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Antisense oligonucleotide therapy has been reported to be associated with renal injury. Here, the mechanism of reversible proteinuria was investigated by combining clinical, pre-clinical, and in vitro data. Urine samples were obtained from Duchenne muscular dystrophy (DMD) patients treated with drisapersen, a modified 2′-O-methyl phosphorothioate antisense oligonucleotide (6 mg/kg). Urine and kidney tissue samples were collected from cynomolgus monkeys (*Macaca fascicularis*) dosed with drisapersen (39 weeks). Cell viability and protein uptake were evaluated in vitro using human conditionally immortalized proximal tubule epithelial cells (ciPTECs). Oligonucleotide treatment in DMD patients was associated with an increase in urinary alpha-1-microglobulin (A1M), which returned to baseline following treatment interruptions. In monkeys, increased urinary A1M correlated with dose-dependent accumulation of oligonucleotide in kidney tissue without evidence of tubular damage. Furthermore, oligonucleotides accumulated in the lysosomes of ciPTECs and reduced the absorption of A1M, albumin, and receptor-associated protein, but did not affect cell viability when incubated for up to 7 days. In conclusion, phosphorothioate oligonucleotides appear to directly compete for receptor-mediated endocytosis in proximal tubules. We postulate that oligonucleotide-induced low molecular weight proteinuria in patients is therefore a transient functional change and not indicative of tubular damage.

INTRODUCTION

Single-stranded antisense oligonucleotides are a promising therapeutic platform to treat a variety of genetic diseases, cancer, and viral infections. Oligonucleotides are excreted primarily, though slowly, via the urine due to high plasma protein binding. Unbound oligonucleotides are filtered by the glomerulus, followed by reabsorption and accumulation into the proximal tubule epithelium. When exceeding critical threshold levels, oligonucleotide accumulation may become associated with tubular dysfunction in animals and man.1,3

Accumulation of antisense oligonucleotides in the kidney was demonstrated in animals by the presence of basophilic granules in the cytoplasm of proximal tubule epithelial cells.4–6 These granules are considered to reflect physiological clearance of oligonucleotides and, in the absence of associated degenerative changes, are not considered of toxicological importance.7,8 To increase resistance to endogenous nuclease breakdown and/or target engagement, the backbone structures of antisense oligonucleotides have been chemically modified using sulfur or hydroxyl groups, resulting in phosphorothioate (PS) and 2′-O-methyl (2OMe) oligonucleotides, respectively.9,10 Dose- and concentration-dependent effects on renal tubule cells were described for the class of 2′-O-methoxymethylribose (OMePS) oligonucleotides. These principles likely also apply to 2OMePS antisense oligonucleotides as they share physicochemical properties.7,11

Antisense oligonucleotides are postulated to be actively reabsorbed from the ultrafiltrate by receptor-mediated endocytosis in proximal tubule cells, as uptake of oligonucleotides is saturable.12,13 Understanding the mechanism of uptake by proximal tubule cells and the effect of antisense oligonucleotides on kidney function is important for their safety assessment. Here, we first describe the reversible nature of the increased urinary excretion of low molecular weight proteins without overt proteinuria, observed during a long-term clinical trial of 2OMePS antisense oligonucleotide treatment in Duchenne muscular dystrophy (DMD) patients.14,15 Next we provide supporting data obtained in monkeys and, finally, we provide mechanistic data from in vitro studies using the human proximal tubule cell model conditionally immortalized proximal tubule epithelial cells (ciPTECs), a cell line previously characterized in terms of intact endocytic machinery16,17 and physiological and pharmacological characteristics,18–20 as well as

Received 15 August 2019; accepted 28 August 2019; https://doi.org/10.1016/j.omtn.2019.08.025.
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toxicological responses. Together, our data provide a molecular model in which 20MePS antisense oligonucleotides competitively inhibit receptor-mediated endocytosis in proximal tubule cells without severe cytotoxic effects or irreversible tubular dysfunction.

