes), and F4/80 (macrophages). Details concerning these Abs can be found in Leenen et al. (24).

After immunohistochemical staining of spleen sections with RA3-6B2 (anti-B220), a mAb specific for mouse B cells*, the relative surface area of B 220-positive spleen cells (%), was determined immediately before bleeding, and relative spleen weight (mg/g) was calculated afterward, and spleens were removed and weighed. Total body weight was determined immediately before bleeding, and relative spleen weight was calculated afterward. A suspension of spleen cells was made over a nylon filter, and cells were counted with a Coulter counter (Coulter Corp., Hialeah, FL). The effect of dietary n-3 FA on spleen weight was determined in normal Swiss mice, normal BALB/c mice, and T cell-deficient nude BALB/c mice (nu/nu).

Spleen analysis

The mice were anesthetized with ether, and blood was taken from the retrobulbar vessels after eye extraction. The animals were killed by neck dislocation, and spleens were removed and weighed. Total body weight was determined immediately before bleeding, and relative spleen weight was calculated afterward. A suspension of spleen cells was made over a nylon filter, and cells were counted with a Coulter counter (Coulter Corp., Hialeah, FL). The effect of dietary n-3 FA on spleen weight was determined in normal Swiss mice, normal BALB/c mice, and T cell-deficient nude BALB/c mice (nu/nu).

Spleen cells were phenotypically characterized with flow cytometry using a set of mAbs specific for mouse cells: 59-AD 2.2 detecting Thy-1 (T lymphocytes), RA3-6B2 detecting B220 (B lymphocytes), R129.19 detecting MT4 (CD4-positive T lymphocytes), 53-6.72 detecting Ly-2 (CD8-positive T lymphocytes), KT3 (CD3-positive T lymphocytes), and F4/80 (macrophages). Details concerning these Abs can be found in Leenen et al. (24).

To analyze the role of mature phagocytic macrophages in the increase in spleen size, mice were fed FO for 0, 2, or 4 wk. Subsequently the spleens were removed, and cells were counted with a Coulter counter (Coulter Corp., Hialeah, FL). The effect of dietary n-3 FA on spleen weight was determined in normal Swiss mice, normal BALB/c mice, and T cell-deficient nude BALB/c mice (nu/nu).

Peripheral blood cell count was performed by separately determining hematocrit and the total white blood cell count (using Coulter counter). Differential white blood cell count was performed by quantifying the light scatter profile of nucleated cells.

Bone marrow analysis

Bone marrow was obtained from both femora of each mouse by flushing the femoral shafts with 3 ml of culture medium (RPMI, Dutch modification; Flow Laboratories, Irvine, Scotland). Nucleated cells were counted with a Coulter counter. Soft agar cultures of bone marrow cells were stimulated with macrophage CSF for 2 wk, and the number of macrophage precursors in the original preparation was quantitated by counting the number of macrophage colonies and clusters (<50 cells); macrophage colony-forming cells and macrophage cluster-forming cells are given per 10⁴ nucleated bone marrow cells (26). Nucleated bone marrow cells were phenotypically characterized with flow cytometry using mAbs detecting Ags of mouse macrophage and granulocyte precursors. The mAbs were produced at the Department of Immunology (Erasmus University, Rotterdam, The Netherlands), as described (27). The following mAbs were used: ER-MP12 and ER-MP20, identifying subpopulations of macrophage precursors; ER-MP58, identifying myeloid cells; and ER-MP21, identifying transferrin receptor-expressing cells (28).

Postendotoxin circulating TNF

At a time period of 1.5 h after i.p. administration of 10 μg of LPS (Escherichia coli, serotype O55:B5; Sigma Chemical Co., St. Louis, MO), mice were anesthetized with ether, and blood was taken from the retrobulbar vessels after eye extraction. Blood samples (approximately 1 ml) were mixed with 100 μl of EDTA solution (21.4 mg/ml of EDTA in H2O) and kept on ice until centrifugation at 1500 g for 5 min. Resultant plasma was isolated and stored at −20°C until assay.

