Temporal Cortex Activation to Audiovisual Speech in Normal-Hearing and Cochlear Implant Users Measured with Functional Near-Infrared Spectroscopy

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Background: Speech understanding may rely not only on auditory, but also on visual information. Non-invasive functional neuroimaging techniques can expose the neural processes underlying the integration of multisensory processes required for speech understanding in humans. Nevertheless, noise (from functional MRI, fMRI) limits the usefulness in auditory experiments, and electromagnetic artifacts caused by electronic implants worn by subjects can severely distort the scans (EEG, fMRI). Therefore, we assessed audio-visual activation of temporal cortex with a silent, optical neuroimaging technique: functional near-infrared spectroscopy (fNIRS).

Methods: We studied temporal cortical activation as represented by concentration changes of oxy- and deoxy-hemoglobin in four, easy-to-apply fNIRS optical channels of 33 normal-hearing adult subjects and five post-lingually deaf cochlear implant (CI) users in response to supra-threshold unisensory auditory and visual, as well as to congruent auditory-visual speech stimuli.

Results: Activation effects were not visible from single fNIRS channels. However, by discounting physiological noise through reference channel subtraction (RCS), auditory, visual and audiovisual (AV) speech stimuli evoked concentration changes for all sensory modalities in both cohorts ($p < 0.001$). Auditory stimulation evoked larger concentration changes than visual stimuli ($p < 0.001$). A saturation effect was observed for the AV condition.

Conclusions: Physiological, systemic noise can be removed from fNIRS signals by RCS. The observed multisensory enhancement of an auditory cortical channel can be plausibly described by a simple addition of the auditory and visual signals with saturation.

Keywords: functional near-infrared spectroscopy (fNIRS), audiovisual, auditory cortex, cochlear implants, speech
INTRODUCTION

Viewing a talking person’s face and mouth may enhance speech understanding in noisy environments (MacLeod and Summerfield, 1990; Helfer, 1997). This effect is due to multisensory integration, in which congruent unisensory signals from multiple modalities are merged to form a coherent and enhanced percept (Stein and Meredith, 1993). The mechanisms underlying multisensory integration have been studied extensively at the single-neuron level in animals (review on seminal work in anesthetized cat; Stein and Meredith, 1993), and in psychophysical eye movement studies in humans (Cornell et al., 2002; Van Barneveld and Van Wanrooij, 2013). How these mechanisms relate to the neural underpinnings of human speech recognition has been studied with neuroimaging and electrophysiological techniques (Calvert et al., 2004; Beauchamp, 2005; Stein, 2012). In individual neurons, the multisensory responses can be much greater than the linear sum of individual unisensory responses. In contrast, for fMRI data, integrating across millions of neurons, super-additivity is typically not found, although multisensory responses are slightly greater than the maximum or mean of the individual unisensory responses (Laurienti et al., 2005).

Here, we attempt to characterize multisensory speech processing by applying an alternative, non-invasive method to record neural activity: functional near-infrared spectroscopy (fNIRS). fNIRS assesses cortical hemodynamic changes in blood oxygenation based on changes in the transmission of near-infrared light through biological tissue and its absorption by oxygenated (HbO2) and deoxygenated (HbR) hemoglobin (Jobsis, 1977; Cope and Delpy, 1988; Huppert et al., 2006, 2009a; Abdelnour and Huppert, 2009). As fNIRS is a non-invasive, minimally-restrictive and quiet optical technique [as opposed to PET (Johnsrude et al., 2002) and fMRI (Hali et al., 2000)], it is ideally suited for auditory studies (Plichta et al., 2011; Pollonini et al., 2014; Santos et al., 2014; Chen et al., 2015b) on human subjects of all ages. Furthermore, this technique does not suffer from the severe limitations imposed by electromagnetic implants (e.g., cochlear implant, CI; Gilley et al., 2006). Therefore, it has been successfully used to study human auditory cortex activation by speech stimuli in normal-hearing adults (Pollonini et al., 2014) and deaf adults and children using a CI (Sevy et al., 2010; Chen et al., 2015a; Dewey and Hartley, 2015).

