Radioactive holmium phosphate microspheres for cancer treatment

A.G. Arranjaa,b,c,d, W.E. Henninkb, A.G. Denkovac, R.W.A. Hendrikxe, J.F.W. Nijsena,d,f,*

a Department of Radiology and Nuclear Medicine, University Medical Center Utrecht, Heidelberglaan 100, 3584 CX Utrecht, The Netherlands
b Department of Pharmaceutics, Utrecht Institute for Pharmaceutical Sciences (UIPS), Science for Life, Faculty of Science, Utrecht University, 3508 TB Utrecht, The Netherlands
c Radiation Science and Technology, Delft University of Technology, Mekelweg 15, 2629 JB Delft, The Netherlands
d Radboudumc, Department of Radiology and Nuclear Medicine, Geert Grooteplein Zuid 10, 6525 GA Nijmegen, The Netherlands
e X-Ray Facilities, Department of Materials Science and Engineering, Delft University of Technology, Faculty of 3mE, Mekelweg 2, 2628 CD Delft, The Netherlands
f Quiem Medical B.V., Zuiphenseweg 55, 7418 AH Deventer, The Netherlands

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ABSTRACT

The aim of this study was the development of radioactive holmium phosphate microspheres (HoPO4-MS) with a high holmium content and that are stable in human serum for selective internal radiation therapy (SIRT) of liver cancer. To this end, holmium acetylacetonate microspheres (HoAcAc-MS) were prepared (34.2 ± 1.0 µm in diameter, holmium content of 46.2 ± 0.8 and density of 1.7 g/cm³) via an emulsification and solvent evaporation method. The concentration of HoAcAc in the organic solvent, the temperature of emulsification and the stirring speed were varied for the preparation of the HoAcAc-MS to obtain microspheres with different diameters ranging from 11 to 35 µm. Subsequently, the AcAc ligands of the HoAcAc-MS were replaced by phosphate ions by simply incubating neutron irradiated HoAcAc-MS in a phosphate buffer solution (0.116 M, pH 4.2) to yield radioactive HoPO4-MS. The obtained microspheres were analyzed using different techniques such as SEM-EDS, ICP-OES and HPLC. The prepared HoPO4-MS (29.5 ± 1.2 µm in diameter and a density of 3.1 g/cm³) present an even higher holmium content (52 wt%) than the HoAcAc-MS precursor (46 wt%). Finally, the stability of the HoPO4-MS was tested by incubation in human serum at 37 °C which showed no visible changes of the microspheres morphology and only 0.1% of holmium release was observed during the 2 weeks period of incubation. In conclusion, this study shows that stable radioactive HoPO4-MS can be prepared with suitable properties to be used for cancer therapy.

1. Introduction

Liver cancer is the fifth most diagnosed cancer type in men and the second most common cause of death from cancer in the world (Ferlay et al., 2015). Actually, more than 700,000 people are diagnosed with primary liver tumors every year worldwide (American Cancer Society) and almost 50% of the patients with colorectal carcinoma will develop metastatic liver cancer (over 1 million patients) (Kelly and Kemeny, 2017). Particularly in these patients, spreading of the tumor to the liver is the major cause of death due to the low response to chemotherapy and external radiotherapy (Valderrama-Treviño et al., 2017; Zarour et al., 2017). Moreover, very few treatment alternatives are available for these group of patients. Hepatic resection is used in up to 15% of the cases with reported 5-year survival rates ranging from 25% to 51%. Unfortunately, most of these patients are not eligible for hepatic resection due to the characteristics of the metastatic lesions (size, number and location) and the presence of extra hepatic disease (Misiakos et al., 2011). Due to the extremely aggressive nature and poor prognosis of primary and metastatic liver cancer, it remains an important public health issue (Soerjomataram et al., 2012; Zarour et al., 2017). Thus, different alternative therapies have been proposed over the last years.

Selective internal radiation therapy (SIRT) of the liver is a type of radionuclide therapy consisting in placing radioactive microspheres with a diameter between 20 and 50 µm in the direct vicinity of the liver tumors in order to deliver high radiation doses directly to malignant cells leaving healthy tissue unaffected (Anderson et al., 1991; Dendy et al., 2017; Meade et al., 1987; Rognoni et al., 2016). This local radionuclide therapy using microspheres loaded with beta-emitting isotopes is a very promising therapeutic modality for inoperable patients suffering from liver malignances. Therefore, several generations of microspheres for SIRT have been developed in recent years comprising different radionuclides, such as yttrium-90 (90Y) and holmium-166 (166Ho), embedded in matrix materials such as biodegradable polymers, ion exchange resins and glass (Meade et al., 1987; Nijsen...
et al., 2002a; Pasciak, 2017). Currently, two 90Y and one 166Ho-based microspheres formulations are commercially available and in clinical use for liver radioembolization: ceramic-based TheraSphere® (MDS Nordion, Canada) and resin-based SIR-Spheres® (SIRTex, Medical Ltd., Australia) containing 90Y, and polymer-based microspheres QuiremSpheres® (Quirem Medical B.V., The Netherlands) loaded with 166Ho (Nijssen et al., 2002a; Pasciak, 2017; Prince et al., 2017).

