PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.
http://hdl.handle.net/2066/195208

Please be advised that this information was generated on 2020-02-13 and may be subject to change.
Variant in genes encoding small GTPases and association with epithelial ovarian cancer susceptibility


1 Department of Health Sciences Research, Mayo Clinic, Rochester, MN, United States of America, 2 Department of Oncology, University of Cambridge, Strangeways Research Laboratory, Cambridge, United Kingdom, 3 Department of Biostatistics and Bioinformatics, Moffitt Cancer Center, Tampa, FL, United States of America, 4 School of Public Health, Louisiana State University Health Sciences Center, New Orleans, LA, United States of America, 5 Division of Population Sciences, Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, FL, United States of America, 6 Netherlands Comprehensive Cancer Organization, Utrecht, The Netherlands, 7 Radboud University Medical Center, Radboud Institute for Health Sciences, Nijmegen, The Netherlands, 8 Genetic Epidemiology Research Institute, UCI Center for Cancer Genetics Research and Prevention, School of Medicine, Department of Epidemiology, University of California Irvine, Irvine, CA, United States of America, 9 Byelorussian Institute for Oncology and Medical Radiology, Aleksandrov N.N., Minsk, Belarus, 10 Cancer Prevention and Control, Rutgers Cancer Institute of New Jersey, New Brunswick, NJ, United States of America, 11 Department of Obstetrics and Gynecology, Oregon
Epithelial ovarian cancer (EOC) is the fifth leading cause of cancer mortality in American women. Normal ovarian physiology is intricately connected to small GTP binding proteins of the Ras superfamily (Ras, Rho, Rab, Arf, and Ran) which govern processes such as signal transduction, cell proliferation, cell motility, and vesicle transport. We hypothesized that common germline variation in genes encoding small GTPases is associated with EOC risk. We investigated 322 variants in 88 small GTPase genes in germline DNA of 18,736 EOC patients and 26,138 controls of European ancestry using a custom genotype array and
logistic regression fitting log-additive models. Functional annotation was used to identify bio-
features and expression quantitative trait loci that intersect with risk variants. One variant,
\textit{ARHGEF10L} (Rho guanine nucleotide exchange factor 10 like) rs2256787, was associated
with increased endometrioid EOC risk (OR = 1.33, \( p = 4.46 \times 10^{-6} \)). Other variants of interest
included another in \textit{ARHGEF10L}, rs10788679, which was associated with invasive serous
EOC risk (OR = 1.07, \( p = 0.00026 \)) and two variants in \textit{AKAP6} (A-kinase anchoring protein 6) which
were associated with risk of invasive EOC (rs1955513, OR = 0.90, \( p = 0.00033 \);
rs927062, OR = 0.94, \( p = 0.00059 \)). Functional annotation revealed that the two \textit{ARHGEF10L}
variants were located in super-enhancer regions and that \textit{AKAP6} rs927062 was associated
with expression of GTPase gene \textit{ARHGAP5} (Rho GTPase activating protein 5). Inherited var-
iants in \textit{ARHGEF10L} and \textit{AKAP6}, with potential transcriptional regulatory function and asso-
ciation with EOC risk, warrant investigation in independent EOC study populations.

\section*{Introduction}
In 2017, in the United States, more than 21,000 women were expected to be diagnosed with
epithelial ovarian cancer (EOC), and more than 14,000 women were predicted to die from the
disease.\cite{1} EOC is heterogeneous and therefore classified into major histological subtypes of
invasive disease—serous, endometrioid, clear cell, and mucinous—and two histological sub-
types of borderline disease—serous and mucinous. These histological subtypes have differences
in genetic and epidemiologic risk factors, molecular events during oncogenesis, response to
chemotherapy, and prognosis.\cite{2}

Approximately 20\% of the familial component of EOC risk is attributable to high-to-inter-
mediate risk gene mutations.\cite{3} In European populations, genome-wide association studies
(GWAS) have identified more than 30 EOC susceptibility alleles, as reviewed previously.\cite{4}
Known common genetic variants explain 3.9\% of the inherited component of EOC risk, and
additional susceptibility loci are likely to exist, particularly for the less common, non-serous
histological subtypes.

