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Prednisolone induces the Wnt signalling pathway in 3T3-L1 adipocytes

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Abstract

Synthetic glucocorticoids are potent anti-inflammatory drugs but show dose-dependent metabolic side effects such as the development of insulin resistance and obesity. The precise mechanisms involved in these glucocorticoid-induced side effects, and especially the participation of adipose tissue in this, are not completely understood. We used a combination of transcriptomics, antibody arrays and bioinformatics approaches to characterize prednisolone-induced alterations in gene expression and adipokine secretion, which could underlie metabolic dysfunction in 3T3-L1 adipocytes. Several pathways, including cytokine signalling, Akt signalling, and Wnt signalling were found to be regulated at multiple levels, showing that these processes are targeted by prednisolone. These results suggest that mechanisms by which prednisolone induce insulin resistance include dysregulation of Wnt signalling and immune response processes. These pathways may provide interesting targets for the development of improved glucocorticoids.

Introduction

Synthetic glucocorticoids (GCs) such as prednisolone and dexamethasone are widely used for the treatment of inflammatory diseases such as rheumatoid arthritis, asthma, inflammatory bowel disease and psoriasis (Del Rosso Do, 2006; Hillier, 2007; Schwartz & Cohen, 2008). Despite their excellent efficacy, GC usage is hampered because of adverse (metabolic) side effects, such as insulin resistance, glucose intolerance, diabetes, central adiposity, dyslipidaemia, skeletal muscle wasting and osteoporosis (De Bosscher & Haegeman, 2009; Rhen & Cidlowski, 2005; Rockall et al., 2003; Schacke et al., 2002, 2006). The precise mechanisms involved in these GC-induced side effects are not completely understood and are tissue specific (Chrousos & Kino, 2009; Kino, 2007; Lu & Cidlowski, 2006; van Raalte et al., 2009). One of the key tissues thought to be involved in metabolic side effects is the adipose tissue. Several studies have shown that the side-effects of GCs in adipose tissue include the development of central adiposity (Rockall et al., 2003), dyslipidaemia (Taskinen et al., 1983; Wajchenberg, 2000) and the inhibition of insulin-stimulated glucose uptake (Bazuine et al., 2004; Buren et al., 2002, 2008; Sakoda et al., 2000).

It is now well established that adipose tissue is not only involved in energy storage, but also functions as an endocrine organ that secretes hundreds of bioactive substances known as adipokines (Kershaw & Flier, 2004; Ouchi et al., 2011; Lehr et al., 2011). Previously, GCs have been found to affect the secretion of several adipokines, including TNF-α, IL-6, Resistin, Adiponectin and Leptin in rodents (Fasshauer & Paschke, 2003). Because recent proteomic approaches have led to the characterization of numerous novel adipokines, we applied a combination of genomics, antibody arrays and bioinformatics approaches to identify prednisolone-induced alterations in (adipokine) gene expression and secretion which could potentially underlie prednisolone-induced metabolic dysfunction of 3T3-L1 adipocytes.

Materials and methods

Set up of the study

In this study, differentiated 3T3-L1 adipocytes were treated with prednisolone or DMSO for 0, 1, 6, 24 and 48 h and the effects of prednisolone were studied at gene expression and protein secretion level.
**Cell culture**

3T3-L1 pre-adipocytes obtained from ATCC were cultured and differentiated in adipocytes as previously described (van den Berghe et al., 1994). Cells were used seven days after completion of the differentiation process. Only cultures in which >95% of cells displayed adipocyte morphology were used. Prior to use, adipocytes were serum starved for 16 h with DMEM supplemented with 0.5% foetal bovine serum.