The holmium loaded microspheres have shown similar results to the other products for SIRT in unrespectable liver metastases (Prince et al., 2017; Smits et al., 2012, 2010) and their efficacy in neuroendocrine and head and neck tumors is currently being investigated (van Nimwegen et al., 2017). These microspheres loaded with the radioactive 166Ho are produced upon neutron activation of 166Ho-based microspheres (166Ho → +n → 166Ho, cross section 66 barn) (Nijssen et al., 2001; Zielhuis et al., 2006). 166Ho emits high-energy beta-minus particles (β⁻) (E_{max} = 1.74 (48.7%) and 1.85 (50%) MeV) with a relatively short half-life of 26.8 h (166Ho → 166Er + β⁻) and a maximum soft-tissue range of 8.4 mm, making this isotope very suitable for use as a therapeutic radionuclide. Importantly, 166Ho also emits low-energy gamma photons (6.2% 81 keV, 0.93% 1.38 keV), that enables the visualization of its distribution and quantification by single-photon emission computed tomography (SPECT) imaging after administration of the microspheres. Moreover, 165Ho is paramagnetic which allows its visualization using magnetic resonance imaging (MRI) for personalized treatment and monitoring of treatment progression and efficacy (Smits et al., 2013). Therefore, a medical device for SIRT based on 166Ho has considerable advantages of using common imaging modalities such as SPECT and high resolution MRI when compared to the 90Y isotope based products.

The commercially available formulation of 166Ho loaded microspheres (QuiremSpheres®) has a mean of 19 wt% (range 16.6–20.4%) of 166Ho content and this radioactive element is stably incorporated in a polymeric matrix of the biodegradable poly-L-lactic acid (PLLA). In the present work, the microspheres with a high holmium content (HoPO4-MS). The stability of these 166HoPO4-MS was finally examined in human serum.

2. Materials and methods

2.1. Materials

Holmium chloride (HoCl₃·6H₂O; MW 379.38; 99.9%) was obtained from Metall Rare Earth Limited. Acetylacetonate (AcAc; ReagentPlus®; MW 100.45; 99.9%) was obtained from ACROS Organics. Ammonium hydroxide (NH₄OH; EMSURE®; MW 35.05; 30–30%), hydrochloric acid (HCl; EMSURE®; MW 37.36; 37%), nitric acid (EMPROVE® and nitric acid (SUPRAPUR®; 65%) were supplied by Millipore.

Fig. 1. Schematic overview of the preparation of radioactive holmium phosphate microspheres: (1) holmium acetylacetonate microspheres (HoAcAc-MS) were prepared by dissolution of holmium acetylacetonate crystals in chloroform followed by solvent evaporation. The obtained microspheres were (2) neutron activated in a nuclear research reactor where the 165HoAcAc-MS were converted into radioactive 166HoAcAc-MS. Finally, (3) the AcAc ligands were exchanged by phosphate ions by simply incubation of the 166HoAcAc-MS in a phosphate buffer solution to yield radioactive holmium phosphate microspheres (HoPO₄-MS).
2.1. Methods

2.1.1. Preparation of HoAcAc microspheres

A solvent evaporation method was used to prepare HoAcAc microspheres (HoAcAc-MS) (Fig. 1, step 1). For this purpose, crystals of holmium acetylacetonate (HoAcAc) were prepared as previously reported by Nijsen et al. (Nijsen et al., 1999). Briefly, an aqueous solution of holmium chloride was mixed with a diluted solution of acetyl acetone and the pH of this solution was adjusted to 8.5 with ammonium hydroxide solution. Complexes of holmium and acetylacetonate were formed overnight, which were subsequently collected and washed with water three times and dried under vacuum at 48 °C for at least 48 h. For the preparation of the HoAcAc-MS, 10 g of HoAcAc crystals were dissolved in chloroform (186 g) and the obtained homogeneous solution was then added to an aqueous solution of PVA (1000 g water with 2% w/w PVA). Overhead four blades propeller stirrers (IKA Eurostar power digi-visc) were used to vigorously stir the mixture in two liters baffled beakers to obtain an oil-in-water (o/w) emulsion which was stirred at a constant speed and temperature. The temperature of the emulsion was controlled using jacketed beakers and a constant flow of nitrogen (12 L/min) was applied for 72 h. Different parameters in this procedure were varied to assess their influence on the final properties of the microspheres: (1) concentration of HoAcAc in the organic solvent (5.4, 6.3 and 7.5% w/w), (2) temperature of emulsification (15, 25 and 35 °C) and (3) stirring speed (300, 400, 500 and 600 rpm). After the emulsification/solvent evaporation procedure, the HoAcAc microspheres were formed (Fig. 1, step 1). The microspheres were collected by centrifugation, washed three times with water and sieved according to the desired size (typically 20–50 μm) using an electronic sieve vibrator (TOPAS EMS 755) and an ultrasonic processor (Hielscher UP200S). Finally, the sieved microspheres were dried at room temperature for 5 h under ambient pressure followed by vacuum drying at room temperature for 72 h.

