The following full text is a publisher's version.

For additional information about this publication click this link.
http://hdl.handle.net/2066/167966

Please be advised that this information was generated on 2019-11-27 and may be subject to change.
Mutations in AGBL5, Encoding α-Tubulin Deglutamylase, Are Associated With Autosomal Recessive Retinitis Pigmentosa

Galuh D. N. Astuti,1–3 Gavin Arno,4 Sarah Hull,3 Laurence Pierrache,5–8 Hanka Venselaar,9 Keren Carss,10,11 F. Lucy Raymond,11,12 Rob W. J. Collin,1,13 Sultana M. H. Faradz,3 L. Ingeborgh van den Born,5 Andrew R. Webster,4 and Frans P. M. Cremers1,13

1Department of Human Genetics, Radboud University Medical Center, Nijmegen, The Netherlands
2Radboud Institute for Molecular Life Sciences, Radboud University, Nijmegen, The Netherlands
3Division of Human Genetics, Center for Biomedical Research, Faculty of Medicine, Diponegoro University, Semarang, Indonesia
4University College London, Institute of Ophthalmology, London, United Kingdom
5The Rotterdam Eye Hospital, Rotterdam, The Netherlands
6Rotterdam Ophthalmic Institute, Rotterdam, The Netherlands
7Department of Ophthalmology, Erasmus Medical Center, Rotterdam, The Netherlands
8Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands
9Centre for Molecular and Biomolecular Informatics, Radboud University, Nijmegen, The Netherlands
10Department of Haematology, University of Cambridge, Cambridge, United Kingdom
11National Institute for Health Research England BioResource–Rare Diseases, Cambridge University Hospitals, Cambridge Biomedical Campus, Cambridge, United Kingdom
12Department of Medical Genetics, Cambridge Institute for Medical Research, University of Cambridge, Cambridge, United Kingdom
13Donders Institute for Brain, Cognition and Behaviour, Radboud University Nijmegen, The Netherlands

Correspondence: Frans P. M. Cremers, Department of Human Genetics, Radboud University Medical Center, P.O. Box 9101, 6500 HB, Nijmegen, The Netherlands; Frans.Cremers@radboudumc.nl.

AGBL5, encoding ATP/GTP binding protein-like 5, was previously proposed as an autosomal recessive retinitis pigmentosa (arRP) candidate gene based on the identification of missense variants in two families. In this study, we performed next-generation sequencing to reveal additional RP cases with AGBL5 variants, including protein-truncating variants.

METHODS. Whole-genome sequencing (WGS) or whole-exome sequencing (WES) was performed in three probands. Subsequent Sanger sequencing and segregation analysis were performed in the selected candidate genes. The medical history of individuals carrying AGBL5 variants was reviewed and additional ophthalmic examinations were performed, including fundus photography, fundus autofluorescence imaging, and optical coherence tomography.

RESULTS. AGBL5 variants were identified in three unrelated arRP families, comprising homozygous variants in family 1 (c.1775G>A:p.(Trp592*)) and family 2 (complex allele: c.[323C>G; 2659T>C]; p.[(Pro108Arg; 887Argext*1)]), and compound heterozygous variants (c.752T>C:p.(Val251Gly) and c.1504dupG:p.(Ala502Glyfs*15)) in family 3. All affected individuals displayed typical RP phenotypes.

CONCLUSIONS. Our study convincingly shows that variants in AGBL5 are associated with arRP. The identification of AGBL5 and TTLL5, a previously described RP-associated gene encoding the tubulin tyrosine ligase-like family, member 5 protein, highlights the importance of poly- and deglutamylation in retinal homeostasis. Further studies are required to investigate the underlying disease mechanism associated with AGBL5 variants.

