Validation of smoking cessation self-reported by patients with chronic obstructive pulmonary disease

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Purpose: The present study reports on the biochemical validation of the self-reported smoking status of patients with chronic obstructive pulmonary disease (COPD). The objective is to establish the proportion of overestimation of self-reported success rates.

Methods: A cross-sectional smoking-status validation study including 60 patients with COPD who reported that they had stopped smoking. In the analysis of urine samples, a cut-off point of 50 ng/mL of cotinine was used.

Results: At the time of biochemical validation, 55 patients reported that they had quit smoking while five patients resumed smoking. Smoking status was biochemically confirmed for 43 patients (78%) and 12 patients (22%) were classified as smokers. The sensitivity of the self-report of smoking was 29% and the specificity was 100%.

Conclusion: Many primary care patients with COPD do not provide valid information on their smoking status, which hampers adequate therapeutic interventions. Integration of biochemical validation in daily care could overcome this problem, but may harm the doctor–patient relationship.

Keywords: chronic obstructive pulmonary disease, smoking cessation, biochemical validation, general practice, outcome measurement

Introduction

Smoking cessation interventions provided in general practice have proven to be effective.¹,² The role of general practitioners (GPs) in supporting their patients who want to quit smoking is documented in international guidelines.³ Addressing the smoking status of patients can be difficult, but GPs are more likely to do so if their patient suffers from smoking-related complaints.⁴ The GPs’ advice and support encourages patients to quit smoking. How do physicians learn whether their support was successful? Are they able to rely on the patients’ self-reports?

Chronic obstructive pulmonary disease (COPD) is illustrative of a smoking-related disease. This slowly progressive disease is characterized by airflow obstruction with related symptoms such as cough, sputum production, dyspnea, and wheeze and it is mainly caused by smoking.⁵-⁷ Among patients with COPD, smoking prevalence varies from 38%–52% in seven countries.⁸ Because smoking accounts for 80%–90% of COPD, one may assume that many patients find it difficult to quit smoking. Smoking cessation slows down the rate of deterioration of lung function, improves COPD prognosis, and may prolong life expectancy for these patients.⁹-¹² Consequently, smoking cessation is the most important element of therapy for COPD,¹³ and it is necessary to measure the results of smoking cessation interventions during the therapeutic process.
The self-report of smoking cessation in community surveys among the general population is reasonably valid. For smokers specially at risk (eg, coronary disease, COPD, pregnancy), the self-reported smoking status is less reliable. Biochemical testing is recommended for specific populations where quitting is very desirable. Cotinine, the major proximate metabolite of nicotine, has been widely used as a biomarker of tobacco exposure. The relatively long half-life of cotinine (16–20 hours) enables detection for a few days after cessation of tobacco use. The sensitivity and specificity of cotinine are high, 97% and 99%–100%, respectively, and it is therefore the preferred substance to measure.

Although the rationale for biochemical validation of the smoking status of patients with increased health risks is evident, the Lung Health Study found little discrepancy between self-reports and cotinine measurement of patients with early-staged COPD. Nevertheless, the patients who had received smoking cessation support misreported their smoking status more often than patients who had not (6% versus 1%). This may suggest that participation in a smoking cessation program increases the social desirability bias. Participation of COPD patients in an intensive smoking cessation program had even higher rates (52%) of misreporting after biochemical testing of their smoking status. Smoking cessation interventions, as carried out in general practice during routine follow-up, are generally less intensive than these programs. The question is, then, how accurate are the self-reports of COPD patients if they have been subjected to a less intensive smoking cessation intervention in the patients’ own general practice? As smoking cessation support is recommended in routine general practice for patients with COPD, it is important for GPs to obtain valid information about their patients’ smoking status. This is necessary to use smoking cessation programs effectively and to adjust the medical therapy to their results. Since the literature is inconclusive about the extent of misreporting of the smoking status of COPD smokers in general practice, the present study examines the validity of self-reported smoking status of COPD patients participating in a study testing a moderately intensive smoking cessation program in daily general practice.

Methods and materials
The present study is a validation study of self-reported smoking status assessed by urine cotinine analysis. The data were collected from all patients who reported that they had not smoked in the past 7 days.

Subjects and procedure
A software program using Anatomical Therapeutical Chemical (ATC) prescription codes and International Classification of Primary Care (ICPC) diagnosis codes selected potential patients with COPD from general practices. The criteria included age of 35 years or more and a diagnosis recorded as COPD, or as ICPC code R95/96, or at least three prescriptions of bronchodilators (ATC code R03a/bc), and/or at least two prescriptions of inhaled anti-inflammatory medication in the last year (ATC code R03). The GPs had to confirm the diagnosis of the selected patients. Patients were eligible to participate if they were: 1) currently smoking; 2) suffering from COPD according to the GP’s diagnosis; 3) not being treated by a chest physician; 4) in command of the Dutch language; and 5) not suffering from any serious physical or psychological co-morbidity. The participants consented to participate in an intervention study in general practice and were allocated to one of the three study groups (including a control group). The patient-directed intervention tailored to general practice patients with COPD was based on the minimal intervention strategy. GPs applied a stage-based protocol, providing information, smoking cessation counseling, and advised the use of pharmacological cessation aids (ie, nicotine replacement therapy or bupropion-SR). The patient education tools consisted of a leaflet especially developed for COPD smokers and a videotape. The patients included in the present study self-reported to have quit smoking during the 12-month follow-up. More information about patient inclusion and the smoking cessation program can be found elsewhere. Figure 1 shows that of the 667 enrolled patients, 86 patients said at the 12-month follow-up that they were not smoking: however, 22 patients (26%) did not enter the present study (17 patients refused and five patients were too ill), three patients were excluded because of their current nicotine replacement therapy, and one patient was excluded due to missing data. Hence, our sample consisted of 60 patients. The procedures followed were approved by the Committee on Human Experimentation, Maastricht University, The Netherlands.