TNF-α was measured by ELISA using TN3, a hamster mAb specific for murine TNF-α, and lymphotoxin, as described (29, 30).

In separate experiments, the effect of splenectomy on postendotoxin circulating TNF was studied. Splenectomy was performed after ether anesthesia in 4-wk-old mice, 2 wk before the animals were started on the experimental diet.

Statistical analysis

Differences between groups were analyzed using the Kruskal-Wallis nonparametric ANOVA test, corrected for ties. Results were considered statistically significant at p < 0.05.

Results

Effects of FO supplementation on spleen cellularity

The effects of dietary n-3 FA on spleen, peripheral blood, and bone marrow cells are summarized in Table I. The following experiments were done to elucidate the mechanisms of the increased spleen weight in FO-fed mice. After 4 wk of dietary supplementation with FO or CO, experimental Swiss mice were compared with control animals on normal lab chow. FO-fed mice had significantly increased relative spleen weights (p = 0.01) and spleen cellularity.

Table I. Effect of dietary n-3 FA on spleen, peripheral blood, and bone marrow cells

<table>
<thead>
<tr>
<th>Diet Group</th>
<th>Fish oil</th>
<th>Corn oil</th>
<th>Normal</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spleen</td>
<td>Relative spleen weight (mg/g)</td>
<td>5.9 ± 0.5</td>
<td>4.8 ± 0.3</td>
<td>4.6 ± 0.5</td>
</tr>
<tr>
<td>Spleen</td>
<td>Splenocyte cellularity (x 10⁶)</td>
<td>2.7 ± 0.2</td>
<td>2.0 ± 0.2</td>
<td>1.4 ± 0.5</td>
</tr>
<tr>
<td>B 220 positive spleen cells (%)</td>
<td>41 ± 4</td>
<td>36 ± 2</td>
<td>31 ± 6</td>
<td>&lt;0.04</td>
</tr>
<tr>
<td>Peripheral blood</td>
<td>Hematocrit</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>White blood cell count (x 10⁹/ml)</td>
<td>12.2 ± 4.0</td>
<td>9.1 ± 1.9</td>
<td>7.9 ± 1.3</td>
</tr>
<tr>
<td></td>
<td>Lymphocytes (%)</td>
<td>55 ± 8</td>
<td>69 ± 4</td>
<td>64 ± 5</td>
</tr>
<tr>
<td></td>
<td>Granulocytes (%)</td>
<td>34 ± 8</td>
<td>25 ± 4</td>
<td>31 ± 6</td>
</tr>
<tr>
<td></td>
<td>Monocytes (%)</td>
<td>11 ± 2</td>
<td>6 ± 1</td>
<td>5 ± 1</td>
</tr>
<tr>
<td>Bone marrow</td>
<td>Cellularity (x 10⁶)</td>
<td>24 ± 5</td>
<td>21 ± 7</td>
<td>13 ± 5</td>
</tr>
<tr>
<td></td>
<td>M-CFC (No. macrophage precursors/10⁴ NBMC)</td>
<td>58 ± 15</td>
<td>51 ± 17</td>
<td>77 ± 20</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 50 high (myeloid cells) (%)</td>
<td>22 ± 5</td>
<td>25 ± 3</td>
<td>22 ± 5</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 12 high/20 neg (committed progenitors) (%)</td>
<td>50 ± 7</td>
<td>50 ± 2</td>
<td>54 ± 6</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 12 high/20 neg (committed progenitors) (%)</td>
<td>1.2 ± 0.3</td>
<td>1.7 ± 0.2</td>
<td>1.6 ± 0.5</td>
</tr>
<tr>
<td></td>
<td>ER-MP 12 pos/20 pos (immature myeloid precursors) (%)</td>
<td>5.4 ± 1.2</td>
<td>5.6 ± 0.5</td>
<td>5.0 ± 1.2</td>
</tr>
<tr>
<td></td>
<td>ER-MP 12 neg/20 high (monocytes) (%)</td>
<td>5.7 ± 0.9</td>
<td>6.2 ± 0.8</td>
<td>5.7 ± 1.2</td>
</tr>
</tbody>
</table>

