Dendritic Cells and Programmed Death-1 Blockade: A Joint Venture to Combat Cancer

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INTRODUCTION

Dendritic cells (DCs) are key antigen-presenting cells capable of presenting tumor antigens to T lymphocytes (1) and promoting innate immunity via, e.g., natural killer (NK) cells (2) and γδ T cells (3). To obtain and engineer DCs for therapeutic approaches, they can be generated ex vivo from multiple sources such as monocytes [monocyte-derived DCs (moDCs)] and CD34+ hematopoietic progenitor cells, or they can be enriched from peripheral and cord blood (4–7). Exploiting their
antigen-specific and immunoregulatory qualities, DCs can be furnished with tumor antigens and other targeted molecules via different techniques (7–9). More than two decades after the first implementation of DCs as an immunotherapy to treat cancer (10), it can be ascertained that DC-based vaccination is safe, well tolerated, and capable of inducing antitumoral immune responses. Objective clinical responses, however, are amenable to substantial improvement (11). To date, scientists believe that the full potential of DC-based immunotherapy has not yet been reached (11–13). This is evidenced by the profound and multidimensional exploration of ways to invigorate the immunotherapeutic potential of DCs, both at the level of DC vaccine engineering and combining DC therapy with other synergistic antitumor (immuno)therapies (14–20). Core objectives of this common quest are to improve DC immunopotency to promote cytotoxic and long-lasting antitumor immunity and to overcome the tumor-mediated immunosuppressive environment (9, 20). In relation to this, interfering with immune checkpoint inhibitory pathways has been on the rise. Since its second-place ranking as a potential target for immunotherapy at the Immunotherapy Agent Workshop of the National Cancer Institute in 2007 research on the inhibitory checkpoint programmed death-1 (PD-1)/programmed death ligand (PD-L) pathway has boosted massively. Due to superior antitumor effects of anti-PD-1 and anti-PD-L1-blocking antibodies, these molecules even climbed to the first position as potential targets for immunotherapy at the 29th Annual meeting of the Society for Immunotherapy of Cancer in 2015 (21). Next to exploiting the systemic monoclonal antibody (mAB) strategy, other promising PD-1/PD-L-targeted approaches are under development. As acknowledged for DC-based vaccination, combination strategies of PD-1-targeted inhibitors with other immune (checkpoint) modulators, cell vaccines, or standard-of-care therapies will likely hold the future (22). In this review, we discuss the role of the PD-1/PD-L pathway in DC-mediated antitumor immunity and the progress of emerging strategies combining DC-based therapy with PD-1/PD-L pathway interference.

**PD-1/PD-L IN HEALTH AND DISEASE**

The PD-1/PD-L axis is one of the most studied pathways to gain understanding of immunoregulatory signals delivered by immune checkpoint receptor/ligand interaction the past few years (23, 24). Originally discovered as a mechanism of the organism to protect itself against T cell reactions toward self-antigens, interaction of PD-1 with one of its ligands (PD-L1 or PD-L2) can induce peripheral tolerance by limiting T cell activity, contributing to protection against tissue damage in case of an inflammatory response (25), prevention of autoimmune diabetes (26), or promotion of the fetal–maternal tolerance (27). Infected and malignant cells that evade immune surveillance have been ascribed to employ the inhibitory PD-1/PD-L pathway (24). Indispensable in healthy immune responses (28, 29), overexpression or induction of PD-1 and its ligands PD-L1 and PD-L2 on both immune and target cells, has been associated with immune deficiency, such as exhausted T cells, dysfunctional NK cells, expanded functional regulatory T (Treg) cells, and immune evasion and suppression (30, 31). PD-L expression can also be indispensable for the establishment of T cell immunity in other settings (28, 29). This ambiguity could be explained by findings that PD-L2 also possesses a costimulatory role (32, 33), possibly through interaction with repulsive guidance molecule b (34). Arising from either intrinsic or adaptive immune resistance (35), PD-1 and PD-L1 surface expression or secretion in different malignancies has been mostly related to poor prognosis (36–42), although discordant data have been reported (43, 44), reflecting the need to improve our understanding of the host immune system and disease-specific microenvironment.

