INTRODUCTION

The clinical course of dengue virus infection ranges from asymptomatic infection to severe disease, known as dengue hemorrhagic fever/dengue shock syndrome (DHF/DSS). The latter is characterized by a transient endothelial hyper-permeability, of which the pathogenesis is still incompletely understood.\(^1\)\(^,\)\(^2\)

Inflammatory cytokines and angiogenic proteins are important mediators of vascular integrity.\(^3\) The angiogenic protein vascular endothelial growth factor (VEGF) is a strong inducer of vascular permeability and several studies have reported circulating VEGF levels in dengue patients with contradicting results.\(^4\)\(^–\)\(^8\) However, the role of another class of angiogenic proteins, known as angiopoietins, is unknown in dengue.

Angiopoietin-1 (Ang-1) and angiopoietin-2 (Ang-2) and their endothelial tyrosine kinase receptor Tie-2 form a central signaling system in endothelial permeability.\(^9\) Angiopoietin-1–mediated Tie2 activation maintains the quiescent state of the endothelium by stabilizing endothelial cell-cell junctions and by countering the permeabilizing effects of VEGF.\(^10\) Angiopoietin-2 antagonizes the effects of Ang-1; it stabilizes the endothelium by disrupting cell-cell adhesion and primes the endothelial cells to the effects of pro-inflammatory cytokines and VEGF.\(^11\) Angiopoietin-2 is almost exclusively produced in endothelial cells and stored in Weibel-Palade bodies (WPBs), from which it can be rapidly released upon activation of the endothelium.\(^12\) Angiopoietin-1 is produced in pericytes and smooth muscle cells, but platelets also contain high quantities of Ang-1.\(^13\) Thus, the number and activation status of circulating platelets may influence plasma Ang-1 levels. Evidence is increasing that platelets are important cells for maintaining vascular stability and platelet-derived Ang-1 may be one of the factors involved.\(^14\)

Dengue hemorrhagic fever/dengue shock syndrome is associated with thrombocytopenia, inflammation, and endothelial cell activation, and these processes might lead to significant alterations in the balance between Ang-1 and Ang-2, favoring plasma leakage and hemorrhage.\(^15\) We therefore studied Ang-1 and Ang-2 levels children with DHF/DSS and related these levels to markers of plasma leakage.

METHODS

This study was part of a larger cohort study that investigated pathophysiologic mechanisms of severe dengue. Consecutive children with DHF/DSS \(\leq\) 15 years of age who were admitted to the pediatric ward or intensive care unit of the Dr. Kariadi University Hospital in Semarang, Indonesia, during 2005–2006 were enrolled in the study. Study details and clinical characteristics have been reported.\(^16\) All patients had positive results for dengue-specific IgM, as determined by enzyme-linked immunosorbent assay (Focus Technologies, Chanhassen, MN). The Ethics Committee of Diponegoro University in Indonesia approved the study. Written informed consent was obtained from the parents or legal guardians of the children.

Forty-nine children, for whom sufficient blood sample volume was available, were randomly selected from this larger cohort and classified as DHF or DSS according to World Health Organization criteria.\(^17\) Twenty-five healthy children from Indonesia served as controls. Demographic and clinical characteristics of study participants are shown in Table 1.

Fluid resuscitation was performed in all patients according to World Health Organization–based protocols. Blood was drawn the days of enrollment and of discharge by using a syringe and directly transferred to vacutainers containing sodium citrate as an anticoagulant. Sodium citrate was used because other anticoagulants or serum result in platelet activation and release of platelet-derived molecules such as Ang-1.
Short application of a tourniquet for blood drawing was often inevitable. Blood samples were processed as soon as possible and handled with care to avoid in vitro platelet activation. Blood was centrifuged for 20 minutes at 1,600 × g, and plasma was stored at −80°C. Plasma levels of Ang-1, Ang-2, and P-selectin were measured by using commercial enzyme-linked immunosorbent assay kits (R&D Systems, Minneapolis, MN).

### RESULTS

Patients with DHF/DSS had lower Ang-1 levels at enrollment compared with corresponding discharge levels and levels in controls, and had the lowest median level in children with DSS (Figure 1A). Nineteen patients had an Ang-1 level below the detection limit of 0.1 ng/mL at enrollment. In contrast, children with DHF/DSS had higher Ang-2 levels at enrollment than at discharge and compared with healthy controls (Figure 1B). Taken together, DHF/DSS was associated with a clear increase in Ang-2/Ang-1 ratios, and those with DSS had the highest ratios (Figure 1C). Five children died and although their Ang-1 levels were in the lowest range, their corresponding Ang-2 levels were more evenly distributed (Figure 1). There was no difference in Ang-1 and Ang-2 levels between children with and without bleeding.

