High-risk human papillomavirus detection in self-sampling compared to physician-taken smear in a responder population of the Dutch cervical screening: Results of the VERA study


A R T I C L E   I N F O
Article history:
Received 26 January 2017
Received in revised form 22 May 2017
Accepted 27 May 2017
Available online 1 June 2017

Keywords:
Human papillomavirus
Population-based screening
HPV-self-sampling
Cervical neoplasms
Questionnaire

A B S T R A C T
In 2017 the cervical cancer screening program in The Netherlands will be revised. Cervical smears will primarily be tested for the presence of high-risk human papillomavirus (hrHPV) instead of cytology, and vaginal self-sampling will be offered to non-responders. This includes a potential risk that part of the women who would otherwise opt for a cervical smear will wait for self-sampling. However, self-sampling for hrHPV in a responder population has never been studied yet. The aim of this study was to investigate the applicability and accuracy of self-sampling in detecting hrHPV in a screening responder population. A total of 2049 women, aged 30–60 years, participating in the screening program in The Netherlands were included from April 2013 to May 2015. After they had their cervical smear taken, women self-collected a cervicovaginal sample with a brush-based device, the Evalyn Brush. Both the cervical smear and self-sample specimen were tested with the COBAS 4800 HPV platform. The hrHPV prevalence was 8.0% (95% CI 6.9–9.2) among the physician-taken samples, and 10.0% (95% CI 8.7–11.3) among the self-samples. There was 96.8% (95% CI 96.0–97.5) concordance of hrHPV prevalence between self-samples and physician-taken samples. Women in our study evaluated self-sampling as convenient (97.1%), user-friendly (98.5%), and 62.8% preferred self-sampling over a physician-taken sampling for the next screening round. In conclusion, self-sampling showed high concordance with physician-taken sampling for hrHPV detection in a responder screening population and highly acceptable to women. Implementation of HPV-self-sampling for the responder population as a primary screening tool may be considered.

© 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Cervical cancer is the fourth most common cancer in women worldwide, with an estimated 530,000 new cases in 2012 (Globocan, n.d.), and approximately 270,000 deaths annually. Nationwide, cytology based cervical cancer screening programs have proven to be effective in reducing both the incidence and mortality of cervical cancer (Arbyn et al., 2009). Cervical cancer screening was introduced in The Netherlands in 1988, and revised in 1996 in a nationwide cytology based 5-yearly screening program for women aged 30–60 years. Despite the significant decrease in incidence of cervical carcinoma. However, cytology based screening has resulted in many false negative results with a sensitivity of only 30–87% (Nanda et al., 2000).

HPV DNA testing can be performed on physician-taken material, and depending on the particular HPV test and workflow used, on self-sampled material (Eperon et al., 2013; Stanczuk et al., 2015; Arbyn et al., 2014). Moreover, primary high-risk human papillomavirus (hrHPV) testing has been shown to be more sensitive than cytology, and performs better in a population entering the screening program (Ronco et al., 2010; Ronco et al., 2014; Rijkaart et al., 2012; Naulel et al., 2007; Kitchener et al., 2009). Based on extensive and accumulating scientific evidence, the Ministry of Health, Welfare and Sports in the Netherlands has decided to revise the screening program in 2017. Cervical smears
will primarily be tested for the presence of hrHPV DNA instead of cytology, with reflex cytology in case of a positive hrHPV test (Standpunt minister van VWS screening op baarmoederhalskanker, 2013).

Previous studies have shown that vaginal self-sampling is a well-accepted alternative for a physician-taken cervical smear for non-responders in organized primary hrHPV based screening, and it increases the screening participation rate (Bosgraaf et al., 2014; Gok et al., 2010; Gok et al., 2012a; Gok et al., 2012b; Bosgraaf et al., 2015; Schmeink et al., 2011). Vaginal self-samples and physician-taken samples show similar test accuracy in detecting cervical intraepithelial neoplasia grade 2 or worse (CIN2+), provided that the test and the self-sampling device have been validated both individually and in a combined method (Arbyn et al., 2014; Schmeink et al., 2011; Snijders et al., 2013; Stanczuk et al., 2016). In the revised Dutch screening program vaginal self-sampling will be available for non-responders (Ministry of Health WaS, n.d.).