**Analysis of insulin signalling**

Differentiated 3T3-L1 adipocytes, grown in 12-well plates, were incubated with prednisolone (1 μM) or DMSO at day 8 for 48 h. After incubation with prednisolone or DMSO, cells were serum-starved for 2 h, and then stimulated with insulin (100 nM) for 10 min. Following insulin stimulation, cells were washed twice with ice-cold PBS, and lysed as described previously (Linssen et al., 2011). Protein expression and phosphorylation were determined by Western blotting as described (Linssen et al., 2011) using the following antibodies: phospho-insulin receptor substrate 1-Tyr1222, phospho-Akt-Thr308, phospho-Akt-Ser473 (all from Cell Signalling Technology, Danvers, MA, USA), total Akt, phospho-Akt substrate of 160-kDa (AS160), total AS160 (all from Millipore, MA, USA), glucose transporter 4 (GLUT4), and β-actin (both from Abcam, Cambridge, UK).

**Assay of 2-deoxy-D-glucose (2DOG) uptake**

Differentiated 3T3-L1 adipocytes, grown in 12-wells plates, were incubated with prednisolone (1 μM) or DMSO at day 8 for 6, 24 and 48 h. After incubation with prednisolone or DMSO for the indicated times, the cells were washed once with HEPES buffer, consisting of 50 mM HEPES, 0.14 M NaCl, 1.85 mM CaCl₂, 1.3 mM MgSO₄, and 4.8 mM KCl [pH 7.4], and then incubated in this buffer for an additional hour at 37°C. Then, cells were stimulated with insulin (100 nM) or kept untreated. After 15 min, 2DOG uptake was initiated by the addition of 2-deoxy-D-[¹⁴C] glucose (0.075 μCi per well) in 3 mM 2DOG. After 10 min, the assay was terminated by three quick washes with ice-cold PBS. Cells were lysed in 0.1 M NaOH, 0.2% SDS, whereafter incorporated 2-deoxy-D-[¹⁴C] glucose was determined by liquid scintillation counting.

**RNA isolation**

Differentiated 3T3-L1 adipocytes grown in 6-well plates were incubated with prednisolone (1 μM) for 1, 6, 24 and 48 h and used for RNA isolation and whole-genome expression profiling. RNA was isolated using TRIzol (Roche), and subsequently purified using the RNeasy mini kit according to the manufacturer’s protocol (Qiagen Benelux B.V. Venlo, The Netherlands). To remove residual traces of genomic DNA, the RNA was treated with DNase I (Invitrogen, Leek, The Netherlands) while bound to the RNeasy column. Quality and quantity of the purified RNA was controlled using a NanoDrop spectrophotometer (Nanodrop Technologies, Montchanin, DE, USA). RNA integrity was investigated by using the 2100 Bioanalyzer (Agilent technologies, Philadelphia, PA, USA).

**Whole-genome expression analysis using Affymetrix GeneChip Mouse Genome 430 2.0 Array**

Amplification of 20 ng total RNA was performed with the Two-Cycle Eukaryotic Target Labeling kit (Affymetrix, Santa Clara, CA). Briefly, after the first round of cDNA synthesis, an unlabeled ribonucleotide mix (MEGAscript T7 kit, Ambion) was used to generate unlabelled cRNA according to the protocol of Affymetrix. After cleanup of the cRNA with a GeneChip Sample Cleanup Module IVT Column (Affymetrix, Santa Clara, CA), the unlabelled cRNA concentration was determined and 150 ng was reverse transcribed using random primers. Subsequently, the T7-Oligo(dT) Promoter Primer was used in the second-strand cDNA synthesis to generate double-stranded cDNA containing T7 promoter sequences. The resulting double-stranded cDNA was then amplified and biotin labelled using the IVT Labeling kit (Affymetrix, Santa Clara, CA). Biotin-labelled cRNA was fragmented at 1 μg/μl following the manufacturer’s protocol. After fragmentation, cRNA (10 μg) was hybridized at 45°C for 16 h to the Human Genome U133 2.0 Plus array (Affymetrix, Santa Clara, CA). Following hybridization, the arrays were washed, stained with phycoerythrin-streptavidin conjugate (Molecular Probes, Eugene, OR), and the signals were amplified by staining the array with biotin-labelled anti-streptavidin antibody (Vector Laboratories, Burlingame, CA) followed by phycoerythrin-streptavidin. The arrays were laser scanned with a GeneChip Scanner 3000 7G (Affymetrix, Santa Clara, CA) according to the manufacturer’s instructions. Data was saved as raw image file and quantified using AGCC v 1.0 (Affymetrix, Santa Clara, CA).

