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Male pseudohermaphroditism due to a homozygous missense mutation of the luteinizing hormone receptor gene

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Leydig cell hypoplasia is a rare autosomal recessive condition that interferes with normal development of male external genitalia in 46,XY individuals. We have studied two Leydig cell hypoplasia patients (siblings born to consanguineous parents), and found them to be homozygous for a missense mutation (Ala593Pro) in the sixth transmembrane domain of the luteinizing hormone (LH) receptor gene. In vitro expression studies showed that this mutated receptor binds human choriongonadotropin with a normal KD, but the ligand binding does not result in increased production of cAMP. We conclude that a homozygous LH receptor gene mutation underlies the syndrome of autosomal recessive congenital Leydig cell hypoplasia in this family. These results have implications for the understanding of the development of the male genitalia.

The process that leads to the development of male sex characteristics is generally thought to consist of three sequential steps. The first is the establishment of male genetic sex by the XY chromosome pattern at conception. The second involves the induction of male gonadal sex, that is testis formation, through the action of SRY (sex determining region of the Y chromosome). The third step consists of the development of internal and external male genitalia, a process which is basically controlled by two factors, testosterone and anti-Müllerian hormone (AMH), both produced by the fetal testis. The Sertoli cell product, AMH, prevents the formation of the müllerian duct derivatives (fallopian tubes, uterus and the upper part of the vagina), whereas the development of the male internal and external genitalia is dependent on the Leydig cell product, testosterone, which is converted to dihydrotestosterone in certain target tissues. Various genetic defects in this pathway have been reported, supporting the concept of sequential determination of male genetic, gonadal and phenotypic sex.

An important step in this differentiation cascade is the development of the Leydig cells. This apparently begins independent of stimulation by gonadotropins at approximately the same time that the formation of the testicular cords occurs, although the process still occurs in cases where testicular cords do not develop. Leydig cell differentiation is crucial for sufficient androgen production by the testis. Absence of Leydig cells or insufficient Leydig cell differentiation can occur as an autosomal recessive condition. The phenotype of inherited Leydig cell hypoplasia ranges from extreme forms presenting as 46,XY females to milder forms in which males present with hypergonadotropic hypogonadism and a micropenis.

Testicular luteinizing hormone (LH) binding was decreased or absent in some studies, which could be either the cause or the consequence of hypoplasia of Leydig cells. Here, we report studies of the LH receptor gene in siblings born to consanguineous parents who have male pseudohermaphroditism due to Leydig cell hypoplasia.

Clinical details
Our study examined two 46,XY siblings who presented with female external genitalia, primary amenorrhea, and lack of breast development. Their parents are first cousins.
and there are 14 additional siblings (Fig. 1). Both cases had a short blind ending vagina, without uterus or fallopian tubes. Serum levels of testosterone and testosterone precursors were abnormally low, and did not respond to stimulation by hCG. Basal levels of LH were markedly increased, but follicle-stimulating hormone (FSH) was within the normal range. The gonads were removed, and upon histological examination found to be testes with normal Sertoli cells, but no mature Leydig cells (Fig. 2).

Homozygous mutation in the LH receptor gene

The LH receptor gene of the two patients with male pseudohermaphroditism was studied for the presence of mutations by single strand conformational polymorphism (SSCP) analysis. We began by studying exon 11 which contains the transmembrane and cytoplasmic domains, with two sets of primers (see Methodology). Amplification with primer set 1 produced a fragment showing abnormal migration. This fragment comprises the third extracellular loop and the seventh transmembrane segment, as well as parts of the sixth transmembrane segment and the intracellular tail.

This part of the LH receptor gene was then sequenced in the two affected siblings and in two unaffected controls with primers la and lb as sequencing primers. A GCC to CCC transversion was detected at position 1787 in the patients (Fig. 3a), producing an Ala593Pro substitution in the sixth transmembrane domain (Fig. 3b). The normal sequence could not be detected, indicating homozygosity of the mutated allele. The presence of two copies of the LH receptor gene was confirmed by dosage analysis (data not shown).