Table I. Effect of dietary n-3 FA on spleen, peripheral blood, and bone marrow cells

<table>
<thead>
<tr>
<th>Diet Group</th>
<th>Fish oil</th>
<th>Corn oil</th>
<th>Normal</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spleen</td>
<td>Relative spleen weight (mg/g)</td>
<td>5.9 ± 0.5</td>
<td>4.8 ± 0.3</td>
<td>4.6 ± 0.5</td>
</tr>
<tr>
<td>Spleen</td>
<td>Splenocyte cellularity (x 10⁶)</td>
<td>2.7 ± 0.2</td>
<td>2.0 ± 0.2</td>
<td>1.4 ± 0.5</td>
</tr>
<tr>
<td>B 220 positive spleen cells (%)</td>
<td>41 ± 4</td>
<td>36 ± 2</td>
<td>31 ± 6</td>
<td>&lt;0.04</td>
</tr>
<tr>
<td>Peripheral blood</td>
<td>Hematocrit</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>White blood cell count (x 10⁹/ml)</td>
<td>12.2 ± 4.0</td>
<td>9.1 ± 1.9</td>
<td>7.9 ± 1.3</td>
</tr>
<tr>
<td></td>
<td>Lymphocytes (%)</td>
<td>55 ± 8</td>
<td>69 ± 4</td>
<td>64 ± 5</td>
</tr>
<tr>
<td></td>
<td>Granulocytes (%)</td>
<td>34 ± 8</td>
<td>25 ± 4</td>
<td>31 ± 6</td>
</tr>
<tr>
<td></td>
<td>Monocytes (%)</td>
<td>11 ± 2</td>
<td>6 ± 1</td>
<td>5 ± 1</td>
</tr>
<tr>
<td>Bone marrow</td>
<td>Cellularity (x 10⁶)</td>
<td>24 ± 5</td>
<td>21 ± 7</td>
<td>13 ± 5</td>
</tr>
<tr>
<td></td>
<td>M-CFC (No. macrophage precursors/10⁴ NBMC)</td>
<td>58 ± 15</td>
<td>51 ± 17</td>
<td>77 ± 20</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 50 high (myeloid cells) (%)</td>
<td>22 ± 5</td>
<td>25 ± 3</td>
<td>22 ± 5</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 12 high/20 neg (committed progenitors) (%)</td>
<td>50 ± 7</td>
<td>50 ± 2</td>
<td>54 ± 6</td>
</tr>
<tr>
<td></td>
<td>ER-MP 58 12 high/20 neg (committed progenitors) (%)</td>
<td>1.2 ± 0.3</td>
<td>1.7 ± 0.2</td>
<td>1.6 ± 0.5</td>
</tr>
<tr>
<td></td>
<td>ER-MP 12 pos/20 pos (immature myeloid precursors) (%)</td>
<td>5.4 ± 1.2</td>
<td>5.6 ± 0.5</td>
<td>5.0 ± 1.2</td>
</tr>
<tr>
<td></td>
<td>ER-MP 12 neg/20 high (monocytes) (%)</td>
<td>5.7 ± 0.9</td>
<td>6.2 ± 0.8</td>
<td>5.7 ± 1.2</td>
</tr>
</tbody>
</table>
FIGURE 1. Time-effect relationships of dietary n-3 FA on relative spleen weight (A) and on postendotoxin circulating TNF (B). Relative spleen weight was increased significantly from 2 wk on FO diet, as was postendotoxin circulating TNF (n = 12).

Phenotypic characterization of spleen cells using a set of mAbs revealed no percentual differences in T lymphocytes or macrophages between the various dietary groups. However, FO-fed mice had a small but significant increase in B220-positive B lymphocytes (Table I).