**Murine adipokine antibody arrays**

This array allows simultaneous detection of 308 secreted mouse proteins in cell culture supernatants (AAM-BLM, L-series, Raybiotech, Inc, Norcross, GA). Differentiated 3T3-L1 adipocytes were subjected to 4 h pred/dmso, followed by two rinsing steps and culture medium containing 0.2% FBS and Prednisolone or vehicle (1 μM) for 44 h. To prepare samples for the antibody array, samples were subjected to centrifotec tubes (Millipore amicon ultra-15 ultrace1 3 K) and washed three times with PBS, pH 8.0. Finally proteins were recovered in 2 ml PBS. Samples were biotin-labelled according to manufacturer’s protocol. Then, membranes were blocked and incubated with the biotinylated samples overnight at 4°C, and finally incubated with HRP conjugated streptavidin. Signals were detected using enhanced chemiluminescence and quantified on a LumiMager system using LumiAnalyst software version 3.1 (Roche Diagnostics, Mannheim, Germany).

**Gene expression data**

Microarray data were analysed with packages from the BioConductor library. Gene expression data was normalized using gcrma. Identification of regulated probes was done using limma package from Bioconductor. For identifying differentially expressed probes after incubation with prednisolone, the following selection criteria were used: fold change >2, intensity >20, and p value <0.05 after correction for multiple testing using the Benjamini-Hochberg correction.
All statistical analysis was performed in R. Significance of regulation of the Wnt pathway has been calculated using globaltest.

**Secreted protein data obtained from murine adipokine antibody arrays**

Regulated proteins as a result of prednisolone treatment have been selected using a p value <0.05 (calculated using a standard Student’s t-test).

**Keyword enrichment analysis**

Keyword enrichment analysis on the microarray data was performed using CoPub, a text mining algorithm that detects co-occurring biomedical concepts in abstracts from the MedLine literature database (Fleuren *et al.*, 2011), using the following settings for the threshold values: p value <0.05, R-scaled score >35 and abstract count >3. A heat map of the pathway terms was been generated using a transformation of the p value in the following way:

\[
\text{Pval}_{\text{transform}} = \frac{1}{(1 + 6 \times \exp(-0.35 \times \log_{10}(p \text{ value}))}
\]

Additional gene annotation enrichment analysis was done using the functional annotation module of Database for Annotation, Visualization and Integrated Discovery (DAVID) tool (Huang *da et al.*, 2009a, b) with default settings against the KEGG pathway database.

**Results**

**Effect of prednisolone on insulin action in 3T3-L1 adipocytes**

Exposure of differentiated 3T3-L1 adipocytes to 1 μM prednisolone had no effect on the expression of key signalling molecules, like the insulin receptor β-subunit, Akt, AS160 and GLUT4. Prednisolone significantly impaired the phosphorylation of Akt-Thr308 and Akt-Ser473 in response to insulin, whereas no effects were observed on IRS1-Tyr1222 (Figure 1). Intriguingly, but in line with a study on the effects of dexamethasone on insulin action in 3T3-L1 adipocytes (Hoehn *et al.*, 2008), prednisolone did not impact on insulin-induced phosphorylation of AS160-Thr642 (Figure 1). The mechanism(s) underlying this discordance in signalling from Akt to AS160 is not understood. Nevertheless, prednisolone was found to impair insulin-stimulated glucose uptake already after 6 h of incubation, and this inhibition was still present at 48 h after incubation (Figure 1). These findings are in line with earlier observations using dexamethasone in 3T3-L1 adipocytes (Bazuine *et al.*, 2004; Hoehn *et al.*, 2008) and confirm the induction of insulin resistance by prednisolone in our experimental conditions.

