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# Brain serotonergic activation in growth-stunted farmed salmon: adaption versus pathology

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Signalling systems activated under stress are highly conserved, suggesting adaptive effects of their function. Pathologies arising from continued activation of such systems may represent a mismatch between evolutionary programming and current environments. Here, we use Atlantic salmon (*Salmo salar*) in aquaculture as a model to explore this stance of evolutionary-based medicine, for which empirical evidence has been lacking. Growth-stunted (GS) farmed fish were characterized by elevated brain serotonergic activation, increased cortisol production and behavioural inhibition. We make the novel observation that the serotonergic system in GS fish is unresponsive to additional stressors, yet a cortisol response is maintained. The inability of the serotonergic system to respond to additional stress, while a cortisol response is present, probably leads to both imbalance in energy metabolism and attenuated neural plasticity. Hence, we propose that serotonin-mediated behavioural inhibition may have evolved in vertebrates to minimize stress exposure in vulnerable individuals.

# 1. Background

The neurobiology of farmed fish is subject to expanding interest due to increasing concern for the welfare of production animals [1], and fishes are also progressively replacing mammalian models in biomedical research [2]. Neural signalling systems activated under stress are highly conserved [3–6], suggesting adaptive effects of their function. It is therefore a conundrum for evolutionary-based medicine that pathologies often arise from continued activation of the same systems [7]. One possibility is that pathologies may have evolved from behavioural responses that were adaptive in more primordial environments [8–11]. It is assumed that a mismatch between the historic and current environment leads to normally adaptive responses overriding self-correcting tendencies of emotional mechanisms, and this leads to pathologies [12]. Empirical investigation into this stance of evolutionary-based medicine is, however, scarce.

Here, we exploit a model in which such mismatch is prone to occur, namely, Atlantic salmon (*Salmo salar*) undergoing rapid domestication [13]. Under aquaculture conditions, it is common to find a proportion of farmed salmon, which are easily catchable at the surface and exhibit a small size, anorexia and a behaviourally inhibited profile. Such moribund fish, known as ‘drop outs’ or ‘loser fish’, are well known to farmers but very little is known about the aetiology leading up to this phenomenon [14,15]. We investigated serotonin (5-hydroxytryptamine, 5-HT) activity in the brain stem, which contains important monoaminergic nuclei innervating large parts of the brain [4], and plasma cortisol levels of surface-dwelling growth-stunted (GS) fish as well as healthy individuals occurring in a commercial aquaculture salmon farm. In the vertebrate brain, 5-HT-mediated signalling has a crucial role in energy regulation, neural plasticity, behavioural and emotional control, as well as neuroendocrine responses to stress [16,17]. Sustained serotonergic activation is associated with chronic stress and stress-induced pathologies, such as depression-like states, in several animal species [4,16,18–20]. We here report the novel observation that the serotonergic system in GS fish is not responsive to additional acute stress, while a significant corticosteroid response is maintained.

## 2. Material and methods

### 2.1. Experimental animals and facilities

GS and healthy fish (commercial strain, Aquagen AS) were selected during two sampling trials at a commercial salmon farm in the Langenuen Straight, Western Norway (60°N). Fish were sampled from two sea cages (25 × 25 × 30 m depth; approx. 18 750 m<sup>3</sup>) containing spring smolts. The fish were reared according to production standards, under a natural light regime and were fed formulated food given over two equally sized meals from 09.00 to 12.00 and 13.00 to 16.00. We sampled fish two (sampling 1; S1) and five (sampling 2; S2) months after seawater transfer to control for repeatability and production phase differences. In this study, we have categorized these two groups as: (i) *healthy*, normal schooling individuals with healthy behaviour and feeding responses, displaying high reactivity to environmental stimuli; and (ii) *GS fish*, which tended to position themselves towards the edge of the sea cage, close to the surface and without much reaction to environmental stimuli, including food pellets. Representative phenotypes are shown in figure 1, and number of individuals collected and their average length and weight are presented in table 1.