In this study, we use fNIRS to record supra-threshold auditory, visual and audiovisual (AV) speech-evoked activity from temporal cortex of normal-hearing adults and post-lingually deaf unilateral CI users. We use a limited number of fNIRS channels in order to reduce the time and complexity of applying the optodes. Figure 1 illustrates the rationale and possible outcomes of our experiments. Pure auditory stimulation is expected to produce a typical hemodynamic response profile (blue, Smith, 2004; Malinen et al., 2007) in line with the BOLD response (for review, see Steinbrink et al., 2006; Cui et al., 2011), that reaches its peak at about 6–10 s after a transient stimulus onset. In contrast, pure visual stimulation may produce at best a lower response (red) for a predominantly auditory-responsive area, which could be due to the expectation of a sound being produced by the moving lips (Calvert et al., 1997). Evidence for clear audiovisual integration would be found if the AV response exceeds mere linear summation of the two unmodal responses, i.e., the additive response, or when it falls below the unisensory auditory response (inhibition; Stein et al., 2009). A sub-additive response might be due to either a multisensory or nonlinear saturation effect.

We also tested a limited number of post-lingually deaf unilateral CI users mainly to examine the feasibility of recording multisensory speech processing at the level of temporal cortex with easily-applied, 4-channel fNIRS in the presence of electrical innervation of the auditory nerve by a CI.

MATERIALS AND METHODS

Subjects

Thirty-three adult native Dutch-speaking normal-hearing subjects (age: 18–62 years, median 29, 15 male, 18 female) and five adult Dutch-speaking post-lingually deaf unilateral CI users (age: 55–59 years, median 57, all female, Table 1) were recruited to participate in this study. All normal-hearing subjects (within 20 dB of audiometric zero, range 0.5–8 kHz) and all CI users...
TABLE 1 | Subject demographics of post-lingually deaf cochlear implant (CI) users.

<table>
<thead>
<tr>
<th>CI user</th>
<th>Implant ear</th>
<th>Etiology</th>
<th>Cochlear implant use (years)</th>
<th>Device</th>
</tr>
</thead>
<tbody>
<tr>
<td>P1</td>
<td>Left</td>
<td>Cogan syndrome</td>
<td>12</td>
<td>C2HgFocus²</td>
</tr>
<tr>
<td>P2</td>
<td>Right</td>
<td>Progressive</td>
<td>5</td>
<td>Nucleus2/RCA²</td>
</tr>
<tr>
<td>P3</td>
<td>Left</td>
<td>Progressive</td>
<td>8</td>
<td>CI¹</td>
</tr>
<tr>
<td>P4</td>
<td>Left</td>
<td>Sudden deafness</td>
<td>19</td>
<td>Nucleus 22²</td>
</tr>
<tr>
<td>P5</td>
<td>Left</td>
<td>Progressive</td>
<td>7</td>
<td>Nucleus2/RCS²</td>
</tr>
</tbody>
</table>

¹Advanced Bionics, Stäfa, Switzerland. ²Cochlear Headquarters, Sydney, Australia.

had normal or corrected to normal vision. Experiments were conducted after obtaining written consent from the subject. The experiments were approved by the Ethics Committee of Arnhem-Nijmegen (project number NL24364.091.08, October 18, 2011) and were carried out in accordance with the relevant institutional and national regulations and with the World Medical Association Helsinki Declaration as revised in October 2008¹.

Experimental Setup

Subjects sat comfortably in a reclining chair, to reduce head movements and to minimize low-frequency so-called Mayer waves, that are presumably caused by slow variations in blood pressure (Julien, 2006). The experiment was performed in a darkened experimental room (3.2 × 3.2 × 3.5 m) in which the walls and the ceiling were covered with black acoustic foam that eliminated echoes for sound frequencies ≥500 Hz (Agterberg et al., 2011). Background noise level was less than 30 dB, A-weighted (dBA; Bremen et al., 2010).

fNIRS data were collected with a pulsed continuous-wave NIRS instrument with four optical sources and two photodetectors (Oxymon MKIII Near-Infrared Spectrophotometer, Artinis Medical Systems BV, Elst, Netherlands; for a comprehensive review of the principles and practicalities of continuous-wave fNIRS, see e.g., Scholkmann et al., 2014). Each optical source consisted of two lasers with emission wavelengths of 858 or 861 nm and 765 nm.

The fNIRS probe template (Figures 2A,B) consisted of two optical sources and a single detector, typically on both sides of the head (see below), with source-detector distances of 25 and 35 mm, termed reference or shallow and deep channel, respectively. Sources and detectors were embedded in plastic molds, which were secured in place on the skull by adjustable straps. The temporal cortex was located based on the 10–20 system (Jasper, 1958), which roughly estimates its location at T7 for the left hemisphere and T8 for the right hemisphere (Herwig et al., 2003; Figures 2A,B). As fNIRS measures brain activity over a diffuse area, we did not pinpoint the exact cortical areas per subject: based on Monte Carlo simulations by others (Fukui et al., 2003; Haeussinger et al., 2011; Strangman et al., 2014; Brigadoi and Cooper, 2015), the average photon path from source to photodetector is estimated to be an ellipsoid with a penetration depth of approximately 2–3 cm. Specifically, the current fNIRS probe template is expected to cover a large area of the temporal cortex (c.f. Sevy et al., 2010).