Inhibitory PD-1/PD-L signaling not only occurs between immune cells interacting with malignant cells, but is also effective between different immune cell types shaping the tumor immune environment. This provides a strong impetus to target this inhibitory axis to breach immune inhibition and promote durable immunity. In various solid and hematological tumors, blockade of the PD-1/PD-L1 pathway has proven to reverse this immune inhibition by restoring both antitumor function and number of tumor-infiltrating CD8+ effector T cells, resulting in reduced tumor size and increased overall survival (45–49). While PD-1/PD-L-targeted research predominantly focuses on effector T cells, interest in other cell types is growing. A study in metastatic melanoma patients showed that, in addition to CD8+ T cells, tumor-infiltrating B cells and myeloid-derived suppressor cells (MDSCs) were increased by PD-1 therapy (50). With regard to innate immunity, it has been evidenced that also NK cells are negatively regulated by PD-1 signaling during chronic infections (Mycobacterium tuberculosis and HIV-1) (51, 52) and in cancer (multiple myeloma, glioblastoma multiforme, ovarian carcinoma, digestive cancers) (53–59), directly relating to NK cell cytotoxic and regulatory dysfunction, immune suppression, and poor prognosis. As for T cells, blockade of this inhibitory pathway by means of mAbs could restore dampened NK cell functions, at the level of both interferon (IFN)-γ response (52) and cytotoxic capacity (57). In addition, antitumor immunity mediated by invariant NK T (iNKT) cells was also shown to be improved by blockade of the PD-1/PD-L pathway (60, 61). Expression of PD-1 is also demonstrated on γδ T cells (62) and resulted in γδ T cell exhaustion that could be overcome by administration of a blocking anti-PD-L1 antibody (63, 64). A subset of γδ T cells also expresses PD-L1 conferring them with tumor-promoting characteristics by inhibiting qβ T cells (65). Therefore, PD-L1-blocking antibodies could also restore antitumor immunity by inhibiting PD-1/PD-L1 interactions between γδ and qβ T cells. With regard to immunoregulatory cells, PD-1 is also highly expressed on Treg cells (66). As shown by Sauer et al. (67) and Francisco et al. (68), interaction between PD-1 and its ligands blocks the Akt/mTOR pathway leading to an increased FoxP3 expression, resulting in Treg cell differentiation and maintenance. Furthermore, blocking the PD-1 pathway combined with antitumor vaccination showed a significant decrease in the number of intratumoral Treg cells and reduced tumor growth (69). In addition to Treg cells, a role for the PD-1/PD-L pathway has been put forward for other regulatory cells including tumor-associated macrophages (TAMs), MDSCs, and mucosal-associated invariant T (MAIT) cells (61, 70–75). While research into the effect of PD-1/PD-L blockade in these cells is limited, preclinical anti-PD-1 therapy has been shown to
reduce the number of immune suppressive TAMs and MDSCs (73) and was able to increase the IFN-γ production by MAIT cells (71), indicating the valuable effect of PD-1/PD-L blockade on immune cells beyond the immune-activating CD8+ CTLs.

THE ROLE OF PD-1/PD-L IN DC-MEDIATED ANTITUMOR IMMUNITY

As orchestrators of the immune system bridging innate and adaptive immunity, DCs are key players in directing antitumor immunity. Capable of expressing both the PD-1 receptor and its ligands, DCs can virtually interact with any PD-1 and PD-L-positive cell (Figure 1). In this context, the most acknowledged interaction is between DCs and T cells. PD-L surface expression on DCs [myeloid DC (mDC), plasmacytoid DC (pDC), and in vitro generated vaccine DC] is highest upon maturation with pro-inflammatory cytokines, Toll-like receptor (TLR) ligands, or (parts of) bacterial strains, often used to enhance the expression of costimulatory molecules on DCs (76–78). This PD-L surface expression has been demonstrated to suppress CD4+ and CD8+ T cell activity in various disease models, such as tuberculosis (79–81), HIV (82), and cancer (76, 83–88). Comparably, PD-1 expression on tumor-infiltrating mDCs has also been shown to suppress CD8+ T cell activity and decrease T cell infiltration in mouse models for advanced ovarian cancer (89) and hepatocellular carcinoma (90). In addition to suppression of immune activation, DC PD-L expression was also shown to be involved in the promotion of CD4+CD25+FoxP3+ Treg cell expansion and function (68). Tumor growth factor-beta in the tumor microenvironment promotes PD-L1 expression on DCs, further maintaining Treg cell populations (87, 91) and de novo generation of Treg cells (92) in favor of the immunosuppressive tumor microenvironment (84).