Keywords: retinitis pigmentosa, whole exome sequencing, AGBL5, post-translational modification

Retinitis pigmentosa (RP) (Mendelian Inheritance in Man [MIM]268000) encompasses a clinically and genetically heterogeneous group of inherited retinal dystrophies (IRDs) with a worldwide prevalence of approximately 1 in 4000 individuals. Retinitis pigmentosa is characterized by initial rod photoreceptor degeneration resulting in night blindness, followed by midperipheral visual field loss and deterioration of central vision due to cone photoreceptor degeneration.1 The clinical presentation of RP is highly variable in terms of onset, disease progression, fundus appearance, and associated ocular features. The ophthalmic features indicative for RP include bone spicule pigmentation with atrophy of the retinal pigment epithelium (RPE), waxy disc pallor, and retinal arteriole attenuation.2–5

To date, 82 genes have been implicated in RP (https://sph.uth.edu/retnet; in the public domain). These genes encode proteins involved in different retinal pathways, including the phototransduction cascade, the retinoid cycle, ciliary transport,
AGBL5 Mutations Are Associated With arRP

Subjects and Methods

Subjects and Clinical Evaluation

The study protocol adhered to the tenets of the Declaration of Helsinki and received approval from the respective local ethics committees. Written informed consent was obtained from all participants or parents of children prior to their inclusion in this study. Patients were ascertained from the inherited retinal disease clinics at the Rotterdam Eye Hospital (The Netherlands) and Moorfields Eye Hospital (London, UK).

Each patient underwent a full clinical examination including visual acuity and dilated fundus examination. Retinal fundus imaging was obtained by 35° color fundus photography (Topcon Great Britain Ltd, Berkshire, UK), 55° fundus autofluorescence (FAF) imaging (Spectralis; Heidelberg Engineering Ltd, Heidelberg, Germany), and Spectralis optical coherence tomography (OCT).

Full-field and pattern electroretinography (ERG, PERG) were available from three patients, performed with gold foil electrodes, which incorporated the International Society for Clinical Electrophysiology of Vision (ISCEV) standards.9,10 We also studied all retrospective data including Stratus (Carl Zeiss Meditec, Dublin, CA, USA) OCTs.

Genetic Analysis

Genomic DNA was isolated from peripheral blood lymphocytes according to standard salting-out procedures.11 In one unaffected individual from family 1, DNA was extracted from saliva material using the Oragene system (OG-500; Gentek, Ottawa, ON, Canada).

Whole-exome sequencing of the proband from family 1 was performed using Agilent’s SureSelect All Human Exome version 2 kit (50 Mb; Agilent Technologies, Santa Clara, CA, USA), followed by sequencing on a SOLiD4 sequencing platform (Life Technologies, Carlsbad, CA, USA). Reads were mapped against the University of California, Santa Cruz (UCSC) genome browser, human Hg19 assembly (build 37) using Lifescope v2.1 software (Life Technologies), and variants were called using the Genome Analysis Toolkit (GATK) v2 according to the recommended guidelines (https://software.broadinstitute.org/gatk/best-practices/; in the public domain). Identified variants were filtered based on minor allele frequency (MAF)~<0.005 in the 1000 genomes database, the NHLBI GO Exome Sequencing Project (http://evs.gs.washington.edu/EVS/ release 20130513; in the public domain), the UK10K database (http://www.uk10k.org; in the public domain), the ExAC database (http://exac.broadinstitute.org; in the public domain), and allele frequency <0.02 in ~6000 internal control genomes.

Sanger Sequencing

Primers for amplification of coding exons and flanking exon-intron boundaries of AGBL5 (NM_021831.5) were designed with Primer3 plus (http://wwwbioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi; in the public domain). Primer sequences and PCR conditions are available upon request.