The association of Ang-1 and Ang-2 levels with plasma leakage was determined by using serum albumin level and the pleural effusion index (PEI). This index was defined as 100 times the maximum width of the (right or left) pleural effusion on a chest radiograph (right lateral decubitus position) divided by the maximum width of the ipsilateral hemithorax. The Ang-1 levels at enrollment correlated positively with albumin levels (Spearman R \( R_s = 0.55, P < 0.0001 \)) and negatively with the PEI (\( R_s = -0.39, P = 0.005 \)). The Ang-2 levels correlated inversely with serum albumin \( R_s = -0.38; P = 0.009 \) but not with the PEI. The Ang-2:Ang-1 ratio was related with the PEI (\( R_s = 0.43, P = 0.002 \)) and albumin levels (\( R_s = -0.60, P < 0.0001 \)). The Ang-1 or Ang-2 levels did not correlate significantly with hematocrit values. Although platelet numbers correlated with serum albumin levels (\( R_s = 0.44, P < 0.05 \)), there was no significant correlation with the PEI (\( R_s = -0.26, P = 0.07 \)).

As mentioned, platelets contain high quantities of Ang-1 and release Ang-1 during platelet activation.\(^{13}\) The Ang-1 levels correlated with the platelet count (\( R_s = 0.44, P < 0.002 \)). Plasma levels of platelet activation marker P-selectin followed the trend of Ang-1 levels and showed a low median level at admission (45.3 ng/mL; interquartile range = 29.4–66.3 ng/mL) that had increased by the day of discharge (120.7 ng/mL; interquartile range = 91.6–187.5 ng/mL). Nonetheless, Ang-1 levels only weakly correlated with P-selectin levels (\( R_s = 0.30, P < 0.04 \)) at enrollment, but when the 19 samples with Ang-1 levels below the detection limit were not analyzed, this correlation improved considerably (\( R_s = 0.69, P < 0.0001 \)).

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**Table 1**

 Demographic and clinical characteristics of the patients studied†

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>DHF (DHF 1 and II), ( n = 26 ) (55%)</th>
<th>DSS (DHF III and IV), ( n = 23 ) (47%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male sex</td>
<td>12 (46)</td>
<td>7 (30)</td>
</tr>
<tr>
<td>Age, years</td>
<td>8 (6–10)</td>
<td>8 (6–10)</td>
</tr>
<tr>
<td>Body weight, kg</td>
<td>20 (17–28)</td>
<td>24 (18–30)</td>
</tr>
<tr>
<td>Body height, cm</td>
<td>125 (111–135)</td>
<td>120 (113–140)</td>
</tr>
<tr>
<td>Body temperature, °C</td>
<td>37.6 (37.0–38.4)</td>
<td>38.0 (37.0–38.5)</td>
</tr>
<tr>
<td>Duration of fever until admission, days</td>
<td>4.0 (3.0–5.0)</td>
<td>4.0 (4.0–5.0)</td>
</tr>
<tr>
<td>Tourniquet test positive</td>
<td>17/23 (74)</td>
<td>10/16 (63)</td>
</tr>
<tr>
<td>Petechiae</td>
<td>1/17 (6)</td>
<td>1/15 (7)</td>
</tr>
<tr>
<td>Epistaxis</td>
<td>3 (12)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Gum bleeding</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Hematemesis</td>
<td>2 (8)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Melena</td>
<td>0 (0)</td>
<td>1 (4)</td>
</tr>
<tr>
<td>Hemoglobin, g/dL</td>
<td>13.1 (12.7–14.0)</td>
<td>13.5 (12.3–14.2)</td>
</tr>
<tr>
<td>Hematocrit, %</td>
<td>38.0 (37.9–41.3)</td>
<td>40.0 (36.0–43.5)</td>
</tr>
<tr>
<td>Platelet count ( \times 10^9/L )</td>
<td>62 (39–88)</td>
<td>39 (26–68)</td>
</tr>
<tr>
<td>Leukocyte count, ( \times 10^3 ) cells/mL</td>
<td>5.0 (3.1–7.8)</td>
<td>5.0 (3.0–9.5)</td>
</tr>
<tr>
<td>Albumin serum, g/dL</td>
<td>3.3 (2.9–3.8)</td>
<td>2.8 (2.5–3.4)†</td>
</tr>
<tr>
<td>Alanine aminotransferase, U/dL</td>
<td>48 (31–82)</td>
<td>46 (32–71)</td>
</tr>
<tr>
<td>Pleural effusion index</td>
<td>8 (0–22)</td>
<td>18 (11–28)†</td>
</tr>
</tbody>
</table>

*Values are medians (interquartile ranges) or absolute no. (%). All data are data at enrollment, except for bleeding manifestations, which are presented for the whole duration of admission. DHF = dengue hemorrhagic fever; DSS = dengue shock syndrome.

