

## PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.

<http://hdl.handle.net/2066/16718>

Please be advised that this information was generated on 2019-10-22 and may be subject to change.

# 4

## Calcium Regulation in Fish

S.E. Wendelaar Bonga and G. Flik

Calcium is important for growth, reproduction, and many other physiological processes such as muscle contraction, nerve signal transduction, and control of membrane permeability and cellular metabolism. These processes are dependent on an effective control of the ionic calcium levels of the extracellular fluid (ECF). Although fishes seem to be slightly more resistant to changes of ionic calcium of the ECF than the terrestrial vertebrates, deviations from the normal physiological concentration ("setpoint") have essentially the same effects in all vertebrates: hypocalcemia leads to increased excitability of cellular membranes and may result in tetanic seizures, whereas hypercalcemia reduces membrane excitability with lethargy and cardiovascular dysfunction as consequences. Precise control of the setpoint of ionic calcium is the rule in fish as well as in other vertebrates.

The maintenance of constant calcium levels in the ECF is effected in fish by the continuous exchange of calcium between the animals and the environment, mainly via the gills. In full-grown animals influx and efflux of calcium are in equilibrium, whereas during periods of growth and reproduction net calcium accumulation occurs. During such periods large amounts of calcium are transported from the sites of entry through the blood and other compartments of the ECF to bones or ovaria. This represents a special challenge for the mechanisms that control the calcium concentration of the ECF.

There are several important differences between fish and the terrestrial vertebrates with respect to the organs involved in the maintenance of the calcium balance of the body, and in the endocrine mechanisms controlling these organs. Firstly, whereas

in terrestrial vertebrates calcium is obtained exclusively from the food via the gut, in fish the gills are in general the main route of uptake of calcium with the gut in a secondary role. The ambient water seems sufficient as a calcium source in fish. Our unpublished studies on the African cichlid *Oreochromis mossambicus* have shown that calcium-deficient food does not have any effect on the calcium balance of tilapia, which confirmed similar observations on goldfish by Ichii and Mugiya (1983). The uptake of calcium in fish then is an essentially continuous process, in contrast to the calcium uptake in terrestrial vertebrates, which is episodic and restricted by the calcium content of the food. The second important difference between fish and terrestrial vertebrates is the role played by the skeleton. In the absence of a continuous access to external calcium sources, terrestrial animals have developed the bone tissue as a calcium store which is used for the minute-to-minute control of the calcium level of the body fluids. We will discuss below that a calcium exchange capacity of the skeleton in fish is not required because the control of calcium homeostasis in fish is mainly effected at the level of the gills, in particular by regulation of the calcium influx from the water. Thirdly, and not surprisingly, given the above-mentioned characteristics of calcium metabolism in fish, there are essential differences in the hormonal mechanisms regulating the calcium balance between fish and the terrestrial vertebrates. In this paper we will concentrate on these regulatory aspects.

#### A. Terrestrial Vertebrates

In the terrestrial vertebrates two hormones are specifically involved in the control of calcium metabolism: parathyroid hormone (PTH) and calcitonin. PTH is the dominant hormonal factor. It stimulates the intestinal uptake and reduces the renal excretion of calcium, processes mediated by a vitamin-D<sub>3</sub>-metabolite 1,25-dihydroxycholecalciferol. PTH further mobilizes calcium from the skeleton. When the ionic calcium concentration of the ECF falls below the physiological setpoint (about 1.24 mM Ca<sup>2+</sup>) the

PTH-cells respond within seconds with the enhanced secretion of PTH (Brown *et al.* 1987). The hormone stimulates the osteoclasts, and this leads to a rapid rise of the extracellular  $\text{Ca}^{2+}$  level. Calcium concentrations above the setpoint inhibit PTH secretion. The rapid response of the PTH cells is mediated by the special  $\text{Ca}^{2+}$ -sensing capacity of these cells (Jones and Fitzpatrick 1990).