Pathogenicity Interpretation of Missense Variant

The Combined Annotation Dependent Depletion (CADD) (http://cadd.gs.washington.edu/; in the public domain) score was used to predict the pathogenicity of missense variants. This scoring system incorporates several widely used in silico tools, such as SIFT (http://sift.jcvi.org/; in the public domain), PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2/; in the public domain), Mutation Taster (http://www.mutationtaster.org/; in the public domain), PhyloP, and Grantham score. Minor allele frequencies were obtained from the Exome Aggregation Consortium (ExAC) database containing ~60,000 exomes of retinal development, and RNA splicing factors.4 Recent whole-exome sequencing (WES) studies demonstrated that approximately 60% of RP cases can be explained by variants in these genes (Haer-Wigman L, personal communication, 2016),5–8 suggesting that several genes are yet to be identified. This study aimed to identify novel genes associated with RP using next-generation sequencing (NGS) technology.

Here, we describe the identification of biallelic variants in AGBL5 in four autosomal recessive retinitis pigmentosa (arRP) patients from three unrelated families.

were filtered based on minor allele frequency (MAF)~<0.005 in our Exome Aggregation Consortium (ExAC) database containing 60,000 internal control genomes. Calling was performed using the haplotype caller module of GATK (https://www.broadinstitute.org/gatk, version 3.3.0; in the public domain), creating gVCF formatted files for each sample. The individual gVCF files for the exomes discussed in this study, in combination with ~3000 clinical exomes (ULC-exomes consortium, University College London, London, UK), were combined into merged VCF files for each chromosome, on average containing 100 samples each. The final variant calling was performed using the GATK Genotype gVCFs module jointly for all samples (cases and controls). Variant quality scores were then recalibrated according to GATK best practices separately for indels and single nucleotide polymorphisms (SNPs). Resulting variants were annotated using ANNOVAR (http://annovar.openbioinformatics.org/; in the public domain) based on Ensembl (http://www.ensembl.org; in the public domain) gene and transcript definitions. Candidate variants were filtered based on function (non-synonymous, presumed loss of function or splicing, defined as intronic sites within 5 bp of an exon–intron junction) and minor allele frequency (<0.005 minor allele frequency in our internal control group, as well as the National Heart, Lung, and Blood Institute [NHLBI] exome sequencing dataset)6000 internal control genomes.

subjects were filtered only on the single nucleotide variants (SNVs) and indels that met the following criteria: passed standard quality filters, predicted to alter the sequence of a protein, and had an allele frequency <0.01 in the 1000 genomes database, the NHLBI GO Exome Sequencing Project (http://evs.gs.washington.edu/EVS/ release 20130513; in the public domain), the UK10K database (http://www.uk10k.org; in the public domain), the ExAC database (http://exac.broadinstitute.org; in the public domain), and allele frequency <0.02 in ~6000 internal control genomes.

Pathogenicity Interpretation of Missense Variant

The Combined Annotation Dependent Depletion (CADD) (http://cadd.gs.washington.edu/; in the public domain) score was used to predict the pathogenicity of missense variants. This scoring system incorporates several widely used in silico tools, such as SIFT (http://sift.jcvi.org/; in the public domain), PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2/; in the public domain), Mutation Taster (http://www.mutationtaster.org/; in the public domain), PhyloP, and Grantham score. Minor allele frequencies were obtained from the Exome Aggregation Consortium (ExAC) database containing ~60,000 exomes of
Mutations Are Associated With arRP

Patients from families 2 and 3 initially developed nyctalopia, with a range of onset from 9 to 30 years before subsequent development of peripheral visual field loss. At last review (age range, 24–45 years), there was mild visual acuity loss with best-corrected visual acuity mean 0.34 logMAR (range, 0.30–0.40). All patients developed posterior subcapsular cataract. Patient F2-II:1 had learning difficulties but no other symptoms suggestive of syndromic disease. There were fundus abnormalities consistent with RP including attenuated vessels, macular and midperipheral RPE mottling/atrophy, and peripheral intraretinal pigment migration, most prominent in the proband of family 3 (Fig. 2I). Fundus autofluorescence imaging demonstrated a ring of increased autofluorescence parafocally in F2-II:1, focally increased autofluorescence corresponding to macular edema in F2-II:3, and a partial ring of reduced autofluorescence in F3-II:1 (Figs. 2D, 2G, 2J). All patients had midperipheral reduced autofluorescence in keeping with regions of RPE atrophy and increased pigmentation. On OCT, there was outer retinal atrophy with variable preservation of photoreceptors most apparent in F2-II:1 with a centrally preserved inner segment ellipsoid band (Fig. 2D). Optical coherence tomography imaging demonstrated intraretinal cysts of the inner nuclear layer, which responded to topical carbonic anhydrase inhibitors (Figs. 2E, 2H, 2K). Electrophysiology demonstrated rod and cone system dysfunction (Table 1).