In the responder screening population the preferred method of screening is still unknown, and it remains to be determined whether self-sampling is comparable to physician sampling in detecting cervical abnormalities. Therefore, the VERA (Validation of the Evalyn brush with the Roche cobas 4800 hrHPV Test) study has been designed. The objectives of the VERA study were to investigate concordance in hrHPV positivity and HPV 16/18 genotypes between self-collected samples, using the Evalyn brush, and physician-taken samples, and the acceptability of self-sampling among a responder population of the Dutch cervical screening program.

2. Materials and methods

2.1. Study population

The Dutch screening program is a nationwide program targeting women aged 30–60 years. Women are invited at 5-year intervals for a cervical smear, generally taken by their physician. In the current study, 2460 women, aged 30–60 years and living in the regions of Nijmegen and ’s-Hertogenbosch in the Netherlands, participated in the VERA study.

2.2. Clinical specimen collection

The participants had their regular cervical smear taken by their physician as part of the nationwide program. A trained physician obtained a liquid-based cytology sample using a Rovers Cervex-Brush (Rovers Medical Devices B.V., Oss, Netherlands). The Cervex-Brush was rinsed in ThinPrep medium (Hologic, Marlborough, MA) in the Nijmegen region and in SurePath medium (Klinipath BV, Duiven, Netherlands) in the ’s-Hertogenbosch region. Cytological examination and classification were performed at the local laboratory according to the CISOE-A (composition, inflammation, squamous epithelium, other and endometrium, endocervical columnar epithelium, and adequacy of the smear) classification, which can easily be translated into the Bethesda 2001 classification (Bulk et al., 2004). The VERA study was not allowed to interfere in the cervical cancer screening program, according to the Dutch law. Therefore, the self sample was always taken after the physician taken sample, and referral was based on the outcome of the cytology assessment, and not of the obtained hrHPV result from the study.

2.3. Self-sampling procedures

After informed consent, and at the time of the appointment with their physician for their scheduled cervical smear, the participants also received a self-sampling kit including a self-sampling device (Evalyn Brush, Rovers Medical Devices B.V., Oss, Netherlands). The Evalyn Brush is about 20-cm long and consists of a transparent case with wings that control the depth of insertion into the vagina. After the device has been inserted up to its wings, pushing the plunger toward the casing will push the brush out into the vagina. The brush needs to be rotated five times; each rotation generates an audible click. After rotation, the brush can be removed from the vagina, and pulled back into the case. A cap is to be clicked onto the case and the brush can be directly sent to the laboratory. The self-sampling kit also includes an explanatory letter, an informed consent form, user instructions (written and drawn), a short questionnaire, and a return envelope with the address of the laboratory. Women self-collected a cervicovaginal sample with the Evalyn Brush either at home or in the physician’s practice, in either case after the physician collected sample was taken. The women were asked to return the dry brush with the self-sampled material, the signed informed consent form, and the questionnaire by regular mail.

2.4. Specimens preparation

Upon arrival of the dry brush devices at the laboratory, the brush tips were suspended in 4.5 ml of Preservcyt medium (Hologic, Marlborough, MA). The vials were vortexed for 3 × 15 s, stored overnight at 4 °C, and again vortexed for 2 × 15 s, before the brushes were removed and discarded.

A total of 3 ml was collected from the physician obtained samples, and stored and transported at room temperature to the department of Medical Microbiology, Radboudumc, Nijmegen, Netherlands, for molecular hrHPV DNA testing.

2.5. Cobas 4800 HPV test

The physician-collected residual liquid-based cytology (LBC) specimens and the self-sampled specimens were vortexed before being placed in the cobas 4800 for hrHPV testing using the clinically validated Roche cobas 4800 HPV Test to detect 12 hrHPV types (hrHPV31, -33, -35, -39, -45, -51, -52, -56, -58, -59, -66, and -68) as a pooled result, and simultaneously provide specific genotyping information for HPV-16 and HPV-18. The cobas 4800 HPV Test features automated sample preparation combined with real-time PCR technology. β-globin from cellular input is used as an internal control to assess specimen adequacy and identify specimens containing factors that inhibit the amplification process.