Calcitonin has long been considered the hormonal antagonist of PTH, because of its reportedly hypocalcemic action. Although hypocalcemic effects have been shown after injection of the hormone in young and growing mammals, birds and reptiles, consistent hypocalcemia could not be demonstrated in adult animals (Copp and Kline 1989). Neither chronically elevated calcitonin levels, nor removal of the calcitonin-producing C-cells by thyroidectomy (in mammals) or ultimobranchialectomy (in birds and amphibians) are inducing persistent imbalance of the extracellular  $\text{Ca}^{2+}$  levels (see Wendelaar Bonga and Pang 1991). This sharply contrasts with the effects of parathyroidectomy, which include hypocalcemia, tetany and death. One role of calcitonin is the reduction of hypercalcemia that occurs after intestinal absorption of calcium following a meal (VanderWiel and Talmage 1981). It promotes the deposition of ingested calcium into the bone. It further protects the skeleton against excessive demineralization during periods of increased calcium demand, such as pregnancy and lactation in mammals (Stevenson *et al.* 1979). In conclusion, in the terrestrial vertebrates the exchange of calcium with the environment is mainly regulated by PTH, that also controls the  $\text{Ca}^{2+}$  concentration of the ECF. The bone plays an essential role as a calcium buffer. Compared to PTH, calcitonin only has a minor function in the homeostatic control of extracellular  $\text{Ca}^{2+}$ .

## B. Fish

Our knowledge of the calcium metabolism and the endocrine control in fish is mainly based on freshwater teleosts and therefore we will mainly deal with this group. Teleost fish maintain plasma total and ionic calcium within narrow limits, but these limits are to some extent dependent on the growth rate and on the cal-

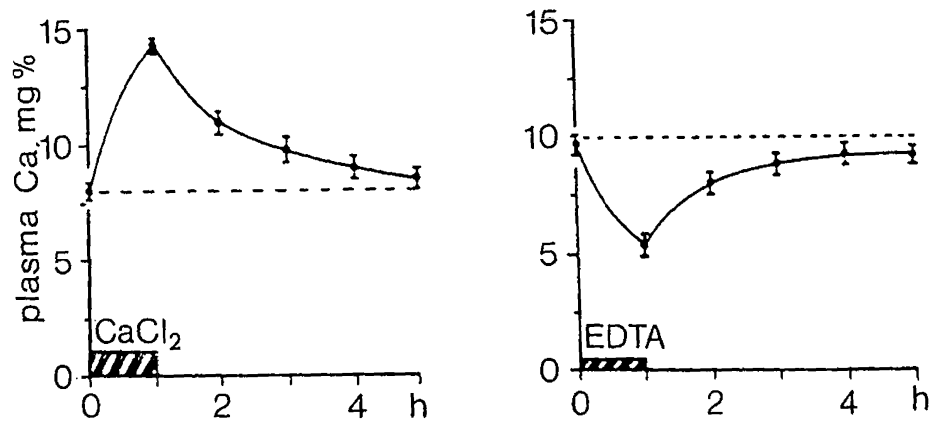


Fig. 1. Recovery from induced hyper- or hypocalcemia in rainbow trout (Copp *et al.* 1985).

cium concentration of the ambient water (Urasa and Wendelaar Bonga 1987). When plasma calcium levels of teleost fish are changed experimentally, normal levels are rapidly restored. In rainbow trout (Copp *et al.* 1985) and American eels (Fenwick and Bresseur 1991) it took 3 to 6 h before a twofold increase of plasma total calcium had decreased to control values (Fig. 1). Restoration of a 50% drop in plasma calcium induced by EGTA infusion in rainbow trout lasted a similar period (Copp *et al.* 1985; Fig. 1). These data show that teleost fish can successfully restore changes in the calcium concentration of the extracellular fluid. The response may be similar to or even more rapid than in the higher vertebrates. The changes in calcium levels in the terrestrial animals are compensated by modulating the exchange of calcium between the bone and the ECF. How is this compensation effected in fish?