Genetic Findings

Biallelic variants in AGBL5 were identified in four patients from three families. These variants are summarized in Tables 1 and 2. None of them have been previously reported in RP patients. Whole-exome sequencing data analysis did not yield other biallelic variants in the genes known to be mutated in IRD (Retinal Information Network, https://sph.uth.edu/retinet/); in the public domain). The proband of family 1 was homozygous for the c.1775G>A.p.(Trp592*) nonsense variant in the AGBL5 gene (Fig. 3). The resulting transcript is expected to undergo nonsense-mediated decay (NMD), suggesting that this variant is a true loss-of-function allele. This variant was reported in 1/121,398 alleles (0.0008%) in the ExAC dataset.

Two affected siblings from family 2 (GC15894) were homozygous for the missense variant c.323C>G.p.(Pro108Arg). All in silico tools predict this variant to be pathogenic (Table 2). The altered proline residue is fully conserved throughout AGBL5 orthologues up to Tetraodon nigroviridis (Fig. 3A) and is located in the core of AGBL5 (Fig. 3B). A change to arginine introduces a larger amino acid, which may destabilize the protein structure. Moreover, proline provides a rigid structure important for interaction with ligands or other proteins. A second missense variant, c.2659T>C.p.(887Argext*1), was also identified in the two affected members of this family. This stop-loss variant is predicted to result in a single amino acid residue extension of the protein. Two rare stop-loss variants affecting other nucleotides of the AGBL5 stop codon, c.2661A>G.p.(887Trpext*1) and c.2661A>T.p.(887Cysext*1), were found in one and seven alleles in the ExAC dataset, respectively.

The compound heterozygous variants c.1504dupG.p.(Ala502fs*15) and c.752T>G.p.(Val251Gly) were identified in a simplex RP patient of family 3. The p.Val251Gly variant alters the valine residue flanking the consensus zinc-binding residues of the M14-like peptidase domain. This valine is conserved throughout paralogues and orthologues of AGBL5 (up to T. nigroviridis). Furthermore, it affects a highly conserved nucleotide with a PhyloP score of 4.75. According to Vissers et al.,12 by using PhyloP score distribution of dbSNP and HGMD variants, this variant can be considered as...
Mutations Are Associated With arRP

### TABLE 1. Clinical Phenotype of Patients Carrying AGBL5 Variants

<table>
<thead>
<tr>
<th>Family ID, Sex, Origin</th>
<th>Age at Diagnosis, Recent Exam, y</th>
<th>Visual Acuity (Snellen, logMAR) Refraction</th>
<th>Ophthalmoscopy</th>
<th>OCT</th>
<th>FAF</th>
<th>Other Symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td>F1-IV:3 (DNA121985, M, Turkey)</td>
<td>16, 43</td>
<td>RE: LP+/C0 +16, 43 Central vision loss, nyctalopia (2nd decade)</td>
<td>NA (Pseudoaphakia)</td>
<td>NA</td>
<td>Cystoid maculopathy progressing into complete loss of outer retinal layers and thinning inner retinal layers at fovea</td>
<td>NP</td>
</tr>
<tr>
<td>F2-II:1 (GC15894.1, F, UK)</td>
<td>9, 24</td>
<td>RE: 6/18 (0.48)</td>
<td>LE: 6/12 (0.3)</td>
<td>LE: 0.50/1.50 × 0.50/1.50 × 100</td>
<td></td>
<td></td>
</tr>
<tr>
<td>F2-II:2 (GC15894.2, F, UK)</td>
<td>20, 27</td>
<td>RE: 6/18 (0.48)</td>
<td>LE: 6/12 (0.3)</td>
<td>LE: 0.50/1.00 × 0.50/1.00 × 1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>F3-II:1 (GC3687, F, UK)</td>
<td>39, 65</td>
<td>RE: 6/12 (0.3)</td>
<td>LE: 6/12 (0.3)</td>
<td>LE: 0.25 DS</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