**Whole genome expression profiling**

We next studied prednisolone-induced changes in gene expression following exposure of 3T3-L1 adipocytes to prednisolone or DMSO for 1, 6, 24 and 48 h. Principle Component Analysis (PCA) showed a separation between prednisolone- and DMSO-treated samples after 6, 24 and 48 h (Supplementary figure 1). At each time-point, the significantly differentially expressed probes were identified between cells treated with prednisolone and DMSO (Supplementary Table 2). Comparison of probes significantly affected by prednisolone at 6 h with probes significantly affected by prednisolone at 48 h showed that probes with a high fold change are found to be regulated at both time points (Figure 2). Probes which are only regulated at 6 h or at 48 h have a lower fold change.

The same trend was observed when comparing prednisolone-induced probes at 6 h with prednisolone induced probes at 24 h (Figure 2B). At all time points, 25 genes represented by 34 probes (in fact 26 genes were found, but for probe 1434025_at we could not find annotation), were found to be significantly regulated by prednisolone (Table 1). Most of these genes were up-regulated and showed a sustained response to prednisolone at all time points, starting after 1 h of incubation, with the highest response after 6 h, and still present after 48 h of incubation.

**Biological pathways targeted by prednisolone**

To identify biological pathways that were affected by prednisolone, we mapped the regulated genes to biological pathway terms using the text mining algorithm CoPub (Fleuren *et al.*, 2011) (Figure 3). It appeared that the most significant pathway terms could be divided into three categories:

- immune system/inflammation (eicosanoid metabolism, prostaglandin metabolism, cytokine receptors, cytokine network, immune system, platelet activation),
- general metabolism (fatty acid metabolism, glucose metabolism, gluconeogenesis, lipid metabolism, fatty acid metabolism, glycolysis) and
- signalling (insulin signalling, TGF-β signalling, Akt signalling, Wnt signalling, MAPK signalling).

Additional functional analysis of these regulated genes using DAVID, an annotation server based on GeneOntology and KEGG pathways (Huang *da et al.*, 2009b), gave similar results (results not shown).

We are interested in mechanisms and pathways that could underlie prednisolone-induced metabolic dysfunction and more specific prednisolone-induced insulin resistance in adipocytes. Therefore for the remainder of the paper, we focused on pathways that are known to play a role in the disturbance of insulin signalling that eventually could lead to insulin resistance, i.e. Akt/insulin signalling, cytokine signalling, TGF-β signalling and MAPK signalling. Furthermore, we also focused on pathways that play a role in the disturbance of adipocyte differentiation that could lead to dyslipidaemia and obesity, i.e. Wnt signalling and cytokine signalling. In Figure 4 the prednisolone-induced genes have been categorized according to these pathways. Genes that have an effect on insulin signalling, e.g. growth-factor insulin-like growth factor 1 (Igf1), insulin receptor substrate 1 (Irs1) and platelet derived growth factor receptor, beta polypeptide (Pdgfrb) and genes from the TGF-β pathway such as MAD homolog 3 (Drosophila) (Smad3), gremlin 1 (Grem1), transforming growth factor, beta 2 (Tgfb2) and transforming growth factor, beta induced (Tgfb1) were down-regulated.
Figure 1. Effect of prednisolone on insulin action in 3T3-L1 adipocyte cells. Effects on protein expression of the insulin receptor (IR) β-subunit, Akt, AS160, and GLUT4, as well as the phosphorylation of insulin receptor substrate 1 (IRS1-Tyr1222), Akt-Thr308, Akt-Ser473, and AS160-Thr642 were determined by Western blotting in cells exposed to 1 μM prednisolone for 48 h prior to stimulation with insulin (10 min, 100 nM). Data are presented as mean ± standard error of the mean of 4 independent experiments (A–D), and representative Western blots (F). E. Effect of prednisolone on 2-deoxyglucose uptake in 3T3-L1 adipocytes after 6, 24 and 48 h of incubation with prednisolone. Data are mean ± standard error of the mean of 4 independent experiments. In all bar graphs, open columns represent the basal condition, and filled bars depict insulin-stimulated cells. The effects of prednisolone on insulin action were analysed using a two-way ANOVA followed by Bonferroni analysis for multiple comparisons. ***Indicates a $p < 0.001$; **$p < 0.01$. Effects of insulin versus basal were analysed using a Student’s t-test. ###$p < 0.001$; ##$p < 0.01$; #$p < 0.05$. DOI: 10.3109/13813455.2013.774022