#### B. 1. Bone

In amphibians and fishes, the calcium-exchange capacity of the bones has been frequently considered as limited. This does certainly not imply that calcium mobilization from bone does not occur. There are several reports for teleost fish with cellular as well as acellular bone indicating that the calcium concentration of bone varies in response to environmental or physiological changes (Fleming *et al.* 1973). In tilapia, a species with

acellular bone, reduction of the water calcium concentration or of the water pH - both stimulate calcium efflux across the gills - causes a transient decrease of the calcium concentration of the bone (Wendelaar Bonga and Dederen 1985). Induction of scale regeneration by removal of mature scales is accompanied by a transient reduction of the calcium concentration of the remaining scales (Weiss and Watabe 1978). Forced gonadal maturation by injection of estrogens may lead to calcium mobilization from the bone and/or the scales (Mugiya 1982). Thus, calcium can be mobilized from skeleton and scales in fish. The contribution of calcium mobilization from the bone to the calcium homeostasis of the ECF is unclear, however. Except for estrogens no hormone is known that stimulates bone calcium release in fish. The function of bone as a readily accessible and hormonally controlled calcium store seems to have been evolved after the water-to-land transition of the vertebrates, in conjunction with the appearance of PTH and a system of bone cells with calcium-releasing activity that rapidly responds to this hormone (Pang *et al.* 1980; Wendelaar Bonga and Pang 1991).

## B.2. Gills

If the bone does not function as a fast-acting calcium buffer, the most likely mechanism for the minute-to-minute control of the extracellular calcium concentration is the regulation of the calcium-exchange between the fish and its environment. The exchange of calcium is much higher in fish than in terrestrial vertebrates. This is mainly caused by the calcium fluxes across the branchial epithelium. Although this epithelium is very tight, in particular in fresh water, there is still a substantial passive efflux of calcium ions. This loss of calcium ions is compensated by the active uptake of calcium ions through the gills and, to a lesser extent, the intestine. Whereas the branchial calcium efflux most likely follows a paracellular route, the calcium influx is an active, transcellular process. We have investigated this process in detail, and these studies have resulted in a

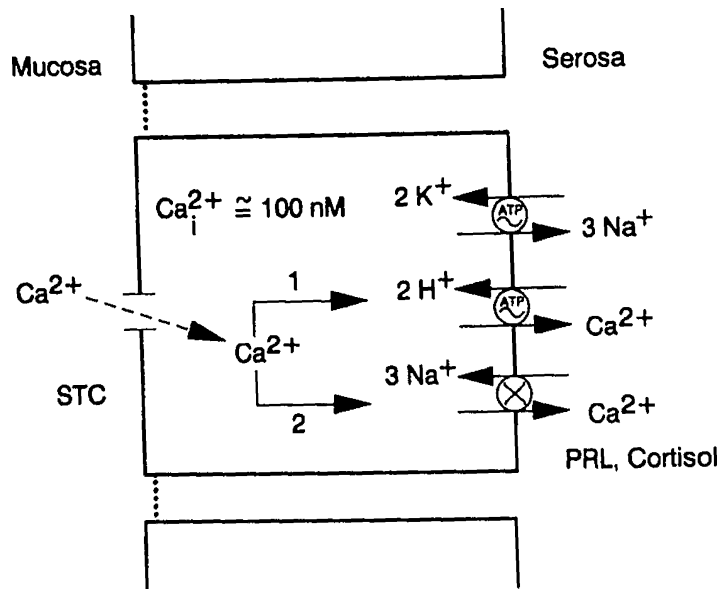


Fig. 2. Transcellular transport of  $\text{Ca}^{2+}$  in fish gills.  $\text{Ca}^{2+}$  from the water may enter the cell passively, down an electrochemical gradient. Stanniocalcin (STC) regulates calcium channels in the apical membrane; STC signal transduction requires second (cAMP, DAG,  $\text{IP}_3$ ) and third ( $\text{Ca}^{2+}$ ) messengers. Extrusion of  $\text{Ca}^{2+}$  from the cell could occur via a  $\text{Ca}^{2+}$ -ATPase (route 1) or via a  $\text{Na}^+/\text{Ca}^{2+}$  exchanger (route 2). Both extrusion mechanisms are present in branchial epithelium. In gill cells route 1 probably dominates. Hormones with hypercalcemic effects such as prolactin (PRL) in several species and cortisol (in trout) determine the relative abundance of the ATPases and by doing so may determine the calcium transporting capacity of the epithelium (Flik *et al.* 1992).