The role of PD-1/PD-L signaling in the crosstalk between DCs and NK cells remains largely unexplored. It has been shown that disruption of the PD-1/PD-L pathway is able to restore NK cell functions, mostly, but not exclusively in multiple myeloma (53, 55, 57, 93). Only few studies suggest a role of this pathway in DC-NK cell crosstalk and controversy remains. Ray et al. (57) demonstrated that NK cell function was abrogated by PD-L1 interactions on pDCs and PD-1 on NK cells and that NK cell functions could be restored by anti-PD-L1 treatment. On the other hand, in a preclinical mouse model, the expression of PD-L1 on NK cells was demonstrated to negatively regulate DC activity via interaction with PD-L1 on DCs (94). To gain more conclusive insights in the contribution of PD-1/PD-L interactions in the crosstalk between DCs and NK cells, more research is warranted. Similar to DC-NK cell crosstalk, little is known about the role of PD-1 signaling in DC-y8 T cell crosstalk (3, 95) and how PD-1/PD-L blockade in combination with DC-based immunotherapies can further empower y8 T cells with antitumor capacities. Other innate immune cells that are able to crosstalk with DCs include iNKT cells, MAIT cells, and MDSCs (96–100). Blockade of PD-1/PD-L interactions between DCs and iNKT cells were shown to increase activation and release of T helper 1 cytokines by the latter resulting in the activation of NK cells and amplified antitumor responses (60, 101). Research on PD-1/PD-L interactions between DCs and MAIT cells or MDSCs is lacking.

Ligation of PD-1 to PD-L1/2 can also exert intrinsic effects on DCs by reverse signaling. Kuipers et al. (102) reported decreased expression of maturation markers in PD-L+ DCs and increased interleukin (IL)-10 production upon treatment with soluble PD-1 (sPD-1), suggesting that through reciprocal signaling a suppressive DC phenotype is attained. In another study, upregulation of PD-1 on DCs was found to be a consequence of DC maturation, especially after TLR-mediated DC activation. Blocking PD-1 during DC maturation resulted in enhanced DC survival and increased immunostimulatory properties (103). In both studies, interference with the PD-1/PD-L pathway increased the immunostimulatory properties of the DCs toward T cell activation.

The interplay of PD-1 and PD-L in DC crosstalk with (virtually all) activating and regulatory adaptive and innate immune cells impacts the productivity of antitumor immunity (Figure 1). Other than monitoring PD-L expression on tumor cells, it has been suggested that monitoring PD-L expression on infiltrating myeloid cells is more predictive for response to blockade of PD-1 signaling (104). Building on the successes of DC-based therapy (11) and PD-1-blocking strategies (105), the exploration of its combinatorial therapeutic use is rationalized to empower the clinical response rates and efficacy of these targeted approaches (7, 16).

STRATEGIES TO LEVERAGE DC IMMUNOPOTENCY BY INTERCEDING PD-1/PD-L SIGNALING

It is generally agreed that the therapeutic potential of DC-based immunotherapy could be improved by tackling the immunosuppressive tumor microenvironment that contributes to ineffective or suboptimal responses (106, 107). Employing intrinsic and adaptive immune resistance mechanisms, PD-1 is a top-ranked checkpoint contributor to blunting immune responses. In a comprehensive review on the molecular and immunological hallmarks and prerequisites for next-generation DC vaccines, Garg et al. (20) discourses its combinatorial use with immune checkpoint inhibitors to enforce efficient antitumor activity. Based on the expression pattern of PD-1 and PD-L on immune cells and cellular contacts between DC and a myriad of immune effector and regulatory cells, blocking PD-1/PD-L interactions will likely impede tumor cell-mediated immune suppression, enhance T cell and NK cell activation and effector functions, and inhibit conversion or activation of Treg cells. However, these actions depend also on the way of implementation of PD-1/PD-L blockade with DC vaccination. Here, we elaborate on the currently applicable strategies (Figure 2) and clinical trials (Tables 1 and 2) that particularly interfere with the PD-1/PD-L pathway in the context of DC-based immunotherapies.

