Inverse Relationship of Serum Hepcidin Levels with CD4 Cell Counts in HIV-Infected Patients Selected from an Indonesian Prospective Cohort Study

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Abstract

Background: Distortion of iron homeostasis may contribute to the pathogenesis of human immunodeficiency virus (HIV) infection and tuberculosis (TB). We studied the association of the central iron-regulatory hormone hepcidin with the severity of HIV and the association between hepcidin and other markers of iron homeostasis with development of TB.

Methods: Three groups of patients were selected from a prospective cohort of HIV-infected subjects in Bandung, Indonesia. The first group consisted of HIV-infected patients who started TB treatment more than 30 days after cohort enrollment (cases). The second group consisted of HIV-infected patients who were matched for age, gender and CD4 cell count to the cases group (matched controls). The third group consisted of HIV-infected patients with CD4 cell counts above 200 cells/mm³ (unmatched controls). Iron parameters including hepcidin were compared using samples collected at cohort enrollment, and compared with recently published reference values for serum hepcidin.

Results: A total of 127 HIV-infected patients were included, 42 cases together with 42 matched controls and 43 unmatched controls. Patients with advanced HIV infection had elevated serum hepcidin and ferritin levels. Hepcidin levels correlated inversely with CD4 cells and hemoglobin. Cases had significantly higher hepcidin and ferritin concentrations at cohort enrollment compared to matched controls, but these differences were fully accounted for by the cases who started TB treatment between day 31 and 60 after enrollment. Hepcidin levels were not different in those with or without hepatitis C infection.

Conclusion: Iron metabolism is distorted in advanced HIV infection with CD4 cell counts correlating inversely with serum hepcidin levels. High serum hepcidin levels and hyperferritinemia were found in patients starting TB treatment shortly after cohort enrollment, suggesting that these parameters have a predictive value for development of manifest TB in HIV-infected patients.

Introduction

Alterations in iron distribution are common in infectious diseases and many of these alterations may be attributable to actions of the iron-regulatory hormone hepcidin [1]. Hepcidin degrades the sole cellular iron exporter ferroportin leading to reduced iron absorption in the intestine and iron retention in monocytes and macrophages and the spleen [2]. Changes in iron homeostasis have been described in HIV-infected patients. Epidemiological studies have found an association between elevated iron status, HIV progression and the risk for opportunistic infections [3,4]. HIV replication involves several iron-dependent steps [5,6], and as a central determinant of macrophage iron contents, hepcidin may play a distinct role in HIV pathogenesis. Indeed, hepcidin was recently shown to increase HIV-1 transcription in cultured monocytes and T-cells by degradation of ferroportin with a secondary increase in intracellular iron [7]. Hepcidin may also be involved in two important complications of human immunodeficiency virus infection/acquired immune deficiency syndrome (HIV/AIDS). First, elevated hepcidin levels limit iron supply to the bone marrow. This may contribute to HIV-associated anemia, which is a common complication of advanced HIV infection with negative impact on clinical outcome and quality of life [8–11]. Second, hepcidin-mediated iron accumulation in macrophages may
increase the risk for outgrowth of intracellular pathogens like *Mycobacterium tuberculosis*. Worldwide, tuberculosis (TB) is the most important infectious disease complication among HIV-infected patients, and several lines of evidence suggest that macrophage iron content is related to the risk for developing active TB [12,13]. Interestingly, hepcidin itself has antibacterial activity and hepcidin was shown to inhibit *M. tuberculosis* growth in vitro [14].

Data on hepcidin levels in HIV infected patients are rarely reported and were found to be related to ferroportin mutations [15]. Apart from that, the pro-inflammatory cytokine interleukin (IL)-6 is a dominant regulator of hepatic hepcidin production in bacterial infections and other inflammatory conditions, but IL-6 concentrations are often only mildly elevated in viral infections. Indeed, recent studies have shown that hepcidin levels are reduced in hepatitis C virus (HCV) infection, which may contribute to pathological liver iron storage in patients with chronic HCV infection [16,17].