2.6. Questionnaires

To investigate the acceptability of the Evalyn Brush, all women were asked to fill out a short questionnaire to record their experience, their response to the instructions, and their assessment of the convenience of the Evalyn Brush. Participants were also asked whether they preferred self-sampling or physician sampling for the next screening round. All questionnaires were collected and analyzed centrally. Cardiff Teleform Software (version 10.1, 2010; Cambridge, UK) was used to design the questionnaire and record the data.

2.7. Outcome measures

The primary outcome measure of the VERA-study was concordance in hrHPV positivity between self-collected samples, using the Evalyn brush, and physician-taken samples. Furthermore, we investigated hrHPV genotypes 16 and 18, and the acceptability of self-sampling in a Dutch cervical screening responder population. For comparison of the presence of HPV-16, HPV-18 and 12 other hrHPV genotypes between the two samples, we used the following terminology; identical, concordant or discordant. An identical result is determined if results from all three cobas HPV channels (16, 18, and 12 other hrHPV genotypes) were equal; concordance is determined as at least one identical genotype in both samples; and discordance is determined as no similarities in the genotypes.

Follow-up outcomes of all participants were retrieved from the nationwide network and registry of histology and cytology database.
(PALGA). The latest diagnosis was registered. When histological and cytological diagnoses were both available in the follow-up period until June 2015, only histological results were included in analysis. All cytological and histological findings recorded before June 2015 were included in our analysis.

2.8. Statistical analysis

We aimed to include 135 hrHPV positive cases in order to reliably assess concordance between physician-taken samples and self-samples. The overall study aim was to target a responder population of 3000 women, with an estimated hrHPV positivity of 4.5%. Analysis of the first 1500 samples revealed an hrHPV positivity rate of 8%, leading to an adjustment of the sample size calculation. With 8% hrHPV positivity only 2000 samples were needed in order to detect concordance with the same reliability. In the analysis of the questionnaires the percentages were calculated for the women who answered the question. Concordance was calculated, and a McNemar test for correlated proportions comparing the hrHPV positive portion of self-sampling with the hrHPV positive portion of physician sampling, was performed on the total group as well as per age group.

3. Results

3.1. Patient characteristics

Samples were collected between April 2013 and September 2014. In total 2460 women participated. From 316 women no cervical smear and/or self-sample was received. Twenty-eight women were excluded because of an invalid test result because of too much blood or mucus. Therefore, in total 2049 women with both a self-sample and physician-taken test result were enrolled.

3.2. HPV prevalence

hrHPV prevalence was 8.0% (163/2049; 95% confidence interval [95% CI], 6.9 to 9.2) among the physician-taken samples, and 10.0% (204/2049; 95% CI, 8.7 to 11.3) among the self-samples (Table 1). The hrHPV prevalence in both the physician-taken samples and the self-samples decreased with age from respectively 18.4%, and 20.0% in women aged 29–33 to 4.7%, and 5.9% in women aged 59–63. (Table 2).

Overall concordance between self-samples and physician-taken samples was 96.8% (Table 1). Using the physician samples result as reference, the proportion of true positive self-samples was 92.6% (151/163; 95% CI, 87.6 to 95.8), while the proportion of true negative self-samples was 97.2% (1886/1984; 95% CI, 96.3 to 97.8). The McNemar test for correlated proportions was highly significant (p = 0.000002) indicating that the proportions are not comparable, although this differed between the age groups (Table 2).

3.3. HPV genotyping

The HPV genotypes detected by the cobas 4800 HPV Test in the physician-taken smear were hierarchically based on oncogenicity for cervical cancer. A total of 43 out of 2049 samples (2.1%) showed single or multiple infections with genotype HPV-16; 12 samples (0.6%) showed single or multiple infections with HPV-18 excluding any coinfections with HPV-16. The remaining 108 samples (5.3%) showed single or multiple infections with non-16/18-HPV genotypes.

In 51 self-samples (2.5%), single or multiple infections with genotype HPV-16 were found, and 11 (0.5%) single or multiple infections with HPV-18 (excluding coinfections with HPV-16). In the non-16/18 HPV genotypes 142 (6.9%) samples were detected.