Systemic Receptor-Ligand Blockade

The use of mAbs that block immune checkpoints, particularly cytotoxic T lymphocyte antigen-4 (CTLA-4), PD-1, and PD-L1, has made a profound impact in the field of cancer immunotherapy
How the PD-1/PD-L signaling axis plays a role in DC-mediated orchestration of innate and adaptive immunity. DCs are renowned for their pivotal role in regulating the immune response through interaction with a variety of immune cells. DC-moderated PD-1 signaling has been demonstrated to prototypically result in an inhibitory crosstalk with effector cells, evidenced by (1) reduced infiltration and activation capacities, decreased pro-inflammatory, and increased inhibitory cytokine release by CD8+ and CD4+ T cells; (2) impaired killing, regulatory and reciprocal DC activation properties of NK cells; and (3) impaired activation, Th1-cytokine secretion, and downstream NK cell activation by iNKT cells. On the opposite, a costimulatory role for particular interactions promoting CD4+ T cell memory has been described as well. In crosstalk with Tregs, PD-1 engagement was shown to mediate their proliferation, regulatory function, and de novo generation, contributing to an immune suppressive environment. The role of PD-1-signaling in DC crosstalk with other emerging PD-1-sensitive effector (γδ T cells) and regulatory cells (MDSC, TAM) remains to be elucidated. Abbreviations: DC, dendritic cell; IFN-γ, interferon-γ; iNKT, invariant NK T cell; MDSC, myeloid-derived suppressor cell; NK, natural killer cell; PD-1, programmed death-1; PD-L1, programmed cell death ligand 1; PD-L2, programmed cell death ligand 2; sPD-1, soluble PD-L1, soluble PD-L1; TAM, tumor-associated macrophage; Treg, regulatory T cell.
anti-lymphocyte activating gene-3 (LAG-3), anti-CTLA-4, and anti-BTLA were able to further increase the IFN-γ-producing and proliferative capacity of T cells, while ineffective on their own (117, 118). These findings further underscore the strength of the PD-1/PD-L-signaling axis relative to other immune checkpoint pathways.

Over the past 8 years, a select number of phase I/II clinical trials combining DC vaccination with anti-PD-1 or anti-PD-L1 antibodies in a range of malignancies have been initiated and are currently all ongoing (Table 1). With the first clinical results expected in the near future, the challenges of conceptualization of such combination therapy are already subject of discussion (20). The growing portfolio of both next-generation DC vaccines and available PD-1 and PD-L targeting mABs makes the possible treatment regimens infinite. Moreover, knowledge is growing that tumors are differentially sensitive to either DC therapy or antibody-mediated checkpoint blockade, either intrinsically or dependent on the stage of the disease. While DC-mediated therapy is consistently proven safe (7), systemic mAB therapy has to deal with several immune-related adverse effects such as skin and mucosal irritation, diarrhea, hepatotoxicity, and endocrinopathy (110, 119). Today, we are learning how to recognize and manage immune-related adverse events and toxicities and gaining knowledge on which therapeutic combinations could be applied best at what time point (120, 121). As an alternative to human(ized) mABs, different blocking moieties with advanced target specificity and affinity and reduced toxicity profiles are under investigation, including chimeric fusion proteins (AMP-224, extracellular domain of PD-L2, and an Fc portion of IgG) and nanotechnologies [nanoparticles (122) and nanobodies ((123), Theravectys, Ablynx)]. Although research in this area is limited, these alternative blockers have interesting features because of their size, stability, and pharmacodynamical properties (124), which might pave the way for implementation in combination therapy with DCs.

**Soluble PD-(L)1**

Comparable to the systemic antibody approach is the use of sPD-1 receptor, which only contains the extracellular domain of the PD-1 molecule and can ligate to PD-Ls, making them inaccessible for interaction with PD-1 molecules on immune effector cells. Binding of sPD-1 to surface PD-L on DCs was demonstrated to enhance proliferation of lymphocytes in vitro. In addition, after administration of a vector encoding for sPD-1, tumor growth was inhibited or delayed in a murine model of hepatocarcinoma (125). Similar results were found by Song et al. (126) who additionally

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**TABLE 1**

<table>
<thead>
<tr>
<th>Phase I/II Clinical Trials</th>
<th>Anti-PD-1</th>
<th>Anti-PD-L1</th>
<th>Available mABs</th>
<th>Possible Treatment Regimens</th>
<th>Knowledge on Tumor Sensitivity</th>
<th>Knowledge on Adverse Events and Toxicities</th>
<th>Alternative Blockers</th>
<th>Implementation in Combination Therapy with DCs</th>
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</tbody>
</table>
### TABLE 1: Active clinical trials combining DC-based anticancer immunotherapy with PD-1/PD-L-targeted therapy (clinicaltrials.gov, January 14, 2018).