2.1.2. Neutron activation of HoAcAc microspheres

Dry 166HoAcAc-MS were neutron activated in the pneumatic rabbit system (PRS) of the research nuclear reactor facility operational at the Department of Radiation Science and Technology of the Delft University of Technology, The Netherlands. Irradiation was performed in polyethylene vials (Vente et al., 2009; Vente et al., 2010) and irradiated with a thermal flux of 4.97 × 10^16 n m^-2 s^-1, epithermal flux of 8.13 × 10^14 n m^-2 s^-1 and fast neutrons flux of 3.48 × 10^15 n m^-2 s^-1. Different amounts of 166HoAcAc-MS (50, 100 and 600 mg) were irradiated for 2, 4, 6 or 8 h to yield radioactive 166HoAcAc-MS (Fig. 1, step 2). The maximum temperature during irradiation was monitored with temperature indicator strips that were attached to the vials immediately prior to irradiation (Digi-Sense, Cole-Parmer). The radioactive 166HoAcAc-MS were allowed to decay for at least one month before handling the samples to reduce exposure of the operator to radiation. The decayed neutron irradiated HoAcAc-MS were analyzed by Scanning Electron Microscope-Energy Dispersive X-ray Spectroscopy (SEM-EDS), x-ray powder diffraction (XRD), Inductively Coupled Plasma-Optical Emission spectroscopy (ICP-OES), High Performance Liquid Chromatography (HPLC) and Gas Chromatography-Mass Spectrometry (GC-MS) to determine the effects of neutron irradiation on the surface properties and chemical composition of the HoAcAc-MS (methods described below).

2.1.3. Stability of HoAcAc microspheres in water after neutron activation

After neutron activation, the stability of the irradiated HoAcAc-MS was evaluated by incubating the microspheres in water for injection (100 mg in 10 mL). The morphological properties of the microspheres were analyzed by optical microscopy and the possible holmium leakage was determined by gamma-scintillation counting of the metastable isotope 166mHo (2480 Wizard2 Automatic Gamma Counter, Perkin Elmer) over a period of 48 h. The metastable isotope 166mHo has a very long half-life (~1200 y) and is formed in a fixed ratio of 7 ppm relative to 166Ho during irradiation of 165Ho, enabling an accurate measurement of the 166Ho release from the microspheres even after the 166mHo non-metastable isotope has been decayed (Seppenwoolde et al., 2005).

2.1.4. Formation of HoPO4 microspheres after neutron activation of HoAcAc microspheres

To form HoPO4-MS (Fig. 1, step 3), the irradiated HoAcAc-MS were incubated (approximately 100 mg per 10 mL) in a phosphate buffer solution (0.116 M NaH2PO4, pH 4.2) with gentle shaking (tube roller shaker) at room temperature. The formation kinetics of the HoPO4-MS was determined by HPLC analysis by measuring the acetylacetonate released from the microspheres at different time points. Light microscopy was used to observe the microspheres during exchange of the AcAc ligands by the phosphate ions. The final HoPO4-MS were characterized by size distribution analysis, light and scanning electron microscopy and the holmium and phosphorus contents were determined by ICP-OES. Finally, the chemical composition of the HoPO4-MS was determined by XRD.

2.1.5. Holmium release during formation of HoPO4 microspheres and stability in human serum

After forming of the HoPO4-MS (Fig. 1, step 3), the phosphate buffer was removed by centrifugation and replaced by human serum (approximately 100 mg in 10 mL). Human serum was collected from blood donors of the University Medical Center of Utrecht after incubation of whole blood in Clot Activator Tubes and centrifugation to collect serum. The stability of the HoPO4-MS was tested at 37 °C under constant shaking in a C24 incubator shaker (New Brunswick Scientific, Edison, USA). The microspheres were centrifuged and samples of human serum were collected at different time points to quantify the amount of 166mHo in the supernatant (8 mL). After incubation, the microspheres were washed with water three times, dried under vacuum and the holmium and phosphorus contents were determined by ICP-OES.