Measurement
The participants were briefed at the beginning of the study (1 year prior to the assessment of their smoking status) about possible self-report crosschecks with biochemical validation. They received a baseline questionnaire (Q0) to complete at home to provide the baseline characteristics of the sample including their motivation to quit smoking, severity of nicotine dependence (Fagerström Test of Nicotine
Dependence), and COPD symptoms (Medical Research Council). Twelve months later, they received another questionnaire (Q1) regarding their smoking status (7-day point prevalence). The patients who reported that they had quit smoking were included in the validation study and received a letter in which they were asked to visit the general practice within a week to produce and hand over a urine sample to be sent to the researchers for analysis. If they were unable to visit the practice, a research assistant collected the sample during a home visit. Furthermore, they were asked to complete an additional short questionnaire at the exact time of the sample production (Q2), which included their current smoking behavior and use of nicotine substitution. All questionnaires were self-administered and were used for

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**Figure 1** Flow of the biochemical validation study.

**Notes:** *Self-report, Biochemically verified, Including the 5 self-reported resumed smokers.*
research only. The results of the biochemical validation of the self-reported smoking status were not communicated to the patients’ GPs.

The amount of urinary cotinine was measured by radioimmunoassay. Radioimmunoassay involves the use of antibodies to cotinine. The double-antibody technique was used to separate free-labeled antigen from antibody-bound antigen. A reaction mixture containing 0.1 mL 3H-cotinine, appropriately diluted antisera and buffer (0.14 M NaCl, 0.01 M Tris-HCl, and 0.1% gelatin) was incubated at 37°C. For inhibition experiments, 0.1 mL-aliquots containing known amounts of standard or dilution of the urine-samples were added in place of buffer. Goat anti-rabbit immunoglobulin was added and the mixture was incubated at 4°C overnight. To control for nonspecific binding, normal rabbit serum was used in place of immune serum. The precipitate was collected by centrifugation at 4200 rpm for 45 minutes at 4°C, the supernatant decanted, and the walls of the tubes wiped dry. To count the 3H-cotinine, the precipitate was dissolved in 0.1 mL 0.1 N sodium hydroxide before adding 2.5 mL scintillation fluid. Eventually the rate of radioactivity was counted and the amount of cotinine could be calculated from the linear portion of the standard curve. The results were reported in nanograms per milliliter.

Analyses
The cut-off point for both smokers and nonsmokers was 50 ng/mL.21 We used the contingency coefficient (two-tailed test) to analyze the association of the self-reported status and the biochemically confirmed smoking status. Moreover, the sensitivity and specificity of the self-report were calculated. Differences in cotinine levels were analyzed by means of the Mann–Whitney U test (two-tailed). The data were analyzed with SPSS software (v. 14.0; SPSS Inc., Chicago, IL).

Results
Table 1 shows the baseline characteristics (Q0) of the 60 participants. Seven patients were allocated to the control group. The sample included 59% men, and the participants had a mean age of almost 60 years. At baseline, 38% indicated they wanted to quit smoking within 1 month, 34% wanted to quit within 6 months, and 28% were not motivated to quit smoking. Thirty percent suffered from chronic sputum production and 35% from chronic coughing.

Forty-three patients (72%) of the 60 patients were classified by the urine cotinine test as nonsmokers (median cotinine level 13.0 ng/mL, quartile range 6.0–22.0), and 17 patients (28%) were classified as smokers (median cotinine level 134.0 ng/mL, quartile range 68.0–194.0) (z=6.00; P<0.001).

During the time of the biochemical validation, 12 of the 17 patients, who were classified as smokers, reported that they did not smoke (11 had taken part in the smoking cessation intervention) and five said that they had smoked recently (three had attended the intervention). Thus, the self-reporting of 43 (78%) of the 55 patients who claimed that they were not smoking at the time of taking the urine sample (Q2) corresponded with the classification by the cotinine level and 12 (22%) patients did not correspond (see Table 2) (contingency coefficient 0.43; P<0.001). The self-report had 29% sensitivity for detecting actual smoking, and a specificity of 100%.