An identical result was found in 96.4% (1976/2049) of the samples. Analysis showed a concordance of 96.8% (1984/2049), and a discordance of 3.2% (65/2049). Forty-two out of 65 discordant samples tested non-16/18 HPV genotype positive in self-sampling, and hrHPV negative in the physician sample. Table 3 shows the results for HPV detection separating HPV 16/18 from the other hrHPV types. The concordance for HPV16 and/or 18 positives is 99.1% (62 (3.0%) HPV 16/18 positives in self-sampling, and 55 (2.7%) in physician sampling), while for hrHPV positives non-HPV16/18 the concordance is 97.0% (164 (8.0%) HPV 16/18 positives in self-sampling, and 128 (6.3%) in physician sampling). The McNemar test for correlated proportions was not significant for HPV16 and/or 18 positives, but highly significant for non-HPV16/18 positive samples, indicating that regarding HPV 16/18 detection, the proportions are equal, and that the differences in proportions are mainly caused by non-HPV 16/18 types.

3.4. Detection rate of CIN2 or worse (CIN2+)

Follow-up outcomes were retrieved from the nationwide network and registry of histology and cytology database (PALGA). All 21 women with HSIL/CIN2+, had hrHPV identified on their physician-obtained sample, whereas 19 (90.5%) also had hrHPV detected on their self-sampled material (Table 4). Two CIN3 results in group D were missed by hrHPV testing on self-sampled material (both women between 30 and 35 years). Both women did not report any problems with performing the self-sample.

3.5. Questionnaires

A total of 2194 out of 2460 questionnaires (89.2%) were returned. The mean age of the women who returned a questionnaire was 43.4 years (range, 29–61 years).

Women had the opportunity to do the self-sample at the physician’s practice (7.1%; 153/2166) or at home (92.9%; 2013/2166). The user friendliness of the self-sampling device was rated good to excellent by 99.2% of the cases women were confident in performing the self-sampling correctly.

Table 1

<table>
<thead>
<tr>
<th></th>
<th>Self-sampling</th>
<th>Total (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>hrHPV positive (%)</td>
<td>hrHPV negative (%)</td>
</tr>
<tr>
<td>Physician sampling</td>
<td>151 (7.4)</td>
<td>12 (0.6)</td>
</tr>
<tr>
<td></td>
<td>53 (2.6)</td>
<td>1833 (89.5)</td>
</tr>
<tr>
<td></td>
<td>204 (10.0)</td>
<td>1845 (90.0)</td>
</tr>
<tr>
<td></td>
<td>2049 (100.0)</td>
<td></td>
</tr>
</tbody>
</table>

Note: This study was conducted in the Netherlands from April 2013 to May 2015. McNemar test for correlated proportions comparing hr-HPV positive portion of self sampling with positive portion of physician sampling, p < 0.000001.
Note: This study was conducted in the Netherlands from April 2013 to May 2015.

Previous large self-sampling study among non-responders that 80.5% in the regular cervical screening program. Bosgraaf et al. showed in a DNA is a well-accepted screening method for women non-participating in the responder screening population with a clinically validated and automated brush-based self-sampling device and physician-taken sample in a re-

other studies investigated hrHPV concordance between a validated self-sampling material and physician sampling material, especially with graphic factors or to a possible selection bias in the population analyzed.

The difference in higher prevalence of hrHPV in self-samples (10%) compared with physician-taken samples (8%) is in line with previous studies showing higher HPV prevalence in self-sampling compared to physician collected sample (Salmeron et al., 2003; Holanda et al., 2006; Girianelli et al., 2006). Indeed, the significant result of the McNemar test for correlated proportions indicate a true difference in HPV positivity. Additional analysis showed that this difference in proportion is mainly caused by detection of non-HPV 16/18 types in self samples. With self-sampling both the cervical and vaginal area are sampled. This may result in a higher non-HPV 16/18 detection rate and thus self-sampling may not always be a true reflection of the cervical hrHPV flora.

Overall, most women (62.7%) preferred the self-sampling over a physician-taken smear, 24.8% preferred a physician-taken smear for the next screening round and 11.9% had no preference for either self-sampling or physician sampling. There were no notable differences between the age groups (Table 5).

The convenience of self-sampling in comparison with a physician-taken sample was rated good to excellent by 97.1% of the women (2100/2163).