2.1.6. Characterization of HoAcAc crystals, (radioactive) HoAcAc microspheres and radioactive HoPO4 microspheres

The HoAcAc crystals, HoAcAc-MS (before and after neutron irradiation) and radioactive HoPO4-MS were characterized for their holmium (Ho) and phosphorous (P) contents using ICP-OES. Samples of 20 to 50 mg were dissolved in 50 mL of 2% nitric acid and the holmium concentration of the solutions was measured at three different wavelength (339.9, 345.6 and 347.4 nm) and the phosphorous concentrations at two wavelength (213.6 and 214.9 nm) using an Optima 4300 CV (PerkinElmer, Norwalk, USA).

The HoAcAc-MS (5 to 15 mg) were dissolved in methanol (10 mL) and the acetylacetonate (AcAc) content was determined by HPLC analysis using an Alliance HPLC system using a C18 column (XSelect CSH C18 3.5 μm 4.6 × 150 mm, Waters) at 40 °C and a 70:30 mixture of methanol and water with 0.1% perchloric acid as the mobile phase (0.5 mL/min). Detection was done at 280 nm.

The water content of vacuum dried HoAcAc crystals and HoAcAc-MS was determined using a Karl Fisher Coulometer (831, Metrohm) by dissolving samples of approximately 20 to 50 mg in 1 mL of methanol. The Karl Fisher method could not be applied to the HoPO4-MS because these microspheres are not soluble in methanol. Therefore, the water content in the HoPO4-MS after vacuum drying the microspheres at room temperature was determined by thermogravimetric analysis (TGA, TA Instruments Q-50) using a samples of approximately 25 mg.

The size distribution of HoAcAc-MS (before and after sieving) and one month decayed radioactive HoPO4-MS was determined using a Coulter counter equipped with an orifice of 100 μm (Multisizer 3, Beckman Coulter, Mijdrecht, The Netherlands).

The density of the HoAcAc crystals and the different microspheres with a sample amount of approximately 250 mg was determined in...
water using a 25 cm$^3$ specific gravity bottle (Blaubrand NS10/19, DIN ISO 3507, Wertheim, Germany).

For identification of the chemical compounds present in non-irradiated and neutron irradiated HoAcAc-MS, 5 mg of samples were dissolved in 1 mL of methanol ($n = 4$ for each condition tested) and analyzed by GC-MS in a Shimadzu Gas Chromatograph-Mass Spectrometer GCMS-QP2010 system equipped with a VF-5 ms column (35 m $\times$ 0.25 mm $\times$ 0.25 µm film thickness). The injection temperature was 265 °C and the column oven was set at 50 °C (8 min hold) and increased to 290 °C at 100 °C/min (6 min hold). The carrier gas was He at a constant flow rate of 1.0 mL/min. The total ion chromatograms (TIC) were obtained and analyzed using the GC-MS Solution v272 software.

The identification of the compounds of the different peaks was performed by MS using the mass spectral library NIST 2011. The MS was operated in full scan mode over 35–500 m/z. The MS transfer line temperature was held at 300 °C and the ion source temperature at 200 °C. After identification, the area under the peak from the TIC was used for quantification of the acetylacetonate present in the samples.

XRD patterns of the HoAcAc crystals, HoAcAc-MS before irradiation, decayed neutron activated HoAcAc-MS and decayed HoPO$_4$-MS were obtained by depositing a small amount (around 5 mg) of the different sample on a Si-510 wafer and analyzed using a Bruker D8 Advance diffractometer in Bragg-Brentano geometry with a Lynxeye position sensitive detector.

Optical microscopy (AE2000 Motic) was used to investigate the morphological properties of the microspheres (sphericity and surface damages) obtained under the various preparation conditions. The surface composition of the HoAcAc crystals, HoAcAc-MS, decayed radioactive HoAcAc-MS and HoPO$_4$-MS was studied using a SEM-EDS (JEOL JSM-T100, InTouchScope™, Tokyo, Japan).

3. Results and discussion

3.1. Preparation of holmium acetylacetonate microspheres (HoAcAc-MS)

Microspheres with a holmium content of 45 wt% were previously prepared using a conventional emulsification and solvent evaporation process (Bult et al., 2012, 2007, 2009). These holmium acetylacetonate microspheres (HoAcAc-MS, Fig. 2B) were prepared by dissolution of holmium acetylacetonate crystals (Fig. 2A) in chloroform followed by solvent evaporation (Fig. 1, step 1).

