Controlled Human Malaria Infection with Graded Numbers of Plasmodium falciparum NF135.C10- or NF166.C8-Infected Mosquitoes

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Abstract. Controlled human malaria infections (CHMIs) with Plasmodium falciparum (Pf) parasites are well established. Exposure to five Pf (NF54)-infected Anopheles mosquitoes results in 100% infection rates in malaria-naive volunteers. Recently Pf clones NF135.C10 and NF166.C8 were generated for application in CHMIs. Here, we tested the clinical infection rates of these clones, using graded numbers of Pf-infected mosquitoes. In a double-blind randomized trial, we exposed 24 malaria-naive volunteers to bites from one, two, or five mosquitoes infected with NF135.C10 or NF166.C8. The primary endpoint was parasitemia by quantitative polymerase chain reaction. For both strains, bites by five infected mosquitoes resulted in parasitemia in 4/4 volunteers; 3/4 volunteers developed parasitemia after exposure to one or two infected mosquitoes infected with either clone. The prepatent period was 7.25 ± 4.0 days (median ± range). There were no serious adverse events and comparable clinical symptoms between all groups. These data confirm the eligibility of NF135.C10 and NF166.C8 for use in CHMI studies.

INTRODUCTION

Controlled human malaria infections (CHMIs) are a well-accepted tool used since the 1980s for the exploration of immunology and pathophysiology of malaria infections and for evaluation of candidate vaccines and drugs. The majority of studies have been conducted with 3D7 and its parental strain NF54.1 Most volunteers have been infected using five laboratory-reared Plasmodium falciparum (Pf)-infected Anopheles mosquitoes, which reproducibly result in optimal infection rates in malaria-naive volunteers.2,3 Reducing the number of NF54 or 3D7 Pf-infected mosquitoes to one or two reduces the infection rate.4,5

Given the diversity of Pf isolates in the field, the value of CHMI trials can be increased by expanding the portfolio with Pf clonal isolates from different geographical origins.6 The Pf isolate NF54, most likely originates from Africa.7 The more recently generated clones are NF135.C10 and NF166.C8, originating from Cambodia and Guinea, respectively. Initial studies with NF135.C10 and NF166.C8 show that these clones are safe and give high infection rates with five infectious mosquitoes (80% [8/10] and 100% [5/5], respectively).6,8 However, these clones show more effective hepatocyte invasion and liver merozoite development as compared with NF54, which results in higher first peak of parasitemia and shorter prepatent period.9 In order to establish an NF135.C10- or NF166.C8 CHMI model with comparable parasite dynamics as obtained in NF54-infected volunteers, the objective of the current study was to compare infection rates and dynamics of parasitemia as well as clinical manifestations using one, two or five NF135.C10- or NF166.C8- infected mosquitoes.

MATERIALS AND METHODS

The study is a single-center, double-blind, randomized trial. The flowchart, screening procedures, and inclusion and exclusion criteria were described previously.10 Briefly, 24 healthy male and female volunteers, aged 18–35 years, were randomly assigned to six groups and exposed to bites from five mosquitoes of which five, two, or one were infected with either NF135.C10 or NF166.C8. Uninfected mosquitoes were added to preserve blinding of all trial personnel except the staff members preparing and aliquotting infected mosquitoes.

Anopheles stephensi mosquitoes were reared and infected according to standardized protocols.11,12 The average number of sporozoites per mosquito used in this study was 69,000 for NF135.C10 and 51,000 for NF166.C8. Mosquito feeding was allowed for 10 minutes and infectivity of mosquitoes was assessed after feeding by dissecting salivary glands for the presence of sporozoites. Feeding was repeated until the predefined number of infected bites was reached. Eight volunteers needed two sessions, one needed three sessions (all in the five- or two-mosquito bite groups).

After CHMI, volunteers were followed-up twice daily from day 5 post-infection until two consecutive blood samples were positive by quantitative polymerase chain reaction (qPCR) (≥500 parasites/mL), or, when they remained negative, until day 13. At this point, volunteers were treated with atovaquone/proguanil and followed on days 1, 2, 3, and 7 after treatment and on day 35 post-infection. Follow-up was prolonged if volunteers remained qPCR-positive or symptomatic. Throughout the study, high-sensitive troponin T, lactate dehydrogenase (LDH), and platelet counts were measured as safety parameters. The first peak of parasitemia was calculated as the geometric mean of Pf parasites per milliliter between day 6.4 and 8.4.13

The trial was registered at www.clinicaltrials.gov, identifier NCT02149550. Ethical approval from the Central Committee on Research Involving Human Subjects (NL48704.000.14) was obtained.

All statistical analyses were performed with IBM SPSS statistics for Windows (Version 23.0; IBM Corp., Armonk, NY). Mean prepatent period, first peak of parasitemia, and the frequency of adverse events (AEs) between the groups were assessed by using unpaired t test, one-way analysis of
The number of volunteers developing parasitemia and the prepatent period in days after infection with graded numbers of NF135.C10- or NF166.C8-infected mosquitoes

<table>
<thead>
<tr>
<th>Strain</th>
<th>No. of infectious bites</th>
<th>Five, two, or one</th>
<th>Five</th>
<th>Two</th>
<th>One</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>NF135.C10</td>
<td>Subjects PCR positive (n/total)</td>
<td>10/12</td>
<td>4/4</td>
<td>3/4</td>
<td>3/4</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td>Prepatent period (days [median [range]])</td>
<td>7.0 [7.0–9.0]</td>
<td>7.5 [7.0–9.0]</td>
<td>9.0 [7.5–9.0]</td>
<td>7.0 [7.0–7.0]</td>
<td>0.06</td>
</tr>
<tr>
<td>NF166.C8</td>
<td>Subjects PCR positive (n/total)</td>
<td>10/12</td>
<td>4/4</td>
<td>3/4</td>
<td>3/4</td>
<td>--</td>
</tr>
<tr>
<td></td>
<td>Prepatent period (days [median [range]])</td>
<td>7.0 [7.0–11.0]</td>
<td>7.0 [7.0–7.0]</td>
<td>7.0 [7.0–11.0]</td>
<td>9.0 [7.5–9.0]</td>
<td>0.09</td>
</tr>
</tbody>
</table>

Differences between groups were tested using the Kruskal-Wallis method.
result in zero successful events. Such bottlenecks may include the frequency of mosquito probing, the number of sporozoites entering the blood stream, and the number that invade hepatocytes. After hepatocyte invasion, the efficiency of parasite multiplication may differ between strains with a potential higher first peak of parasitemia and correspondingly shorter prepatent periods for NF135.C10 and NF166.C8. This difference in multiplication inside hepatocytes is unlikely related to differences in fitness of these clones, as the IC50 of CSP-antibodies blocking in vitro hepatocyte development is comparable.9

Data from this and other studies with NF135.C10- and NF166.C8-infected volunteers showed that there is no increase in the number or severity of AEs compared with NF54-infected volunteers, despite the higher first peak of parasitemia.10 The prevalence and severity of AEs will likely further reduce if antimalarial treatment is initiated upon more stringent qPCR cut-off values.20

In conclusion, this study with limited numbers of volunteers per group shows that bites of five mosquitoes of NF135.C10 and NF166.C8 of either clone consistently gives 80–100% patent parasitemia in the three studies performed with these heterologous clones. In future CHMI studies using clones NF135.C10 and NF166.C8 and lower numbers of mosquito bites may suffice to achieve blood-stage dynamics which are more similar to NF54, albeit reaching lower infection rates.

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REFERENCES