For 21 normal-hearing subjects the optodes were positioned by aligning the mid-point of the long-distance (35 mm) source-detector pairs above the preauricular point at the T7 and T8 location of the International 10/20 system on the left and right hemisphere, respectively (Niedermeyer and Lopes da Silva, 2005). For the other 12 normal-hearing subjects, who were measured prior to the other subjects, only one side was recorded (left hemisphere, T7). For the CI users, only the hemisphere contralateral to the implant was measured with two sources and one detector, to avoid placement problems of the optodes over the implant. The straps were adjusted to guarantee secure coupling between optodes and scalp at acceptable comfort levels of the subject. Secure coupling was verified online by the presence of a detectable photon count and of a clear cardiac oscillatory response in the raw NIRS trace measured before the experiment. The optodes were connected via optical fibers to the NIRS instrument. The company’s software Oxysoft controlled data acquisition, and allowed for online observation of the data. The data were stored at a sampling rate of either 10 (for the early measurements, which included 12 normal-hearing subjects and all 5 CI users) or 250 Hz (for later measurements on 21 normal-hearing subjects). For data analysis, the latter data were downsampled to 10 Hz.

Stimuli

The stimuli were composed of digital video recordings of a female speaker reading aloud children’s stories in Dutch (Figures 2C,D). In the auditory-only condition, the voice was presented without visual input (Figure 2D). In the visual-only condition, the video of the woman reading the story was presented on the screen without the auditory signal (Figure 2C). In the auditory-visual condition, the video was presented with the corresponding auditory input. The recordings were digitally edited into 36 20.5 s segments, each consisting of a single vignette from one of three stories (in Dutch: “De boer, de geit, de wolf en de kool”, “De professor”, and “De prinses”). The three stimulus conditions were presented interleaved in pseudorandom order within a single block. Stimulus generation was controlled by a Dell PC (Dell Inc., Round Rock, TX, USA) running Matlab version 2009b (The Mathworks, Natick, MA, USA) using Psychophysics Toolbox 3 extensions (Brainard, 1997; Pelli, 1997; Kleiner et al., 2007). Sounds were presented through headphones (Sennheiser PCX 350 NoiseGuard, Sennheiser electronic GmbH & CO KG, Wedemark, Lower Saxony, Germany, noise cancellation off) at a comfortable listening volume of 55 dBA, while the video was presented on the Dell PC’s monitor. As the implant interfered with placement of headphones for three out of five CI users, the acoustic stimuli to these CI users was alternatively presented via the direct input to the CI or via a free-field speaker.

Paradigm

The 36 segments were played in chronological order, each followed by a silent, dark period ranging from 25 to 50 s (randomly drawn from a uniform distribution). Even the shortest
intermittent period of 25 s allowed the hemodynamic response to return to baseline, while the randomization limited time locking of any periodic physiological signal to stimulus onsets. The segments were presented in three blocks of 12 stimuli each. A single session consisted of these three blocks with intermittent breaks of about 4–5 min wherein the light was turned on. Every block started with a baseline measurement (in silence and darkness) of 2 min. A session of three blocks (36 segments) took about 45 min to complete.

For every block, the 12 segments were pseudo-randomly assigned to an experimental condition (four segments auditory-only, four segments visual-only, four segments auditory-visual). Subjects were instructed to pay attention to the segments (watching, listening, both), and were asked afterwards whether they understood the gist of the storyline. Other than that, subjects were not given further task instructions.

Analysis

Signal Processing

The optical densities for each channel and wavelength were stored on disk (in the native oxy-format from the Artinis system) for offline analysis in Matlab (Release 2014b, the Mathworks, Inc., Natick, MA, USA). Data was read into Matlab via Artinis’ proprietary function `oxysoft2matlab`. The 250-Hz sample-rate data were downsampled to 10 Hz (using the `resample` function from Matlab’s Signal Processing Toolbox), for computational efficiency.