model that has been verified for a number of teleost species, including trout, tilapia and two eel species (Fig. 2). The calcium-influx is effected by the chloride cells, the cells specialized for ion transport in fish gills. Calcium ions from the water are entering the cytoplasm of the chloride cells via the apical membrane of these cells, probably through voltage-independent calcium channels, and following the electrochemical gradient (Verbost *et al.* 1989). The cytoplasmic ionic calcium concentration is two to three orders of magnitude lower than in the ambient water or in the ECF. This gradient favours the entrance of calcium into the cell. However, special mechanisms are required for the extrusion of calcium ions from the cytoplasm across the basolateral membrane into the ECF. We have identified these mech-

anisms as ATP-driven  $\text{Ca}^{2+}$ -pumps and  $\text{Na}^+/\text{Ca}^{2+}$ -exchangers. Studies on the activity of both mechanisms as a function of the  $\text{Ca}^{2+}$  concentration have shown that the  $\text{Ca}^{2+}$ -pump is more important than the  $\text{Na}^+/\text{Ca}^{2+}$ -exchanger for the extrusion of calcium from the chloride cells. The reverse is indicated for the intestine (for a review of our studies see Flik *et al.* 1992).

### B. 3. Endocrine Control of Calcium Metabolism.

#### a. Prolactin

The absence of parathyroid glands from purely aquatic amphibians and fish has now been well established. This absence has stimulated the search for a hormone with a similar hypercalcemic action. Several studies have indicated that the pituitary gland is the source of such a hormone, with prolactin as the most likely candidate. In the terrestrial vertebrates prolactin has some effects on calcium metabolism, in particular during the female reproductive period: it stimulates intestinal calcium uptake and the transfer of calcium to the offspring via the placenta or the mammary glands (see Wendelaar Bonga and Pang 1991).

In fish the role of prolactin in calcium metabolism seems more important than in the terrestrial animals. Removal of the pituitary gland in freshwater fish usually leads to mild hypocalcemia although data in the literature are not always consistent (Wendelaar Bonga and Pang 1989). Pang *et al.* (1978) have shown that the hypocalcemia of hypophysectomized killifish from low-calcium freshwater can be corrected by injections of pituitary homogenates or mammalian prolactin preparations. The latter produced hypercalcemia in e.g. intact sticklebacks, tilapia, trout, salmon or eel (Wendelaar Bonga and Pang 1989). Recently, using recombinant hormone preparations, we have shown that both forms of tilapia prolactin induce hypercalcemia in intact freshwater tilapia (Swennen *et al.* 1991). The most likely mechanism of action of prolactin is the stimulation of the activity of the ATP-dependent  $\text{Ca}^{2+}$ -pumps of the chloride cells: prolactins of mam-



malian or teleostean origin, including the recombinant tilapia preparations, increase the  $\text{Ca}^{2+}$ -ATPase activity and stimulate the  $\text{Ca}^{2+}$ -influx of the gills (Flik *et al.* 1986, 1989a). It takes several days for these effects to develop, and this contrasts sharply with the immediate responses to PTH in terrestrial animals. It is therefore unlikely that in teleosts prolactin accounts for the rapid restoration of experimentally induced hypocalcemia such as shown in fig. 1, or for the minute-to-minute control of the calcium concentration of the ECF in general. The primary function of prolactin in fish most likely is the control of integumental permeability to water and ions (Hirano 1986; Wendelaar Bonga and Pang 1989). Hypercalcemic actions have been ascribed to some other pituitary hormones of teleosts, but these have not been confirmed (see Wendelaar Bonga and Pang 1989). In trout cortisol was shown to produce hypercalcemia by increasing total body calcium uptake (Flik and Perry 1989). This hormone had no effect on tilapia (our unpublished observations) and it is unlikely to be a hormone with a specific hypercalcemic function.