### 2.2. Acute stress treatment

All fish were captured by quickly netting them out of the cage with a dip net from 2 m depth close to the net wall. Healthy fish were lured towards the net wall by throwing out feed-pellets and were caught while feeding. Meanwhile, GS fish remained close to the net wall the whole time and were simply netted out of the cage. Fish from both groups were either sampled directly after capture or subjected to a 30 min acute stress test before sampling (see table 1 for *n*/group). Acute stress consisted of individual confinement in a circular 121 plastic bucket containing seawater from a depth of 1 m (at approx. 12°C) for a period of 30 min after Pottinger & Carrick [21]. Oxygen levels were constantly monitored with an oxygen probe (Storvik HQ40, Storvik Aqua AS, Sunndalsøra, Norway) and kept above 6 mg l<sup>-1</sup> to ensure that fish were not exposed to hypoxia [22].



**Figure 1.** Representative pictures of healthy (a) and GS (b) fish from an aquaculture farm in the Langenuen Strait, Western Norway. (Photos: Ole Folkedal.)

**Table 1.** Average length and weight ( $\pm$ s.d.) and number ( $n$ ) of healthy and GS Atlantic salmon collected during first (S1) and second (S2) samplings at basal and acute stress conditions.

	unstressed		stressed	
	healthy	GS	healthy	GS
S1				
length (cm)	24.7 $\pm$ 0.6	18.9 $\pm$ 0.6	25.1 $\pm$ 0.7	17.8 $\pm$ 0.8
weight (g)	176 $\pm$ 13.1	50.3 $\pm$ 7.1	179 $\pm$ 16	42.4 $\pm$ 7.4
$n$	15	14	14	12
S2				
length (cm)	38.2 $\pm$ 1.2	25.4 $\pm$ 0.8	38.2 $\pm$ 1.1	26.1 $\pm$ 0.9
weight (g)	721 $\pm$ 81.4	145 $\pm$ 16.8	716 $\pm$ 54.9	162 $\pm$ 18.3
$n$	10	10	10	10

### 2.3. Sampling protocol

Fish were euthanized with a strong overdose of 1 g l<sup>-1</sup> MS-222 (Finquel<sup>®</sup>, Argent Chemical Laboratories, Redmond, WA, USA), which rendered them completely motionless (no opercular movement) within 10 s of immersion. Fish were rapidly weighed, fork length measured and a blood sample was taken from the caudal vessels with 23 G, 1 ml syringes containing the anticoagulant ethylene diamine tetra acetic acid (EDTA). Following centrifugation for 5 min at 9.289 rcf and 4°C, plasma samples were frozen and stored at -80°C for later analysis. Fish were decapitated and the brain stem was quickly excised within 2 min. Brain stems were wrapped in individually marked aluminium foil packets, snap-frozen in liquid nitrogen and stored at -80°C for later analysis of 5-HT neurochemistry.

### 2.4. Brain stem serotonergic neurochemistry

Frozen brain stems were homogenized in 4% (w/v) ice cold perchloric acid (PCA) containing 0.2% EDTA and 3,4-dihydroxybenzyl amine hydrobromide (DHBA, 40 ng ml<sup>-1</sup>) as an internal standard using a Potter-Elvehjem homogenizer. After spinning samples for 10 min at 15.493 rcf and 4°C, the supernatant was analysed by means of high-performance liquid chromatography (HPLC). The mobile phase was made up of 12  $\mu$ M EDTA, 86 mM sodium phosphate and 1.4 mM sodium octyl sulfate in deionized water (resistance 18.2 MW), containing 7% acetonitrile set to pH 3.1 using phosphoric acid. The system contains a solvent delivery system (Shimadzu, LC-10AD), an auto-injector (Famos, Spark), a reverse-phase column (4.6 mm 100 mm, Hichrom, C18, 3.5 mm) and an ESA Coulochem II detector (ESA, Bedford, MA, USA) with two electrodes at -40 mV and +320 mV. A conditioning electrode with a potential

of +40 mV was used to oxidize possible contaminants before analysis. Brain stem concentrations of 5-HT and the 5-HT catabolite 5-hydroxyindoleacetic acid (5-HIAA) were quantified by comparison with standards and corrected for recovery of the internal standard using HPLC software (CSW, Data Apex Ltd, the Czech Republic).