RESULTS

Reversible Low Molecular Weight Proteinuria upon Oligonucleotide Treatment

Urine samples from patients were collected as part of a clinical trial in which DMD patients were treated with the antisense oligonucleotide drisapersen to change splicing of the dystrophin gene (DMD). Weekly subcutaneous (s.c.) administration of 6 mg/kg drisapersen for 72 weeks was associated with a slight but statistically significant increase in the concentration of both total protein and alpha-1-microglobulin (A1M) in the urine of DMD patients (Figures 1A and 1D). Median protein levels in urine increased slightly from 80 [55–115] mg/L to 210 [123–270] mg/L (p < 0.001) and remained elevated after an 8-week treatment interruption (recovery 190 [150–300] mg/L) (Figure 1A). Although the variation between and within subjects was considerable, elevations were generally mild and subclinical (i.e., urinary protein below 300 mg/L). The median levels of A1M in urine before treatment were below the detection limit of the assay (<4 mg/L) and after continuous treatment reached 36.4 [18.1–71.8] mg/L (p < 0.001). After treatment interruption, the A1M decreased toward baseline levels (5.65 [4.00–21.20] mg/L).

For the intermittent treatment, urine samples were collected for analysis before each treatment switch, and this revealed a clear pattern of reversible low molecular weight proteinuria for A1M (Figures 1D and 1E). The pooled median value after 8 weeks on-treatment (33.4 [26.2–70.8] mg/L) significantly decreased after 4 weeks off-treatment (7.11 [5.54–14.67] mg/L, p < 0.001) (Figure 1E). This pattern was also observed in individual patients (Figure 1F). Total protein did not change significantly between the on- and off-treatment cycles (Figures 1D and 1E). The A1M-FITC uptake assay was performed in absence of AON to evaluate whether long-term AON incubation had reduced the overall protein reabsorption capacity of these cells. This showed that the 7-day pre-incubation with AON1 did not affect BSA uptake by cPTECs (Figure 4B). Also no changes in cell viability were found, both with and without 24 h recovery (Figure 4C).

Low Molecular Weight Proteinuria Correlates with Oligonucleotide Accumulation in Monkey Kidney

Cynomolgus monkeys treated weekly with s.c. drisapersen at 0, 2, 6, and 12 mg/kg for 39 weeks demonstrated a dose-dependent increase of A1M (i.e., median values increased 4, 6, and 21-fold over untreated controls, at doses of 2, 6, and 12 mg/kg, respectively). Urinary total protein concentrations were only slightly elevated (<2-fold). The increase in A1M correlated well (R² = 0.98, p < 0.01) with the accumulation of drisapersen in the kidney cortex (Figure 2C). Histopathological evaluation of the kidney did not reveal any tubular abnormalities apart from a dose-related increase of the antisense oligonucleotide manifested as the presence of basophilic granules in proximal tubules (data not shown).

Endocytic Uptake and Limited Degradation of Oligonucleotides in Proximal Tubular Cells In Vitro

To evaluate uptake of antisense oligonucleotides in vitro, we incubated human cPTECs with drisapersen, antisense oligonucleotide 1 (AON1), or antisense oligonucleotide 2 (AON2) (oligonucleotides with similar 20MePS chemistry to drisapersen but with a different nucleotide sequence) (Figure 3A). Incubation at different time points (4 and 24 h) and different concentrations (ranging from 0.3 to 10× Cmax of drisapersen in clinical trials) showed a time- and concentration-dependent uptake of AON in cPTECs (Figures 3B and 3C). Recovery for 20 or 48 h following AON uptake did not reduce levels in the cell, indicating that the intracellular antisense oligonucleotides are not sensitive to degradation within this time period. Confocal microscopy using fluorescently labeled AONs (Figures 3D and 3E) demonstrated that the AONs accumulate in a vesicular pattern, suggesting sequestration in endosomes. Co-localization with fluorescently labeled albumin (BSA-Alexa 657) (Figure 3D) or GST-tagged receptor-associated protein (RAG-GST) (Figure 3E), two known ligands of receptor-mediated endocytosis, confirmed that the uptake of AONs in tubule cells occurs via the same endocytic pathway.