#### b. Calcitonin

Calcitonin has been demonstrated in all major groups of fishes with exception of the agnathans. Its function in fish is still in dispute. Administration of mammalian or teleost calcitonin has only occasionally resulted in hypocalcemia. The doses required were high. In many experiments the hormone had no effect on plasma calcium (see Wendelaar Bonga and Pang 1991). An antihypercalcemic action of the hormone was demonstrated by Fenwick and Lam (1988) in mudskippers that were made hypercalcemic by taking them out of the water. On the other hand, removal of the Stannius bodies leads to a marked hypercalcemia, due to elevated total and ionic calcium levels (Hanssen *et al.* 1989) and develop in the presence of the ultimobranchial bodies. These bodies even show signs of hypersecretion of calcitonin in stanniectomized fish (Wendelaar Bonga and Greven 1978). Thus, the antihypercalcemic potency of the hormone is limited, in particular when compared with stanniocalcin, the hormone of the Stannius bodies present in teleostean and holostean fish. Enhanced plasma calcitonin levels

were reported for salmonids during gonadal maturation (Björnsson *et al.* 1989), and enlarged ultimobranchial glands were reported for several female fish during the reproductive period. This is consistent with the elevated plasma calcitonin levels that occur during the female reproductive period of some terrestrial vertebrates, and that have led to the hypothesis that calcitonin protects the skeleton during reproduction (see under A). Calcitonin may have a similar function in fish.

### c. Stanniocalcin

Stanniocalcin is a glycoprotein that has recently been isolated from the Stannius bodies of some teleost species. It is most likely identical with the proteinaceous hypocalcemic factor called hypocalcin by Pang *et al.* (1974), and teleocalcin, purified and partially sequenced by Wagner *et al.* (1986). Stanniocalcin is different from the small glycoprotein isolated from salmon that was originally called teleocalcin (Ma and Copp, 1978), and also different from the glycoprotein called parathyrin of the corpuscles of Stannius isolated from European eels by Milet *et al.* (1980). With the identification (Wagner *et al.* 1986; Lafeber *et al.* 1988) and sequencing (Butkus *et al.* 1987) of stanniocalcin, the confusion about the nature of the major calcium-regulating factor of the Stannius bodies has been solved, and stanniocalcin has now been established as a hormone with unique structural and functional characteristics. It is a dimeric glycoprotein with monomeric constituents with a molecular radius of about 30 kDa (Wagner *et al.* 1986; Flik *et al.* 1989b). The 30 kDa form is dominating in the circulation (Wagner *et al.* 1991).

Injection of stanniocalcin effectively reduces the hypercalcemia that occurs after operative removal of the Stannius bodies. It acts by reducing the branchial influx of calcium, and its effects become noticeable within 15 min after injection (Lafeber *et al.* 1988b). Because it leaves the passive efflux of calcium unaffected, the result is a rapid decrease of the calcium concentration of the blood plasma (Wagner *et al.* 1986; Lafeber *et al.* 1988b). Verbost *et al.* (1989) have shown that stanniocalcin inhibits  $^{45}\text{Ca}^{2+}$  uptake from the water into the branchial