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(A) pIRS1-Tyr1222 (AU)

(B) pAkt-Thr308 (AU)

(C) pAkt-Ser473 (AU)

(D) pAS160-Thr642 (AU)

(E) Glucose uptake (AU)

(F) Western blots

- IRβ-subunit
- pIRS1-Tyr122
- pAkt-Thr308
- pAkt-Ser473
- tAkt
- pAS160-Thr642
- tAS160
- GLUT4
- β-actin
Phosphatase and tensin homolog (Pten) an important regulator of the Akt pathway was up-regulated. The majority of the genes coding for cytokines, such as Interleukin-6 (IL-6), chemokine (C-X-C motif) ligand 5 (CXCL5), chemokine (C-X-C motif) ligand 10 (CXCL10) and chemokine (C-C motif) ligand 7 (CCL7), were down regulated by prednisolone. Other cytokines such as interleukin 1 receptor, type I (IL1r1) and interleukin 1 receptor accessory protein (IL1rap) were up regulated.

Many Wnt signalling pathway members were significant down regulated by prednisolone in our study. We observed down regulation of frizzled receptors Fzd1, Fzd2, Fzd4, Fzd5 together with ligand wingless-related MMTV integration site 5A (Wnt5A) and antagonist secreted frizzled-related protein 2 (Sfrp2). Also several down-stream molecules such as Wisp1, Wisp2, Id2 were down regulated (Figure 4).

Regulation of adipokines measured after 48 h of incubation with prednisolone

The microarray data showed several transcripts that encode hormones secreted by adipocytes, like cytokines, and regulators of the Wnt, Akt and TGF-β pathway. We therefore decided to study to what extend the altered levels of gene expression would lead to changes in protein expression, by analyzing the effects of prednisolone on adipokine secretion by 3T3-L1 adipocytes treated with DMSO or prednisolone for 48 h. For this purpose, the supernatants of the 3T3-L1 adipocytes that were used in the gene expression study were analyzed. Out of the 308 proteins on the array, 25 proteins were significant regulated by prednisolone (Table 2, p value <0.05. For all proteins, see Supplementary Table 1). The majority of these proteins are cytokines involved in inflammatory and immune response related processes (indicated with a purple background in Table 2). Cytokines such as CXCL2, CCL3, CCL9, IL31 were down regulated by prednisolone while CCR6, IL12B, TNFSF8, TNFRSF7 and TNFRSF17 showed an up regulation.

Also several components of the Wnt signalling pathway were significantly regulated at protein level (Table 2, indicated with a blue background). DKK4 an inhibitor of the Wnt signalling pathways was up-regulated. Furthermore downstream regulators of the Wnt signalling pathway, LRP6 and WISP1 are down regulated. Additionally, initiation of insulin signalling is represented by the down regulation of IGFBP5, IGFBP3 and Resistin.

These results indicate that the regulation of the Akt/insulin signalling, cytokine signalling and Wnt signalling pathways by prednisolone is not only found on the transcriptome level, but is also translated into altered proteins levels in these pathways.

Regulation of the Wnt signalling pathway by prednisolone

Individual Wnt signalling components were found to be significantly regulated by prednisolone at both gene and protein levels. Therefore, we next investigated whether the entire Wnt signalling pathway was affected by prednisolone. The KEGG database was used to select genes upstream in the canonical Wnt signalling pathway that are more exclusively linked to the Wnt signalling in comparison with down-stream genes like Smad3, Lef, Tak1 and Pparδ, which are also linked to TGF-β signalling, MAPK signalling and Cell cycle (Supplementary Table 3). We used the expression profiles of these 51 Wnt signalling genes to test whether there is a difference in expression for the entire Wnt signalling pathway between prednisolone and DMSO treated samples. In this method, genes that do not meet the FC ≥2-fold and Pval <0.05 cut offs but may have nevertheless a changed expression level were taken into consideration. A significant difference in expression for the entire Wnt signalling pathway was observed after 6 h (p value = 0.031). In Figure 5, the
Table 1. Regulated probes (genes) by prednisolone after 1, 6, 24 and 48 hours of incubation. For each probe the log fold change at each time point is shown. Probes that are up-regulated are indicated with a red color and probes that are down-regulation are indicated with a green color.