Physiological noise, such as heart pulsation, respiration, and Mayer waves (Huppert et al., 2009b) is mixed with cortical activity in the fNIRS signal. A clear cardiac oscillation is regarded as evidence for a proper contact between the optical probes and the scalp (Themelis et al., 2004). Therefore, following Pollonini et al. (2014), we determined the scalp coupling index (SCI) as the correlation between the two photodetected signals at 765 and \( \sim 860 \) nm, band-pass filtered between 0.5 and 2.5 Hz (typical frequency range for heart rate that excludes low-frequency fNIRS activity), for every optode. Typically, the SCI was highly positive (median 0.98), as expected from physiological signals that have no origin in the neural source (Yamada et al., 2012), and only 24 out of 354 channels [(21 normal-hearing subjects \( \times \) 2 hemispheres + 12 normal-hearing subjects \( \times \) 1 hemisphere + 5 CI users \( \times \) 1 hemisphere) \( \times \) 2 channels \( \times \) 3 recording blocks] had an SCI less than 0.9. These 24 low-SCI channels were rejected from further analysis as we deemed those indicative for poor
contact between optode and scalp. Then, to remove cardiac, respiratory, and Mayer wave noise sources, we used the `removeheartbeat` function from the NIRS Analysis Package (Fekete et al., 2011); in short, this algorithm extracts an oscillatory template from a narrow-frequency filtered average of all channels per subject (with the filter band containing the oscillatory frequencies of interest), and subtracts this from each channel. Then, we band-pass filtered the signals between 0.008 and 0.1 Hz (Figures 3A,B, red and yellow curves). Next, the data was de-trended using a 20th-degree polynomial in order to remove slow temporal drifts (Figures 3A,B, black and purple curves; Pei et al., 2007). These processed optical densities were converted to changes in oxygenated and deoxygenated hemoglobin concentration (HbO₂ and HbR, respectively) using the modified Lambert-Beer law (Cope and Delpy, 1988; Kocsis et al., 2006). Subsequently, the preprocessed data were normalized by the variance in each recorded signal for the entire session.

Despite these filtering procedures, a considerable amount of noise originating from non-cortical physiological processes still remained (Scholkmann et al., 2014). To deal with this, we applied reference channel subtraction (RCS; Scarpa et al., 2013; Brigadoi and Cooper, 2015). This assumes that the shallow channel (the signal originating from the shorter-distance optode source-detector) is dominated by non-cortical signals, while the deep channel (the signal arising from the longer-distance optode source-detector) also includes more of the cortical event-related signal of interest. Therefore, we determined the fNIRS signal as the residual signal from a simple linear regression between the deep and shallow channels (Figure 4C). Note that we applied the normalization of data in the graphs both before and after RCS, so that the signals are scaled with respect to the data variance, and are hence dimensionless. An individual trace of HbO₂ for a single normal-hearing subject (NH1) for the shallow (black line), deep channel (red line) and the residual signal, during presentation of auditory snippets, is plotted in Figure 4A. An example of how RCS can affect the average evoked response at the single subject-level is shown in Figure 4B. Even though we can expect that the shallow channel might contain some cortical signal because of the relatively large distance of 25 mm, the RCS procedure in general improved the beta coefficients (Figure 4D; see also “GLM” Section) and the signal response (Figure 4E). Because the same (systemic, not event related) noise is present in deep and reference channels, it is successfully removed by RCS. As a result, the variance in the deep channel signal decreases, and hence the signal-to-noise ratio increases. Normalization with a smaller variance leads to an increase of the beta coefficient (Figure 4D), and to the appearance of a clear average activation signal (Figure 4E).

**Statistics**

**Average**

Functional signals were averaged across the 12 repeats of each stimulus modality to calculate the average hemodynamic response for each participant and hemisphere. These traces were averaged across participants and hemispheres (no significant hemispheric differences were observed for the bilaterally-measured 21 normal-hearing subjects according to a Wilcoxon signed-rank test, p > 0.05 for both HbO₂ and HbR) to determine the mean response for both cohorts.
We compared both the measured concentration changes of HbO$_2$ and HbR to a predicted hemodynamic response function (HRF). The HRF consists of a canonical impulse response function $h$ (as used by the SPM toolbox; Henson and Friston, 2007; Lindquist et al., 2009):

$$h(t) = \frac{1}{\Gamma(6)} \cdot 5^5 e^{-5t} - \frac{1}{6\Gamma(16)} \cdot 15^3 e^{-15t}$$  (1)

(with $t$ time and $\Gamma$ the gamma function), which peaks at $\sim 5$ s, convolved with the boxcar function (1 during stimulus presentation, 0 otherwise). After convolution the HRF signal is expected to peak at $\sim 12$ s. All pre-processing steps performed on the data were also applied to the HRF signal.