Normal ovarian physiology is intricately connected to tightly regulated small GTP binding
proteins of the Ras superfamily (Ras, Rho, Rab, Ral, Arf, and Ran) which regulate key cellular
processes such as signal transduction, cell proliferation, cell motility, and vesicle transport.\cite{5}
These proteins function in a highly coordinated manner through signaling networks and feed-
back loops within and among the small GTPase subfamilies.\cite{6} The Rab and Rab GTPases are
thought to function in membrane trafficking in exocyst assembly and vesicle-tethering pro-
cesses;\cite{7, 8} Rho-related proteins function to integrate extracellular signals with specific targets
regulating cell morphology, cell aggregation, tissue polarity, cell motility and cytokinesis.\cite{5}
Ras family genes cycle between their inactive GDP forms in the cytoplasm and the active GTP-
bound forms on the plasma membrane and are associated with signaling pathways contribut-
ing to normal and aberrant cell growth.\cite{9}

As regulation of the RAS signal transduction pathway involves a highly complex, highly
polymorphic machinery of genes, we conducted a large-scale candidate pathway association
study, hypothesizing that variation in small GTPase genes is associated with EOC risk.

\section*{Materials and methods}
\subsection*{Variant selection}
RAS pathway genes were selected based on the Cancer Genome Anatomy Project and review
of the published literature (\url{www.pubmed.gov}). Within 115 candidate genes, 6103 single
nucleotide polymorphism (SNPs) were interrogated in early GWAS analysis of 7931 EOC patients and 9206 controls;[10] 339 SNPs in 88 of these genes showed nominal evidence of association with risk of EOC or of serous EOC (p < 0.05 using all participants or North American participants only)[10] and were targeted in the present analysis (S1 Table).

Study participants and genotyping
We studied 18,736 EOC patients (10,316 of serous histology) and 26,138 controls who participated in Ovarian Cancer Association Consortium studies; all participants were of European ancestry.[11] This included participants from the GWAS which was used for variant selection (described above)[10] and an additional 10,243 patients and 16,932 controls. Genotyping used a custom Illumina Infinium array. [11] SNPs were excluded according to the following criteria: no genotype call; monomorphism; call rate less than 95% and minor allele frequency > 0.05 or call rate less than 99% with minor allele frequency < 0.05; evidence of deviation of genotype frequencies from Hardy-Weinberg equilibrium (p < 10^{-7}); greater than 2% discordance in duplicate pairs. Overall, 322 small GTPase gene SNPs were genotyped and passed QC; numbers of participants with data for each SNP vary, as some DNA samples failed QC for particular SNPs. This study was reviewed and approved by the Mayo Clinic Institutional Review Board as protocol 1367–05.

Genetic association
We followed STREGA guidelines for genetic association studies.[12] Unconditional logistic regression treating the number of minor alleles carried as an ordinal variable (log-additive model) was used to evaluate the association between each SNP and EOC risk adjusted for age, study site, and principal components to account for residual differences in European ancestry. Six series of analyses were conducted considering the following groups: all invasive EOC combined, each of the four main invasive histological subtypes (serous, endometrioid, clear cell and mucinous), and all borderline tumors combined. No corrections were made for multiple testing.

Functional annotation
For SNPs of interest, dbSUPER [13] and Haploreg v4.1[14] were used to evaluate publicly available data for variant overlap with human super-enhancers,[15] known expression quantitative trait loci (eQTL), GWAS hits, and other regulatory marks. In addition, we assessed correlations between germline genotype with tumor expression levels (eQTL analysis) using 312 Mayo Clinic patients (226 serous, 54 endometrioid, 22 clear cell, 5 mucinous, and 5 of other histological subtypes). Expression data were obtained using fresh frozen tumor RNA and Agilent whole human genome 4x44 expression arrays and were analyzed in the form of log ratios of signals from individual tumors compared to signals from a reference mix of 106 tumor samples[16, 17] versus signals from a reference mix of 106 tumor samples[16, 17]. Expression levels for minor allele carriers versus non-carriers were compared using the Wilcoxon rank sum statistic.