## 2.5. Cortisol radioimmunoassay

Plasma samples were diluted 2.2 times in assay buffer and cortisol assayed by radioimmunoassay (RIA) following the procedure described by Gorissen *et al.* [23]. Intra- and interassay variations were 12.5% and 3.5%, respectively, and cross-reactivity of the cortisol antibody (antibody [xm210]; Abcam, Cambridge, MA, USA) was as follows: cortisol 100%, 11-deoxycortisol 0.9%, prednisolone 5.6%, corticosterone 0.6%, 11-deoxycorticosterone, progesterone, 17-hydroxyprogesterone, testosterone, oestradiol and oestriol all less than 0.01%.

## 2.6. Statistical analyses

All datasets were tested by Levene's test for variance homogeneity, and values were either log- (cortisol, 5-HT and 5-HIAA) or arcsine-transformed (5-HIAA/5-HT ratios) when necessary. A two-way ANOVA was used to compare concentrations of 5-HT, 5-HIAA, cortisol and the 5-HIAA/5-HT ratios for each sampling, with fish type (GS versus healthy control) and treatment (basal versus acute stress) as independent variables, followed by a Tukey–Kramer honestly significant difference (HSD) post hoc test when a significant interaction effect was indicated.

# 3. Results

## 3.1. Brain stem 5-hydroxytryptamine neurochemistry

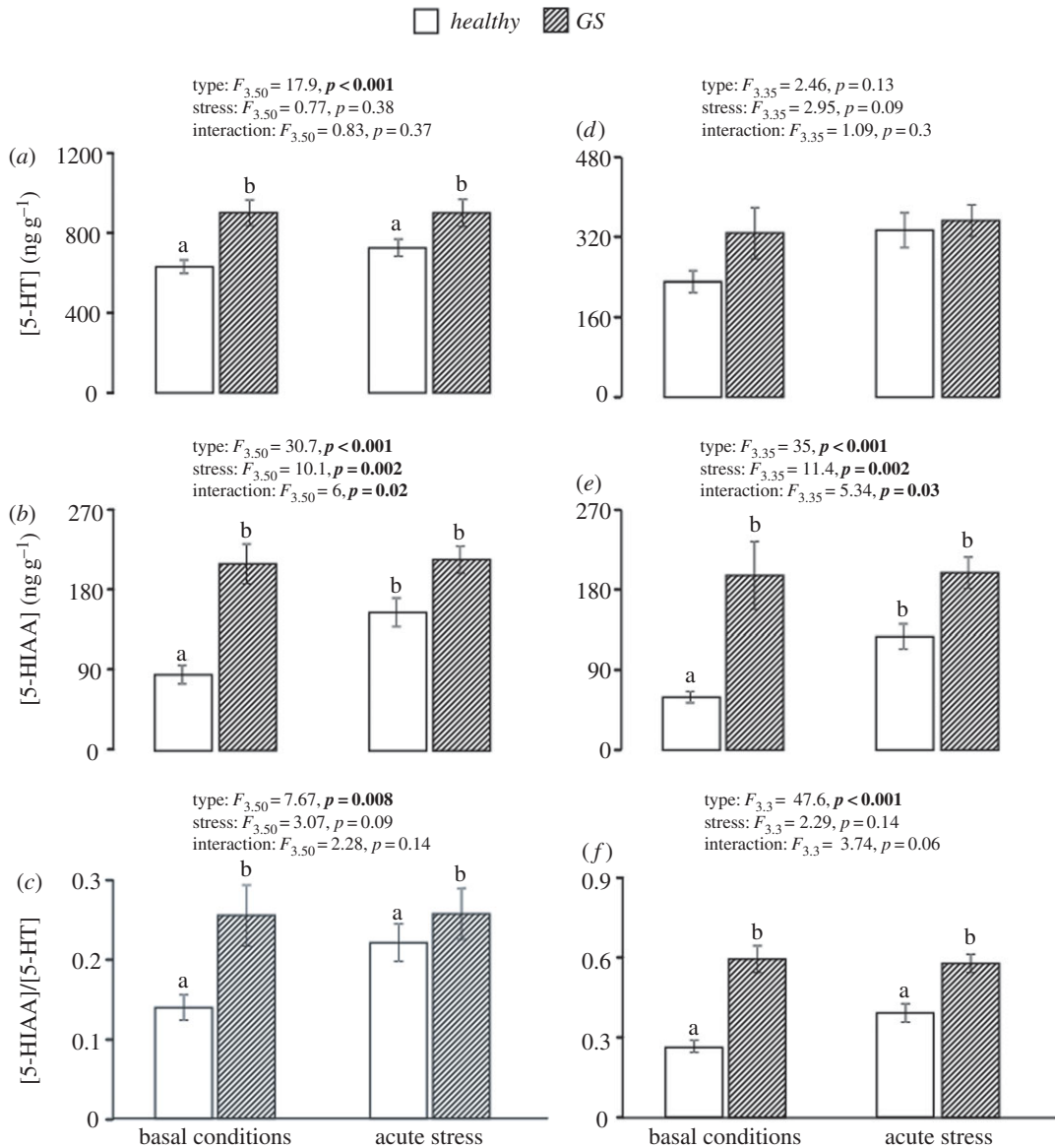