The present study was performed in Indonesia, which has one of the fastest growing HIV epidemics in Asia with a high rate of TB and hepatitis C co-infection. Our primary aim was to study the effect HIV infection on serum hepcidin levels and other markers of iron homeostasis and to compare hepcidin levels with recently determined reference levels for hepcidin in healthy Dutch volunteers [18]. The secondary aim was to identify whether hepcidin and other markers of iron homeostasis were associated with development of TB more than 30 days after inclusion in the study. Finally, we studied whether iron parameters were influenced by factors such as gender, anemia, the use and kind of antiretroviral treatment (ART) and HCV co-infection.

**Methods**

**Patients and setting**

This study was designed as a nested case control study in a cohort of HIV-infected patients in Hasan Sadikin Hospital in Bandung, the referral hospital for HIV care in West-Java [11]. Free anti-retroviral treatment (ART) is delivered since December 2004. Following the 2006 World Health Organization (WHO) guidelines [19], indications for start of ART during the study period were: a) HIV stage IV irrespective of CD4 cell count; b) HIV stage III with a CD4 cell count below 500 cells/mm^3^; and c) HIV stage I or II with a CD4 cell count less than 200 cells/mm^3^.

First-line ART includes the nucleoside reverse transcriptase inhibitors (NRTI) zidovudine, stavudine and lamivudine, and the non-nucleoside reverse transcriptase inhibitors (NNRTI) efavirenz and nevirapine.

Starting in September 2007, all patients, aged 14 years and above are being enrolled in a prospective cohort study. Baseline enrollment consists of a structured interview, physical examination, chest X-ray and blood examination with storage of serum at −80°C. The latter is performed within one week from initial presentation and enrollment in the cohort. Injecting drug use is an important route of HIV transmission in West Java and HCV co-infection [20].

**Methods**

**Hepcidin in Indonesian HIV Patients**

Serum hepcidin and iron parameters were measured in archived serum collected at time of cohort enrollment in the Department of Laboratory Medicine of the Radboud University Medical Centre using a combination of weak cation exchange chromatography and time-of-flight mass spectrometry (TOF MS). This technique allows discrimination between the three naturally occurring isoforms of hepcidin (hepcidin-20, -25, and -26), of which only the 25 amino acid form can block the iron transporter ferroportin and is referred to as “bioactive” hepcidin. An internal standard (synthetic hepcidin-24; Peptide International Inc.) was used for quantification [22,23]. Peptide spectra were generated on a Micromass LT matrix-enhanced laser desorption/ionization TOF MS platform (Bruker Daltonics). Serum hepcidin-25 (further referred to as hepcidin) concentrations were expressed in nanomoles. The lower limit of detection of this method was 0.5 nM; average coefficients of variation were 2.7% (intra-run) and 6.5% (inter-run). The median level of serum hepcidin-25 concentrations in healthy Dutch male adults is 4.5 nM, while the reference range (P2.5–P97.5) is <0.5–14.7 nM; for premenopausal women these values are 2.0 nM (<0.5–12.3) [17,24].

**Laboratory methods**

Serum concentrations of total iron, total iron binding capacity (TIBC), transferrin saturation (TS), ferritin, sTfR and C-reactive protein (CRP) were determined as described previously [25]. The following reference ranges were used: serum iron: 10–25 μmol/L;
transferrin saturation (TS): 30–60%; total iron binding capacity (TIBC): 45–80 μmol/L; serum ferritin: 15–280 μg/L for males and 6–80 μg/L for premenopausal females; soluble transferrin receptor (sTfR): 0.76–1.76 mg/L. For statistical analyses, the lower limit of detection for CRP (5 mg/L) was used in samples with values below this detection limit. A full blood count was measured on a Cell Dyne 3000 (Abbott) hematology analyzer. Anemia was defined according to World Health Organization/ Aids Clinical Trial Group (WHO/ACTG) criteria as a hemoglobin level below 13.0 g/dL for men and below 12.0 g/dL for women. Anti-hepatitis C antibodies were tested by an electro-chemiluminescence assay (ECLIA; Elecsys, Roche Diagnostics); CD4-cell counts were determined by flow cytometry (BD Biosciences, Jakarta, Indonesia). Due to limited volume of archived samples, serum levels of ferritin and sTfR were available in only 107 patients and serum iron, TIBC, TS and CRP in 38 patients.