<table>
<thead>
<tr>
<th>Intervention</th>
<th>Type of DC vaccine</th>
<th>Therapy schedule</th>
<th>Comparator(s)</th>
<th>Condition</th>
<th>Phase</th>
<th>N</th>
<th>Trial identifier</th>
<th>Status</th>
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<tbody>
<tr>
<td><strong>PD-1-/PD-L-targeted therapy</strong></td>
<td></td>
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<tr>
<td>Anti-PD-1 Ab (nivolumab)</td>
<td>Autologous DC loaded with CMV pp65 mRNA</td>
<td>Neoadjuvant + adjuvant DC vaccination with anti-PD-1 therapy</td>
<td>Without neoadjuvant DC vaccination</td>
<td>Recurrent brain tumors</td>
<td>I</td>
<td>7</td>
<td>NCT02529072</td>
<td>Active, not recruiting</td>
</tr>
<tr>
<td></td>
<td>Autologous DC loaded with NY-ESO-1 peptide</td>
<td>Therapy cycles of cyclophosphamide, TCR-transduced PBMC, anti-PD-1 therapy, DC vaccination, and rhIL-2</td>
<td>Single group</td>
<td>NY-ESO-1+ solid tumors</td>
<td>I</td>
<td>12</td>
<td>NCT02775292</td>
<td>Recruiting</td>
</tr>
<tr>
<td></td>
<td>Autologous DC loaded with autologous tumor lysate</td>
<td>Therapy cycles of i.d. DC vaccination with anti-PD-1 therapy</td>
<td>DC therapy alone</td>
<td>Recurrent glioblastoma</td>
<td>II</td>
<td>30</td>
<td>NCT03014804</td>
<td>Not yet recruiting</td>
</tr>
<tr>
<td>Anti-PD-1 Ab (pembrolizumab)</td>
<td>Autologous DC loaded with peptide</td>
<td>Anti-PD-1 SoC post-DC therapy</td>
<td>Single group</td>
<td>Advanced melanoma</td>
<td>I</td>
<td>12</td>
<td>NCT03092453</td>
<td>Recruiting</td>
</tr>
<tr>
<td></td>
<td>Autologous DC loaded with autologous tumor antigens</td>
<td>Therapy cycles of anti-PD-1 and cryosurgery plus i.t. DC vaccination</td>
<td>Single group</td>
<td>Non-Hodgkin lymphoma</td>
<td>I/II</td>
<td>44</td>
<td>NCT03035331</td>
<td>Recruiting</td>
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<tr>
<td></td>
<td>Autologous DC</td>
<td>Therapy cycles of i.n. DC vaccination with anti-PD-1 therapy, radiotherapy, GM-CSF and anti-TNF-alpha therapy</td>
<td>Single group</td>
<td>Follicular lymphoma</td>
<td>II</td>
<td>20</td>
<td>NCT02677155</td>
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</tr>
<tr>
<td>DC-CIK</td>
<td></td>
<td></td>
<td>Anti-PD-1 Ab alone</td>
<td>Advanced solid tumors</td>
<td>I/II</td>
<td>100</td>
<td>NCT03190811</td>
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</tr>
<tr>
<td>DC-CIK</td>
<td></td>
<td></td>
<td>Anti-PD-1 Ab alone</td>
<td>NSCLC</td>
<td>I/II</td>
<td>60</td>
<td>NCT03360630</td>
<td>Recruiting</td>
</tr>
<tr>
<td>Anti-PD-1 Ab</td>
<td>DC-CIK</td>
<td>i.v. anti-PD-1 Ab-treated DC vaccination</td>
<td>Single group</td>
<td>Refractory solid tumors</td>
<td>I/II</td>
<td>50</td>
<td>NCT02886897</td>
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</tr>
<tr>
<td>Anti-PD-1 Ab (CT-011)</td>
<td>DC/tumor cell fusion vaccine</td>
<td>Therapy cycles of anti-PD-1 therapy with DC vaccination post-auto-SCT</td>
<td>Anti-PD-1 Ab alone</td>
<td>Multiple myeloma</td>
<td>II</td>
<td>35</td>
<td>NCT01067287</td>
<td>Active, not recruiting</td>
</tr>
<tr>
<td>SoC CPI therapy</td>
<td>Autologous TLPLDC vaccine</td>
<td>DC vaccination (tumor lysate + yeast cell wall particles + DC) following CPI monotherapy (comparison based on response to CPI therapy)</td>
<td>CPI non-responder, progressive disease following initial response to CPI, stable disease after CPI</td>
<td>Metastatic melanoma</td>
<td>I/II</td>
<td>45</td>
<td>NCT02677411</td>
<td>Recruiting</td>
</tr>
<tr>
<td><strong>PD-L1/PD-L2-targeted therapy</strong></td>
<td></td>
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<tr>
<td>Anti-PD-L1 Ab (avelumab)</td>
<td>Autologous DC vaccine</td>
<td>Therapy cycles of DC vaccination with anti-PD-L1 therapy</td>
<td>Single group</td>
<td>Metastatic colorectal cancer</td>
<td>I/II</td>
<td>33</td>
<td>NCT03152565</td>
<td>Not yet recruiting</td>
</tr>
<tr>
<td>Anti-PD-L1 Ab (durvalumab)</td>
<td>DC/AML fusion vaccine</td>
<td>Not specified</td>
<td>DC therapy alone, traditional care</td>
<td>Acute myeloid leukemia</td>
<td>II</td>
<td>105</td>
<td>NCT03059485</td>
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</tr>
<tr>
<td>PD-L siRNA lipofection of the DC vaccine</td>
<td>MiHa-loaded DC</td>
<td>Post-auto-HSCT</td>
<td>Single group</td>
<td>Hematological malignancies</td>
<td>I/II</td>
<td>10</td>
<td>NCT02528682</td>
<td>Recruiting</td>
</tr>
</tbody>
</table>