We investigated 5-HT neurochemistry at basal conditions and after acute stress in groups of healthy and GS salmon at two different time points (two and five months following transfer to seawater rearing, i.e. S1 and S2, respectively). At both time points, GS fish sustained significantly higher levels of the principal 5-HT catabolite 5-HIAA compared with healthy controls ( $p < 0.001$  in both samplings). Interestingly, whereas 5-HIAA levels were significantly increased by confinement stress in controls (post hoc  $p_{S1} = 0.001$ ,  $p_{S2} = 0.002$ ), 5-HIAA levels did not differ between basal and confinement stress ( $p_{S1} = 0.96$ ,  $p_{S2} = 0.87$ ) in the GS fish (figure 2*b,e*), indicating a blunted serotonergic response to stress in these individuals. Meanwhile, there was a significant effect of phenotype on the 5-HIAA/5-HT ratio ( $p_{S1} = 0.008$  and  $p_{S2} < 0.001$ ) in both samplings, where healthy fish displayed lower ratios than GS fish (figure 2*c,f*). For 5-HT concentrations, there was a discrepancy between the two samplings, whereas there was a fish-type effect during S1 in which healthy fish had significantly lower 5-HT levels compared with GS fish ( $p < 0.01$ ), this difference was not statistically significant in S2 ( $p = 0.13$ ). Nevertheless, we found that on both samplings there was neither a stress nor an interaction effect on 5-HT (figure 2*a,d*).