Results and discussion
Demographic and clinical characteristics of the study sample (18,736 EOC patients and 26,138 controls) have been described previously.[11] In brief, compared to controls, patients were older, attained menarche at older ages, and had higher body mass index. As expected, most tumors (57.6%) were of serous histology with 14.2% endometrioid, 7.1% clear cell, 6.5% mucinous, and 14.6% other/unknown.
From among 322 SNPs in 88 RAS pathway small GTPase genes, we observed that 99 SNPs in 43 genes were nominally associated with EOC risk (p<0.05) (S2 Table). These associations were from six separate analyses that evaluated all patients with invasive disease, patients with one of the four main invasive histological subtypes, serous [n = 8,372], endometrioid [n = 2,068], clear cell [n = 1,025] and mucinous [n = 943], as well as patients with borderline tumors.

In ARHGEF10L, which encodes the Rho guanine nucleotide exchange factor 10-like protein, SNP rs2256787 was associated with invasive endometrioid EOC risk (OR = 1.33, 95% CI: 1.18–1.50, p = 4.5x10^{-6}) (Table 1). (Fig 1) shows the ORs and 95% CIs associated with the G allele at this SNP overall and by contributing study.

Three other variants were associated at p-value<10^{-4} (Table 1, S1, S2 and S3 Figs). rs10788679 in an intron of ARHGEF10L was associated with risk of invasive serous EOC (OR = 1.07, 95% CI: 1.03–1.11, p = 2.6x10^{-4}); ARHGEF10L SNPs rs2256787 and rs10788679 are independent (r^2 = 0.02, 1000 Genomes Project EUR). In addition, rs1955513 was most strongly associated with all invasive EOC risk (OR = 0.90, 95% CI: 0.85–0.95, p = 3.3x10^{-5}). This variant lies in an intron of A-kinase (PRKA) anchor protein 6 (AKAP6). Another variant in AKAP6, intronic SNP rs927062, was also associated with all invasive EOC risk (p = 5.9x10^{-4}); AKAP6SNPs rs1955513 and rs927062 are in modest linkage disequilibrium (r^2 = 0.15, 1000 Genomes Project EUR).

We investigated whether the four variants of interest, rs2256787, rs10788679, rs1955513, rs927062, which are all intronic, alter expression of their proximal GTPases, or coincide with regulatory marks that may affect expression (Table 1). In publicly available databases,[13, 14] the ARHGEF10L SNPs rs2256787 and rs10788679 coincide with a human ovary super-enhancer, a region of the genome with unusually strong enrichment for the binding of

<table>
<thead>
<tr>
<th>Gene</th>
<th>SNP</th>
<th>Chr:Position</th>
<th>Alleles</th>
<th>MAF</th>
<th>Histology</th>
<th>OR  (95% CI)</th>
<th>P-value</th>
<th>Conserved site</th>
<th>eQTL</th>
<th>Tissues with enhancer histone mark</th>
<th>Tissues with DNAse site</th>
<th>In super-enhancer</th>
</tr>
</thead>
<tbody>
<tr>
<td>ARHGEF10L</td>
<td>rs2256787</td>
<td>1:17,765,403</td>
<td>A/C</td>
<td>0.07</td>
<td>Endometrioid</td>
<td>1.33 (1.18–1.50)</td>
<td>4.5 x 10^{-6}</td>
<td>No</td>
<td>No</td>
<td>ESC, ESDR, IPSC, FAT, STRM, BRST, BRN, SKIN, VAS, LIV, GI, HRT, MUS, LNG, OVRY, PANC</td>
<td>None</td>
<td>Yes</td>
</tr>
<tr>
<td></td>
<td>rs10788679</td>
<td>1:17,789,549</td>
<td>A/G</td>
<td>0.42</td>
<td>Serous</td>
<td>1.07 (1.03–1.11)</td>
<td>2.6 x 10^{-4}</td>
<td>No</td>
<td>No</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>AKAP6</td>
<td>rs1955513</td>
<td>14:32,245,693</td>
<td>C/A</td>
<td>0.07</td>
<td>All invasive</td>
<td>0.90 (0.85–0.95)</td>
<td>3.3 x 10^{-3}</td>
<td>Yes</td>
<td>No</td>
<td>FAT, SKIN, VAS, BRN, MUS, GI, BLD</td>
<td>SKIN, MUS, MUS, THYM, BLD</td>
<td>No</td>
</tr>
<tr>
<td></td>
<td>rs927062</td>
<td>14:32,164,800</td>
<td>G/A</td>
<td>0.21</td>
<td>All invasive</td>
<td>0.94 (0.90–0.97)</td>
<td>5.9 x 10^{-4}</td>
<td>No</td>
<td>Yes, ARHGAP5</td>
<td>None</td>
<td>None</td>
<td>GI</td>
</tr>
</tbody>
</table>