CAI, Carbonic Anhydrase Inhibitor; DS, Dioptres Sphere; FAF fundus autofluorescence; L, left; LE, left eye; logMAR, Logarithm of the Minimum Angle of Resolution, to measure metric visual acuity; LP, light perception; NA, not applicable; NP, not performed; OCT, optical coherence tomography; R, right; RE, right eye; RPE, retinal pigmented epithelium.
Pathogenic as it is close to the mean PhyloP score (4.7) of the pathogenic de novo variants identified in persons with intellectual disability. All in silico prediction tools indicate that this variant is pathogenic (Table 2). Alteration of this hydrophobic valine to glycine may destabilize the secondary structure of AGBL5 in this domain (Fig. 3B). All variants cosegregated with disease in the family members available for testing (Fig. 1).

**DISCUSSION**

*AGBL5* Is a Novel Gene Associated With RP

Previous studies by Patel et al.5 and Kastner et al.13 discovered *AGBL5* as a novel candidate gene associated with RP. These studies identified homozygous missense variants (p.(Arg276Trp) and p.(Asp295Asn)) in arRP cases from Saudi and Turkish origin.

**Table 2.** *AGBL5* Variants Identified in This Study and Their In Silico Analyses

<table>
<thead>
<tr>
<th>DNA Variant</th>
<th>Protein Variant</th>
<th>ExAC Allele Frequency, %</th>
<th>SIFT</th>
<th>PolyPhen-2</th>
<th>Mutation Taster</th>
<th>CADD Phred</th>
<th>PhyloP</th>
</tr>
</thead>
<tbody>
<tr>
<td>c.323C&gt;G</td>
<td>p.(Pro108Arg)</td>
<td>0</td>
<td>Deleterious</td>
<td>Probably damaging</td>
<td>Disease causing</td>
<td>24.0</td>
<td>5.45</td>
</tr>
<tr>
<td>c.752T&gt;G</td>
<td>p.(Val251Gly)</td>
<td>0</td>
<td>Deleterious</td>
<td>Probably damaging</td>
<td>Disease causing</td>
<td>27.1</td>
<td>4.73</td>
</tr>
<tr>
<td>c.1775G&gt;A</td>
<td>p.(Trp592*)</td>
<td>0.0008</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>c.1504dupG</td>
<td>p.(Ala502Glyfs*15)</td>
<td>0</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>c.2659T&gt;C</td>
<td>p.(888Argext*1)</td>
<td>0</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
</tbody>
</table>

NA, not applicable.
In our study, we identified five novel variants in *AGBL5* in three unrelated RP cases that further confirm the causality of this gene. The novel variants consist of two missense, two protein-truncating, and one stop-loss variant. To date, all missense variants in *AGBL5* with the exception of p.(Pro108Arg) are located within the M14-like carboxypeptidase A domain (Fig. 3A), which highlights the importance of this protein domain for *AGBL5* function. This protein domain encodes cytosolic carboxypeptidase-like protein 5; ATP/GTP-binding protein-like 5. Wild-type and mutant residues are shown and colored in cyan and green, respectively. The p.(Pro108Arg) variant, alter proline in position 108 to arginine, is located in the core of the *AGBL5* protein. This alteration created a larger amino acid, which may destabilize the protein structure. The p.(Val251Gly) missense variant changed the hydrophobic valine to glycine and may destabilize the structure of *AGBL5*. *These two variants were identified from previous studies by Patel et al.*\(^5\) and Kastner et al.,\(^13\) respectively.