## 3.2. Plasma cortisol levels

At both time points of sampling, plasma cortisol was affected by both type ( $p_{S1} < 0.001$  and  $p_{S2} = 0.02$ ) and stress ( $p \leq 0.001$  in both samplings), with no interaction effect between the two variables ( $p = 0.1$  in both). Plasma cortisol levels were significantly lower in healthy fish compared with GS fish. However, both groups responded with increased plasma cortisol levels to confinement stress (figure 3).

# 4. Discussion

The occurrence of GS fish is a common and costly occurrence in salmon aquaculture, affecting up to a quarter of the stock, but little has been done to elucidate the aetiology of this phenotype. We here present, for the first time, physiological data for GS fish, showing that elevated serotonergic activity is a main characteristic of the GS phenotype. That is, compared with healthy fish, GS salmon are characterized by increased basal brain serotonergic activity and cortisol production measured over two samplings several months apart, which suggest chronic activation of the serotonergic and hypothalamic-pituitary interrenal (HPI) axis systems [6,24,25]. Our findings support the view that research focusing on genetic markers

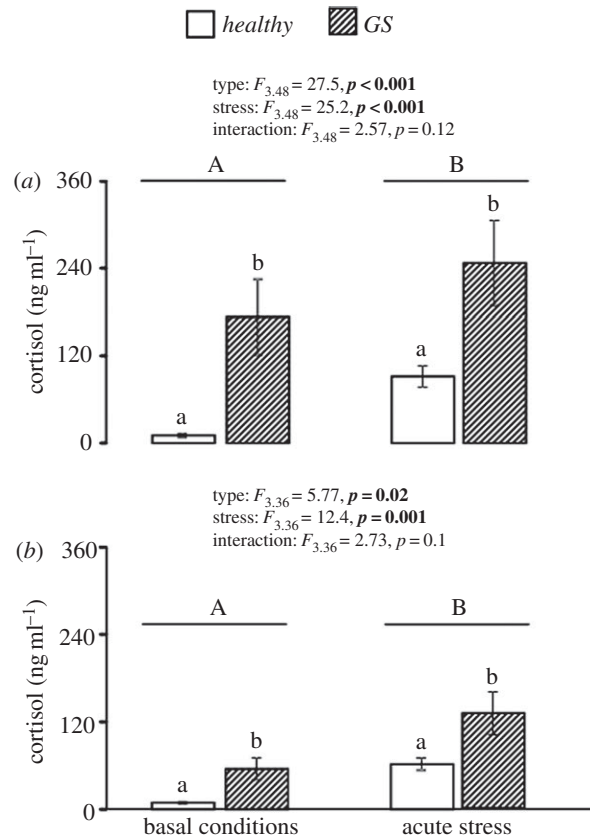


**Figure 2.** Effect of fish type (i.e. healthy versus GS) and stress (basal versus acute stress) on serotonin (5-HT) neurochemistry in the brain stem of Atlantic salmon at S1 (a–c) and S2 (d–f). Two-way ANOVA statistics are given in figure for each panel. Small letters indicate a fish-type effect or Tukey–Kramer HSD post hoc differences following a significant interaction effect. Data represent mean  $\pm$  s.e.m.

for stress coping abilities in farmed fish is necessary to improve welfare and selection for breeding. Intriguingly, a growing body of evidence indicates that several depression-like syndromes are associated with increased serotonergic signalling (for a review, see [17]). In this context, it is interesting to note that the behavioural and serotonergic profile exhibited by GS fish is reminiscent of a depressed state, similar to those described in mammals [17,26–28]. However, further research is needed in order to corroborate this possibility.

Both 5-HIAA levels and the 5-HIAA/5-HT ratios as well as plasma cortisol levels observed during both samplings in GS fish at basal conditions indicate chronic serotonergic and HPI axis activation [6,24,25]. Notably, there were two discrepancies between both samplings. First, measured plasma cortisol and 5-HT levels were overall higher in all groups during S1, compared with S2. Both cortisol and 5-HT concentrations are known to vary with the life stage of the fish and most notably cortisol and 5-HT increase during smoltification (the metamorphosis that prepares salmon for the saltwater environment). That is, an increase during smoltification in brain 5-HT and plasma cortisol concentrations of up to 50% [29] and up to 100% [30], respectively, has been reported. In our experiment, fish were sampled shortly after smoltification (S1) and five months later (S2). In line with these reports, both 5-HT and





**Figure 3.** Effect of fish type (i.e. healthy versus GS) and stress (basal versus acute stress) on plasma cortisol concentrations in Atlantic salmon, at S1 (a) and S2 (b). Two-way ANOVA statistics are given in figure for each panel. Small and big letters indicate a type and treatment effect, respectively. Data represent mean  $\pm$  s.e.m.

plasma cortisol levels were higher at the sampling point closest to smoltification. Second, while basal serotonergic activity was found to be consistently higher, at both sampling points, in GS fish compared with healthy individuals, 5-HT concentrations were similarly higher in GS fish compared with healthy fish in S1 only. However, 5-HT concentrations do not directly reflect serotonergic signalling. Firstly, 5-HT is rapidly replaced by newly synthesized 5-HT intracellularly following its release at the synapse [31] and secondly, extracellular 5-HT concentrations reflect both 5-HT release and clearance from the synapse [17]. Instead, 5-HIAA levels and the 5-HIAA/5-HT ratio are used as a proxy for serotonergic activity [6,17,31,32], as the formation of the 5-HT catabolite 5-HIAA is almost exclusively a result of monoamine oxidase metabolizing 5-HT after release into the synapse and reuptake into the presynaptic neuron or surrounding cells. Our most important finding is that the serotonergic system of GS fish is unresponsive to acute novel stress. This phenomenon has, to the best of our knowledge, never been shown before, but is a classic example of allostatic overload in a physiological system (i.e. the inability of regulatory mechanisms to react to further challenge). The inability of the serotonergic system to respond to novel acute stressors, while a cortisol response is maintained, probably leads to both an imbalance in energy metabolism [17] and attenuated neural plasticity [5,33]. Therefore, the passive-behavioural phenotype characteristic of GS fish may be mediated by sustained serotonergic activation in order to minimize further stress exposure.