SNP, single nucleotide polymorphism; alleles show minor/major; MAF, minor allele frequency; OR, odds ratio; CI, confidence interval; eQTL, expression quantitative locus with p<0.05 in EOC tumors; histone marks and DNAse I hypersensitive sites from HaploReg v 4.1 indicating tissue types as defined therein; super enhancer information based on the human super-enhancer database available at http://bioinfo.au.tsinghua.edu.cn/dbsuper/index.php; none of these SNPs had previous GWAS associations with any phenotype based on the EBI GWAS catalog or resided within promoter histone marks; all SNPs are intronic to the gene indicated.

https://doi.org/10.1371/journal.pone.0197561.t001
<table>
<thead>
<tr>
<th>Source</th>
<th>Case/Control</th>
<th>MAF</th>
<th>OR (95% CI)</th>
<th>PVal</th>
</tr>
</thead>
<tbody>
<tr>
<td>AUS</td>
<td>109/951</td>
<td>0.07</td>
<td>1.37 (0.81, 2.32)</td>
<td>0.2454</td>
</tr>
<tr>
<td>BAV</td>
<td>13/142</td>
<td>0.08</td>
<td>1.78 (0.47, 6.66)</td>
<td>0.3933</td>
</tr>
<tr>
<td>BEL</td>
<td>21/1256</td>
<td>0.07</td>
<td>2.38 (0.94, 6.05)</td>
<td>0.0677</td>
</tr>
<tr>
<td>DAN</td>
<td>68/810</td>
<td>0.06</td>
<td>1.50 (0.83, 2.71)</td>
<td>0.18</td>
</tr>
<tr>
<td>DOV</td>
<td>150/1346</td>
<td>0.07</td>
<td>0.97 (0.60, 1.57)</td>
<td>0.916</td>
</tr>
<tr>
<td>GER</td>
<td>37/411</td>
<td>0.07</td>
<td>0.73 (0.25, 2.13)</td>
<td>0.5656</td>
</tr>
<tr>
<td>HAW</td>
<td>12/156</td>
<td>0.04</td>
<td>2.92 (0.51, 16.58)</td>
<td>0.2277</td>
</tr>
<tr>
<td>HJO</td>
<td>24/269</td>
<td>0.06</td>
<td>2.24 (0.74, 6.75)</td>
<td>0.1514</td>
</tr>
<tr>
<td>HMO</td>
<td>12/131</td>
<td>0.06</td>
<td>0.73 (0.09, 6.07)</td>
<td>0.7681</td>
</tr>
<tr>
<td>HPE</td>
<td>102/1465</td>
<td>0.07</td>
<td>1.46 (0.90, 2.39)</td>
<td>0.1289</td>
</tr>
<tr>
<td>LA2</td>
<td>87/984</td>
<td>0.07</td>
<td>1.94 (1.21, 3.11)</td>
<td>0.006</td>
</tr>
<tr>
<td>MAY</td>
<td>96/743</td>
<td>0.06</td>
<td>2.10 (1.25, 3.52)</td>
<td>0.0047</td>
</tr>
<tr>
<td>MCC</td>
<td>6/58</td>
<td>0.07</td>
<td>7.54 (0.64, 88.82)</td>
<td>0.1085</td>
</tr>
<tr>
<td>MDA</td>
<td>28/383</td>
<td>0.08</td>
<td>0.51 (0.12, 2.15)</td>
<td>0.3566</td>
</tr>
<tr>