Long-Term Exposure to Antisense Oligonucleotides Does Not Result in Cytotoxicity

To evaluate long-term adverse effects of renal tubule cell exposure to antisense oligonucleotides, we pre-incubated cPTECs with AON1 at Cmax levels for 7 days, followed by 24 h recovery in absence of AON (Figure 4). Next, the BSA uptake assay was performed in absence of AON to evaluate whether long-term AON incubation had reduced the overall protein reabsorption capacity of these cells. This showed that the 7-day pre-incubation with AON1 did not affect BSA uptake by cPTECs (Figure 4B). Also no changes in cell viability were found, both with and without 24 h recovery (Figure 4C).

Antisense Oligonucleotides Compete with Protein Reabsorption In Vitro

The reversible low molecular weight proteinuria observed in patients may be the result of competition for receptor-mediated endocytosis rather than a consequence of direct tubular toxicity. The receptor proteins megalin and cubilin at the apical membrane of the proximal tubule mediate reabsorption by binding and uptake of ligands, including A1M, receptor associated protein (RAG), and albumin. RAP (gene name LRPAP1) is a 39 kDa chaperone protein that binds to megalin with high affinity, whereas albumin is a large ~68 kDa plasma protein that in small amounts can pass the glomerular filter and is efficiently reabsorbed by the proximal tubule. Therefore we examined the uptake of fluorescently labeled A1M (A1M-FITC) (Figures 5A–5D), fluorescently labeled BSA-FITC (Figures 5E–5G), and RAP-GST (Figures 4H–4L) in cPTECs in the absence or presence of antisense oligonucleotides. At 37°C, A1M-FITC was actively taken up over time by cPTECs (Figure 5A) and could be inhibited by excess unlabeled A1M (Figure 5B). Antisense oligonucleotides (AON1 or drisapersen) reduced A1M-FITC uptake in cPTECs in a time- and concentration-dependent manner (Figures 5C and 5D), suggesting competitive interaction with the same receptor.
Co-incubation with AON1 or AON2 significantly inhibited BSA-FITC uptake (up to 50%) in ciPTECs at both 30 min and 4 h in a concentration-dependent manner (Figures 5E–5G). The presence of antisense oligonucleotides also interfered with RAP-GST cell surface binding at 4°C (up to 70% reduction in binding) and inhibited active uptake of RAP-GST at 37°C (up to 25% reduction) (Figures 5H–5J).

Pre-incubation with oligonucleotides for 24 or 48 h prior to incubation with the labeled substrate did not significantly inhibit the uptake of RAP-GST at 37°C (Figures 5H and 5I), indicating that antisense oligonucleotide incubation directly competes with receptor binding capacity and has no cytotoxic effect that affects the megalin- or cubilin-mediated endocytosis rate. Some residual inhibition was seen at 4°C after pre-incubation, which is likely due to occupation of ligand binding sites at the cell surface that were not washed away at 4°C.

Together these data indicate that antisense oligonucleotides directly compete with receptor-mediated protein reabsorption in ciPTECs, without adversely affecting the reabsorption capacity of the cells.

DISCUSSION
Antisense oligonucleotide therapeutics accumulate dose dependently in renal proximal tubular cells, and renal tubular effects are commonly associated with oligonucleotide treatment in both animal studies and in the clinic. Little is known, however, about the cellular mechanism and the potential adverse effects. In this study, we investigated how 2OMePS oligonucleotides affect urinary protein excretion, epithelial reabsorption, and cytotoxicity by combining clinical,
pre-clinical, and in vitro data. Our data support the hypothesis that 2OMePS oligonucleotides compete with receptor-mediated endocytosis in the proximal tubule epithelium and reduce the uptake of endogenous ligands such as A1M, resulting in reversible low molecular weight proteinuria in patients and animals.