Although we have established that GS fish suffer from an elevated activation of the serotonergic system, the underlying causes of the increased serotonergic activity remains to be determined. There are multiple potential contributors to increased serotonergic activity in farmed salmon. For example, subordinate social rank within aquaculture populations is associated with chronically increased serotonergic activity [34]. Furthermore, the strains of Atlantic salmon used in aquaculture have gone through a rapid and intense domestication, and the aquaculture environment challenges individuals with a series of stressors that do not occur in nature [13]. For example, vaccinating fish shortly after smoltification (a common practice in aquaculture [35,36]) challenges homeostatic balance, induces autoimmunity [37] and decreases feed intake for several weeks [36]. Interestingly, both vaccination [38]

and smoltification [29] alter serotonergic signalling and could lead to prolonged serotonergic activation. Thus, an environmental mismatch between natural (e.g. smoltification) and 'artificial' stressors (e.g. grading, vaccination) might leave some individuals more vulnerable to disease and behavioural syndromes [13]. Further investigations are, however, needed to pinpoint the proximate causes of the GS phenotype, e.g. the possible interaction between genetic effects, immune status and social hierarchies.

Future studies should focus on the environmental factors and genetic, behavioural and physiological characteristics underpinning GS fish, especially considering that individual differences in vulnerability are still unresolved [17,39]. We believe that these findings should encourage further exploration of both the evolutionary background and causative molecular mechanisms for 5-HT-induced behavioural inhibition. Such knowledge may be pivotal in advancing the development of new clinical approaches to stress-induced pathologies and may ultimately facilitate a shift in focus to the medical ideal of prevention rather than cure.

**Ethics.** This work was approved by the Norwegian Animal Research Authority (NARA), following the Norwegian Regulation on Animal Experimentation of 1996.

**Data accessibility.** All relevant data are within the paper and its electronic supplementary material.

**Authors' contributions.** M.A.V., O.F., I.B.J. and Ø.Ø. planned the research. M.A.V., O.F. and M.G. collected and analysed data. M.A.V., I.B.J., E.H., M.G., G.F., T.S.K. and Ø.Ø. wrote the paper. All authors gave the final approval for publication.