Data analysis and statistics

Serum concentrations of hepcidin and other markers of iron homeostasis were compared between cases and the two control groups. Subsequently, the effect of HIV infection on iron homeostasis was analyzed by comparing iron parameters in matched and unmatched controls with recently determined reference levels of serum hepcidin and by determination of the correlation of serum hepcidin levels with CD4 cell count. Finally, in the whole group of HIV-infected patients, associations were examined between iron parameters and gender, anemia, ART and the presence of HCV antibodies. Data are presented as medians with interquartile range (IQR) unless otherwise stated. Paired analysis for cases and the matched control group was performed using the Wilcoxon signed-rank test for continuous variables and the McNemar test for categorical variables. Unpaired comparisons between cases and unmatched controls and between matched and unmatched controls were performed using the Mann-Whitney test and Chi-squared used. Comparison of median hepcidin levels in our study population with reference values was performed using the Wilcoxon signed-rank sum test. A two-sided P value of less than 0.05 was considered statistically significant. All statistical analyses were done by software SPSS version 17.0.

Results

Patient characteristics

A total number of 127 HIV-infected patients were included in our study: 42 cases who developed TB during follow-up and two control groups, consisting of 42 controls matched to cases for age, sex and CD4 cell count (matched controls), and 43 unmatched controls with a CD4 cell count of more than 200 cells/mm3 (unmatched controls). The majority of cases and matched controls had an advanced HIV infection with a median CD4 cell count of 30 cells/mm3 and 70 cells/mm3, respectively; the unmatched controls were moderately immunocompromised with a median CD4 cell count of 365 cells/mm3. Matched analysis did not find any difference between cases and matched control group for age, sex and CD4. Table 1 summarizes clinical and routine laboratory characteristics in study subjects at the time of cohort enrollment. In the cases, TB treatment was started at a median of 125 days (IQR 44–238 days) after cohort enrollment. Thirty-three patients (78%) of the cases had pulmonary TB; including 12 patients with AFB smear positive TB. Patients without bacteriological confirmation had a good response to TB treatment (by definition). Nine patients were diagnosed with extra-pulmonary TB of whom seven with lymphadenitis. The diagnosis of extra-pulmonary TB was made by histopathology examination in three cases and by a combination of clinical and radiological features, combined with a good response to anti-TB treatment in the remaining six. Ten of the cases had a history of TB-treatment, a median of 512 days (IQR 357–587 days) before which only one case did not finished 6 months TB treatment due to side effect. By definition, the patients in the control groups had neither a history of TB, nor any signs or symptoms suggesting development of active TB. After cohort enrollment, 27 out of 31 cases and 20 out of 31 matched controls initiated ART. CRP levels were only moderately elevated and mostly limited to the severely immunocompromised patients in the cases and matched controls group.

Iron parameters in relation to HIV status and CD4 cell count

Median serum hepcidin levels in the whole group of HIV-infected patients were above reference values for healthy Dutch adults (7.3 vs. 4.5 nM; p<0.001) and premenopausal females (3.1 vs. 2.0 nM; p = 0.028) [16,22]. However, these higher hepcidin were largely restricted to cases and matched controls, i.e. those with advanced HIV infection (table 2). The unmatched male controls had a similar median hepcidin level value as healthy Dutch adults (4.4 [2.1–8.1] nM), but female unmatched controls had a higher median hepcidin level (4.4 [1.5–8.2] nM). The higher hepcidin levels in cases compared to unmatched controls were associated with higher serum levels of the iron storage protein ferritin and lower serum iron, TIBC and mean corpuscular volume (MCV) values (table 2). Serum hepcidin levels also correlated inversely with CD4 cell counts, hemoglobin levels and MCV, and positively with serum ferritin levels (Fig. 1A–D).