Low molecular weight proteins pass the glomerular filter and are prevented from excretion through active tubular reabsorption. The presence of A1M in urine is, therefore, often considered indicative of impaired tubular function. Our clinical data show that the levels of A1M in urine correlated strongly to antisense oligonucleotide therapy in DMD patients, but normalized during planned off-treatment periods. Total protein remained well below levels associated with renal disease, suggesting reversible proximal tubule function impairment rather than cytotoxicity. Of note, urinary protein was not normalized to urinary creatinine, because this is highly dependent on skeletal muscle mass and unsuitable as a reference in DMD patients. Although the level of oligonucleotide accumulation in renal tissue from patients is not known, the tubular effects are thought to be closely linked to the degree of uptake into proximal tubular cells. This was also seen in the monkeys where drisapersen accumulation in the kidney closely correlated with the urinary protein (without any histopathological evidence of tubular toxicity after 39 weeks of treatment). Elimination of oligonucleotides is known to follow first-order kinetics; after a steady state is reached, the oligonucleotide concentration in cells does not increase further with increasing duration of treatment. The kinetics of tissue uptake and elimination are primarily determined by the PS backbone of the oligonucleotide, but species differences challenge prediction of tubular accumulation in humans. However, when patients were treated with drisapersen for 72 weeks, levels of A1M and total protein increased in the first weeks of treatment and remained stable thereafter, reflecting steady-state kinetics. It is also important to note that the reversible effects in the tubular compartment described in this paper are distinct from glomerular effects that have occasionally been seen with PS antisense oligonucleotides. Glomerular effects are related to the inflammatory potential of oligonucleotides, for which animals are thought to be more sensitive than humans.

In the current study, antisense oligonucleotides accumulated in human proximal tubule cells in vitro without cytotoxic effects after 7 days of exposure to antisense oligonucleotide at concentrations equivalent to or exceeding peak plasma levels of drisapersen in humans. Consistent with the long tissue half-life of 2OMePS oligonucleotides (estimated in humans to be over 5 weeks), no significant degradation was seen after 20 h or 48 h recovery. To investigate the link between oligonucleotide treatment and proteinuria, we evaluated receptor-mediated protein reabsorption by proximal tubule cells in vitro, using the well-characterized ciPTEC model. This cell line is human-derived and has demonstrated tubular epithelial features, including functional basolateral and apical drug transporters such as organic cation transporter 2, P-glycoprotein 1, and multidrug resistance proteins, as well as intact receptor-mediated endocytosis, including expression of megalin and cubilin. Dysfunction of proximal tubular protein reabsorption by megalin and cubilin causes low molecular weight proteinuria in both animal models and human disease, and as we hypothesize, so does competitive inhibition via saturation by oligonucleotide treatment.

A1M and, to a lower extent, albumin are both present in the glomerular filtrate and are almost completely reabsorbed by receptor-mediated endocytosis in proximal tubule cells. Incubation of ciPTECs with antisense oligonucleotides indeed reduced the binding and uptake of A1M, albumin, and RAP, whereas pre-incubation with antisense oligonucleotide for 24 h and up to 7 days did not diminish protein reabsorption. This suggests a direct competition at the receptor level, rather than a reduction in reabsorption capacity due to intracellular oligonucleotide accumulation. The oligonucleotides may block protein receptor binding sites at the cell surface, or directly interact with A1M, albumin, and RAP in the solution that can prevent their binding and internalization. Both hypotheses would explain the reduced uptake of proteins from the filtrate and their release in the urine.

In this study we focused on the effect of oligonucleotide exposure on protein reabsorption and cell viability in proximal tubule cells. Although we find that the 2OMePS oligonucleotides interfere with protein uptake, this does not directly imply that the proteins can also block oligonucleotide uptake, or that the bulk of oligonucleotides are taken up by the same pathway as the protein ligands. So far, several different pathways have been described for the uptake of PS-modified oligonucleotides, which vary greatly among different tissues and organisms and can even depend on the oligonucleotide sequence.