AML, acute myeloid leukemia; CPI, checkpoint inhibitor therapy; CIK, cytokine-induced killer cells; DC, dendritic cell; HSCT, hematopoietic stem cell transplantation; IL-2, interleukin 2; i.d., intradermal; i.n., intranodal; i.t., intratumoral; i.v., intravenous; MiHa, minor histocompatibility antigens; NSCLC, non-small-cell lung cancer; PBMC, peripheral blood mononuclear cells; PD-1, programmed death-1; PD-L1, programmed death ligand 1; siRNA, small interfering RNA; SoC, standard of care; TCR, T cell receptor; TLPLDC, tumor lysate particle-loaded dendritic cell.
demonstrated increased expression of activation markers on DC in mice treated with sPD-1. Kuipers et al. (102), however, demonstrated a decrease in the expression of maturation markers on DCs treated with sPD-1. These discrepancies might be ascribed to different experimental settings such as the use of other sPD-1 encoding vectors. Applying the sPD-1 approach in human moDCs, Pen et al. (127) transfected mRNA encoding for sPD-1 or sPD-L1 in DC for transient local expression, thereby limiting possible adverse effects seen with systemic PD-1/PD-L blockade. With this approach, they demonstrated an upregulation of CD80 on sPD-1- or sPD-L1-expressing DCs and an increase in both CD4+ and CD8+ T cell effector functions without influencing the induction of Treg cells. Today, clinical trials evaluating this approach have not been registered.

**Chemo-Immunotherapy**

Anticancer chemotherapeutics remain an important systemic treatment modality to arrest or eliminate rapidly growing cancer cells. Besides lowering the tumor burden, evidence is growing that these cytotoxic drugs also rely on several off-target immunological effects, including enhancement of the immunogenicity of malignant cells and, at least for some chemotherapeutics, suppression of inhibitory mechanisms (128, 129). Complementing conventional chemotherapy regimens with DC-targeted immunotherapy is therefore a promising strategy, actively investigated in clinical trials for a range of malignancies (>140 registered trials at Clinicaltrials.gov based on “DC and chemo” search). DC vaccine efficacy can avail from chemotherapy-induced immunologic effects, including enhancement of the immunogenicity of malignant cells and, at least for some chemotherapeutics, suppression of inhibitory mechanisms (128, 129). Complementing conventional chemotherapy regimens with DC-targeted immunotherapy is therefore a promising strategy, actively investigated in clinical trials for a range of malignancies (>140 registered trials at Clinicaltrials.gov based on “DC and chemo” search). DC vaccine efficacy can avail from chemotherapy-induced immunologic effects, including enhancement of the immunogenicity of malignant cells and, at least for some chemotherapeutics, suppression of inhibitory mechanisms (128, 129). Complementing conventional chemotherapy regimens with DC-targeted immunotherapy is therefore a promising strategy, actively investigated in clinical trials for a range of malignancies (>140 registered trials at Clinicaltrials.gov based on “DC and chemo” search).

### Table 2 | Clinical trials combining DC vaccination strategies with PD-1/PD-L1-modulating chemotherapeutics (clinicaltrials.gov, January 14, 2018).