Discussion
We found a misreport rate of 22%. For COPD patients both lower24 and higher20 misreport rates have been described. It is important to bear in mind that the self-reporting of smoking status was combined with providing a urine sample for biochemical validation. This combination might result in a lower misreport rate, because the patients are aware that their self-reports will be checked. If this is true, then the misreporting of 22% could be an underestimation. If the patients who refused to be tested (n=22) were still smoking, then the misreport rate could be as high as 42%. The real misreporting

### Table 1 Characteristics of 60 self-reported quitters

<table>
<thead>
<tr>
<th>Variable</th>
<th>Sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>Men (%)</td>
<td>59</td>
</tr>
<tr>
<td>Mean age (sD)</td>
<td>59.9 (11.0)</td>
</tr>
<tr>
<td>Stages of change (%)</td>
<td></td>
</tr>
<tr>
<td>Preparer</td>
<td>38</td>
</tr>
<tr>
<td>Contemplator</td>
<td>34</td>
</tr>
<tr>
<td>Precontemplator</td>
<td>28</td>
</tr>
<tr>
<td>Mean number of cigarettes per day (SD)</td>
<td>16.4 (8.2)</td>
</tr>
<tr>
<td>Mean FTND (SD)</td>
<td>4.2 (2.4)</td>
</tr>
<tr>
<td>Chronic sputum (%)</td>
<td>30</td>
</tr>
<tr>
<td>Chronic coughing (%)</td>
<td>35</td>
</tr>
<tr>
<td>Mean dyspnea, range 0–3 (SD)</td>
<td>1.3 (1.1)</td>
</tr>
</tbody>
</table>

Note: Percentages are rounded.

Abbreviations: FTND, Fagerström Test of Nicotine Dependence; SD, standard deviation.

### Table 2 Association between self-reported quit rates and biochemically validated quit rates

<table>
<thead>
<tr>
<th>Biochemical validation</th>
<th>Self-report (questionnaire 2)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Not smoking</td>
</tr>
<tr>
<td>Not smoking</td>
<td>43</td>
</tr>
<tr>
<td>Smoking</td>
<td>12</td>
</tr>
</tbody>
</table>
figures will probably lie somewhere between these figures. The low sensitivity of self-reports (29%), even in a situation where the report may be biochemically verified, shows their restricted value in clinical smoking cessation programs for high-risk populations.

There may be several explanations for the difference between self-reported and biochemically determined smoking status. First, the time elapsed between the two measurements (Q1 and Q2) might be a factor. One of the reasons for the great percentage of misreports might be that patients had relapsed to smoking after Q1. We tried to overcome this problem by reassessing the smoking status (Q2), which showed that some of the COPD patients had indeed resumed smoking. Second, the results did not account for the influence of environmental tobacco smoke, which is known to influence cotinine levels. However, since our cut-off point was 50 ng/mL this is not likely. Third, misreporting might be triggered by the fact that patients participated in a smoking cessation intervention. Of the 12 patients who misreported their smoking status, 11 participated in the intervention. Interestingly, while Monninkhof et al report a misreport rate of 52% for COPD patients participating in an intensive smoking cessation intervention, they found a misreport rate of 13.6% at baseline (ie, for patients not participating in a smoking cessation intervention). This suggests that although biochemical validation of self-reported smoking status of COPD patients is recommendable in any situation to obtain valid data, participation in an intensive smoking cessation program may increase these misreport rates.

Although the number of participating patients might limit the external validity of our results, our population is a reflection of the self-reported quitters with COPD from a large sample of patients treated in general practice. However, the small numbers of patients that misreported their smoking status hampered in-depth analyses of factors contributing to (in)valid self-reports.

Our results show that the self-reporting of quitting by COPD smokers is not always trustworthy. A general question remains how should medical professionals deal with patients who need ongoing medical attention for chronic smoking-related diseases and who may lie about their smoking behavior? In our study we did not communicate the results of the biochemical validation to the GPs because they were used for research purposes, but some argue that biochemical validation should be integrated in the therapeutic process.

Clinical practice teaches that some physicians are reluctant to treat patients with COPD who persist in smoking. They argue that treating symptoms that are consciously caused by the patient’s persistence in an unhealthy lifestyle is a waste of valuable time and effort. In this case, biochemical validation might be a good instrument for assessing smoking and for increasing the pressure on the patient to quit smoking. Apart from the questions of which measurement is feasible in routine daily general practice and which measurement has good sensitivity and specificity (eg, carbon monoxide or cotinine measures), this harsh approach might damage the relationship with the patient and it questions the ethical codes for health professionals. Misreporting one’s smoking behavior, lying about it, social desirability bias; whatever we call it, it is probably a result of feeling ashamed or guilty, and therefore one might ask whether it is wise to apply biochemical validation tests as a sort of lie detector. Besides the ethical problem, Bize et al found in their systematic review no sufficient evidence that providing this kind of biomedical feedback contributes to successful smoking cessation. Therefore, in spite of the importance of getting a reliable view of the patients’ smoking status in the context of therapeutic general practice interventions, we do not have a clear-cut answer on how best to obtain this information in a real-life general practice setting. Ethical issues concerning the patient’s own responsibility and the effectiveness of biomedical feedback should be weighed against the professional need of getting reliable information about therapeutic patient outcomes.

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Disclosure
The authors declared no conflicts of interest in this work.

References