Iron parameters and development of TB

Compared to matched controls, cases starting TB treatment (‘cases’) more than 30 days after enrollment had significantly higher serum hepcidin (p = 0.035) and ferritin levels (p = 0.006)(Table 2). However, this difference was fully accounted for by the cases starting TB treatment between day 31 and 60. Compared to those starting TB treatment more than 60 days after cohort enrollment, the cases had significantly higher serum hepcidin and ferritin levels at cohort enrollment with median (IQR) serum levels for hepcidin of 16.1 nM [8.4–20.6 nM] vs. 5.8 nM (4.1–14.1 nM; p = 0.01) and for ferritin of 1206 μg/L (591–1799 μg/L) vs. 310 μg/L (190–887 μg/L; p = 0.004), despite the fact that clinical and radiological examination showed no signs for TB at cohort enrollment. There were no significant differences in either hepcidin or other iron indicators between cases starting TB treatment more than 60 days after cohort enrollment and their matched controls. Combining all cases of pulmonary TB, smear positive pulmonary TB was associated with a significantly higher serum hepcidin level at cohort enrollment than smear negative TB (15.9 nM [13.1–21.9 nM] vs. 5.3 nM [4.1–5.3 nM] nM; p = 0.003). In addition, cases developing extrapulmonary TB had significantly higher ferritin levels at cohort enrollment than cases with pulmonary TB (1351 μg/L [725–1800 μg/L] vs. 491 μg/L [193–1250 μg/L]; p = 0.03), but similar hepcidin levels (9.4 nM [6.0–11.8 nM] vs. 11.5 nM [4.9–17.0 nM]; p = 0.47).

Other factors affecting iron parameters

Apart from HIV and/or TB infection, iron parameters may be influenced by other factors such as gender, anemia, the use and kind of ART and HCV co-infection. As shown in Table 3, women had significantly lower serum hepcidin and ferritin levels than men.
Table 1. Characteristics of HIV-infected patients who were diagnosed with tuberculosis after cohort enrollment and control groups.