**Competing interests.** The authors declare no competing financial interests.

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## References

- Braithwaite VA, Ebbesson LOE. 2014 Pain and stress responses in farmed fish. *Rev. Sci. Tech.* **33**, 245–253. (doi:10.20506/rst.33.1.2285)
- Schartl M. 2014 Beyond the zebrafish: diverse fish species for modeling human disease. *Dis. Model Mech.* **7**, 181–192. (doi:10.1242/dmm.012245)
- Herculano AM, Maximino C. 2014 Serotonergic modulation of zebrafish behavior: towards a paradox. *Prog. Neuro-Psychopharmacol. Biol. Psych.* **55**, 50–66. (doi:10.1016/j.pnpbp.2014.03.008)
- Lillesaar C. 2011 The serotonergic system in fish. *J. Chem. Neuroanat.* **41**, 294–308. (doi:10.1016/j.jchemneu.2011.05.009)
- Sørensen C, Johansen IB, Øverli Ø. 2013 Neural plasticity and stress coping in teleost fishes. *Gen. Comp. Endocrinol.* **181**, 25–34. (doi:10.1016/j.ygcen.2012.12.003)
- Winberg S, Nilsson GE. 1993 Roles of brain monoamine neurotransmitters in agonistic behaviour and stress reactions, with particular reference to fish. *Comp. Biochem. Physiol.* **106C**, 597–614. (doi:10.1016/0742-8413(93)90216-8)
- Sapolsky RM. 2004 *Why zebras don't get ulcers*. New York, NY: Henry Holt and Company (Holt paperbacks).
- Henriques GR. 2000 Depression: disease or behavioral shutdown mechanism. *J. Sci. Health Policy* **1**, 152–165.
- Nesse RM. 2000 Is depression an adaptation? *Arch. Gen. Psych.* **57**, 14–20. (doi:10.1001/archpsyc.57.1.14)
- Price J, Sloman L, Gardner R, Gilbert P, Rohde P. 1994 The social competition hypothesis of depression. *Br. J. Psychol.* **164**, 309–315. (doi:10.1192/bjp.164.3.309)
- Raison CL, Miller AH. 2013 The evolutionary significance of depression in pathogen host defense (PATHOS-D). *Mol. Psychiatry* **18**, 15–37. (doi:10.1038/mp.2012.2)
- Nettle D. 2004 Evolutionary origins of depression: a review and reformulation. *J. Affect Disord.* **81**, 91–102. (doi:10.1016/j.jad.2003.08.009)
- Huntingford FA, Adams C. 2005 Behavioural syndromes in farmed fish: implications for production and welfare. *Behaviour* **142**, 1207–1221. (doi:10.1163/156853905774539382)
- Stephen C, Ribble CS. 1995 An evaluation of surface moribund salmon as indicators of seapen disease status. *Aquaculture* **133**, 1–8. (doi:10.1016/0044-8486(94)00400-1)
- Stien LH *et al.* 2013 Salmon welfare index model (SWIM 1.0): a semantic model for overall welfare assessment of caged Atlantic salmon: review of the selected welfare indicators and model presentation. *Rev. Aquacult.* **5**, 33–57. (doi:10.1111/j.1753-5131.2012.01083.x)
- Langfume L, Mongeau R, Cohen-Salmon C, Hamon M. 2008 Corticosteroid–serotonin interactions in the neurobiological mechanisms of stress-related disorders. *Neurosci. Biobehav. Rev.* **32**, 1174–1184. (doi:10.1016/j.neubiorev.2008.04.006)
- Andrews PW, Bharwani A, Lee KR, Fox M, Thomson Jr JA. 2015 Is serotonin an upper or a downer? The evolution of the serotonergic system and its role in depression and the antidepressant response. *Neurosci. Biobehav. Rev.* **51**, 164–188. (doi:10.1016/j.neubiorev.2015.01.018)
- Graeff FG, Guimarães FS, De Andrade TGCS, Deakin JFW. 1996 Role of 5-HT in stress, anxiety, and depression. *Pharmacol. Biochem. Behav.* **54**, 129–141. (doi:10.1016/0091-3057(95)02135-3)
- Anderson G, Maes M, Berk M. 2012 Biological underpinnings of the commonalities in depression, somatization, and chronic fatigue syndrome. *Med. Hypotheses* **78**, 752–756. (doi:10.1016/j.mehy.2012.02.023)
- Krishnan V, Nestler EJ. 2011 Animal models of depression: molecular perspectives. In *Molecular and functional models in neuropsychiatry* (ed. JJ Hagan), pp. 121–147. Berlin, Germany: Springer.
- Pottinger TG, Carrick TR. 1999 Modification of the plasma cortisol response to stress in rainbow trout by selective breeding. *Gen. Comp. Endocrinol.* **116**, 122–132. (doi:10.1006/gcen.1999.7355)
- Jobling M. 1994 *Fish bioenergetics*. London, UK: Chapman & Hall.
- Gorissen M, Bernier NJ, Manuel R, de Gelder S, Metz JR, Huising MO, Flik G. 2012 Recombinant human leptin attenuates stress axis activity in common carp (*Cyprinus carpio* L.). *Gen. Comp. Endocrinol.* **178**, 75–81. (doi:10.1016/j.ygcen.2012.04.004)
- Sørensen C, Johansen IB, Øverli Ø. 2013 Physiology of social stress in fishes. In *The physiology of fishes* (eds DH Evans, JB Clairborne, S Currie), pp. 289. Boca Raton, FL: Taylor and Francis Group.
- Wendelaar Bonga SE. 1997 The stress response in fish. *Physiol. Rev.* **77**, 591–625.
- Blanchard RJ, McKittick CR, Blanchard DC. 2001 Animal models of social stress: effects on behavior and brain neurochemical systems. *Physiol. Behav.* **73**, 261–271. (doi:10.1016/S0031-9384(01)00449-8)
- Sapolsky RM. 2011 Physiological and pathophysiological implications of social stress in mammals. In *Comprehensive physiology* (ed. DM Pollock), pp. 517–532. New York, NY: John Wiley & Sons, Inc.
- Shively CA, Willard SL. 2012 Behavioral and neurobiological characteristics of social stress versus depression in nonhuman primates. *Exp. Neurol.* **233**, 87–94. (doi:10.1016/j.expneurol.2011.09.026)
- Ebbesson SOE, Smith J, Co C, Ebbesson LOE. 1996 Transient alterations in neurotransmitter levels