4. Discussion

This study shows a high concordance in hrHPV prevalence between self-sampling material and physician sampling material, especially with regard to HPV 16 and/or 18 positivity. Furthermore, the brush-based Eavalyn device provided a well-accepted self-sampling method for women participating in the cervical screening program. We showed that self-sampling for hrHPV DNA in a Dutch responder screening population is well accepted, and is rated as highly user-friendly. Over 95% of enrolled women found self-sampling convenient and user-friendly. Most women (62.8%) preferred self-sampling over a physician-taken smear for the next screening round. To the best of our knowledge, no other studies investigated hrHPV concordance between a validated brush-based self-sampling device and physician-taken sample in a responder screening population with a clinically validated and automated PCR platform (Meijer et al., 2009).

Current and prior studies have shown that self-sampling for hrHPV DNA is a well-accepted screening method for women non-participating in the regular cervical screening program. Bosgraaf et al. showed in a previous large self-sampling study among non-responders that 80.5%

preferred self-sampling with a brush-based device over a physician-taken smear (Bosgraaf et al., 2015). Szarewski et al. demonstrated in an English screening population that 73% preferred to use the self-sampling at home rather than come to the clinic (Szarewski et al., 2007). The only limitation of self-sampling is the uncertainty about performing the self-sampling correctly.

In previous studies, HPV self-sampling was offered to non-responder women, i.e. women who were never or irregularly screened. In these studies the hrHPV prevalence was 8.3 to 10.3% (Gok et al., 2010; Gok et al., 2012b; Bosgraaf et al., 2015). The hrHPV prevalence of 8.0% in the physician-taken smears in the current study is similar to the prevalence in the non-responder studies in the Netherlands (Gok et al., 2010; Gok et al., 2012b; Bosgraaf et al., 2015; Verhoef et al., 2014a). This is surprising as former studies found generally a prevalence of 5% in a responder population (Rijkarth et al., 2012). This can be due to the technique used (GP5+/6 + -PCR versus cobas hrHPV Test), to demographic factors or to a possible selection bias in the population analyzed.

The difference in higher prevalence of hrHPV in self-samples (10%) compared with physician-taken samples (8%) is in line with previous studies showing higher HPV prevalence in self-sampling compared to physician collected sample (Salmeron et al., 2003; Holanda et al., 2006; Girianelli et al., 2006). Indeed, the significant result of the McNemar test for correlated proportions indicate a true difference in HPV positivity. Additional analysis showed that this difference in proportion is mainly caused by detection of non-HPV 16/18 types in self samples. With self-sampling both the cervical and vaginal area are sampled. This may result in a higher non-HPV 16/18 detection rate and thus self-sampling may not always be a true reflection of the cervical hrHPV flora.

Table 2
Participation rate and prevalence of hrHPV, categorized by age.

<table>
<thead>
<tr>
<th>Age category (in years)</th>
<th>Participants per age group</th>
<th>Physician sampling</th>
<th>Self-sampling</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>hrHPV Positive (%)</td>
<td>hrHPV Negative (%)</td>
<td>hrHPV Positive (%)</td>
</tr>
<tr>
<td>29–33</td>
<td>45 (18.4)</td>
<td>200 (81.6)</td>
<td>49 (20.0)</td>
</tr>
<tr>
<td>34–38</td>
<td>20 (8.3)</td>
<td>219 (91.3)</td>
<td>28 (11.7)</td>
</tr>
<tr>
<td>39–43</td>
<td>30 (7.0)</td>
<td>288 (91.1)</td>
<td>30 (9.5)</td>
</tr>
<tr>
<td>44–48</td>
<td>31 (7.7)</td>
<td>370 (92.3)</td>
<td>40 (10.0)</td>
</tr>
<tr>
<td>49–53</td>
<td>24 (5.8)</td>
<td>392 (94.2)</td>
<td>30 (7.2)</td>
</tr>
<tr>
<td>54–58</td>
<td>16 (4.9)</td>
<td>311 (95.1)</td>
<td>22 (6.7)</td>
</tr>
<tr>
<td>59–63</td>
<td>4 (1.7)</td>
<td>81 (95.3)</td>
<td>5 (5.9)</td>
</tr>
<tr>
<td>Unknown</td>
<td>19</td>
<td>–</td>
<td>19 (100.0)</td>
</tr>
<tr>
<td>Total</td>
<td>2049</td>
<td>1886 (92.0)</td>
<td>1845 (90.0)</td>
</tr>
</tbody>
</table>

Note: This study was conducted in the Netherlands from April 2013 to May 2015.