Several parameters of the preparation process can influence the properties of the obtained microspheres. To this end, the influence of the (1) concentration of HoAcAc in the organic solvent (5.4, 6.3 or 7.5 wt%), the (2) temperature of emulsification (15, 25 and 35 °C) and the (3) stirring speed (300, 400, 500 and 600 rpm) on the size of the HoAcAc-MS were studied. It was observed that the concentration of HoAcAc in the organic phase is critical to obtain microspheres in the final dispersion without contamination with HoAcAc crystals. When relatively high concentrations of HoAcAc in the organic phase were used (6.3 or 7.5 wt%), the HoAcAc-MS were contaminated with HoAcAc crystals (Fig. S1). Using 5.4 wt% of HoAcAc in chloroform resulted in the formation of only HoAcAc-MS (Fig. 2B). The mean size, chemical composition and density of the obtained HoAcAc-MS are reported in Table 2. The density of the HoAcAc crystals precursor (1.6 g/cm$^3$) is very similar to that of the HoAcAc-MS (1.7 g/cm$^3$), demonstrating that the microspheres are non-porous.

HoAcAc-MS were also prepared at different temperatures (15, 25 and 35 °C; Table 1). An increasing diameter of the microspheres was observed with higher temperature of the mixture: the average mean diameter for microspheres prepared at 15 °C was 27 µm whereas at 35 °C the formed microspheres had a diameter of 33 µm (Table 1). Similar findings were reported in previous studies where the temperature of emulsification was also varied to prepare polymeric microspheres (Mateovic-Rojnik et al., 2005; Sahoo et al., 2007; Yang et al., 2000a; Yang et al., 2000b). The vapor pressure of chloroform is 16.8, 26.2 and 39.5 kPa at 15, 25 and 35 °C (http://www.ddbst.com/ddb.html). Thus, higher temperatures provide a greater driving force for evaporation leading to a faster hardening of the droplets to finally yield into microspheres. This rapid solvent removal at higher temperatures will shorten the time to further decrease the size of the droplets by the applied shear forces during stirring (Scheme S1) (Li et al., 1995; Maia et al., 2004; Mateovic-Rojnik et al., 2005).

The effect of the stirring speed on the size of the microspheres was also studied and it was observed that the average diameter of the microspheres decreased from 31.2 ± 1.2 µm to 11.1 ± 1.6 µm with increasing stirring speed from 300 to 600 rpm respectively (Fig. 3). Likely, a higher stirring speed yielded due to the higher shear forces an emulsion with smaller chloroform droplets finally resulting after evaporation of the solvent in smaller HoAcAc microspheres.

The research on the examined parameters resulted in a standard and reproducible method for the preparation of HoAcAc-MS under the following conditions: 5.4 wt% of HoAcAc in chloroform as the dispersed phase, 25 °C and 300 rpm at a fixed concentration of emulsifier (2%)
and 72 h of emulsification/solvent evaporation. The so obtained microspheres (n = 12) had a mean size of 31.2 ± 1.2 µm and, after sieving through 50 and 20 µm sieves, a mean size of 34.2 ± 1.0 µm. The used parameters yields 3.4 ± 0.3 g of microspheres after sieving.

The recovery of holmium was ca. 45% from 10 g HoAcAc crystals (density 1.6 g/cm3 and holmium content 35.2 ± 1.5% (w/w)) to HoAcAc-MS (density of 1.7 g/cm3 and a holmium content of 46.2 ± 0.8 (w/w)).

The HoAcAc crystals and the HoAcAc-MS were characterized using different techniques. The results show that HoAcAc crystals are formed through the complexation of one holmium atom with three acetylacetonate molecules and one molecule of water (Table 2), resulting in a highly crystalline complex (Fig. 4A). These results are in line with previous findings (Kooijman et al., 2000; Nijsen et al., 2001). After dissolving this complex in chloroform and emulsification of obtained solution in an aqueous solution, the obtained HoAcAc-MS have a composition of one and a half acetylacetonate molecule and one water molecule per holmium atom and per water molecule. The new complex of the holmium and AcAc is not crystalline but amorphous as observed by XRD (Fig. 4B). As suggested by Bult et al., a network is formed in which one and a half acetylacetonate molecule is complexed with one holmium ion in an amorphous state (Bult et al., 2009).

### Table 1

Influence of temperature of emulsification on the average diameter, density and yield of holmium acetylacetonate microspheres (HoAcAc-MS, n = 6 for each temperature). HoAcAc-MS were prepared at different temperatures using an o/w emulsification and solvent evaporation followed by sieving through 50 and 20 µm sieves.