epithelial cells, and this indicates that it acts by inhibiting the entrance of  $\text{Ca}^{2+}$  across the apical membranes of the chloride cells. The authors therefore suggested that stanniocalcin is a specific blocker of voltage-independent  $\text{Ca}^{2+}$ -channels in these membranes. The hormone did not influence the  $\text{Ca}^{2+}$ -extruding mechanisms in the basolateral membranes of the chloride cells (Flik *et al.*, unpublished). Because stanniocalcin is unlikely to influence the  $\text{Ca}^{2+}$ -channels of the apical membranes directly, we have postulated that the receptors for stanniocalcin are located in the basolateral membrane, and that the signal is mediated to the channels by second messenger. The hormone indeed modifies the second messenger systems in these cells: it reduces the branchial cAMP content and inhibits adenylate cyclase activity and stimulates the production of 1,2-sn-diacylglycerol in tilapia branchial cells (Flik *et al.* 1992). The latter effect indicates that stanniocalcin acts via the phosphoinositol pathway.

Our present concept of the endocrine control of calcium influx in teleost fish is illustrated in Fig. 2. As discussed above (see under B 2)  $\text{Ca}^{2+}$  can enter the fish - at least its chloride cells - passively, as long as the  $\text{Ca}^{2+}$ -channels are open and the electrochemical gradient is favourable. The latter is likely to be the case in most fresh waters, even with very low  $\text{Ca}^{2+}$  levels. This means that under normal freshwater conditions teleost fish may face hypercalcemia rather than hypocalcemia, even when the water  $\text{Ca}^{2+}$ -concentration is well below that of the ECF, unless the  $\text{Ca}^{2+}$  channels are closed by stanniocalcin. This is the essence of our model. It explains why the  $\text{Ca}^{2+}$  concentration of the blood plasma is rising to extremely high levels when the Stannius bodies are removed. This effect has been consistently observed in freshwater and seawater species and is caused by increased  $\text{Ca}^{2+}$  influx (So and Fenwick 1979). The total and ionic  $\text{Ca}^{2+}$  levels produced as a result of the absence of stanniocalcin are much higher than those that can be obtained by injection of hormones with hypercalcemic actions such as prolactin or cortisol. Apparently, in freshwater as well as seawater, the total body uptake of calcium and the control of the  $\text{Ca}^{2+}$  concentration of the ECF are under inhibitory control by stanniocalcin.

In conclusion then, the endocrine control of the calcium concentration of the ECF is most likely dominated by one hormone only in terrestrial vertebrates and in teleost fish. This contrasts with the earlier concept of two antagonistically acting hormones. In the terrestrial vertebrates it is PTH, which increases the  $\text{Ca}^{2+}$ -concentration of the ECF, mainly by stimulating the intestinal uptake of calcium and by mobilizing calcium from the skeleton. In teleost fish it is stanniocalcin, which lowers the  $\text{Ca}^{2+}$ -concentration of the ECF by inhibiting the passive influx of calcium from the ambient water into the branchial chloride cells and, possibly, the epithelial cells of the intestine.

## REFERENCES

- Brown EM, Leboff MS, Oetting M, Posillico JF, Chen C (1987) Secretory control in normal and abnormal parathyroid tissue. *Recent Prog Horm Res* 43: 337-382
- Björnsson BT, Haux C, Bern HA, Deftos LJ (1989) 17-Beta-estradiol increases plasma calcitonin levels in salmonid fish. *Endocrinology (Baltimore)* 125: 1754-1760
- Butkus A, Roche PJ, Fernley RT, Haralambidis J, Penschow JD, Ryan JB, Trahair JF, Tregéar GW, Coghlan JP (1987) Purification and cloning of a corpuscle of Stannius protein from *Anguilla australis*. *Mol Cell Endocrinol* 54: 123-133
- Copp DH, Kline LW (1989) Calcitonin. In: Pang PKT, Schreibman MP (eds), *Vertebrate endocrinology: fundamentals and biomedical applications*, Vol 3. Academic Press, San Diego, pp 79-103
- Copp DH, Ma SWY, Qiu MC, Sim M, Szeto K (1985) Role of the pituitary and the corpuscles of Stannius in calcium regulation in teleosts. In: Lofts B, Holmes WN (eds), *Current trends in comparative endocrinology*. Hong Kong University Press, Hong Kong, pp 819-821
- Fenwick JC, Lam TJ (1988) Effects of calcitonin on plasma calcium and phosphate in the mudskipper, *Periophthalmodon schlosseri* (Teleostei), in water and during exposure to air. *Gen Comp Endocrinol* 70: 224-230
- Fenwick JC, Brasseur JG (1991) Effects of stanniectomy and experimental hypercalcemia on plasma calcium levels and calcium influx in American eels, *Anguilla rostrata*, Lesueur. *Gen Comp Endocrinol* 82: 459-465
- Fleming WR, Brehe JE, Hanson R (1973) Some complicating factors in the study of the Ca metabolism of teleosts. *Amer Zool* 13: 793-797
- Flik G, Perry SF (1989) Cortisol stimulates whole body calcium uptake and the branchial calcium pump in freshwater rainbow trout. *J Endocrinol* 120: 75-82
- Flik G, Fenwick JC, Kolar ZI, Mayer-Gostan N, Wendelaar Bonga SE (1986) Effects of ovine prolactin on calcium uptake and distribution in *Oreochromis mossambicus* *Am J Physiol* 250: R161-

