Creatinine and Surveys: An Assessment

By Berti G. Blijenberg1, Rita J. Brouwer1, Henk Baadenhuijzen2 and Geert J. M. Boerma3

1 Department of Clinical Chemistry, University Hospital Rotterdam-Dijkzigt, Rotterdam, The Netherlands
2 Department of Clinical Chemistry, University Hospital Nijmegen-St. Radboud, Nijmegen, The Netherlands
3 Department of Clinical Chemistry, University Hospital Rotterdam-Sophia Children's Hospital, Rotterdam, The Netherlands

(Received May 15/July 21, 1995)

Summary: We analysed the results of surveys on creatinine held in The Netherlands during the years 1992, 1993 and 1994. Assay results of 113 samples were reviewed: 88 human sera and 25 samples of animal origin. The results of 5 creatinine assays, 4 based on the Jaffé reaction and 1 enzymatic procedure, are discussed.

The enzymatic assay showed by far the best performance, while some of the Jaffé methods differed considerably. All results were evaluated by reference to a HPLC-based selected method for creatinine.

Our study shows the need for caution when applying survey performance criteria for creatinine.

Introduction

In two earlier publications we described the influence of a number of factors on the accuracy of various creatinine assays developed for routine use in clinical chemistry (1, 2). We also questioned the value of quality assessment specimens for creatinine assays.

Since these earlier studies provided only limited data on accuracy-influencing factors and the value of quality assessment specimens, and in view of the large variation of results, especially in some modifications of the Jaffé reaction, we decided to study these aspects in greater detail in a wider setting.

Here, we describe the cooperation between three laboratories, all participating in the serum chemistry section of the Dutch Quality Assessment Foundation (SKZL) (3). The results submitted during the years 1992, 1993 and 1994 were compared with those obtained with our high performance liquid chromatography (HPLC) based selected method for creatinine (4).

In addition, all samples under study were analysed separately with the instruments also used in our previous study i.e. the DuPont Dimension and the Merck ELAN.

Materials and Methods

Materials

One hundred and thirteen control sera were included in this study, 25 of animal origin, 88 of human origin. During the years 1992,

<table>
<thead>
<tr>
<th>Tab. 1 Regression equations of all comparisons</th>
</tr>
</thead>
<tbody>
<tr>
<td>a. Range 0–1000 μmol/l (n = 113) x = HPLC</td>
</tr>
<tr>
<td>Chem-l</td>
</tr>
<tr>
<td>Dimension (enzym.)</td>
</tr>
<tr>
<td>Hitachi</td>
</tr>
<tr>
<td>Dimension (Jaffé)</td>
</tr>
<tr>
<td>ELAN</td>
</tr>
<tr>
<td>y = 1.08x - 4.1</td>
</tr>
<tr>
<td>y = 0.98x + 3.0</td>
</tr>
<tr>
<td>y = 0.96x + 17.2</td>
</tr>
<tr>
<td>y = 1.02x - 4.5</td>
</tr>
<tr>
<td>y = 1.08x - 20.8</td>
</tr>
<tr>
<td>r = 0.97</td>
</tr>
<tr>
<td>r = 0.99</td>
</tr>
<tr>
<td>r = 1.00</td>
</tr>
<tr>
<td>r = 0.97</td>
</tr>
<tr>
<td>r = 0.95</td>
</tr>
</tbody>
</table>

b. Range 0–300 μmol/l (n = 73) x = HPLC

Chem-l
Dimension (enzym.)
Hitachi
Dimension (Jaffé)
ELAN
y = 1.08x - 1.0
y = 0.99x + 2.3
y = 0.97x + 16.3
y = 1.03x - 6.1
r = 0.98
r = 0.99
r = 1.00
r = 0.97

1993 and 1994 they were distributed in the normal way by the official Quality Assessment Foundation in The Netherlands to all participating laboratories.

Methods

Five routine methods for creatinine were applied with the following instruments:

1. Bayer-Technicon Chem-1 with a Jaffe reaction based method
2. Boehringer Mannheim Hitachi 717 and 747 with a Jaffe reaction based method
3. DuPont Dimension with an enzymatic method
4. DuPont Dimension with a Jaffe reaction based method
5. Merck ELAN with a Jaffe reaction based method

Methods 1, 2, 4 and 5 were routine procedures as supplied by the various manufacturers. Method 3 is a home made adaptation of the Boehringer Mannheim enzymatic method (creatininase) to the Dimension (5). All instruments were run according to the manufacturers’ instructions.

Procedure

The results of methods 1, 2 and 3 were taken from the quality control files of the three participating laboratories. Similarly, the

Fig. 1 Residual creatinine values for the various methods (see section Methods) obtained with human control samples with creatinine (HPLC) concentrations lower than 300 μmol/l.