**FIGURE 3.** Exon–intron structure of *AGBL5*, RP-associated variants, and three-dimensional modeling of missense variants. (A) Schematic representation of *AGBL5* variants identified in this and previous studies. Cross-species amino acid comparison showed that all missense variants affect highly conserved residues. Variants identified in this study are colored in red, and variants in previous studies are in black. *These two variants occurred homozgyously in F2. (B) Three-dimensional protein modeling of *AGBL5* missense variants. Three-dimensional structure of cytosolic carboxypeptidase-like protein 5; ATP/GTP-binding protein-like 5. Wild-type and mutant residues are shown and colored in cyan and green, respectively. The p.(Pro108Arg) variant, alter proline in position 108 to arginine, is located in the core of the *AGBL5* protein. This alteration created a larger amino acid, which may destabilize the protein structure. The p.(Val251Gly) missense variant changed the hydrophobic valine to glycine and may destabilize the structure of *AGBL5*. *These two variants were identified from previous studies by Patel et al.*\(^5\) and Kastner et al.,\(^13\) respectively.

In our study, we identified five novel variants in *AGBL5* in three unrelated RP cases that further confirm the causality of this gene. The novel variants consist of two missense, two protein-truncating, and one stop-loss variant. To date, all missense variants in *AGBL5* with the exception of p.(Pro108Arg) are located within the M14-like carboxypeptidase A domain (Fig. 3A), which highlights the importance of this protein domain for *AGBL5* function. This protein domain encodes cytosolic carboxypeptidase-like (CCPs) that have been implicated to play a role in the posttranslational modifications of tubulin. Furthermore, mutation of the active site of the carboxypeptidase domain causes abnormal cilia development in zebrafish.\(^14\) The protein-truncating variants, that is, p.(Trp592*) and p.(Ala502Glyfs*15), are expected to result in complete loss of function due to premature translation termination and NMD.

The stop-loss p.(Arg887ArgExt*) variant found in family 2 results in loss of the canonical termination codon, which leads to the extension of the transcript by one codon and consequent extension of the protein by a single amino acid residue. This finding in addition to the presence of eight alleles in control data (ExAC) with similar stop-loss variants may suggest that the stop-loss variant is a bystander in the pathogenesis of *AGBL5* retinopathy, although a modifier or indeed pathogenic effect cannot be ruled out.

The two arRP families reported by Patel et al.\(^5\) and Kastner et al.\(^13\) display a classical RP phenotype, as do our patients. There was some variability in visual function; our eldest patient (age 63) still had relatively well-preserved visual acuity, whereas one subject was blind at age 32. All patients showed retinal cysts in the macula region at some stage of the disease, but this occurs in 18% to 32% of RP patients.\(^15\)

Interestingly, Kastner et al.\(^13\) reported intellectual disability in an affected male and proposed a possible correlation between sex and brain phenotype. In our study, learning difficulties occurred in a female patient, which does not support a correlation between sex and brain phenotypes. Additional RP patients with *AGBL5* variants need to be identified to gain more insight into a possible association with brain phenotypes.