R166

- Flik G, Fenwick JC, Wendelaar Bonga SE (1989a) Calcitropic actions of prolactin in freshwater North American eel (*Anguilla rostrata* LeSueur). *Am J Physiol* 257: R74-R79
- Flik G, Labedz T, Lafeber FPJG, Wendelaar Bonga SE, Pang PKT (1989b) Studies on teleost corpuscles of Stannius: physiological and biochemical aspects of synthesis and release of hypocalcin in trout, goldfish and eel. *Fish Physiol Biochem* 7: 343-349
- Flik G, van der Velden JA, Dechering KJ, Verbost PM, Schoenmakers TJM, Kolar ZI, Wendelaar Bonga SE (1992)  $Ca^{2+}$  and  $Mg^{2+}$  transport in gills and gut of tilapia, *Oreochromis mossambicus*: a review. *J Exp Zool* (in press)
- Hanssen RGJM, Lafeber FPJG, Flik G, Wendelaar Bonga SE (1989) Ionic and total calcium levels in the blood of the European eel (*Anguilla anguilla*): effects of stanniectomy and hypocalcin replacement therapy. *J Exp Biol* 141: 177-186
- Hirano T (1986) The spectrum of prolactin actions in teleosts. In: Ralph CL (ed), *Comparative endocrinology: developments and directions*. AR Liss, New York, pp 53-74
- Ichii T, Mugiya Y (1983) Effects of dietary deficiency in calcium on growth and calcium uptake from the aquatic environment in the goldfish, *Carassius auratus*. *Comp Biochem Physiol* 74A: 259-262
- Jones JI, Fitzpatrick LA (1990) Binding of [ $^{125}I$ ]iodipine to parathyroid cell membranes: evidence of a dihydropyridine-sensitive calcium channel. *Endocrinology* (Baltimore) 126: 2015-2020
- Lafeber FPJG, Flik G, Wendelaar Bonga SE, Perry SF (1988a) Hypocalcin from Stannius corpuscles inhibits gill calcium uptake in trout. *Am J Physiol* 254: R891-R896
- Lafeber FPJG, Hanssen RGJM, Choy YM, Flik G, Herrmann-Erlee MPM, Pang PKT, Wendelaar Bonga SE (1988b) Identification of hypocalcin (teleocalcin) isolated from trout Stannius corpuscles. *Gen Comp Endocrinol* 69: 19-30
- Ma WY, Copp DH (1978) Purification, properties and action of a glycopeptide from the corpuscles of Stannius which affect calcium metabolism in the teleost. In: Gaillard PH, Boer HH (eds), *Comparative endocrinology*. Elsevier Biomedical Press, Amsterdam, pp 283-286
- Milet C, Hillyard CJ, Martelly E, Girgis S, MacIntyre L, Lopez E (1980) Similitudes structurales entre l'hormone hypocalcémiant des corpuscules de Stannius de l'anguille et l'hormone parathyroïdienne mammalienne. *C R Acad Sci (Paris)* D291: 977-980
- Mugiya y (1982) Calcium metabolism in estrogenized fish. in: Oguro C, Pang PKT (eds), *Comparative endocrinology of calcium regulation*. Japan Scientific Societies Press, Tokyo, pp 109-114
- Pang PKT, Pang RK, Sawyer WH (1974) Environmental calcium and the sensitivity of killifish (*Fundulus heteroclitus*) in bioassays for the hypocalcemic response to Stannius corpuscles from killifish and cod (*Gadus morhua*) *Endocrinology* (Baltimore) 94: 548-555
- Pang PKT, Schreibman MP, Balbontin F, Pang RK (1978) Prolactin