To a lesser extent, B lymphocytes seem to contribute to the increased spleen cellularity in FO-fed mice. Quantification of the relative surface area of red and white pulp in spleen sections indicated a decrease in T and B cell-containing white pulp (Fig. 3).
groups. White blood cell count was higher in FO-fed mice, but this difference did not reach statistical significance. Analysis of the light-scatter profile of peripheral blood cells showed a significant increase in percentage of monocytes in FO-fed mice ($p = 0.02, n = 6$).

The effect of FO supplementation on bone marrow cells

Possible relations between changes in spleen and peripheral blood cellularity and the cellular composition of bone marrow were investigated. Bone marrow cellularity was increased significantly in FO-fed mice compared with mice fed CO or normal lab chow. This increase in bone marrow cellularity was equally distributed over the various hematopoietic lineages, since differential flow-cytometric analysis of bone marrow cells, using ER-MP12 and ER-MP20 mAbs, showed no significant differences between the various experimental groups (Table I). Moreover, soft agar cultures of bone marrow cells did not reveal a relative increase in macrophage precursors, measured as macrophage colony-forming cells or macrophage cluster-forming cells.

The effect of FO supplementation on postendotoxin TNF

To investigate the suggested increase in mononuclear phagocyte functionality, we measured circulating TNF concentrations after endotoxin challenge. Postendotoxin concentrations of circulating TNF were significantly higher in FO-fed mice than in control mice. Postendotoxin TNF was 10.4 ± 6.6 ng/ml in FO-fed mice, 2.2 ± 1.4 in CO-fed mice, and 2.7 ± 1.3 in mice fed normal diet (mean ± SD, $p < 0.05$). In separate experiments, time-effect relationships in TNF production and FO diet were investigated. Similar to the increase in spleen weight, increases in postendotoxin circulating TNF were most prominent after 2 wk of FO diet, and appeared somewhat blunted after 6 wk (Fig. 1B). Postendotoxin TNF in T cell-deficient nude BALB/c mice was increased significantly from 2 wk on, indicating that the increase in postendotoxin circulating TNF was independent of the presence of T cells (results not shown).

Splenectomy, performed 2 wk before starting on the FO-supplemented diet, did not influence the increase in postendotoxin circulating TNF concentrations after 2 and 4 wk of diet: at both time points, postendotoxin circulating TNF concentrations were increased significantly compared with baseline (results not shown). At baseline and at 2 and 4 wk of dietary FO supplementation, postendotoxin circulating TNF was not detectable in liposomal Cl$_2$MDP-treated mice (Fig. 2B). These observations suggest that splenic macrophages, despite their accumulation after dietary FO supplementation, are not the most important contributors to the production of circulating TNF. On the other hand, the macrophage population that is depleted by liposomal Cl$_2$MDP and that does not reside in the spleen appears to be the most important producer of postendotoxin circulating TNF.

The effect of FO supplementation on circulating concentrations of CSF-1

The mononuclear phagocyte growth factor, CSF-1, is required for the development of the majority of macrophages (33). To accurately measure circulating concentrations of CSF-1, serum samples were subjected to a mouse CSF-1-specific RIA. In biological samples, this RIA detects only biologically active CSF-1 (32, 34), including both glycoprotein and proteoglycan forms (35). Circulating concentrations of immunoreactive CSF-1 did not differ among the various diet groups: 702 ± 89 U/ml in FO-fed mice, 753 ± 96 in CO-fed mice, and 802 ± 158 in mice fed normal diet (mean ± SD, $p = 0.40$).

Discussion

The present study shows that dietary n-3 FA supplementation has a significant effect on the generation and distribution of mononuclear phagocytes in mice: n-3 FA induce a generalized increase in bone marrow cellularity, peripheral blood monocytosis, and accumulation of macrophages in the spleen, leading to an increase in spleen size. What could be the mechanism of these n-3 FA-induced changes?