<table>
<thead>
<tr>
<th>DC-based therapy</th>
<th>PD-1-/PD-L1-modulating chemotherapeutics</th>
<th>Indication</th>
</tr>
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<tbody>
<tr>
<td>Autologous DC loaded with TAA-coding RNA(s)</td>
<td>Cisplatin</td>
<td>Melanoma (NCT02285413), malignant pleural mesothelioma (NCT026498629)</td>
</tr>
<tr>
<td>Autologous DC loaded with tumor lysate</td>
<td>Cisplatin</td>
<td>Multiple myeloma (NCT0083538), ovarian cancer (NCT02432378)</td>
</tr>
<tr>
<td>Autologous DC-CIK</td>
<td>Cisplatin</td>
<td>Esophageal cancer (NCT016911625, NCT02644863), NSCLC (NCT02651441)</td>
</tr>
<tr>
<td>Autologous DC loaded with TAA(s) oxaliplatin</td>
<td>Cisplatin</td>
<td>NSCLC (NCT02766348)</td>
</tr>
<tr>
<td>Autologous DC-CIK</td>
<td>Oxaliplatin (as part of FOLFRINOX)</td>
<td>Pancreatic cancer (NCT02548169), colorectal neoplasms (NCT1413295, NCT2503150)</td>
</tr>
<tr>
<td>Autologous DC</td>
<td>Carboplatin</td>
<td>Gastric cancer (NCT2504229, NCT2215837), colorectal cancer (NCT2202928, NCT2415699)</td>
</tr>
<tr>
<td>Autologous DC</td>
<td>Carboplatin</td>
<td>NSCLC (NCT02689719), breast cancer (NCT03387553)</td>
</tr>
</tbody>
</table>

CIK, cytokine-induced killer cell; DC, dendritic cell; DC-CTL, dendritic cytotoxic lymphocyte; NSCLC, non-small-cell lung cancer; TAA, tumor-associated antigen.

**DC-Targeted PD-L RNA Interference (RNAi) Technology**

Taking into account the orchestrating role of DCs, targeted downregulation of PD-L expression on DCs is expected to potentiate DC-mediated T cell and NK cell activation and prevent Treg cell stimulation. RNAi approaches targeting immunosuppressive factors in DCs have been applied to improve immunogenic functions of next-generation DC vaccines (13). This strategy aims at enhancing DC-mediated antigen-targeted T cell responses at the level of the DC/effector cell immunological synapse, irrespective of tumor PD-L expression. Analogous to DCs expressing sPD-1 or sPD-L1 (vide supra), this technique offers attractive safety considerations compared to systemic antibody administration. The targeted nature of this approach shifts the in situ balance between immune stimulatory and inhibitory signals in the DC/effector cell immunological synapse toward immune stimulation, which has been suggested to result in reversal of the PD-1-mediated T cell exhaustion status (135). Various preclinical studies demonstrated feasibility and effectiveness of introducing small interfering RNAs or short hairpin
RNAs interfering with inhibitory immune-related pathways in DCs, such as suppressor of cytokine signaling (136), indoleamine 2,3-dioxygenase (137), and PD-L1/PD-L2 (138–142). Focusing on the PD-1/PD-L pathway, silencing of PD-L1 and/or PD-L2 in DCs has been evaluated with different RNAi introduction techniques, including viral transduction and non-integrating electroporation, lipid nanoparticle transfection, and the cGMP-compliant transfection reagent SAINT-RED (77, 138, 141, 143, 144). Preclinical data demonstrated that PD-L-silenced DCs could (1) increase expansion, promote pro-inflammatory cytokine secretion and degranulation, and augment antitumor function of antigen-specific CD8+ T cells in human in vitro models (138, 140, 142) and (2) induce significant antitumor immunity in vivo in different malignant mouse models (139, 141). Alternatively, in situ PD-L silencing can also be achieved through the use of small molecules. Dorsomorphin, a small molecule inhibitor of the bone morphogenetic protein signaling pathway, was shown to efficiently downregulate PD-L1 and PD-L2 expressions on treated DCs resulting in increased T cell proliferation and enhanced NK cell-mediated killing of target cells (145).

Today, few DC-associated RNAi approaches are currently being tested in early-phase clinical trials, including one trial evaluating PD-L1/2-silenced DC vaccines (NCT02528682). Results of this trial are awaited.

**CLINICAL TRIALS**

Based on the general appreciation that DC vaccination can be improved by blockade of the PD-1/PD-L pathway, as shown by both in vitro experiments and in vivo animal models, most of these combination approaches are embedded in various clinical trials (146). With the exception of sPD-1, autologous DC vaccines are combined with (i) systemic mAbs targeting PD-1 or PD-L1, (ii) platinum-based chemotherapeutics, and (iii) in situ PD-L RNAi to treat patients with both hematological cancers [multiple myeloma, acute myeloid leukemia (AML)] and solid tumors (renal cell carcinoma, mesothelioma, lymphoma, colon cancer, melanoma, ovarian cancer, pancreatic cancer, nasopharyngeal cancer, and glioblastoma). Clinical trials combining DC vaccination with PD-1/PD-L interference, registered by January 2018, are listed in Tables 1 and 2 and discussed in the corresponding paragraphs. The fast-growing number of clinical studies combining DC-based therapy with PD-1/PD-L blockade strategies emphasizes the potential of this combinatorial approach in the future treatment of cancer patients.

