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First case of *Mycobacterium heckeshornense* cavitary lung disease in the Latin America and Caribbean region

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Abstract

A case of cavitary pulmonary disease caused by *Mycobacterium heckeshornense* in Uruguay is described. This is the first case reported in the Latin America and Caribbean region, showing that this species is a worldwide opportunistic human pathogen.

Keywords: Epidemiology, infection, mycobacterium, *Mycobacterium heckeshornense*, pulmonary

Original Submission: 30 October 2015; Accepted: 9 December 2015

Article published online: 18 December 2015

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In July 2013 a 53-year-old white homeless man with a history of alcoholism was admitted to the hospital with long-standing cough, fever and fatigue. Physical examination revealed malnutrition with severe weight loss, dyspnea and decreased lung sounds primarily in the upper lobes. Chest radiograph and computed tomographic imaging (Fig. 1) revealed bilateral upper lobe infiltrates with cavitation. Haematological analysis revealed a hemoglobin level of 9.8 g/dL and a leukocyte count of 6.9 × 10⁹/L. HIV serology was negative. With a presumptive diagnosis of tuberculosis, two serial sputum samples and a bronchoalveolar lavage sample were sent to the national tuberculosis reference laboratory. All direct smears (stained with Auramine O and Ziehl-Neelsen) were positive for acid-fast bacilli. Antituberculosis treatment was started, with isoniazid, rifampicin, pyrazinamide and ethambutol (HRZE) provided according to the guidelines of the National Program of Tuberculosis of Uruguay. The bronchoalveolar lavage sample was decontaminated with the N-acetyl-L-cysteine–sodium hydroxide (NALC-NaOH) method and inoculated into mycobacterium growth indicator tubes (MGIT; BD Biosciences, Sparks, MD, USA) automated liquid culture system. After 26 days of incubation, the MGIT culture flagged positive; Ziehl-Neelsen stain of liquid culture was positive for acid-fast bacilli, and immunochromatographic test for the identification of *Mycobacterium tuberculosis* complex (TBC ID; Becton Dickinson, Franklin Lakes, NJ, USA) was negative. Cells were pelletted and DNA extracted. The GenoType Mycobacterium CM (common mycobacteria) and AS (additional species) assays (Hain Lifesciences, Nehren, Germany) for identification of nontuberculous mycobacteria were performed. The results were positive for *M. heckeshornense*.

Subcultures on Ogawa medium, as well as the solid cultures from the two sputum samples, yielded small, hemispheric, smooth and yellow-pigmented colonies after 4 weeks of incubation.

Phenotypic characterization was performed for all isolates (Table 1), and DNA was submitted to the Institut Pasteur at Montevideo for sequencing studies. To identify the isolates to the species level, the complete 16S rDNA gene and a fragment of *hsp65* and *rpoB* genes were sequenced (GenBank accession numbers KP636957, KP636958 and KP636959, respectively). The DNA sequences obtained were compared with the GenBank/European Molecular Biology Laboratory sequence database. Gene sequencing unambiguously confirmed the identification of *M. heckeshornense* (100%
similarity with *M. heckeshornense* type strain sequences for all targets. With these clinical, radiologic and microbiologic findings, the patient met the American Thoracic Society/Infectious Diseases Society of America diagnostic criteria of nontuberculous mycobacterial lung disease [1]. Treatment was thus changed to isoniazid, rifampicin, ethambutol, levofloxacin and clarithromycin, all provided daily. Ethambutol had to be stopped owing to ocular toxicity. The patient improved clinically, and a regression of the lesions was observed radiologically. After 12 months of treatment with the macrolide-containing regimen, the patient had converted to negative cultures, and treatment was stopped. After 12 months of follow-up, his disease is clinically and radiologically stable and culture negative.

*M. heckeshornense* was first reported in 2000 as a pathogenic, slowly growing scotochromogenic mycobacterium, phylogenetically related to *Mycobacterium xenopi*, that caused a bilateral cavitary lung disease in an immunocompetent patient [2]. Since then, a limited number of cases were reported in humans, some of them involving severe pulmonary infections [3–5] often

TABLE I. Phenotypic characteristics of isolates from Uruguay and isolates originally reported as *Mycobacterium heckeshornense* sp.

<table>
<thead>
<tr>
<th>Feature</th>
<th>Mycobacterium xenopi</th>
<th>Isolates from Uruguay (n = 3)</th>
<th>Isolates of <em>M. heckeshornense</em> sp. nov.*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Smooth Scotochromogen Slow</td>
<td>Smooth Scotochromogen Slow</td>
</tr>
<tr>
<td>Morphology</td>
<td></td>
<td>Smooth Scotochromogen Slow</td>
<td>Smooth Scotochromogen Slow</td>
</tr>
<tr>
<td>Pigment</td>
<td>Smooth</td>
<td>Smooth Scotochromogen Slow</td>
<td>Smooth Scotochromogen Slow</td>
</tr>
<tr>
<td>Growth rate</td>
<td>Smooth</td>
<td>Smooth Scotochromogen Slow</td>
<td>Smooth Scotochromogen Slow</td>
</tr>
<tr>
<td>Growth at 37°C</td>
<td>Smooth</td>
<td>Smooth Scotochromogen Slow</td>
<td>Smooth Scotochromogen Slow</td>
</tr>
<tr>
<td>Growth at 42°C</td>
<td>Short</td>
<td>Short</td>
<td>Short</td>
</tr>
<tr>
<td>Niacin production</td>
<td>−</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td>Nitrate reduction</td>
<td>−</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td>Catalase (heat stable)</td>
<td>−</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td>Arylsulfatase 3 days/14 days</td>
<td>+ (weak)/+</td>
<td>+ (weak)/+</td>
<td>+ (weak)/+</td>
</tr>
<tr>
<td>Pyrazinamide</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Tween 80 hydrolysis</td>
<td>−</td>
<td>−</td>
<td>−</td>
</tr>
</tbody>
</table>

*Identification number of isolates obtained by Roth et al. [5].
identified as *M. xenopi* and also involving extrapulmonary infections [6–12]. Furthermore, it was isolated from animal sources such as porcine lymph nodes [13] and from a domestic cat with a severe generalized infection [14]. This could indicate that this species is widely distributed in the environment. The new commercially available molecular biology methods can fast and precisely identify most common *Mycobacterium* species. In our case, we identified correctly *M. heckeshornense* using the Genotype CM/AS HAIN tests in 2 days.

The patient was treated with standard therapy (HRZE) for 2 months, until the diagnosis of *M. heckeshornense* was confirmed. In the absence of guidelines for the treatment of *M. heckeshornense* disease, the patient was treated with isoniazid, rifampicin, ethambutol, levofloxacin and clarithromycin. This led to prolonged culture conversion and no relapse 12 months after treatment cessation. In some patients [5,7], resolution was reached with surgical procedures. In our case, the extent of the pulmonary lesions and the general condition of the patient prevented this approach. Although *M. heckeshornense* is genetically related to *M. xenopi*, its pathogenicity is noticeably higher [4].

In summary, we describe a case of *M. heckeshornense* cavitary pulmonary disease, the first of its kind described in the Latin America and Caribbean region. A treatment regimen of isoniazid, rifampicin, clarithromycin and levofloxacin led to symptomatic improvement. *M. heckeshornense* is a serious nontuberculous mycobacterial pathogen with a worldwide spread.

**Acknowledgements**

Partially funded by FOCEM (MERCOSUR Structural Convergence Fund), grant COF 03/11. GG, CR and CC are researchers from the Sistema Nacional de Investigadores (ANII), Uruguay.

**Conflict of Interest**

None declared.

**References**