The mutation inactivates signal transduction

The Ala593Pro mutation was constructed in vitro, and the resulting mutant LH receptor cDNA transiently expressed in human embryonic kidney 293 cells. The high affinity (mean $K_p = 0.8 \pm 0.15$ nM; $n=2$) binding of $[^{125}I]$-labelled choriogonadotropin (hCG) to partially purified membrane preparations was no different from binding to membrane preparations from cells transfected with the wild type LH receptor cDNA (mean $K_p = 0.5 \pm 0.4$ nM; $n=2$) (Fig. 4). The difference in the values of $B^{max}$ of the different membrane preparations is partly due to experimental variation in transfection efficiency (five-fold less for the mutant LH receptor construct), and partly to reduced expression of the mutant receptor at the cell membrane. The number of mutant or wild-type receptors expressed per transfected cell varied between 1,000 and 3,000, which is similar to the figure found in vivo. In contrast, when hCG-induced cAMP production was determined, no hormonal effect was detected for the mutated LH receptor, even at very high (1000 ng/ml) hCG concentrations (Fig. 4). These results indicate that the missense Ala593Pro mutation completely abolishes signal transduction, probably at the level of coupling to the...
A homozygous missense mutation in the sixth half of the LH receptor molecule. This function resides in the N-terminal to the receptor albeit at lower maximal capacity. No hermaphroditism due to Leydig cell hypoplasia in our inherited Leydig cell hypoplasia is consistent with previous studies of other patients which have indicated reduced LH independent male precocious puberty. The Ala593Pro mutation abolishes LH receptor function in vitro. This is very different from the effect of missense mutations of the LH receptor gene that have been found in patients with autosomal dominant LH independent male precocious puberty. The Ala593Pro mutation, by contrast, might introduce a more drastic conformational change in the receptor caused by the constrained angle conferred by the proline residue. Such mutations are more likely to render the receptor inactive. Many studies indicate that undifferentiated Leydig cells require LH for proliferation and differentiation. In humans, fetal Leydig cell number and testosterone production correlate with plasma hCG levels. This is consistent with experimental studies in rats that have shown that full-length LH receptor mRNA is expressed in testis from day 16.5 of gestation onwards, suggesting an active role for LH/hCG-dependent testosterone production in the determination of male external genitalia. After specific removal of differentiated Leydig cells from adult rat testes, immature Leydig cells reappear more quickly if LH is present. Furthermore, LH can induce morphological differentiation of immature to mature Leydig cells in neonatal rats, compared to the situation in fetal life where the Wolffian duct-derived structures can only have an active role for LH/hCG-dependent testosterone production at a later fetal stage. This differentiation is paralleled by an increase in the activity of the enzymes that are responsible for testicular steroidogenesis.

The initiation of androgen synthesis by Leydig cells early in fetal life may be independent of LH or hCG as indicated by data from rats and rabbits. Our findings may support this notion, because testes with associated epididymis and vas deferens were found in both patients. These wolffian duct-derived structures can only have an active role for LH/hCG-dependent testosterone production at a later fetal stage. This differentiation is paralleled by an increase in the activity of the enzymes that are responsible for testicular steroidogenesis.
cell hypoplasia, the testes contain normal Sertoli cells, with no mature Leydig cells. Sometimes, small clusters of immature Leydig cells are present\(^7\). The histological findings in Leydig cell hypoplasia can now be interpreted as reflecting lack of stimulation through the LH receptor.

No abnormal female sex characteristics have been noted in sisters of patients with this form of male pseudohermaphroditism\(^13\), which is consistent with experimental results that indicate absence of a functional ovarian LH receptor until after birth\(^21\). However, LH receptor function is required for a normal female menstrual cycle. Hence, the presence of primary amenorrhea in a 46,XX sister of the patients described above should be of particular interest. (As this patient lives approximately 4,000 kilometres from our centre in rural Brazil, we have not yet been able to contact the patient and obtain a blood sample.) Further studies should reveal if she is indeed homozygous for the same LH receptor missense mutation.

Primary amenorrhea with ovarian resistance to luteinizing hormone might conceivably be the female counterpart to the pattern of male pseudohermaphroditism studied here. Amenorrhea has been noted in another sister of a patient with no mature Leydig cells, the testes contain normal Sertoli cells, and a short blind ending vaginal pouch. The karyotype was 46,XY.

**Methodology**

**Patients.** The patients are two siblings born to consanguineous first cousin parents. The patients originate from a small village in rural north/northeast Brazil (Fiaui State). They were referred at ages 37 and 42, because of primary amenorrhea and lack of breast development. Both cases had a euthyroid habitus, absence of breast tissue, and sparse axillary hair. The external genitalia were female type, with distinct urethral and vaginal orifices, and triangle-shaped pubic hair. Pelvic ultrasound examination showed no internal genitalia, hypoplasia\(^13\).