<td>MSK</td>
<td>20/555</td>
<td>0.08</td>
<td>1.59 (0.61, 4.17)</td>
<td>0.3443</td>
</tr>
<tr>
<td>NCO</td>
<td>108/781</td>
<td>0.07</td>
<td>1.20 (0.68, 2.09)</td>
<td>0.531</td>
</tr>
<tr>
<td>NEC</td>
<td>126/997</td>
<td>0.06</td>
<td>1.57 (0.98, 2.54)</td>
<td>0.0626</td>
</tr>
<tr>
<td>NHS</td>
<td>13/383</td>
<td>0.07</td>
<td>0.50 (0.06, 3.95)</td>
<td>0.5098</td>
</tr>
<tr>
<td>NJO</td>
<td>27/179</td>
<td>0.07</td>
<td>1.16 (0.36, 3.72)</td>
<td>0.8016</td>
</tr>
<tr>
<td>NOR</td>
<td>27/370</td>
<td>0.06</td>
<td>2.11 (0.80, 5.58)</td>
<td>0.1316</td>
</tr>
<tr>
<td>NTH</td>
<td>64/323</td>
<td>0.06</td>
<td>1.18 (0.55, 2.57)</td>
<td>0.6693</td>
</tr>
<tr>
<td>OVA</td>
<td>101/741</td>
<td>0.07</td>
<td>1.26 (0.75, 2.10)</td>
<td>0.3864</td>
</tr>
<tr>
<td>POC</td>
<td>39/416</td>
<td>0.07</td>
<td>0.76 (0.27, 2.17)</td>
<td>0.6115</td>
</tr>
<tr>
<td>POL</td>
<td>33/211</td>
<td>0.06</td>
<td>1.62 (0.62, 4.21)</td>
<td>0.3247</td>
</tr>
<tr>
<td>SEA</td>
<td>215/5839</td>
<td>0.06</td>
<td>1.05 (0.70, 1.56)</td>
<td>0.8157</td>
</tr>
<tr>
<td>STA</td>
<td>30/334</td>
<td>0.06</td>
<td>1.01 (0.34, 3.01)</td>
<td>0.9793</td>
</tr>
<tr>
<td>TOR</td>
<td>132/440</td>
<td>0.06</td>
<td>1.52 (0.90, 2.56)</td>
<td>0.1152</td>
</tr>
<tr>
<td>UCI</td>
<td>48/366</td>
<td>0.06</td>
<td>1.50 (0.65, 3.46)</td>
<td>0.3385</td>
</tr>
<tr>
<td>UK2</td>
<td>188/1009</td>
<td>0.07</td>
<td>1.32 (0.89, 1.95)</td>
<td>0.173</td>
</tr>
<tr>
<td>WOC</td>
<td>20/204</td>
<td>0.06</td>
<td>1.65 (0.47, 5.79)</td>
<td>0.4365</td>
</tr>
<tr>
<td>Combined</td>
<td>1984/22700</td>
<td>0.07</td>
<td>1.33 (1.18, 1.50)</td>
<td>4.5x10^-6</td>
</tr>
</tbody>
</table>

Odds Ratio
Fig 1. Association of rs2256787 in the ARHGEF10L gene with invasive endometrioid EOC risk by study site and combined. Squares represent the estimated per-allele odds ratio (OR) and are proportional to sample size for each study; lines indicate its 95% confidence interval (CI); source indicates contributing study. MAF, control minor allele frequency; PVal, per-allele p-value adjusted for age, site, and principal components to account for residual differences in European ancestry.