Table 3
Concordance for HPV 16/18, and non-16/18 hrHPV.

<table>
<thead>
<tr>
<th>Physician sampling</th>
<th>HPV 16/18 positive</th>
<th>HPV 16/18 negative</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>50</td>
<td>5</td>
</tr>
<tr>
<td>Total (%)</td>
<td>12</td>
<td>1982</td>
</tr>
<tr>
<td></td>
<td>62 (3.0)</td>
<td>1987 (97.0)</td>
</tr>
<tr>
<td></td>
<td>Non-16/18 hrHPV positive</td>
<td>Non-16/18 hrHPV negative</td>
</tr>
<tr>
<td></td>
<td>117</td>
<td>11</td>
</tr>
<tr>
<td></td>
<td>47</td>
<td>1874</td>
</tr>
<tr>
<td>Total (%)</td>
<td>164 (8.0)</td>
<td>1885 (92.0)</td>
</tr>
</tbody>
</table>

Concordance for HPV 16/18 positive is 2032/2049 = 99.1%.
Concordance for non-16/18 hrHPV positive is 1991/2049 = 97.0%.
McNemar test for correlated proportions HPV 16/18 positive samples p = 0.14.
McNemar test for correlated proportions non-16/18 hrHPV positive samples p < 0.000002.
High hrHPV prevalence rates may lead to a high referral rates. For this reason a triage test is often suggested to reduce referral rates and prevent overtreatment. It has been shown that cytology triage is not possible on self-sampled material (Arbyn et al., 2012). hrHPV genotyping during cervical cancer screening might improve cervical cancer screening to become more effective, because it might identify women with a higher risk of cervical cancer (Kjaer et al., 2010). HPV-16 and HPV-18 are the most oncogenic hrHPV genotypes (Kjaer et al., 2010). Using immediate genotyping with the COBAS platform to make a decision to publish the article. This study was conducted in the Netherlands from April 2013 to May 2015.

### 4.1. Strengths and limitations

Strengths of this study include the fact that hrHPV testing was performed both on self-sampled and physician-sampled material of the same women, with a combination of a clinically validated self-sampling device as well as a clinically validated HPV assay. This study also measured the acceptability of the self-sampling device, and assessed attitudes toward self-sampling among responders in a Dutch real-world cervical cancer screening setting.

A limitation of our study is that the self-sample was always the second sample to be collected. This may have affected the concordance given that the initial physician collected sample could have depleted the amount of available hrHPV. With this sample consecution there was an exclusion of 121 out of 2170 samples, because these samples contained too much blood for reliable testing. In other studies the invalid rate of a brush-based self-sampling device is <0.5%. Additionally, the follow-up was not based on the hrHPV result. Therefore, the follow-up outcomes retrieved from PALGA were unable to show how many physician-taken samples missed CIN2 +.

### 5. Conclusion

Self-sampling with the Evalyn Brush compared with the cobas 4800 platform showed a high concordance with physician-taken sampling for hrHPV detection in women participating in the regular cervical screening program, especially regarding HPV 16/18 detection. With self-sampling more non-hrHPV 16/18 infections are detected. Self-sampling is highly acceptable to women, and a well-accepted alternative to physician-taken samples in a responders screening population. On the basis of the results of this study, implementation of HPV-self-sampling in a responder population as a primary screening tool may be considered.

### Conflict of interests

All the authors declare that they do not have any conflict of interest.

### Details of ethics approval

The Committee for Human Research approved the VERA study. Registration number 2011/331 NL nr.: 37385.091.11.

### Funding

Roche Molecular Diagnostics provided the HPV kits. Rovers Medical Devices provided the Evalyn Brushes. The funding sources had no role in data collection, data analysis, data interpretation, writing of the report or in decision to publish the article.
Acknowledgments

We thank the participating physicians, their assistants, and the women who have participated in this study.

References