<table>
<thead>
<tr>
<th></th>
<th>Cases**</th>
<th>Matched controls</th>
<th>p*</th>
<th>Unmatched controls</th>
<th>p†</th>
<th>p‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>29 (26–33)</td>
<td>30 (28–32)</td>
<td>0.51*</td>
<td>27 (25–29)</td>
<td>0.03</td>
<td>0.002</td>
</tr>
<tr>
<td>Male, %</td>
<td>90.5</td>
<td>90.5</td>
<td>1.00†</td>
<td>76.7</td>
<td>0.95</td>
<td>0.95</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>18.5 (16.9–21.1)</td>
<td>18.1 (17.3–20.2)</td>
<td>0.73</td>
<td>20.5 (18.8–21.8)</td>
<td>0.016</td>
<td>0.005</td>
</tr>
<tr>
<td>Follow-up after cohort enrollment, days</td>
<td>836 (273–1110)</td>
<td>657 (68–1091)</td>
<td>0.257</td>
<td>539 (55–1095)</td>
<td>0.20</td>
<td>0.91</td>
</tr>
<tr>
<td>ART at baseline, %</td>
<td>28.6</td>
<td>26.2</td>
<td>0.81</td>
<td>34.9</td>
<td>0.53</td>
<td>0.38</td>
</tr>
<tr>
<td>History of TB treatment before initial visit, %</td>
<td>23.8</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>CD4 cell count, cells/mm³</td>
<td>30 (14–138)</td>
<td>70 (27–137)</td>
<td>0.50*</td>
<td>365 (324–461)</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>CD4 cell count &lt;200 cells/mm³</td>
<td>81.0</td>
<td>90.5</td>
<td>0.21</td>
<td>0</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Hemoglobin, g/dL</td>
<td>12.0 (10.1–14.1)</td>
<td>13.7 (12.1–14.6)</td>
<td>0.02</td>
<td>14.5 (12.9–15.3)</td>
<td>&lt;0.001</td>
<td>0.02</td>
</tr>
<tr>
<td>Anemia, %</td>
<td>63.4</td>
<td>38.1</td>
<td>0.02</td>
<td>14.0</td>
<td>&lt;0.001</td>
<td>0.01</td>
</tr>
<tr>
<td>C-reactive protein, mg/L</td>
<td>16.0 (5.0–36.0)</td>
<td>5.0 (5.0–15.5)</td>
<td>0.21</td>
<td>5.0 (5.0–5.0)</td>
<td>0.02</td>
<td>0.20</td>
</tr>
<tr>
<td>C-reactive protein &gt;10 mg/L, %</td>
<td>57.1</td>
<td>37.5</td>
<td>0.38</td>
<td>6.7</td>
<td>0.009</td>
<td>0.04</td>
</tr>
<tr>
<td>Alanine transaminase, IU/L</td>
<td>33.5 (20.0–54.5)</td>
<td>33.0 (20.8–66.0)</td>
<td>0.90</td>
<td>47.0 (21.0–69.0)</td>
<td>0.47</td>
<td>0.47</td>
</tr>
<tr>
<td>Random blood glucose, mg/dL</td>
<td>102.0 (87.3–116.5)</td>
<td>92.5 (82.3–103.8)</td>
<td>0.04</td>
<td>91.0 (88.8–98.0)</td>
<td>0.03</td>
<td>0.76</td>
</tr>
<tr>
<td>Total cholesterol, mg/dL</td>
<td>158.5 (120.5–183.5)</td>
<td>129.0 (111.0–167.0)</td>
<td>0.30</td>
<td>158.0 (120.5–178.5)</td>
<td>0.89</td>
<td>0.33</td>
</tr>
<tr>
<td>LDL, mg/dL</td>
<td>84.5 (63.5–114.0)</td>
<td>70.0 (49.0–100.0)</td>
<td>0.24</td>
<td>78.0 (54.5–103.5)</td>
<td>0.76</td>
<td>0.52</td>
</tr>
<tr>
<td>Triglyceride, mg/dL</td>
<td>152.5 (103.0–196.0)</td>
<td>145.0 (114.0–172.0)</td>
<td>0.77</td>
<td>113.0 (96.5–154.0)</td>
<td>0.13</td>
<td>0.11</td>
</tr>
<tr>
<td>Hepatitis C antibodies present, %</td>
<td>72.5</td>
<td>73.7</td>
<td>0.91</td>
<td>66.7</td>
<td>0.58</td>
<td>0.51</td>
</tr>
</tbody>
</table>

Data are presented as medians and interquartile range (IQR) unless stated otherwise. ART, antiretroviral therapy; BMI, body mass index; LDL, low-density lipoprotein. The number of samples available in each group is shown in superscript after the IQR or percentage if less than total number.

**Cases group: HIV-infected patients diagnosed with tuberculosis (TB) more than 1 month after cohort enrollment. Matched controls: HIV-infected patients without diagnosis of TB before or after cohort enrollment, matched for age, gender and CD4 cell count to cases group. Unmatched controls: HIV-infected patients without diagnosis of TB before or after cohort enrollment and CD4 cells >200 cells/mm³. P values were determined for b difference between matched controls and cases, c between unmatched controls and cases and, d between matched and unmatched controls.

*Wilcoxon signed rank test.

Wilcoxon signed rank test.

Anemia defined as hemoglobin level <13 g/dL for men or <12 g/dL for women.