We employed a general linear model (GLM) to quantify the strength between the measured responses to each stimulus condition and the HRF. This model assumes that auditory ($\beta_a$) and visual ($\beta_v$) inputs independently elicit a hemoglobin concentration change. An extra, third component ($\beta_{av}$) is added in this model, which represents the type and amount of multisensory integration during the presentation of audiovisual stimuli:

$$y(t) = X_A(t)\beta_a + X_V(t)\beta_v + X_{AV}(t)\beta_{av} + \varepsilon(t) + C$$  (2)

with fNIRS data $y(t)$, the explanatory variables $X(t)$, constant regression coefficients $\beta$, offset $C$ and Gaussian noise $\varepsilon(t)$.

For each GLM fit, we determined the goodness of fit ($R^2$-value, and the corresponding $F$ and $p$-values). We took the significance of every regression coefficient as a measure of activation compared to baseline, by determining the corresponding $t$- and one-sided $p$-value (larger than 0 for HbO$_2$ and smaller than 0 for HbR).

Comparisons

To determine whether the beta values differed from a distribution with median 0, the Wilcoxon signed-rank test was applied per cohort. Also, we determined the slope between regression coefficients by determining the optimal fit through simple linear regression. The Wilcoxon Rank Sum test differences in regression coefficients between cohorts were determined. Significance was assessed at the 0.05 alpha level.
RESULTS

Functional NIRS—Representative Single Subject Data
We measured fNIRS activity over the temporal cortex of 33 normal-hearing subjects and 5 CI users while they were watching and/or listening to auditory, visual and audiovisual speech stimuli (Figure 5). Individual traces of HbO2 and HbR signals for a single representative normal-hearing subject (NH17) generally increase and decrease, respectively, during the stimulus epochs (Figures 5A,B). Still, despite the extensive pre-processing (see “Materials and Methods” Section), signal drift, typical for fNIRS measurements (Sevy et al., 2010), occurs also during the silent dark periods. To deal with this stimulus-independent noise, we averaged the signals over the 12 trials per stimulus modality (Figures 5C,D). The normalized, average HbO2 over the 12 auditory-only (A) stimuli increases from baseline at sound onset reaching its maximum after about 15 s (Figure 5C, blue), which is slightly more (∼9%) than the average for the 12 AV stimuli (Figure 5C, green). The visual (V) trial average (Figure 5C, red), while also increasing, only reaches a maximum of ∼27% of the A maximum. These increases are mirrored in the HbR decreases, albeit with a slightly lower amplitude (Figure 5D). After stimulus offset, HbO2 and HbR return gradually to baseline (within 10 s). Typically, and as exemplified for this subject, the hemodynamic response corresponds well to the actual signals (cf. Figures 1, 5B).

Hemodynamic Response Shapes to Auditory, Visual and Audiovisual Stimulation for Normal-Hearing Subjects and CI Users
To reveal the shape of the cortical hemodynamic response, we averaged the trial averages over subjects for the A, V, and AV modalities, for the time interval between 10 s before stimulus onset and 10 s after stimulus offset (Figure 6). All modalities demonstrated similar response shapes, albeit with varying amplitudes. The signals changed after stimulus onset (increase for HbO2 and decrease for HbR) followed by a recovery back to baseline after stimulus offset. The data of the CI users (Figures 6B,D) exhibited similar trends (Figures 6A,C), although the standard errors were slightly larger (also due to the lower number of subjects in the CI user cohort). Moreover, the observed response resembled the predicted response shape (cf. Figure 1), at least qualitatively. These similarities in response shapes for all cohorts, modalities and prediction indicate that fNIRS can consistently measure temporal cortical responses to auditory and visual stimuli in normal-hearing and cochlear-implanted adults.

Cortical Hemodynamic Amplitude Changes Reveal Saturation
To quantify the evoked responses, we fitted a GLM (Eqn. 2; see “Materials and Methods” Section) that assumes that auditory and visual inputs independently elicit a hemoglobin amplitude change, also during audiovisual stimulation. An extra, third component is added in this model, which represents the type and amount of multisensory integration during the presentation of audiovisual stimuli. The analysis yields a set of three beta coefficients (Figure 7) representing the modeled amplitude changes for each response component (auditory, visual and audiovisual interaction), for each subject (both cohorts), for both hemispheres (if applicable), and for both HbO2 and HbR. In line with the grand average response for the normal-hearing cohort (Figures 6A,C), significant activation was observed for the majority of single HbO2 and HbR channels in normal-hearing subjects by auditory and/or visual stimulation (A: 52/54 and 50/54; V: 47/54 and 38/54 regression coefficients were larger/smaller than 0, for HbO2 and HbR, respectively). Most channels did not exhibit an additional AV component (AV: 7/54 and 9/54).