<table>
<thead>
<tr>
<th>Temperature [°C]</th>
<th>Size before sieving [µm]</th>
<th>Size after sieving [µm]</th>
<th>Density [g/cm³]</th>
<th>Yield 20–50 µm [g]</th>
<th>Yield 20–50 µm in terms of Ho recovery [%]</th>
</tr>
</thead>
<tbody>
<tr>
<td>15</td>
<td>26.7 ± 0.8</td>
<td>28.6 ± 1.1</td>
<td>1.76 ± 0.06</td>
<td>2.52 ± 0.25</td>
<td>33.8 ± 3.4</td>
</tr>
<tr>
<td>25</td>
<td>31.2 ± 1.2</td>
<td>34.2 ± 1.0</td>
<td>1.71 ± 0.05</td>
<td>3.39 ± 0.33</td>
<td>45.5 ± 4.4</td>
</tr>
<tr>
<td>35</td>
<td>33.2 ± 1.8</td>
<td>34.7 ± 0.9</td>
<td>1.72 ± 0.07</td>
<td>4.26 ± 0.27</td>
<td>57.2 ± 3.6</td>
</tr>
</tbody>
</table>

### Table 2

Characteristics of HoAcAc crystals (n = 6), the standard HoAcAc-MS (n = 12) and radioactive HoPO4-MS (n = 6).

<table>
<thead>
<tr>
<th>Sample</th>
<th>Calculated structure</th>
<th>Ho [wt%]</th>
<th>P [wt%]</th>
<th>AcAc [wt%]</th>
<th>H2O [wt%]</th>
<th>Density [g/cm³]</th>
<th>Mean diameter [µm]</th>
</tr>
</thead>
<tbody>
<tr>
<td>HoAcAc crystals</td>
<td>Ho(AcAc)3·H2O</td>
<td>35.2 ± 1.5</td>
<td>n.a.</td>
<td>60.1 ± 1.6</td>
<td>3.7 ± 0.4</td>
<td>1.61 ± 0.12</td>
<td>n.a.</td>
</tr>
<tr>
<td>HoAcAc-MS</td>
<td>Ho2(AcAc)3·2H2O</td>
<td>46.2 ± 0.8</td>
<td>n.a.</td>
<td>46.8 ± 1.2</td>
<td>5.2 ± 0.7</td>
<td>1.71 ± 0.05</td>
<td>34.2 ± 1.0</td>
</tr>
<tr>
<td>HoPO4-MS</td>
<td>HoPO4·3H2O</td>
<td>52.0 ± 2.3</td>
<td>9.7 ± 0.5</td>
<td>less than 0.01</td>
<td>20 ± 2.1</td>
<td>3.12 ± 0.08</td>
<td>29.5 ± 1.2</td>
</tr>
</tbody>
</table>

n.a. not applicable.

Fig. 3. Effect of emulsion stirring speed on the mean diameter of the obtained holmium acetylacetonate microspheres (HoAcAc-MS, n = 3 for each stirring speed). The temperature of emulsification (25 °C), concentration of HoAcAc in chloroform (5.4 w/v%), concentration of emulsifier (2%) and duration of emulsification (72 h) were fixed.

Fig. 4. X-Ray powder diffraction patterns of (A) HoAcAc crystals, (B) HoAcAc-MS, (C) radioactive HoAcAc-MS (600 mg irradiated for 4 h measured after 2 months) and (D) radioactive HoPO4-MS.

### 3.1. Neutron activation of HoAcAc-MS

HoAcAc-MS prepared under standard conditions were neutron activated to convert the non-radioactive 165Ho into the beta and gamma-emitting 166Ho (Fig. 1, step 2). In previous studies, several irradiation parameters were varied to investigate their influence on the obtained radioactive HoAcAc loaded poly(l-lactic acid) microspheres (Nijsen et al., 2002b; Nijsen et al., 1999; Vente et al., 2009). It was observed that the irradiation facility, duration of irradiation as well the size of the samples influenced the final properties of the microspheres. In the present study, the tested parameters were the irradiation time (2, 4, 6 and 8 h) and the amount of microspheres per vial (50, 100 or 600 mg). The different irradiation times allow to evaluate the effect of increasing doses of neutron irradiation on the microspheres, whereas the amount of sample per vial is important to optimize the amount of microspheres that will be administered to the patients (Smits et al., 2012).