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genes that had the most influence on this difference are shown. In this figure, the gene expression ratios (open dots) and protein ratios (black dots) as a result of prednisolone treatment are also shown. Genes that had an influence on the difference between DMSO and prednisolone treated samples and that showed a higher expression in DMSO treated samples (red bars) were down-regulated because of prednisolone treatment (open dots). The same effect on the Wnt signalling pathway was also seen after 48 h ($p$ value = 0.064) of treatment (Supplementary Figure 3). This analysis indicates that in 3T3-L1 adipocytes, the entire Wnt signalling pathway is down-regulated by prednisolone.

**Discussion and conclusions**

The prolonged use of glucocorticoids is hampered by metabolic side-effects such as insulin resistance, which eventually leads to diabetes and obesity. Adipose tissue is thought to play an important role in GC-induced development of metabolic side effects. The mechanisms behind these side-effects are not completely understood. Key findings of our study indicate that prednisolone reduces insulin stimulated glucose uptake in 3T3-L1 cells, which is reflected by changes in expression of genes and proteins from the Akt/insulin, cytokine and Wnt signalling pathways. To our knowledge, this is the first work that uses a combination of gene expression and antibody arrays to study effects of prednisolone in adipocyte cells.

The set of 25 genes that showed regulation by prednisolone at all time points includes known target genes of the glucocorticoid receptor, such as *Ddit4*, *Dusp1* and *Cebpd* (Toonen et al., 2011; Yang et al., 2005), and genes such as *Fam107a* and *PDK4*, that were also found to be regulated by prednisolone in *in vivo* studies of mouse and human (Bastard et al., 2006; Frijters et al., 2010; Toonen et al., 2011). This indicates that prednisolone acts via its target genes and initiates a GC specific response in these 3T3-L1 cells.

Multiple cytokine molecules were regulated by prednisolone. Increasing evidence suggests that a chronic low-grade state of inflammation in adipose tissue contributes to the development of systemic insulin resistance and diabetes (Bastard et al., 2006; Harford et al., 2011; Heilbronn & Campbell, 2008). In the absence of IL1, mice are protected against high fat diet induced insulin resistance which is accompanied with a reduction in local adipose tissue inflammation (de Roos et al., 2009; McGillicuddy et al., 2011). Our results showed up-regulation of IL1 signalling by means of IL1r1 and IL1rap. This suggests a role of IL-1 signalling in GC induced development of insulin resistance in 3T3-L1 cells.

Also other studies reported about the up-regulation of cytokines such as *Il6*, *Cxc15*, *Ccl2* and *Cxc110* in insulin resistance and obesity (Figure 6) (Bastard et al., 2006; Chavey et al., 2009; Lagathu et al., 2003; Murdolo et al., 2008; Rotter et al., 2003; Tateya et al., 2010). In contrast to...
Figure 4. Prednisolone induced genes after 1 h (T1), 6 h (T6), 24 h (T24), 48 h (T48). Prednisolone-induced up-regulated genes are shown in red, prednisolone induced down-regulated genes are shown in green. For clarity genes have been assigned to major pathways. However, one should keep in mind that genes can belong to multiple pathways depending on the definition of a pathway.
Table 2. Regulated proteins after 48 hours of incubation with prednisolone. Inflammatory related proteins are indicated with a purple background, Wnt signaling related proteins with a blue background and insulin signaling related proteins with a green background. Up-regulated proteins are indicated with a red color. Down-regulated proteins are indicated with a green color.

<table>
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<tr>
<th>Protein Symbol</th>
<th>Protein Name</th>
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these studies, we observed a down regulation of expression of these genes by prednisolone. Based on these results it is difficult to deduce whether prednisolone-induced regulation of cytokines in adipocytes contribute to the development of insulin resistance or are the result of the immune suppressing properties of prednisolone or a combination of both.

