In this communication, we use magnetic birefringence to probe morphological changes in polymersomes during dialysis, while taking samples at regular intervals followed by the \textit{ex situ} imaging of their conformations using electron microscopy. Here, we demonstrate how the morphology of polymersomes during dialysis can be probed in a continuous and non-invasive way using \textit{in situ} magnetic birefringence. These measurements clearly reveal the exact times at which the morphology of the polymersomes change, making it possible to take samples for electron microscopy at the crucial points of dialysis.

Magnetic birefringence can be observed upon the alignment of molecules or aggregates. Even seemingly non-magnetic matter (like the vast majority of polymers and biomolecules) is in fact weakly magnetic (diamagnetic). Molecules used for self-assembly are usually anisotropic in shape, leading to a magnetic response that is also anisotropic. These molecules therefore have a preferential orientation in a magnetic field. The difference in energy between two orthogonal orientations of a molecule is normally quite small and hence the alignment is largely randomized by thermal motion. However, when these molecules form aggregates or self-assemble into supramolecular structures, the total diamagnetic anisotropy can be enhanced significantly. This principle has been used to orient organic nanostructures composed of a wide variety of molecules, including polymers. The diamagnetic anisotropy, and hence the magnetic alignment, of these self-assembled structures is also related to their overall shape. For instance, a sphere with an isotropic orientational distribution of molecules has no preferential axis of alignment. However, when the distribution of molecular orientations is anisotropic magnetic alignment can occur. For example, a disk constructed from diamagnetic anisotropic molecules aligns with its surface either parallel or perpendicular to the magnetic field, depending on the sign of the diamagnetic anisotropy of the molecular building blocks.

Magnetic orientation of supramolecular aggregates in solution results in a difference in the refractive index for light polarized parallel and perpendicular with respect to the magnetic field. This magnetic birefringence has been frequently measured to determine the degree of alignment of various aggregates in magnetic fields. In this communication, we use magnetic birefringence to probe morphological changes in polymersomes during dialysis, while...
Further insight into the mechanism of osmotically induced shape change is presented. To explore the morphological changes we implemented a dialysis cell with a magnetic birefringence setup in a 2 T magnet (Fig. 1). Light of a HeNe laser (1.5 mW, 632.8 nm) was focussed on the upper chamber of a flow cell, which contained a PEG44–PS133 polymersome sample. Initially the sample consisted of spherical polymersomes in pure water, having a radius of 252 nm and a PDI of 0.134, as determined by dynamic light scattering (DLS). A dialysis fluid consisting of 50% water, 40% THF and 10% 1,4-dioxane was pumped through the bottom chamber at a rate of 100 mL h⁻¹, as indicated by the red dots in Fig. 2I (ESI†). At all points, the magnitude of the birefringence reflects the competition with thermal motion tends to randomize the orientation of the molecules resulting in partial alignment with the field.

The birefringence was measured over a time interval of 360 minutes (blue curve in Fig. 2I). At the beginning of the experiment, the birefringence remained zero up to 170 minutes (point c), after which a rapid increase in the signal was observed. A maximum was reached at 230 minutes (point e), after which the birefringence decreased. From point g onwards, the sample showed a small birefringence, which remained constant until the end of the measurement (point h). At certain time points, magnetic field sweeps were performed where the magnetic field was reduced to zero and subsequently brought back to 2 T (indicated in Fig. 2I by the coloured lines 1–5). The field sweeps were sufficiently rapid (about one minute) to allow the subsequent birefringence signal to remain unchanged by the continuous dialysis. The birefringence always decreased to zero when the magnetic field was brought to zero. Restoring the field to 2 T also resulted in recovery of the birefringence to the same value before the field sweep, showing that the measured constant magnetic birefringence. At point d the birefringence increased to half its maximal value. The cryo-SEM images show ellipsoidal polymersomes, which can only be explained by a partial deflation of the spherical polymersomes. When the birefringence reached a maximum (point e), flat disks were observed under a cryo-SEM. Continuation of the dialysis led to the bending of the flat disks (point f) and the formation of stomatocytes (points g and h), where the structures partly inflated again. At this point, the hydrodynamic radius of the polymersomes was decreased to 218 nm with a PDI of 0.12, as determined by DLS (see ESI†).

Measuring magnetic birefringence during dialysis offers the opportunity to stop dialysis at well-defined points to take samples for further investigation by electron microscopy. In this manner, the morphology can be related to the amplitude of the birefringence signal. Samples were taken at points a–h as indicated by the red dots in Fig. 2I (ESI†).

Cryogenic-Scanning Electron Microscopy (cryo-SEM) images of samples (a–h) are shown in Fig. 2II. At the beginning of dialysis, polymersomes were present with the expected spherical morphology (point a) corresponding to zero birefringence. At points b and c, no changes in conformation were observed, in agreement with the measured constant magnetic birefringence. At point d the birefringence increased to half its maximal value. The cryo-SEM images show ellipsoidal polymersomes, which can only be explained by a partial deflation of the spherical polymersomes. When the birefringence reached a maximum (point e), flat disks were observed under a cryo-SEM. Continuation of the dialysis led to the bending of the flat disks (point f) and the formation of stomatocytes (points g and h), where the structures partly inflated again. At this point, the hydrodynamic radius of the polymersomes was decreased to 218 nm with a PDI of 0.12, as determined by DLS (see ESI†).

At all points, the magnitude of the birefringence reflects the shape of the structures. All spherical polymersomes show zero
In conclusion, we have demonstrated that magnetic birefringence can be used as a useful tool to monitor morphological changes in polymersomes resulting from dialysis in a flow cell. This method has the advantage of being non-invasive; while the morphology can be determined without disrupting the dialysis setup for sample investigation by electron microscopy. Also, because the dialysis in the flow cell is rather slow (in the order of several hours), the shape transformations can be determined at very precise points on the birefringence curve, providing samples with very specific and predictable conformations. In principle, this method could be extended to obtain more quantitative information about the rigidity of the structures in all stages of the process. However this would require a more extended theoretical analysis.

We acknowledge the support of the HFML-RU/FOM, member of the European Magnetic Field Laboratory (EMFL). R. S. M. Rijken acknowledges the Graduate School for Molecules and Materials for a PhD position and NWO for its corresponding grant. D. A. Wilson acknowledges funding from the European Research Council under the European Union's Seventh Framework Programme (FP7/2007-2012)/ERC-StG 307679 “StomaMotors”. R. J. M. Nolte acknowledges support from the European Research Council (ERC Advanced grant 290886-ALPROS) and R. J. M. Nolte and J. C. M. van Hest acknowledge support from the Ministry of Education, Culture and Science (Gravity program 024.001.035). Finally, we would like to thank G.-J. Janssen for his assistance with the electron microscopes and P. Walraven for the fabrication of the flow cell.

Notes and references

7. (a) D. A. Wilson, R. J. M. Nolte and J. C. M. van Hest, Nat. Chem., 2012, 4, 266; (b) D. A. Wilson, B. de Nijs, A. van Bladeren, R. J. M. Nolte and J. C. M. van Hest, Nanoscale, 2013, 5, 1315.