Defined as positive anti-HCV antibodies.

doi:10.1371/journal.pone.0079904.t001

(p<0.01) owing to their lower total iron stores. Second, anemic patients had higher ferritin and hepcidin levels (p<0.001 and p = 0.05, respectively) together with a lower value of TIBC (p<0.001), which is consistent with anemia of chronic disease. Third, patients taking stavudine had somewhat higher hepcidin concentrations than those taking zidovudine, although this difference was not statistically significant (p = 0.24). Finally, HCV-infected patients had slightly higher serum hepcidin, ferritin and transferrin saturation levels, although these differences were not statistically significant.

Discussion

Different studies have shown that disturbances in iron homeostasis and anemia are associated with advanced HIV infection and an adverse outcome [3,4,9,26]. To our knowledge, only one study has so far reported data on hepcidin in HIV-infected patients. Masaisa et al. found a mutation (Q248H) in the iron exporter ferroportin to be related to lower serum hepcidin levels, higher ferritin levels and a higher risk for pulmonary TB [15]. Our data show that serum hepcidin levels were increased in Indonesian patients with advanced HIV infection. In contrast, HIV-infected patients without TB and with CD4 cell counts above 200 cells/mm³ (unmatched controls) had similar serum hepcidin levels as healthy Dutch adults. Inflammatory cytokines, especially IL-6, are important inducers of hepatic hepcidin expression. Our observation of an inverse relation of CD4 cell counts with serum hepcidin levels and the fact that elevated CRP levels were more common in those with advanced HIV-infection support the notion that elevated hepcidin levels in advanced HIV are predominantly caused by HIV-associated inflammation. Moreover, only few of the unmatched controls with CD4 cell counts above 200 cells/mm³ had an elevated CRP level (6.7%), and this may explain why this group had similar hepcidin levels as healthy Dutch individuals. Other HIV-specific mechanisms, apart from inflammation, may also contribute to higher hepcidin levels. HIV can directly infect bone marrow progenitor cells leading to bone marrow suppression, which is associated with upregulated hepcidin expression [27,28]. Zidovudine is also known for its suppressive effect on the bone marrow, but use of zidovudine was not associated with higher hepcidin levels in our study.

Our finding of increased hepcidin expression with secondary iron dysregulation in advanced HIV infection may have several clinical consequences. First, increased hepcidin expression leads to macrophage iron loading and this may be associated with a higher risk for intracellular infections. M. tuberculosis intracellular growing bacterium that needs iron for its multiplication, and macrophage iron loading might thus stimulate development of TB. In our present study, we indeed found higher serum hepcidin and ferritin levels at cohort enrollment in those who subsequently started TB
treatment, but these higher levels were restricted to those starting treatment from day 31 to 60 after enrollment. Recent studies have shown that *M. tuberculosis* and mycobacterial components may also induce hepcidin and we speculate that the presence of subclinical TB at cohort enrollment accounted for these higher hepcidin and ferritin levels [14,29]. Alternatively, higher hepcidin and ferritin levels may put HIV-infected patients at an increased risk for TB reactivation of TB immune reconstitution inflammatory syndrome (IRIS). The latter seems less likely, because hepcidin not only functions as an iron regulator, but also possesses antibacterial effects. Indeed, other authors have suggested increased hepcidin production in response to *M. tuberculosis* infection to represent a host defense mechanism [14,29]. A second possible clinical consequence of the high hepcidin levels is its effect on HIV

![Figure 1. Correlation of serum hepcidin levels with hematology and iron parameters.](https://www.plosone.org/doi/abs/10.1371/journal.pone.0079904.g001)

Revised Table 2:

**Hepcidin and iron parameters at cohort enrollment in HIV-infected patients with (cases) or without (controls) incident TB.**