In line with the significance of unsensory individual channel activation, the coefficients for both the auditory and visual components reveal a general positive amplitude change for HbO2 (Wilcoxon signed-rank test; for auditory coefficients: \( p < 0.001, z = 6.8, \text{rank} = 1779 \), and for visual coefficients: \( p < 0.001, z = 6.2, \text{rank} = 1709 \)) on a group-level, although there is a large intra-coefficient variability, with beta values ranging between −0.3 and 1.9. In contrast, comparisons between coefficients show a systematic trend of visual coefficients being smaller than auditory coefficients (Figure 7A, open circles; Wilcoxon signed-rank test: \( p < 0.001, z = 5.8, \text{rank} = 1657 \)). A similar, opposite pattern arises for HbR (Figure 7D). Wilcoxon signed-rank test: \( p < 0.001, z = −6.6, \text{rank} = 11; p < 0.001, z = −5.3, \text{rank} = 182 \), for A and V, respectively; for comparison between A and V: \( p < 0.001, z = −5.1, \text{rank} = 203 \). The auditory data signify that we can reliably obtain auditory responses from temporal cortex with fNIRS, and the slightly weaker visual response data arguably imply that cross-modal, visual activation can arise from the same recording site (see also “Discussion, Multisensory Integration vs. Saturation” Section).

To test for multisensory integration, researchers typically compare the bimodal response to the largest unimodal response (Stein et al., 2009; Figure 1). As the far majority of auditory coefficients are larger than the visual coefficients (HbO2: 49 of 54; HbR: 43 of 54), we chose to compare only the auditory response with the bimodal response for all subjects (Figures 7B,E). Note that the bimodal amplitudes are constituted by the sum of the auditory, visual and audiovisual-interaction coefficients (see Methods). These audiovisual amplitudes are highly similar to the auditory coefficients as all points lie close to the unity line, both for HbO2 and HbR (Figures 7B,E; Wilcoxon signed-rank test: for HbO2 \( p = 0.14, z = −1.5, \text{rank} = 688, \text{slope} = 0.95 \); for HbR \( p = 0.34, z = 0.95, \text{rank} = 1011, \text{slope} = 0.89 \)).

The audiovisual interaction components are almost exactly inversely related to the visual amplitudes (Figures 7C,F; i.e., data points lie close to \( y = −x \) line, regression slopes: \( −0.93 \) and \( −0.87 \) for HbO2 and HbR, respectively). This might be indicative of a saturation effect as the extra
audiovisual interaction effect almost exactly counterbalances any effect a visual component might have (see also “Discussion” Section).

Concentration changes evoked in the five CI users (Figure 7, gray squares) resembled those evoked in the normal-hearing subjects. Specifically, the auditory and visual coefficients for the CI users ranged from 0.02 to 1.4 (Figure 7A) and from −1.5 to 0.7 (Figure 7B) for HbO₂ and HbR, respectively. Significant activation from baseline for auditory components was observed for 4 out of 5 and 5 out of 5 subjects for HbO₂ and HbR, respectively. The visual components were significant for 5 out of 5 and 3 out of 5 subjects, respectively. The audiovisual components were significant for 0 out of 5 and 2 out 5 CI users.

DISCUSSION

Overview
In this study, we assessed audio-visual activation in temporal cortex with fNIRS. Specifically, we studied cortical activation as present in concentration changes of oxy- and deoxy-hemoglobin of normal-hearing adult subjects and post-lingually deaf unilateral CI users in response to auditory, visual and auditory-visual speech stimuli. Sounds
FIGURE 6 | Grand average hemodynamic response of normal-hearing subjects and CI users. Grand average responses for HbO$_2$ of normal-hearing subjects (A) and CI users (B). Grand average responses for HbR of normal-hearing subjects (C) and CI users (D). For the normal-hearing subjects 54 channels (21 bilateral, 12 unilateral) and for the CI users five unilateral channels were recorded. Gray rectangular patch denotes stimulus activation. Colors denote: red—visual; blue—auditory; green—audiovisual stimulation. Shaded areas depict standard error of the mean over subjects.

evoked larger concentration changes than visual stimuli (Figures 7A,D). The audiovisual fNIRS signal resembled the purely auditory response (Figures 7B,E) with the visual component being almost exactly inversely related to the audiovisual component (Figures 7C,F). Interestingly, hemodynamic concentration changes evoked in the CI users strongly resembled those of the normal-hearing subjects (Figure 7).