In comparison with the current clinically used 166Ho loaded microspheres (QuiremSpheres®), the substantially higher holmium content of the HoAcAc-MS (46 wt%) resulted in a higher radioactivity per
mg of sphere by a factor of 2.5. Within 6 h of neutron activation of 600 mg of HoAcAc-MS, an activity of 47 GBq can be produced at the end of bombardment (thermal flux $4.97 \times 10^{16}$ n m$^{-2}$ s$^{-1}$) compared to 19 GBq for the QuiremSpheres®.

It was observed that both the irradiation time and volume of sample in the irradiation vial had an effect on the final appearance of the HoAcAc-MS. After 2 or 4 h irradiation, SEM analysis of the microspheres showed a smooth spherical appearance of the microspheres identical to non-irradiated microspheres regardless the amount of microspheres used per vial (Fig. 5). However, when longer irradiation periods were used (8 h), the microspheres displayed cracks on their surface. Unexpectedly, the amount of microspheres per vial had a substantial effect on the damages of the microspheres: when relatively low amounts of microspheres were irradiated (50 or 100 mg), the microspheres showed surface damages with irradiation times of 6 and 8 h. On the other hand, when a larger amount of microspheres was irradiated (600 mg), no significant damages were observed up to 8 h irradiation.

Due to the fact that some of the neutron activated and thereafter decayed HoAcAc-MS presented cracks on their surface after irradiation, HoAcAc-MS before and after neutron activation were analyzed by GC–MS to detect whether degradation compounds are formed. Fig. 6 shows that indeed several compounds were formed during irradiation of the HoAcAc-MS. Some of the compounds detected were methyl acetate and acetone (mass spectrums are displayed in Fig. S2) suggesting that the radiation induced degradation of the AcAc molecule. The radiation responsible for the degradation of the acetylacetonate molecule are the gamma rays which are present in the reactor during neutron irradiation of the microspheres. This is in line with the literature (Barker, 1963; Hummel, 1995) reporting that the dominating processes in the degradation of ketones upon exposure to ionizing radiation is the break of the C–C bond adjacent to the C=O bond and chemical reactions with radiolytic products of water (e.g. hydrated electrons, $\text{HO}_2$, $\text{H}_3\text{O}^+$, $\text{OH}^-$, $\text{H}_2\text{O}_2$ and $\text{H}_2$) (Čech, 1974; Rama Rao et al., 1970).

The degradation products formed during irradiation have boiling points around 60 °C (boiling points of acetone and methyl acetate are 56 °C and 57 °C respectively) and the average temperature reached during irradiation of the samples was 54–60 °C. It can be therefore expected that the evaporation of these volatile degradation products during irradiation will result in surface cracking of the microspheres. Moreover, the amount of microspheres per vial plays a particular role in the heat transfer and therefore influences the heating degree of the samples. When smaller samples are used, there is less heat conductivity resulting in a faster heating up and thus a higher temperature of the microspheres which in turn will result in a higher driving force for
evaporation of volatile degradation products and subsequently in the formation of cracks. The AcAc content of the HoAcAc-MS after irradiation under different conditions was quantitatively determined by GC-MS and HPLC and it was observed that this content decreased with increasing irradiation time (Fig. S3). The XRD pattern of the irradiated HoAcAc-MS remains similar to the non-irradiated HoAcAc-MS (Fig. 4C) showing that, after neutron activation, the microspheres remained amorphous.

3.2. Stability of HoAcAc-MS in water after neutron activation

The stability of the irradiated HoAcAc-MS was investigated by incubating the particles in water for injection. Non-irradiated HoAcAc-MS are stable in water. On the other hand, it was observed that the HoAcAc-MS irradiated for 4 h in a vial of 600 mg were not stable in water for injection and released more than 90% of their radioactive 166mHo within 2 days (Fig. S4a). Optical microscopy observations revealed a progressive degradation of the microspheres with complete disintegration of the microspheres after 48 h incubation in water for injection (Fig. S4b). The instability of the irradiated microspheres points to irradiation damage.

Interestingly, in previous work from our group, it was observed that when the irradiated HoAcAc-MS were incubated in a phosphate buffer, the microspheres remained spherical andholmium release was not observed (Bult et al., 2012). The reasons for this stability in a phosphate buffer were further investigated in the present study.

3.3. Formation of HoPO4-MS by incubation of irradiated HoAcAc-MS in a phosphate buffer

As pointed out above, irradiated HoAcAc-MS incubated in phosphate buffer were stable (Bult et al., 2012). In this work, after incubation of the irradiated HoAcAc-MS in a phosphate buffer (0.116 M, pH 4.2) for 1 h at room temperature (Fig. 1, step 4), the resulting microspheres were characterized by ICP-OES. It was found that they contain 9.7 wt% of phosphorus, 52 wt% of holmium and no traces of AcAc were detected (less than 0.01%) (Table 2). This revealed that during incubation of the irradiated HoAcAc-MS in the phosphate buffer, the AcAc ligands were replaced by phosphate ions forming new complexes of phosphate and 166Ho.