<table>
<thead>
<tr>
<th></th>
<th>Cases group*</th>
<th>Matched controls</th>
<th></th>
<th>Unmatched controls</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Hepcidin, nM</td>
<td>9.9 (5.0–18.2)</td>
<td>4.4 (2.0–7.9)</td>
<td>0.035</td>
<td>0.06</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ferritin, μg/l</td>
<td>680.5 (245.0–1439.0)</td>
<td>213.5 (74.3–358.0)</td>
<td>0.006</td>
<td>&lt;0.001</td>
<td></td>
<td></td>
</tr>
<tr>
<td>sTfR, mg/l</td>
<td>1.4 (1.1–2.2)</td>
<td>1.3 (1.1–1.6)</td>
<td>0.61</td>
<td>0.28</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Serum iron, μmol/l</td>
<td>7.0 (5.0–11.0)</td>
<td>16.0 (10.0–27.0)</td>
<td>0.27</td>
<td>0.09</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TIBC, μmol/l</td>
<td>39.0 (38.0–54.0)</td>
<td>57.0 (49.0–66.0)</td>
<td>0.26</td>
<td>0.08</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Transferrin saturation, %</td>
<td>16.0 (12.0–27.0)</td>
<td>26.0 (16.0–47.0)</td>
<td>0.26</td>
<td>0.15</td>
<td></td>
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</tr>
<tr>
<td>MCV, fl</td>
<td>83.5 (80.3–92.5)</td>
<td>88.0 (84.0–104.0)</td>
<td>0.53</td>
<td>0.11</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MCH, pg</td>
<td>29.0 (26.3–32.0)</td>
<td>30.0 (28.0–35.0)</td>
<td>0.98</td>
<td>0.14</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Data depicted are median with interquartile range (IQR). TB, tuberculosis; TIBC, total iron binding capacity; sTfR, soluble transferrin receptor; MCV, mean cell volume; MCH, mean cell hemoglobin.

Numbers of samples available in each group is shown in superscript after the IQR if less than total number.

*Cases group: HIV-infected patients diagnosed with tuberculosis (TB) more than 1 month after initial visit. Matched controls: HIV-infected patients without diagnosis of TB before or after initial visit, matched for age, gender and CD4 cell count to cases group. Unmatched controls: HIV-infected patients without diagnosis of TB before or after initial visit not matched for age, gender and CD4 cell count to cases group. P values were determined for \(a\) difference between matched controls and cases by using the Wilcoxon signed-rank test and \(b\) between unmatched controls and cases and \(c\) matched and unmatched controls by using the Mann-Whitney test. doi:10.1371/journal.pone.0079904.t002
Hepcidin in Indonesian HIV Patients