HPLC results were taken from the Academic Hospital Rotterdam, which is part of the National External Quality Assessment Scheme. Extra samples were stored at −80 °C during 1992, 1993 and 1994, and used for the methods 4 and 5.

Statistical analysis

Regression analysis was done according to Passing & Bablok (6).

Results

We started by comparing the routine methods (methods 1, 2 and 3) with the HPLC selected method. All values deviating more than 20% from the corresponding HPLC result were redetermined. This figure of 20% was chosen arbitrarily. In total 7 results proved to be clerical errors.

Then, using the same set of specimens, creatinine was determined on the Dimension (Jaffé method) and the ELAN.

All regression equations are tabulated separately in table 1.

In view of the clinical significance of low to medium range creatinine concentrations, those samples of human origin with HPLC values lower than 300 μmol/l were plotted separately (figs. 1a–e).

The regression equations for the range 0–300 μmol/l are also included in table 1.

Discussion

Our results from the elaborated study generally confirm those found earlier, thus providing a sounder basis for evaluation. The enzymatic procedure for creatinine scores better than the Jaffé reaction based assays with the specimens used in the Dutch Quality Assessment Scheme. There are also differences between the various creatinine (Jaffé) methods.

Many samples showing deviations from the HPLC results are problematic for all Jaffé reaction based creatinine methods. Nevertheless, it is possible to construct a worst-case scenario by comparing the Chem-1 data with the ELAN, as in figure 2.

We assume that the differences in the results of the various Jaffé methods are due to the assay design. They are all kinetic measurements, but they differ considerably with respect to sample and reagent volumes, reagent concentrations and choice of reagents and type of measurement (mono- vs. bichromatic and timing of absorbance readings). The ELAN does not employ a bilirubin correction, the Hitachi and Chem-1 have chromatic cor-

![Graphical presentation of human control serum results obtained with the ELAN and the Chem-1. Regression equation: y = 1.11x - 30.0; r = 0.89 x(mean) = 93 μmol/l; y(mean) = 110 μmol/l x(median) = 87 μmol/l; y(median) = 66 μmol/l](image)

reactions, while the Dimension reagent contains potassium hexacyanoferrate(III) for bilirubin oxidation. Due to the influence of bilirubin, the ELAN showed considerable deviations from the HPLC values, especially with two extreme specimens containing 222 and 343 μmol bilirubin per litre; specimens with bilirubin values of about 100 μmol/l (6 in total) also showed discrepant creatinine values.

Most of the deviating results for all methods, however, could not be attributed to interference by bilirubin. Most of the human serum samples were pooled left-over patient samples, sometimes spiked with additional material like enzymes, bilirubin and sucrose. This makes it nearly impossible to understand the cause of the deviation. However, the findings with these human samples do show again the limitations of the Jaffé reaction based creatinine methods in the daily routine of a laboratory. It is also clear from figure 2 that the expression “Jaffé method” is an inaccurate statement.

The commercial sera used, BioRad (Lyphochek and Liquichek) and Beckman Decision, show an interesting pattern. The various techniques showed particularly substantial differences in the analysis of BioRad products, so that the use of these products in surveys may be questionable. Of course, this argument does not hold for the normal application of these products in precision checks of a procedure.

These results reveal once more the general difficulty with most classical External Quality Assessment (EQA) schemes. Issues of cost, stability and available analyte concentration often necessitate the use of processed lyophilized control serum, which very often lack the required degree of commutability with the native human serum specimens. As correctly pointed out by Thienpont

and Stöckl (7, 8), present-day quality assessment schemes expose the dilemma of which combination of control material and target value to choose. This dilemma is particularly acute when the results of such schemes are to be used for accreditation/licensing purposes.

Summarizing, the enzymatic method for creatinine used in the present survey scores higher than any of the Jaffé reaction based methods, based on comparison with our HPLC based selected method. For the Jaffé reaction based methods, survey material must be chosen carefully and possibly selectively. Results from samples of animal origin have not been described in detail, because of the limited number used (n = 25); however, the results were generally similar to those obtained with the human serum samples.

Acknowledgements

The authors wish to thank Mrs. A. P. Copper-Staamer for clerical support and Mr. R. Leeneman (Rotterdam) and Mrs. M. Hessels (Nijmegen) for help with data collection.

References


Dr. B. G. Blijenbergh
Academic Hospital Rotterdam-Dijkzigt
Department of Clinical Chemistry
Dr. Molewaterplein 40
NL-3015 GD Rotterdam
The Netherlands