The aspecific increase in bone marrow cellularity without preferential stimulation of monocytopoiesis is in accordance with our observation on CSF-1: this specific monocyte growth factor was not increased in the n-3 FA-fed mice. The generalized increase in bone marrow cellularity suggests that earlier, broad spectrum hematopoietic growth factors such as IL-3 and granulocyte-macrophage CSF are involved. The production of these factors is largely regulated by proinflammatory cytokines including IL-1 (36, 37). We have shown previously that IL-1α production capacity is increased in mice after dietary n-3 FA (15). Increased constitutive production of this cytokine may be involved in the increased bone marrow cellularity observed in the present study. Moreover, Pelus and others have shown that PGE$_2$ has an important modulatory effect on hematopoiesis in mice: exogenous administration of PGE$_2$ reduced nucleated bone marrow and splenic cellularity, while blockade of PGE$_2$ biosynthesis increased bone marrow and splenic cellularity, especially in the presence of IL-1α (38–42). These observations may partly explain the results obtained in the present study, since dietary supplementation with n-3 FA in mice basically leads to increased IL-1α and a decreased PGE$_2$ production capacity.

Peripheral blood monocytosis induced by dietary n-3 FA has not been described before, but may be induced through a feedback mechanism following the accumulation of macrophages in the red pulp of the spleen. It is apparently not due to increased concentrations of CSF-1. This is in accordance with the observation that the number of red pulp macrophages is reduced only significantly in CSF-1-deficient op/op mice (43). Other factors, including IL-3, granulocyte-macrophage CSF, and the proinflammatory cytokines, may be considered. The results of our experiments with macrophage depletion using liposomal Cl$_2$MDP suggest that the observed increase in spleen size following dietary FO supplementation is largely due to accumulation of macrophages in the spleen.

The immunohistochemical staining of spleen sections with B220 showed a decrease in the relative surface area of white pulp in the FO-fed mice, thereby supporting the concept of accumulation of red pulp macrophages in FO-fed mice. As regulator cells, macrophages are responsible for the growth of lymphoid organs such as the spleen. Since TNF is an important growth factor for lymphoid organs, increased endogenous TNF production in FO-fed mice may contribute to this effect (44). The phenotypic characterization of spleen cell suspensions using a set of mAbs is not suitable for the assessment of the number of macrophages in the spleen, since especially tissue-fixed macrophages are lost in the process of making a cell suspension over a nylon filter. We therefore regard the concept that dietary FO supplementation leads to macrophage accumulation in the spleen as valid.

The increase in spleen size after dietary n-3 FA appeared to be accompanied by increased circulating concentrations of postendotoxin TNF. We and others have shown previously that peritoneal macrophages of mice fed n-3 FA have increased TNF production capacity at the level of the single cell (15, 20, 45). Since splenectomy did not affect the increased TNF concentrations following dietary n-3 FA, macrophage accumulation in the spleen is not responsible for the increased TNF concentrations. Interestingly, macrophage depletion by liposomal Cl$_2$MDP completely abolished
postendotoxin circulating TNF. It might be expected that Kupffer cells of the liver, due to their sinusoidal location, might be a major target for the uptake of liposomal CLMDP. In fact, recent evidence indicates that macrophage depletion in mice results in a 50 to 70% reduction in TNF mRNA in the liver following endotoxin challenge (46). Therefore, the Kupffer cells may be the most important contributors to the production of postendotoxin TNF. Similar to peritoneal macrophages, Kupffer cells may have increased TNF production capacity at the single cell level following dietary n-3 FA. We did not assess the number of Kupffer cells. The small increase in relative liver weight in FO-fed mice does not rule out a substantial increase in the number of Kupffer cells.

In conclusion, dietary n-3 FA supplementation in mice induces an aspecific increase in bone marrow cellularity, peripheral blood monocytes, accumulation of macrophages in the spleen, and increased postendotoxin circulating concentrations of TNF. The mechanism of these changes remains to be elucidated.

Acknowledgments
We thank Piet Spaan, Yvette Brom, and Reza Zadeh for technical assistance.

References

Email: P. Brignon et al. (1992). The Journal of Immunology, 155:3163.