To be therapeutically active following endocytosis in the tubule cells, the oligonucleotides have to reach the cytoplasm or nucleus without degradation in the endo-lysosomal compartment. Although the
receptors and pathways involved have not been fully elucidated, a distinction is generally made between a productive pathway (oligonucleotides reach their target and lead to gene knockdown or alternative splicing) or a nonproductive pathway (oligonucleotides stay in the endo-lysosomal compartment and are degraded).38,40

The majority of PS-modified oligonucleotides are protein bound in the blood (>90%), which facilitates its uptake into cells and tissues and reduces the free fraction available for glomerular filtration.43 This raises the question whether in vivo proximal tubule cells will encounter the oligonucleotides mostly in its unbound or protein-bound form, because this may change the uptake route and interaction with membrane proteins. Animal studies have also shown that PS-modified oligonucleotides can enter proximal tubule cells from both the tubular and the capillary side, and different uptake mechanisms may be involved in each case.44 Here the in vitro oligonucleotide uptake experiments (Figures 3B and 3C) have been performed in serum-free medium, so uptake has taken place in the absence of serum proteins like albumin. We also evaluated whether coincubation with BSA, A1M, or RAP-GST could block or enhance oligonucleotide uptake, but the presence of these ligands did not affect oligonucleotide uptake over 24 h (data not shown). This suggests that oligonucleotide uptake predominantly takes place in its free form and uptake is not facilitated by binding to serum proteins like albumin.

In conclusion, our data support a model of direct competition of 2OMePS antisense oligonucleotide with protein reabsorption in proximal tubule cells, consistent with the observations of reversible proteinuria in patients and the absence of tubular damage in monkeys. This supports the limited risk for kidney tubular injury at therapeutic doses of PS oligonucleotides.

MATERIALS AND METHODS

Compounds

The oligonucleotides used in this study have a fully modified 2OMePS backbone and were synthesized at BioMarin Nederland B.V. (Leiden, the Netherlands). Sequences of drisapersen (PRO051), AON1, and AON2 are 5'-UCA AGG AAG AUG GCA UUU CU-3' (targets dystrophin exon 51), 5'-UAU GAG UUU CUU CCA AAG CAG CCU C-3' (5'-methylated Cs) and 5'-GAG UUU CUU CCA AAG CAG CCU CUC-3', respectively. AON1-Texas red and drisapersen-Cy3 were synthesized by Biospring (Frankfurt, Germany) and BioMarin Nederland, respectively.

Urinalysis in Human Subjects

Urine samples were collected during an open-label study in 12 DMD patients receiving drisapersen (DMD141673).14 Treatment involved weekly s.c. administration at 6 mg/kg body weight for 72 weeks, followed by an 8-week treatment interruption, and subsequently 9 intermittent treatment cycles of 8 weeks on-treatment/4 weeks off-treatment. First morning urine was used for total protein and A1M analysis, before or 1 week after treatment. Excretion was not normalized to creatinine because DMD patients have reduced muscle mass.24,45

Study in Monkeys

Cynomolgus monkeys (Macaca fascicularis) received drisapersen (in 20 mM phosphate buffer with 0.8% w/v NaCl) for 39 weeks by weekly s.c. administration at doses of 0, 2, 6, and 12 mg/kg body weight. Overnight urine was collected and analyzed for total protein and creatinine using ADVIA 1650 (Siemens), and A1M using Myriad RBM’s Human CustomMAP. At necropsy (39 weeks) kidney cortex was snap-frozen for drisapersen analysis by hybridization-ligation
The in vivo study was ethically reviewed and carried out in accordance with European Directive 86/609/EEC.