Furthermore our results indicated that the reduction in insulin stimulated glucose uptake by prednisolone acts partly via the down regulation of insulin signalling. This regulation might be mediated by Pten, a known suppressor of insulin signalling that acts via the PI3K/Akt signalling pathway (Tang et al., 2005). Additional studies confirm this observation and suggest an important role of Pten in the development of insulin resistance and diabetes (Ikubo et al., 2009; Lazar & Saltiel, 2006; Nakashima et al., 2000).

Again an apparent contrasting effect of prednisolone is the down regulation of proteins like Igfb3 and resistin that reduce glucose uptake in adipose tissue (Chan et al., 2005; Kim et al., 2007).

Inhibition of TGF-β/Smad3 signalling results in diminished adiposity, improved glucose tolerance and insulin sensitivity and in the protection from insulin resistance, diabetes and obesity (Tan et al., 2011; Tsurutani et al., 2011; Yadav et al., 2011). The observed down regulation of Smad3 suggests that disturbance of glucose uptake and adiposity by prednisolone in 3T3-L1 cells acts via additional mechanisms.

Activation of the Wnt signalling pathway has been linked to inhibition of adipogenesis (Bennett et al., 2002; Christodoulides et al., 2006; Longo et al., 2002; Ouchi et al., 2010).

Recent studies toward the characterization of the adipocyte secretome have shown that multiple regulators of the activity the Wnt signalling are secreted by adipocytes, such as WISP2 and SFRP5 (Leht et al., 2012; Ouchi et al., 2010). Interestingly, expression and circulating levels of these factors are altered in insulin resistance and obesity (Dahlman et al., 2012; Hu et al., 2012; Mori et al., 2012; Ouchi et al., 2010), underlying the suggestion that a dysregulated secretion of Wnt regulators could lead to metabolic disturbances and eventually into the development of obesity and diabetes (Ouchi et al., 2010; Jin, 2008; Liu & Habener, 2010; Oh & Olefsky, 2010; Schinner, 2009). Yet, in the context of our study, it remains to be investigated to what extent the factors affected by prednisolone interfere with insulin action in adipocytes.

Prednisolone induced Pten, Smad3 and Lef1 which are able to mediate cross-talk between the signalling pathways (Guo & Wang, 2009). Via the alteration of the Pten function, TGF-β is able to influence the Akt activity, and via Smad3 the Akt pathway is able to restrict the TGF-β pathway. Also Smad activity is involved in the cross-talk between Wnt signalling and TGF-β signalling, as part of the Smad/β-catenin/Lef protein complex in the nucleus. These genes could play a central role in the development of prednisolone induced metabolic effects because of their ability to connect multiple pathways (Figure 6). The sometimes opposing effects of prednisolone on individual pathway members of Smad3 and Akt in comparison with earlier work, requires additional in vivo studies in which the systemic effect of prednisolone on these pathways could be determined.

Figure 5. Wnt signalling genes that have an influence on the difference between prednisolone and DMSO, after 6h of incubation. Genes that are up-regulated in DMSO treated samples are shown in red and genes that are up-regulated in prednisolone treated samples are shown in green. Ratios of prednisolone-induced genes are indicated with open dots. Prednisolone-induced ratios of the proteins are indicated with black dots. Ratios are indicated in log2 fold change.
To our knowledge, for the first time it is shown that prednisolone affects the expression and secretion of Wnt regulators in adipocytes. The fact that Wnt signalling also participates in the development of metabolic disturbances in other tissues (Abiola et al., 2009; Anagnostou & Shepherd, 2008) may direct future work towards a dedicated study of the Wnt signalling pathway in GC-induced metabolic effects. In an in vivo setting the Wnt signalling mediated GC effects in multiple glucose responsive tissues such as muscle tissue, liver tissue and adipose tissue could be determined. Hence new GC compounds with an improved efficacy/side effects ratio should have a reduced effect on components in the Wnt signalling pathway.

Declaration of interest
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Bastard JP, Maachi M, Lagathu C, et al. (2006). Recent advances in the relationship between obesity, inflammation, and insulin resistance. Eur Cytokine Netw, 17:4–12.