- during a critical period of neural development in coho salmon (*Oncorhynchus kisutch*). *Brain Res.* **742**, 339–342. (doi:10.1016/S0006-8993(96)00993-6)
30. Barton BA, Schreck CB, Ewing RD, Hemmingsen AR, Patiño R. 1985 Changes in plasma cortisol during stress and smoltification in coho salmon, *Oncorhynchus kisutch*. *Gen. Comp. Endocrinol.* **59**, 468–471. (doi:10.1016/0016-6480(85)90406-X)
  31. Shannon NJ, Gunnet JW, Moore KE. 1986 A comparison of biochemical indices of 5-hydroxytryptaminergic neuronal activity following electrical stimulation of the dorsal raphe nucleus. *J. Neurochem.* **47**, 958–965. (doi:10.1111/j.1471-4159.1986.tb00704.x)
  32. Fillenz M. 1993 Neurochemistry of stress: introduction to techniques. In *Stress: from synapse to syndrome* (eds SC Stanford, P Salmon), pp. 247–279. San Diego, CA: Academic Press.
  33. Anacker C *et al.* 2013 Glucocorticoid-related molecular signaling pathways regulating hippocampal neurogenesis. *Neuropsychopharmacology* **38**, 872–883. (doi:10.1038/npp.2012.253)
  34. Cubitt KF, Winberg S, Huntingford FA, Kadri S, Crampton VO, Øverli Ø. 2008 Social hierarchies, growth and brain serotonin metabolism in Atlantic salmon (*Salmo salar*) kept under commercial rearing conditions. *Physiol. Behav.* **94**, 529–535. (doi:10.1016/j.physbeh.2008.03.009)
  35. Huntingford FA, Adams C, Braithwaite VA, Kadri S, Pottinger TG, Sandøe P, Turnbull JF. 2006 Current issues in fish welfare. *J. Fish Biol.* **68**, 332–372. (doi:10.1111/j.0022-1112.2006.001046.x)
  36. Sørum U, Damsgård B. 2004 Effects of anaesthetisation and vaccination on feed intake and growth in Atlantic salmon (*Salmo salar* L.). *Aquaculture* **232**, 333–341. (doi:10.1016/S0044-8486(03)00529-5)
  37. Koppang EO *et al.* 2008 Vaccination-induced systemic autoimmunity in farmed Atlantic salmon. *J. Immunol.* **181**, 4807–4814. (doi:10.4049/jimmunol.181.7.4807)
  38. Baganz NL, Blakely RD. 2012 A dialogue between the immune system and brain, spoken in the language of serotonin. *ACS Chem. Neurosci.* **4**, 48–63. (doi:10.1021/cn300186b)
  39. Heim C, Binder EB. 2012 Current research trends in early life stress and depression: review of human studies on sensitive periods, gene–environment interactions, and epigenetics. *Exp. Neurol.* **233**, 102–111. (doi:10.1016/j.expneurol.2011.10.032)