**FUTURE PERSPECTIVES**

Multimodality strategies striving to maximize the efficacy of DC-based cancer immunotherapy are emerging (16, 20, 107). Evidenced by a growing body of preclinical and clinical data, engineering next-generation DC vaccines and redirecting the tumor microenvironment are highly promising (7). The significant role of PD-1-signaling in DC-mediated antitumor immunity rationalizes its therapeutic combinatorial use in the rapidly evolving cancer immunotherapy landscape. The PD-1-/PD-L-blocking industry—and the immune checkpoint industry in general—has expanded drastically in the last years. Leading pharmaceutical companies are putting huge efforts in the development of systemic antibody therapies, with an estimated market value of $35 billion (147). The market for DC-based therapies is as big, with approximately 500 clinical trials registered evaluating DC vaccines, reflecting the immense scientific and pharmaceutical impact of such combinatorial therapy. The growing understanding of the immunological effects of some conventional chemotherapeutics, related to DC activation and PD-1 therapy sensitivity and resistance, provides rationale for the development of synergistic adjuvant combinations and carefully designed chemoinmunotherapy schedules that aim beyond the mere elimination of the suppressive tumor (20, 107). In addition to the pioneering CTLA-4 and PD-1 inhibitors, other immune checkpoints have been attributed to hamper DC-mediated immunity, including LAG-3 and TIM-3 (56, 119, 148). The LAG-3 mAb IMP321 was demonstrated to induce DC maturation (149–151) and is now further tested in clinical trials (NCT00351949, NCT00349934). TIM-3, present on, among others, DCs, was shown to induce T helper 1 cell death when interacting with its ligand galectin-9 on T cells (119, 152), whereas dual blockade of TIM-3 and PD-1 or CTLA-4 was able to suppress tumor growth with possibility of cure in a fibrosarcoma mouse model (153). Overall, targeting multiple immune checkpoints simultaneously with DC therapy is likely to result in synergistic efficacy (107).

Designed to potentiate the patient’s own immune system, unsatisfactory DC-based therapy efficacy led to an era of meticulous vaccine and protocol optimization aiming to enhance vaccine immunogenicity (7, 20). With the approval of immune checkpoint inhibitors, the significance of simultaneously targeting the inhibitory immune mechanisms was clinically established. In search of a balanced treatment, combinatorial DC and PD-1 pathway-targeted immunotherapy has some implications. The lack of specificity of systemic immune checkpoint blockade is prone to eliciting indiscriminate immune activation, resulting in significant immune-mediated adverse reactions and immune-related adverse events. In addition to the frequently observed development of therapy resistance, vigilant immunomonitoring to elucidate these mechanisms and advance early detection is warranted (105, 154, 155). Recently, resistance to anti-PD-1 therapy has been related to disturbance of antigen presentation, DC migration, and DC maturation (156), underscoring the importance of combinatorial treatment schedules. More than 20 years of clinical testing affirms that tumor-specific DC therapy is well tolerated and safe, and overstimulation, autoimmunity, or therapy resistance has been described (11, 20). By robustly breaching PD-1-related inhibitory signaling and demasking immune evasion, DC therapy could get that extra push to prevail durable antitumor immunity while compensating for the lack of specificity of immune checkpoint blockade (107).

Taken apart, it can be concluded that DC therapy and PD-1 blocking approaches will prove best in a combinatorial setting subject to the malignancy and the disease status (157). In this perspective, the search for biomarkers predicting response
CONCLUSION

In this review, we highlighted the role of the PD-1 pathway in DC-mediated antitumor immunity. Aiming to improve DC therapy efficacy, different strategies to invigorate DC immunopotency by impeding PD-1-mediated immune regulation were discussed. From the most advanced research on therapeutic blocking antibodies, lessons learned from chemotherapy-induced immune regulation, and data from more recent developments with gene-silencing techniques, it can be concluded that combinatorial DC and PD-1 pathway-targeted therapy approaches could complement or even synergize under defined circumstances. Five years after the comprehensive review on combination therapy with DC vaccines and immune checkpoint blockade by Vasaturo et al. (107), touching upon the first few preclinical studies on PD-1 combination strategies in particular, we witness that preclinical research has expanded drastically and has been translated into a number of clinical trials. We are now awaiting the first clinical results that will substantially direct future anticancer treatment approaches.

AUTHOR CONTRIBUTIONS

MV, JVDB, EM, and EL wrote the paper. ES, VVT, and WH critically revised the manuscript.

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REFERENCES


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