Feasibility
Since we show robust evoked activity in the temporal cortex for three different sensory conditions in fNIRS data on a group level (Figure 6), fNIRS seems suited to study auditory and visual processing. Furthermore, the responses for the various modalities were consistent when compared against each other within subjects (Figure 7). Nevertheless, a large idiosyncratic variation on single-modality fNIRS responses (Figure 7) may limit the use of this technique on single subject level. The causes for the observed inter-subject variance might be threefold: (1) methodological; (2) analytical; and (3) experimental. First, we will briefly explain and discuss these issues.

A methodological source of inter-subject variability in our data is the placement of the optodes. According to the 10/20 International System, we placed the optodes based on external anatomical landmarks (i.e., nasion and inion; Sevy et al., 2010; Dewey and Hartley, 2015). Alternatively, one could place the optodes based on functional landmarks, by conducting a short functional localizer experiment, such that the location of the maximal response is searched for in a pilot experiment. For example, tone responsiveness could be determined in order to localize basic auditory-responsive regions (Plichta et al., 2011). Another optimization of the current 2-channel optode design would be to use multichannel optode arrays (Pollonini et al., 2014; Chen et al., 2015b; Dewey and Hartley, 2015), so that only the channel(s) with the strongest evoked responses are analyzed (as is current practice, e.g., Chen et al., 2015b), or to determine a clearly localized response (e.g., Kennan et al., 2002; Pollonini et al., 2014; Chen et al., 2015b). In addition, one might consider to determine the individual optode positions in such a
FIGURE 7 | Beta coefficients of the GLM. HbO₂ (A–C) and HbR (D–F) β-coefficients are shown for all subjects for all stimulus modalities. (A,D): visual vs. auditory regression coefficients. (B,E): summed auditory, visual and AV-interaction (representing the AV response amplitude) vs. auditory regression coefficients. (C,F): audiovisual-interaction vs. visual regression coefficients. Open circles indicate normal-hearing subjects—filled squares indicate CI users. The black line depicts the best-fit regression line.

multi-optode array from anatomical MRI scans per subject (Barbour et al., 1995; Pogue and Paulsen, 1998; Barnett et al., 2003).

This study reveals that RCS is a very important factor in the analysis (Figure 4). Typically, this is not performed (Scarpa et al., 2013), although it is considered essential in removing systemic noise (Scholkmann et al., 2014). Without RCS, no effect in any of the sensory modalities would be observed in the current data (not shown here, but see Figures 4D,E). A refinement of the current procedure would be to systematically change the inter-optode distances in order to optimally record purely systemic noise (in the reference channel) and the largest evoked hemodynamic signal (in the deep channel). The use of a multichannel optode array with varying optode distances might be ideally suited to disentangle the systemic noise from the evoked signal.

Finally, the experimental paradigm might in itself explain the variability. In this case, it might turn out that idiosyncratic variability is real, and that the amount of neural or hemodynamic activity varies on an individual basis. Variation might then be reduced if the evoked response is maximized for all subjects by specifically tailored experimental paradigms. For example, in the current paradigm, subjects were passively exposed to the stimuli, while active listening typically results in increased cortical activity in humans (Grady et al., 1997; Vannest et al., 2009; Turner et al., 2013) and non-human primates (Wang et al., 2005; Massoudi et al., 2013, 2014; Osmanski and Wang, 2015). Furthermore, one might refine the stimuli in order to elicit optimal responses from the brain area under consideration. Here, we used speech stimuli, although primary auditory and belt areas might be more responsive to basic acoustic stimuli, such as amplitude-modulated Gaussian white noises, or dynamic spectral-temporal ripples. Yet, higher (belt) auditory cortical regions might respond better to more natural stimuli.