https://doi.org/10.1371/journal.pone.0197561.g001

transcriptional coactivators in this tissue. As ARHGEF10L rs2256787 associated with endometrioid EOC risk, we were particularly interested in eQTLs in the 54 endometrioid patients; however, there was no evidence of association between rs2256787 genotype and ARHGEF10L expression in endometrioid EOC tumors or other tumor subtypes. In 312 invasive EOC tumors, the G allele of AKAP6 rs927062 correlated with reduced expression of Rho GTPase activating protein 5 (ARHGAP5), a GTPase ~150kb upstream of AKAP6 ($\beta = -0.22$, 95% CI: -0.41 to -0.03, $p = 6.6 \times 10^{-3}$). Other unstudied variants may also be associated with expression of ARHGAP5 (or may be more strongly associated than rs927062), thus future genome-wide or pathway-based analysis of GTPase SNP-expression relationships are of great interest. In other histology-specific eQTL analyses, none of the four variants tested were associated with EOC tumor mRNA expression.

**Conclusion**

We investigated 322 SNPs in 88 genes encoding small GTP binding proteins of the Ras superfamily (Ras, Rho, Rab, Rat, Arf, and Ran) in germline DNA of over 17,000 EOC patients and 26,000 controls. The 88 genes were derived from G protein (guanine nucleotide-binding proteins) signaling, Ras-GTPases, regulation of Rho GTPase protein signal transduction and activation of Rac GTPase activity. Ras-GTPases are activated at the plasma membrane by guanine nucleotide exchange factors (GEF) such as: son of sevenless homologs 1 and 2 (Drosophila) (SOS-1 and SOS-2); Ras protein-specific guanine nucleotide-releasing factor 1 (GRF1); Rap guanine nucleotide exchange factor 1 (GRF2); and RasGEF domain family, members 1A, 1B and 1C (RasGRF). They are inactivated by GTPase activating proteins (GAP) which include RAS p21 protein activator (GTPase activating protein) 1 (p120RasGAP). GEF factors are recruited to the plasma membrane by scaffold and adaptor complexes such as SHC/Grb2 that associate with activated tyrosine kinase receptors (TKR). These factors exchange GTP for GDP on the Ras protein. The resulting GTP-Ras protein activates various downstream effectors such as MAP-kinase Raf-1 which activates the MEK/ERK gene regulation cascade, a primary cell growth and anti-apoptosis pathway. Ras-GTPases family members regulate the action of other GTPase pathways involving Rap, Raf, Rac and Rho Ras-GTPase. Ras-GTPases also regulate phosphoinositide 3-kinase (PI3K) and phospholipase C (PLC) activities. Several of these genes are mutated in ovarian tumors.

Overall, analysis at only one SNP yielded a p-value < $10^{-5}$: rs2256787 in ARHGEF10L which was associated with 33% increased endometrioid EOC risk. Of note, the experiment-wide error rate for this SNP, accounting for the initial overall set of 6103 candidate SNPs equals 0.027 (Bonferroni-corrected p-value 4.5 x 10^{-6} x 6103); additionally accounting for six case groups analyzed, this value increases to 0.16 (Bonferroni-corrected p-value 4.5 x 10^{-6} x 6103 x 6). However, as SNPs, as well as case groups, are not independent, simulation studies are necessary to derive an empirical p-value. Another ARHGEF10L SNP, rs10788679, in showed the smallest p-value in analysis of serous EOC and was the second-most strongly associated SNP in all analyses. ARHGEF10L is a member of the RhoGEF family GEFs that activate Rho GTPases. The Rho branch of the Ras super family encompasses 20 genes in humans, of which Rho, Rac and Cdc42 are the best characterized. Rho GTPases regulate the actin cytoskeleton and control changes in cell morphology and cell motility triggered by extracellular stimuli. Rho GTPases are regulated by GDP/GTP exchange factors and GAPs. Members of this
subfamily are activated by specific GEFs and are involved in signal transduction. SNPs in this gene are also associated with obesity[22] and cutaneous basal cell carcinoma.[23]

The SNP most associated with risk of invasive EOC was rs1955513 in the AKAP6 gene. This gene is involved in overall G protein signaling. SNPs in this gene are also associated with neurologic functioning [24] and anorexia. [25] Functionally, rs927062 in AKAP6 was associated with expression of the Rho GTPase activating protein 5, ARHGAP5, also known as p190 RhoGAP, which negatively regulates RHO GTPases. The p190 RhoGAP gene contains a carboxy-terminal domain that functions as a GAP for the Rho family GTPases. In addition to its RhoGAP domain, p190 contains an amino-terminal domain that contains sequence motifs found in all known GTPases.