**Amenorrhea has been noted in another sister of a patient with male pseudohermaphroditism\(^12\), which is consistent with experimental results that indicate absence of a functional ovarian LH receptor until after birth\(^21\). However, LH receptor function is required for a normal female menstrual cycle. Hence, the presence of primary amenorrhea in a 46,XX sister of the patients described above should be of particular interest. (As this patient lives approximately 4,000 kilometres from our centre in rural Brazil, we have not yet been able to contact the patient and obtain a blood sample.) Further studies should reveal if she is indeed homozygous for the same LH receptor missense mutation.

Primary amenorrhea with ovarian resistance to luteinizing hormone might conceivably be the female counterpart to the pattern of male pseudohermaphroditism studied here. Amenorrhea has been noted in another sister of a patient with primary amenorrhea due to Leydig cell hypoplasia\(^13\).

In conclusion, the finding of an LH receptor defect in male pseudohermaphroditism with testicular Leydig cell hypoplasia provides a biological basis for this disorder. Our data support a crucial role for the LH receptor in Leydig cell differentiation, and hence in the regulation of embryonal production of testosterone by the male gonad.

**Mutation analysis.** Genomic DNA was isolated from peripheral blood as described\(^24\). Because other family members live approximately 4,000 kilometres from Sao Paulo, only the index cases were studied.

DNA fragments were amplified with PCR and analysed by SSCP analysis\(^17\) with the following adaptations for primer set I (see below); 1 mM Mg\(_2\)Cl\(_2\) in the PCR and an annealing temperature of 38°C. Two different sets of primers were used in the PCR: (1a) 5'-CGTATTCCAAGCATTGGC-3' (nt 1731-1748), (1b) 5'-CCCCGAGCTTACAGGAGC-3' (nt 1924-1942); (2a) 5'-GATLGCAATGTTGTCG-3' (nt 1834-1854), (2b) 5'-GGATTGAGAAGGCTTATTGG-3' (nt 2054-2078). Nucleotide numbers are according to Minegishi et al.\(^27\).

For sequencing, the DNA fragments were separated on a 1.5% LMP agarose gel BRL Life Technologies and purified with the gelase system (Epicentre Technologies) according to the "fast protocol" given by the manufacturer. About 50 fmol of the purified DNA fragments and the 1a and 1b primers were used for sequencing with the cycle sequencing kit of BRL (BRL Life Technologies).

**In vitro expression.** The wild type human LH receptor expression plasmid (pSG5-hLHR) was constructed by cloning the human LH receptor cDNA (nucleotides -3 to 2374 relative to the translation start site) into the EcoRI site of pSG5, an expression vector that contains the SV40 large T early promoter, intron 11 of the rabbit \(\beta\)-globin gene, and an SV40 polyadenylation signal\(^8\). The protein encoded by this cDNA differs from the published human LH receptor\(^27\) by the insertion of two amino acids (Leu and Gin) at positions 20 and 21 (E.M., unpublished observations). The Ala593Pro mutation was constructed in pSG5-hLHR by site-directed mutagenesis using PCR\(^26\), yielding pSG5-hLHR4. Mutagenesis was confirmed by sequence analysis, and revealed an additional nucleotide change (C to A) at position 1983 in the pSG5-hLHR vector. This change does not alter the amino acid encoded by this codon. Human embryonic kidney 293 (HEK293) cells were transiently transfected as described before\(^8\), 48 h after transfection. 100 ng of the expression plasmid (pRSVluc\(^3\)) and pSG5, pSG5-hLHR or pSG5-hLHR4. 48 h after transfection, [\(^{35}\)S]-hC (Dupont) binding to partially purified membranes was determined according to Ketelslegers & Catt\(^31\).

For the estimation of hCG-dependent cAMP production, HEK293 cells were co-transfected in 12 well tissue culture plates (Costar, Cambridge, MA, USA) with a luciferase expression plasmid pRSVluc\(^3\) and pSG5, pSG5-hLHR or pSG5-hLHR4. 48 h after transfection cells were incubated for 2 h with culture medium containing 0.1% BSA and 0.2 mM isobutylmethylxanthine (Sigma) and different concentrations of hCG (urinary form).
The remaining cells were lysed, and luciferase activity was determined on the same wells. Subsequent correction for transfection efficiency was determined by subtraction of basal cAMP production in pSG5-transfected cells. We thank John Phillips III, of Nashville, Tennessee for his help in organizing this study, and Theo van der Knaap for help with the photomicrographs. Partially supported by FAPESP (94/1316-9,92/254-9).