Table 3. Hepcidin, iron and inflammatory parameters in HIV-infected patients at cohort enrollment.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Hepcidin (nM)</th>
<th>Serum iron (µmol/l)</th>
<th>TIBC (µmol/l)</th>
<th>Transferrin saturation (%)</th>
<th>Ferritin (µg/l)</th>
<th>sTfR (mg/l)</th>
<th>CRP (mg/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Gender</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>7.3 (4.0–13.1)^m</td>
<td>11.5 (6.3–20.0)^a</td>
<td>53.9 (39.0–62.8)^a</td>
<td>27.5 (14.5–36.5)^a</td>
<td>344 (194–795)^a</td>
<td>1.4 (1.2–2.0)^a</td>
<td>5.0 (5.0–16.0)^a</td>
</tr>
<tr>
<td>Female</td>
<td>3.1 (1.5–6.9)^a</td>
<td>9.5 (8.5–12.5)^a</td>
<td>54.0 (48.6–66.8)^a</td>
<td>17.0 (12.8–25.3)^a</td>
<td>60 (46–132)^a</td>
<td>1.5 (1.1–1.9)^a</td>
<td>5.0 (5.0–7.3)^a</td>
</tr>
<tr>
<td><strong>Anemia</strong></td>
<td></td>
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<tr>
<td>Yes</td>
<td>9.2 (4.6–16.4)^a</td>
<td>9.0 (4.5–13.0)^a</td>
<td>42.0 (38.5–49.0)^a</td>
<td>21.0 (11.0–33.0)^a</td>
<td>698 (257–1460)^a</td>
<td>1.4 (1.0–2.2)^a</td>
<td>14.0 (5.0–41.5)^a</td>
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<tr>
<td>No</td>
<td>5.3 (2.5–9.6)^a</td>
<td>16.0 (9.0–20.0)^a</td>
<td>61.0 (55.0–66.0)^a</td>
<td>26.0 (16.0–31.0)^a</td>
<td>217 (112–351)^a</td>
<td>1.4 (1.2–1.8)^a</td>
<td>5.0 (5.0–5.0)^a</td>
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<tr>
<td><strong>Receive ART</strong></td>
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<tr>
<td>Yes</td>
<td>5.6 (2.6–9.8)^a</td>
<td>20.5 (9.8–36.8)^a</td>
<td>45.5 (39.0–53.0)^a</td>
<td>42.5 (19.0–87.0)^a</td>
<td>290 (161–1081)^a</td>
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<td>7.0 (5.0–26.5)^a</td>
</tr>
<tr>
<td>No</td>
<td>5.2 (2.2–9.6)^a</td>
<td>27.0 (4.0–40.0)^a</td>
<td>49.0 (39.0–57.0)^a</td>
<td>47.0 (10.0–93.0)^a</td>
<td>252 (130–741)^a</td>
<td>1.7 (1.2–2.1)^a</td>
<td>7.0 (5.0–14.0)^a</td>
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<tr>
<td><strong>HCV</strong></td>
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<tr>
<td>Yes</td>
<td>6.8 (3.0–11.6)^a</td>
<td>11.0 (6.0–11.0)^a</td>
<td>55.0 (39.0–64.0)^a</td>
<td>27.0 (16.0–32.0)^a</td>
<td>298 (191–596)^a</td>
<td>1.3 (1.1–1.9)^a</td>
<td>5.0 (5.0–16.0)^a</td>
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<tr>
<td>No</td>
<td>5.8 (2.5–13.1)^a</td>
<td>9.0 (7.0–12.0)^a</td>
<td>50.0 (44.0–56.0)^a</td>
<td>16.0 (13.0–25.0)^a</td>
<td>257 (84–698)^a</td>
<td>1.5 (1.2–2.2)^a</td>
<td>5.0 (5.0–14.0)^a</td>
</tr>
</tbody>
</table>

Data are based on 127 HIV-infected patients belonging to case group and the matched and unmatched control groups. Data depicted as median with interquartile range. TIBC, total iron binding capacity; sTfR, soluble transferrin receptor; CRP, C-reactive protein; ART, antiretroviral therapy; HCV, hepatitis C; ZDV, zidovudine; d4T, stavudine.

Hemoglobin and serum hepcidin levels were available for 127 patients; ferritin and sTfR for 107 patients and serum iron, TIBC, transferrin saturation and CRP in 38 patients. Anti HCV antibodies were available in 113 patients. Numbers of samples per groups are shown in superscript after the IQR.

*p<0.01 between male and female group;

*p<0.01 between anemia and non-anemia group;

*p<0.05 between group with and without ART.

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confirmed in our study, which is the reason for not performing a multivariate analysis. However, we feel that none of these issues can account for the significant associations we found between hepcidin and CD4 cell count and development of TB.

In conclusion, our study shows that advanced HIV infection is associated with increased hepcidin expression and characteristic features of iron maldistribution. This may, among others, stimulate HIV progression and contribute to HIV-associated anemia. Patients starting TB treatment shortly after cohort enrollment had significantly higher serum hepcidin and ferritin level at cohort enrollment, but this effect was not present in those starting TB treatment after at least 2 months.

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We would like to thank Dr. Bayu Wahyudi, Director of Hasan Sadikin General Hospital, and Prof. Tri Hanggono Achmad, Dean of the Medical Faculty Padjadjaran University for encouraging and accommodating research at their institutions.

Author Contributions

Conceived and designed the experiments: RW QiIM BA HJ PS RS DS RvC AvdV. Performed the experiments: RW ARI DS. Analyzed the data: RW QiIM DS RvC AvdV. Contributed reagents/materials/analysis tools: ARI DS. Wrote the paper: RW QiIM RvC AvdV.

References