†\( P < 0.05 \), by Mann-Whitney U test.

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**Figure 1.** Plasma levels of (A) angiopoietin-1 and (B) angiopoietin-2 and corresponding angiopoietin-2:angiopoietin-1 ratios (C) in children in Indonesia with dengue hemorrhagic fever (DHF) \( n = 26 \) and dengue shock syndrome (DSS) \( n = 23 \) on day 0 (day of admission), day of discharge \( n = 44 \) and in a control group of healthy children from Indonesia \( n = 25 \). The minimum detection limit of angiopoietin-1 was 0.1 ng/mL. Black points indicate non-survivors. Horizontal lines represent median values. \( P \) values were determined by Wilcoxon signed-rank test for data in time and Mann-Whitney U test for comparison with the control group. *\( P < 0.05 \).
DISCUSSION

This study shows that DHF/DSS is associated with reduced Ang-1 plasma levels and increased Ang-2 levels. These proteins and their endothelial receptor are important mediators of vascular integrity, and we speculate that this imbalance in the Ang/Tie system contributes to the transient increase in vascular permeability. Platelets contain Ang-1, and we hypothesize that dengue-associated thrombocytopenia explains the low Ang-1 levels. Increasing evidence suggests a role for platelets in regulation of vascular integrity with Ang-1 and other platelet-derived angiogenic proteins as central mediators.14 As suggested by others, platelet count and activation status should be taken into account when plasma levels of platelet-derived proteins are measured.18,19

Angiopoietin-2 is produced in endothelial cells and stored in WPBs.12 The finding of high levels of von Willebrand factor, the most abundant WPB constituent, in dengue patients, supports the notion that endothelial cell activation with WPB exocytosis is an important feature of dengue infection.15,20 Although the exact mechanisms of increased WPB exocytosis with subsequent Ang-2 release in dengue are unknown, we hypothesize that endothelial activation caused by increased pro-inflammatory cytokines is centrally involved.21 In addition to the effect of inflammatory cytokines, other mechanisms may also be involved, including direct interaction of dengue virus with endothelial cells, release of mast cell products, and pro-coagulant factors such as thrombin.21–28

No definite conclusion on a causal relationship of Ang-1 and Ang-2 levels and plasma leakage could be made in this observational study. Similar trends in Ang-1 and Ang-2 levels have been found in severe malaria and sepsis,26,27 but in contrast to DHF/DSS and sepsis, plasma leakage is not a prominent feature of malaria. Differences in other mediators of vascular permeability, such as pro-inflammatory cytokines and angiogenic proteins and their soluble receptors, might account for these differences in vascular permeability.

Our study had some limitations. First, no values for the platelet count at discharge and in the control group were available. Second, despite our efforts to limit ex vivo platelet activation, some degree of activation is often inevitable and this should be taken into account when interpreting plasma Ang-1 and P-selectin levels. Angiopoietin-1 levels across different studies seem to vary more than Ang-2 levels in controls.16,28–31 Angiopoietin-1 levels in the controls of our study were comparable with those of other studies, which also found relatively higher Ang-1 levels.29,30 Therefore, we suggest that relative changes in Ang-1 levels within one study may be more informative than comparing absolute Ang-1 levels between studies. Third, the dengue diagnosis was based on the presence of dengue-specific IgM in a single sample. False-positive IgM results caused by cross-reactivity may occur in patients with infectious diseases other than dengue, although the fact that all children in our study population had proven plasma leakage (pleural effusion on chest radiograph) around defervescence, a finding specific for dengue, makes alternative diagnoses unlikely.

In conclusion, DHF/DSS is associated with an imbalance in the Ang/Tie system favoring plasma leakage. Dengue-associated thrombocytopenia may play a so far overlooked role in the transient plasma leakage seen in DHF/DSS. No specific treatment for dengue is available. Interventions aimed at the prevention of thrombocytopenia or the correction of the imbalance in the Ang/Tie system might offer a valuable adjunctive treatment of this devastating disease.

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