---

**AGBL5 Mutations Are Associated With arRP**

*IOVS | November 2016 | Vol. 57 | No. 14 | 6185*
Pathak et al.14 demonstrated that knockdown of Agb5 in zebrafish leads to ciliopathy phenotypes, that is, axis curvature, hydrocephalus, pronephric cysts, and abnormal multicilia motility. However, in humans, variants in AGBL5 are associated with nonsyndromic RP. These phenotypic discrepancies might be caused by homologous genes in humans that might compensate for the effect caused by these mutations. Alternatively, as the NMD event was not functionally assessed, there is a possibility that in the human patients, truncated proteins with residual activity are being produced. Finally, recent studies show that the use of morpholinos for gene knockdown in zebrafish may lead to off-target effects, thereby explaining additional phenotypes observed in the zebrafish studies.16

Imbalance of Poly- and Deglutamylation in Posttranslational Modifications Is Associated With RP

Approximately one-third of IRD-associated genes play roles in maintaining the structure and function of photoreceptor sensory cilium.17 Previous studies have implicated TTLL5, a gene encoding a polyglutamylase (tubulin tyrosine ligase-like family, member 5) in the pathogenesis of autosomal recessive cone–rod dystrophy.18 AGBL5, also known as CCP5, is known to function in polyglutamylation and deglutamylation of tubulin, and perturbations in this pathway may lead to photoreceptor cell degeneration.19 Tubulin glutamylation is an essential posttranslational modification associated with stable microtubules, neuronal axons, mitotic spindles, centrioles, and cilia.20 The tubulin tyrosine ligase-like (TTLL) and CCP deglutamylases are required to maintain the balance of tubulin glutamylation in ciliogenesis.21,22 Any imbalance in poly- and deglutamylation might disrupt cilia function. Hyperglutamylation is known to diminish Tetrahymena cilia motility and to induce axonemal microtubules defects.23 TTLL5 mutant mice displayed a complete loss of retinitis pigmentosa GTPase regulator (RPGR) glutamylation without marked changes in tubulin glutamylation levels. The TTLL5 mutant mouse developed slow photoreceptor degeneration with early mislocalization of cone opsins, features resembling those of Rpgr-null mice.18 CCP5 knockdown in zebrafish led to hyperglutamylation and reduced the disorganized cilia beat pattern. Ttll5, on the other hand, plays a role in polyglutamylation, which is crucial for microtubule sliding and cilia waveform regulation.24 Interestingly, a recent study demonstrates the role of TTLL5 in RPGRQ1R3 glutamylation.24 The opposite function of AGBL5 (CCP5) in the deglutamylation pathway may also play an important role in the balance of glutamate chains in RPGRQ1R3. Posttranslational modifications are regulated by several members of the TTLL and CCP gene families, and we hypothesize that besides AGBL5 and CCP5, other TTLL (TTLL1 through TTLL11) and CCP (CCP1 through CCP6) genes are plausible candidate genes for IRD.

In conclusion, this study strengthens the role of AGBL5 variants in RP. The involvement of both AGBL5 and TTLL5 in IRDs underlines the critical importance of glutamylation in retinal function.

Acknowledgments

The authors thank the patients and family members for participation in the study.

The research of GDN was supported by the Directorate General for Higher Education (DIKTI) of the Ministry for National Education of Indonesia and the Radboud University Medical Center, Nijmegen, The Netherlands. The research of GA, SH, and ARW was supported by the National Institute for Health Research England (NIHR) BioResource–Rare Diseases (RG05906), the NIHR Biomedical Research Centre at Moorfields Eye Hospital, and the University College London Institute of Ophthalmology (BRC2_003); the Foundation Fighting Blindness (USA, CCL/0710-0505-MEH1002); Fight For Sight (1518 and 1801); Moorfields Eye Hospital Special Trustees (ST1109B); Rosetrees Trust (M184); RP Fighting Blindness (GR581). LP is supported by Foundation Combined Ophthalmic Research Rotterdam (CORR).

Disclosure: G.D.N. Astuti, None; G. Arno, None; S. Hull, None; L. Pierrache, None; H. Venselaar, None; K. Carss, None; F.L. Raymond, None; R.W.J. Collin, None; S.M.H. Faradz, None; L.I. van den Born, None; A.R. Webster, None; F.P.M. Cremers, None

References