In conclusion, our study identified potentially functional genetic variants in small GTPase genes that may have roles in EOC susceptibility. To interpret these associations, we suggest consideration of effect sizes and directionality in the context of the sets of histotype-specific analyses conducted; whether a more conservative or liberal statistical significance threshold is applied, the small set of variants highlighted for detailed functional follow-up remain the same. A limitation of this work is that nearby imputed variants were not examined and thus other ungenotyped variants may be driving the reported associations. Nonetheless, four variants in two genes show promising associations that have not been reported previously but point to known pathways that are mutated in ovarian tumors. The results of our investigation suggest that further assessment of this important pathway is warranted in additional collections of densely genotyped EOC patients and controls.

Supporting information

S1 Fig. Association of rs10788679 in the ARHGEF10L gene with invasive serous EOC risk by study site and combined. Squares represent the estimated per-allele odds ratio (OR) and are proportional to sample size for each study; lines indicate its 95% confidence interval (CI); Source indicates contributing study [11]; MAF, control minor allele frequency; PVal, per-allele p-value adjusted for age, site, and residual European principal components.

(TIFF)

S2 Fig. Association of rs1955513 in the AKAP6 gene with invasive EOC risk by study site and combined. Squares represent the estimated per-allele odds ratio (OR) and are proportional to sample size for each study; lines indicate its 95% confidence interval (CI); Source indicates contributing study [11]; MAF, control minor allele frequency; PVal, per-allele p-value adjusted for age, site, and residual European principal components.

(TIFF)

S3 Fig. Association of rs927062 in the AKAP6 gene with invasive EOC risk by study site and combined. Squares represent the estimated per-allele odds ratio (OR) and are proportional to sample size for each study; lines indicate its 95% confidence interval (CI); Source indicates contributing study [11]; MAF, control minor allele frequency; PVal, per-allele p-value adjusted for age, site, and residual European principal components.

(TIFF)

S1 Table. Results from prior published EOC GWAS results on the targeted 339 SNPs in 88 RAS pathway genes. More details are available upon request.

(XLS)

S2 Table. Results from EOC genetic association analysis on 99 SNPs in RAS pathway genes with nominal p-value <0.05 in analysis of all invasive patients, patients with invasive
serous, endometrioid, clear cell, or mucinous subtypes, and patients with borderline
tumors versus controls. More details are available upon request.

(XLSX)

Acknowledgments

We thank all the individuals who took part in this study and all the researchers, clinicians and
technical and administrative staff who have made possible the many studies contributing to this
work. In particular, we thank: D. Bowtell, A. deFazio, D. Gertig, A. Green, P. Parsons, N. Hay-
ward, P. Webb and D. Whiteman (AUS); G. Peuteman, T. Van Brussel and D. Smeets (BEL);
the staff of the genotyping unit, S LaBoissiere and F Robidoux (Genome Quebec); U. Eilber
(GER); L. Gacucova (HMO); P. Schurmann, F. Kramer, W. Zheng, T. W. Park, Simon, K. Beer-
Grondke and D. Schmidt (HJO); S. Windebank, C. Hilker and J. Vollenweider (MAY); the state
cancer registries of AL, AZ, AR, CA, CO, CT, DE, FL, GA, HI, ID, IL, IN, IA, KY, LA, ME, MD,
MA, MI, NE, NH, NJ, NY, NC, ND, OH, OK, OR, PA, RI, SC, TN, TX, VA, WA, and WY.
The authors assume full responsibility for analyses and interpretation of these data (NHS); L.
Paddock, M. King, L. Rodriguez-Rodriguez, A. Samoila, and Y. Bensman (NJO); M. Sherman,
A. Hutchinson, N. Szeszenia—- Dabrowska, B. Peplonska, W. Zatonski, A. Soni, P. Chao and
M. Stagner (POL); C. Luccarini, P. Harrington the SEARCH team and ECRIC (SEA); I. Jacobs,
M. Widschwendter, E. Wozniak, N. Balogun, A. Ryan and J. Ford (UKO); Carole Pye (UKR);
A. Amin Al Olama, K. Michilaidou, K. Kuchenbaker (COGS). The Australian Ovarian Cancer
Study acknowledges the cooperation of the participating institutions in Australia and acknowl-
edges the contribution of the study nurses, research assistants and all clinical and scientific col-
laborators to the study. The complete Australian Ovarian Cancer Study Management Group
can be found at www.aocstudy.org (Georgia.Trench@qimrberghofer.edu.au). We would like to
thank all of the women who participated in these research programs.