Multisensory Integration vs. Saturation

Our data is in line with cross-sensory influences on neuronal activity, as a clear response was evoked by visual trials over an auditory-responsive, temporal cortical area (Figures 6, 7). This is in line with earlier studies that show a cross-sensory influence on neuronal activity at early cortical areas, which have been traditionally held as unisensory (Ghazanfar et al., 2002; Foxe and Schroeder, 2005; Schroeder and Foxe, 2005; Kayser et al., 2007, 2010; Koelewijn et al., 2010). However, we cannot exclude the possibility that recordings may have partially been taken from higher auditory supplementary areas, as fNIRS records signals arising from a large (1–2 cm) diffuse area (Boas et al., 2004). As such visual-evoked signals might potentially originate from areas in the superior temporal gyrus that encode for face recognition, lip-reading, or other higher-cognitive functions (Sams et al., 1991; Calvert et al., 1997; MacSweeney et al., 2000). The data support the idea that we recorded from predominantly auditory areas, as sounds almost invariably elicited the largest responses, and the visual activation...
was nearly completely nulled during audiovisual stimulation (i.e., audiovisual activation was not significantly different from the inverse of visual-only activation, Figures 7C,F).

On a group level, the AV responses hint at auditory dominance (Figure 7), because the visual response, as presented in isolation, does not appear in the AV response. Two distinct mechanisms might underlie this phenomenon. First, a true multisensory integrative effect could have been present (Stein et al., 2009; Figure 1), in which the visual component is effectively counterbalanced by an inhibitory audiovisual integration effect (Figures 7C,F). Alternatively, the hemoglobin response might have reached saturation by the supra-threshold, highly intelligible auditory stimulus. Then, adding a visual stimulus will not lead to a stronger response. It seems unlikely that the nearly exact inverse relationship between the audiovisual and visual components in the audiovisual regression model (Figures 7C,F) would be explained by multisensory integration, as it is precisely expected from a saturation effect. To better dissociate these different interpretations, auditory and visual stimuli should be presented in regimes that prevent neural saturation, and/or better characterize the visual response.

Note that with a one-channel setup it is impossible to decide whether the auditory and visual activations originated from the same area, or from spatially separated areas, when the AV response would equal the sum of the A and V responses. However, if the AV response deviates from the purely additive prediction several possibilities may be dissociated, as explained in Figure 8. Importantly, activation of two distinct, independent brain areas (Figures 8A,E) does not predict the saturation that is observed in our data. The sub-additive AV response observed in our result (a peak activation between the blue and green lines in Figure 8) allows for two possible scenarios: (i) the signals could have originated from true multisensory neurons (Figures 8C,G), or (ii) from two distinct subpopulations of unisensory-responsive neurons within the recorded area (Figures 8B,F). Note, however, that whenever the peak activation exceeds the additive response (green line), or falls below the strongest unimodal response (blue
line), it will be a signature for true multisensory neural integration.

**Post-Lingually Deaf CI Users**

The brain can reorganize after sensory deprivation, such as caused by deafness (Rauschecker, 1995; Lee et al., 2001). The question is whether cross-modal reorganization after deafening might introduce stronger visual effects over auditory cortex in post-lingual deaf subjects. This is not the case in our limited group of CI users (Figures 7A,D), as visual activation was lower than auditory evoked activity.

Our cohort of post-lingual deaf CI users did not differ from the normal-hearing cohort with respect to cortical activation for audio-visual stimuli (Figure 7, gray squares). This is seemingly in contrast to the principle of inverse effectiveness, which suggests that people with sensory impairments might benefit from multisensory integration. Specifically, a larger multisensory enhancement compared to the purely auditory response would be predicted because of the hearing-impairment of the CI users (and thus the weaker auditory percepts). This is not observed (Figures 7B,E), indicating that either the stimuli were still supra-threshold for these subjects, or that saturation still dominated the audiovisual responses. Both possibilities imply a paradigm that aims at near-threshold stimulation in order to study this principle. Moreover, a larger cohort of CI users is desired when the issues of supra-threshold stimuli and response saturation have been overcome.

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**CONCLUSION**

We found increased activation to auditory, visual and audiovisual stimulation in temporal cortex of normal-hearing subjects and post-lingually deaf CI users using NIRS. Our findings demonstrate the potential of NIRS for studying the neural mechanisms of audiovisual integration, both in normal-hearing subjects and in hearing-impaired subjects following cochlear implantation.

**AUTHOR CONTRIBUTIONS**

Conceived and designed the experiments: AFMS, EAMM, LVS, HYH, MMvW. Performed the experiments: LPHvdR, HYH, LVS. Analyzed the data: LPHvdR, MMvW. Wrote the paper: LPHvdR, MMvW, AJvO, AFMS, EAMM.

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van de Rijt et al.

Audio-Visual Temporal fNIRS Activation

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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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