Cell Culture
Development of the ciPTECs used in this studied is described in detail elsewhere.20 In brief, primary renal cells were isolated from the urine of a healthy volunteer and immortalized by transduction with human telomerase (hTERT) and the temperature sensitive SV40 large T antigen (SV40T) to promote respectively genome stability and cell expansion at 33°C. In this way, a stable clone exhibiting robust proximal tubule characteristics was selected and further characterized based on the presence of an intact endocytic machinery and physiological and pharmacological characteristics, as well as toxicological responses.16–23 Cells were seeded 7 days before the experiment at 55,000 cells/cm² and grown for 1 day at 33°C and 5% v/v CO₂, followed by 6 days at 37°C and 5% v/v CO₂ to confluence in culture medium as previously described to stimulate receptor-mediated endocytosis. Experiments are performed in complete medium (containing DMEM HAM’s F12, Life Technologies, Paisley, UK), 5 µg/mL insulin, 5 µg/mL transferrin, 35 ng/mL epidermal growth factor (EGF), 40 µg/mL tri-iodothyronine (Sigma, St. Louis, MO, USA), and 10% fetal calf serum (FCS, Greiner Bio One, Kremsmuenster, Austria), or in culture medium without FCS, named “serum-free medium” (SFM).

Antisense Oligonucleotide Uptake
Cells were washed (HBSS, Invitrogen, Carlsbad, CA, USA) and exposed to 0.32–10.8 µM AON1 or 0.34–11.4 µM AON2 for 4 h or 24 h at 37°C. Samples were either harvested immediately or subsequently incubated in SFM to recover for 20 h (4 h exposure) or 48 h (24 h exposure). Cells were harvested by washing in HBSS, heparin (20 U/mL, Sigma, St. Louis, MO, USA), and HBSS, before lysis in RLT buffer (1 min, room temperature; QIAGEN, Hilden, Germany). Lysates were evaluated as described previously.27

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Alpha-1-microglobulin

A

B

C

D

BSA-FITC

E

F

G

RAP-GST

H

I

J

K

L

(legend on next page)
Functional Uptake Assay upon Long-Term Pre-Incubation
Cells were exposed to 1.08 μM AON1 in complete culture medium for 7 days at 37°C 5% v/v CO₂, refreshed daily. Cells were washed (SFM) and incubated in absence of antisense oligonucleotides for 24 h. BSA-FITC uptake capacity was evaluated by washing cells (SFM) and incubation with 0.15–7.3 μM BSA-FITC in SFM for 4 h at 37°C. After washing 3 times with ice-cold HBSS (Invitrogen, Carlsbad, CA, USA), fluorescence was quantified by plate reader (ex 485 nm; em 535 nm).

Viability Assay upon Long-Term Pre-Incubation
Cells were exposed to 1.08 μM AON1, refreshed daily. Cells were washed (SFM) and either first incubated in SFM in absence of antisense oligonucleotides for 24 h before the tetrazolium dye MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (Sigma) was added (0.5 mg/mL, 3 h), or the MTT assay was performed immediately. Formazan was dissolved in DMSO (Merck, milliQ) (Sigma) was added (0.5 mg/mL, 3 h), or the MTT assay was performed immediately. Formazan was dissolved in DMSO (Merck, Whitehouse Station, NJ, USA) and optical density was measured (560 nm, background at 670 nm was subtracted) using Benchmark Plus (Bio-Rad, Hercules, CA, USA).

Data Analysis and Statistics
For antisense oligonucleotide uptake and inhibition assays, data are either presented as lysate concentration or normalized to samples without inhibitor. For A1M-FITC, BSA-FITC, and RAP-GST uptake and inhibition assays, data are either presented as absolute fluorescence values or normalized to the activity of samples without antisense oligonucleotide and plotted after background subtraction. For MTT viability assay, data were normalized to the viability of control cells. Statistics was performed by one or two-way ANOVA (two-tailed, α = 0.05) using GraphPad Prism. All data are plotted with GraphPad Prism (version 6.07) and are presented as mean ± SD of at least two separate experiments (n = 2) performed in triplicate, unless stated otherwise.

AUTHOR CONTRIBUTIONS

ACKNOWLEDGMENTS
This work was supported by a collaboration research agreement from BioMarin Nederland B.V., Leiden, the Netherlands. We would like to thank Nicole Datson from BioMarin for her continued support and advice on this project and processing of the current manuscript.

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