- and pituitary control of calcium regulation in the killifish, *Fundulus heteroclitus*. Gen Comp Endocrinol: 306-316
- Pang PKT, Kenny AD, Oguro C (1980) The evolution of the endocrine control of calcium regulation. In: Pang PKT, Eppler A (eds), Evolution of vertebrate endocrine systems. Texas Tech University Press, Lubbock, pp 323-356
- So YP, Fenwick JC (1977) Relationship between net  $Ca^{2+}$  influx across a perfused eel gill and the development of poststanniectomy hypercalcemia. J Exp Zool 200: 259-264
- Stevenson JC, Hillyard CJ, MacIntyre I, Cooper H, Whitehead MI (1979) A physiological role for calcitonin: protection of the maternal skeleton. Lancet 2: 769-770
- Swennen D, Rentier-Delrue F, Auperin B, Prunet P, Flik G, Wendelaar Bonga SE, Lion M, Martial JA (1991) Production and purification of biologically active recombinant tilapia (*Oreochromis niloticus*) prolactins. J Endocrinol 131: 219-227
- Urasa FM, Wendelaar Bonga SE (1987) Effects of calcium and phosphate on the corpuscles of Stannius of the teleost fish *Oreochromis mossambicus*. Cell Tiss Res 249: 681-690
- VanderWiel CJ, Talmage RV (1981) Ultrastructural and physiological evidence for calcitonin-induced postprandial calcium storage in bones of rats. Calcif Tiss Int 33: 417-424
- Verboost PM, van Rooij J, Flik G, Lock RAC, Wendelaar Bonga SE (1989) The movement of cadmium through freshwater trout branchial epithelium and its interference with calcium transport. J Exp Biol 145: 185-197
- Wagner GF, Hampong M, Park CM, Copp DH (1986) Purification, characterisation, and bioassay of teleocalcin, glycoprotein from salmon corpuscles of Stannius. Gen Comp Endocrinol 63: 481-491
- Wagner GF, Milliken C, Friesen HG, Copp DH (1991) Studies on the regulation and characterization of plasma stanniocalcin in rainbow trout. Mol Cell Endocrinol 79: 129-138
- Weiss RE, Watabe N (1978) Studies on the biology of fish bone - I. Bone resorption after scale removal. Comp Biochem Physiol 60A: 207-211
- Wendelaar Bonga SE, Greven JAA (1978) The relationship between prolactin cell activity, environmental calcium and plasma calcium in the teleost *Gasterosteus aculeatus*. Observations on stanniectomized fish. Gen Comp Endocrinol 36: 90-101
- Wendelaar Bonga SE, Dederen LHT (1986) Effects of acidified water on fish. Endeavour 10: 198-201
- Wendelaar Bonga SE, Pang PKT (1989) Pituitary hormones. In: Pang PKT, Schreibman MP (eds), Vertebrate endocrinology: fundamental and biomedical implications, vol. 1. Academic Press, San Diego, pp 105-137
- Wendelaar Bonga SE, Pang PKT (1991) Control of calcium regulating hormones in the vertebrates: parathyroid hormone, calcitonin, prolactin, and stanniocalcin. Int Rev Cytol 128: 139-213