The formation kinetics of the 166HoPO4 complexes was also monitored by HPLC through the release of AcAc over time (Fig. 7A). It was observed that 100% of the AcAc present in the HoAcAc-MS was released within 1 h of incubation in phosphate buffer. Interestingly, imaging of the microspheres by optical microscopy showed the formation of a ring-like structure that increased over time and correlated with the amount of AcAc released (Fig. 7B). Likely, the AcAc ligands are exchanged by phosphate ions resulting in HoPO4, which has different refractive index than the HoAcAc.

The exchange reaction was performed at pH 4.2 because acetylacetone is essentially in its non-charged state (pKa = 9.0) decreasing the stability of the HoAcAc complexes (Smith, 2010; Stary and Liljenzin, 1982). This in turn favors the release of the AcAc and subsequent binding of the phosphate ions. During the exchange reaction, it was observed that a maximum of 0.28 ± 0.02% (n = 8) of 166mHo was released during 1 h incubation. Moreover, the geometry of the microspheres was retained (Fig. 7B) while acetylacetone and its degradation products were completely exchanged with phosphate ion to yield radioactive 166HoPO4-MS. Thermogravimetric analysis of the HoPO4-MS revealed a total weight loss of 21.5 wt% which corresponds to water loss (Fig. S5). Altogether, these results show that the final HoPO4-MS have a chemical composition of one holmium atom per phosphate group complexed with three water molecules (Table 2).

After exchange of the AcAc by the PO4 ions, it was observed by optical (Fig. 7B) and scanning electron microscopy (Fig. 2C) that the microspheres retained their spherical shape without alterations of their surface morphology. It was further observed that the mean diameter of the microspheres decreased from 34.2 ± 1.0 µm to 29.5 ± 1.2 µm (Fig. 2B and 2C, Table 2), which corresponds to a reduction of the microspheres volume of approximately 30% and consequently resulted in an increase of the microsphere’s density from 1.7 to 3.1 g/cm3 (Table 2). XRD analysis shows that HoPO4-MS are fully amorphous (Fig. 4D).

3.4. Stability of radioactive HoPO4-MS in human serum

Anticipating the use of the HoPO4-MS for internal radionuclide therapy, the stability of the microspheres was tested in human serum. The microspheres had a very good stability and less than 0.1% of 166Ho was released during their incubation in serum for two weeks (Fig. 8A). This is in line with the literature where it is described that rare earth metal phosphates are practically insoluble in aqueous media (Kijowski and LeGeros, 2005) and explains the high stability of these microspheres in serum. Furthermore, no changes of the microspheres were observed by optical microscopy (Fig. 8B) and the size distribution remained constant during the 2 weeks period of incubation.

4. Conclusions

The study shows that stable radioactive HoPO4 microspheres with a
very high holmium content for internal radionuclide therapy can be produced. The previously described HoAcAc-MS with a 46 wt% holmium content were prepared and the influence of different processing parameters was investigated to define the optimal preparation conditions. The most critical parameter was the stirring speed and the mean diameter of the HoAcAc-MS could be easily tailored from 11 to 31 µm. It was also observed that after irradiation the microspheres are unstable in water for injection. Therefore, holmium phosphate microspheres (HoPO₄-MS) were prepared from the neutron irradiated HoAcAc-MS by incubating the microspheres in a phosphate buffer. The radioactive HoPO₄-MS that have a high holmium content (52 wt%) are very stable in human serum (holmium release less than 0.1%). It is concluded that these HoPO₄-MS are attractive microspheres for application in the SIRT technology for liver malignancies and for intratumoral treatment of solid tumors. Future studies will focus on the pre-clinical testing of these microspheres.

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5. Declaration of interest

JFW Nijsen is inventor on the patents related to the HoAcAc-MS and HoPO₄-MS which are assigned to University Medical Center Utrecht Holding BV and/or Quirem Medical (patent families: USA Patent No. 6,373,068 B1, PCT/NL03/00485, EP07112807.8, 10190254.2, P114198PC00, P112614NL00). He is co-founder and chief scientific officer of Quirem Medical, and has a minority share in the company Quirem Medical. The activities of JFW Nijsen within Quirem Medical are approved and supported by Dirkjan Masman (Director Technology Transfer Office Radboudumc) and Mathias Prokop (Head of Radiology and Nuclear Medicine at Radboudumc). All authors have revised and have approved the final manuscript.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at https://doi.org/10.1016/j.ijpharm.2018.06.036.

References