Author Contributions

**Data curation:** Stacey J. Winham, Katja K. H. Aben, Hoda Anton-Culver, Natalia Antonen-
kova, Elisa V. Bandera, Yukie T. Bean, Matthias W. Beckmann, Line Bjorge, Natalia Bogda-
nova, Louise A. Brinton, Angela Brooks-Wilson, Fiona Bruinsma, Clareann H. Bunker,
Ralf Butzow, Karen Carty, Jenny Chang-Claude, Linda S. Cook, Daniel W Cramer, Julie M.
Cunningham, Cezary Cybulski, Agnieszka Dansonka-Mieszkowska, Evelyn Despierre, Jen-
nifer A. Doherty, Thilo Dörk, Andreas du Bois, Matthias Dürst, Douglas F. Easton, Diana
M. Eccles, Robert P. Edwards, Arif B. Ekici, Peter A. Fasching, Brooke L. Fridley, Aleksan-
dra Gentry-Maharaj, Graham G. Giles, Rosalind Glasspool, Marc T. Goodman, Jacek Gron-
wald, Philipp Harter, Alexander Hein, Florian Heitz, Michelle A. T. Hildebrandt, Peter
Hillemanns, Claus K. Hogdall, Estrid Hogdall, Satoyo Hosono, Edwin S. Iversen, Anna
Jakubowska, Allan Jensen, Bu-Tian Ji, Audrey Y. Jung, Beth Y. Karlan, Melissa Kellar, Lam-
bertus A. Kienemey, Boon Kiong Lim, Susanne K. Kjaer, Camilla Krakstad, Jolanta Kupry-
janczyk, Diether Lambrechts, Sandrina Lambrechts, Nhu D. Le, Shashi Lele, Jenny Lester,
Douglas A. Levine, Zheng Li, Dong Liang, Jolanta Lissowska, Karen Lu, Jan Lubinski, Lene
Lundvall, Leon F. A. G. Massuger, Keitaro Matsuo, Valerie McGuire, John R. McLaughlin,
Iain McNeish, Usha Menon, Roger L. Milne, Francesmary Modugno, Kirsten B. Moysich,
Roberta B. Ness, Heli Nevanlinna, Kunle Oduseni, Sara H. Olson, Irene Orlow, Sandra Orsu-
lic, James Paul, Tanja Pejovic, Liisa M. Pelttari, Jenny B. Permutt, Malcolm C. Pike, Eliza-
beth M. Poole, Barry Rosen, Mary Anne Rossing, Joseph H. Rothstein, Ingo B.
Runnebaum, Iwona K. Rzepecka, Eva Schernhammer, Ira Schwaab, Xiao-Ou Shu, Yurii B.

**Formal analysis:** Joe Dennis.

**Resources:** Thomas A. Sellers, Simon A. Gayther, Linda E. Kelemen, Georgia Chenevix-Trench, Harvey A. Risch, Paul D. P. Pharoah, Ellen L. Goode, Catherine M. Phelan.

**Writing – original draft:** Madalene Earp, Jonathan P. Tyrer.


**References**


