Urethral Reconstruction of Critical Defects in Rabbits Using Molecularly Defined Tubular Type I Collagen Biomatrices: Key Issues in Growth Factor Addition

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Tubular type I collagen biomatrices with and without growth factors (GFs) were constructed and evaluated in a rabbit model for critical urethral defects. Porous tubular biomatrices with an inner diameter of 3 mm were prepared using highly purified collagen fibrils and were crosslinked with or without heparin. Heparinized biomatrices were supplemented with the heparin-binding GFs vascular endothelial GF, fibroblast GF-2, and heparin-binding epidermal GF. Biomatrices with and without GFs were used to replace a critical 1 cm urethral segment in rabbits (n = 32). All animals showed normal urination without urinary retention. General histology and immunohistology of graft areas (2, 4, 12, and 24 weeks after implantation) indicated that all biomatrices were replaced by urethra-like structures with normal appearing cytokeratin-positive urothelium surrounded by vascularized tissue. The GF-containing biomatrices showed an increase in extracellular matrix deposition, neovascularization, urothelium, glands, granulocytes, and fibroblasts, compared with biomatrices without GF. GFs substantially improved molecular features of healing but failed to be superior in functional outcome. Retrograde urethrography indicated a normal urethral caliber in case of biomatrices without GF, but a relative narrowing of the urethra at 2 weeks postsurgery and diverticula after 4 weeks in case of biomatrices with GF. In conclusion, tubular acellular type I collagen biomatrices were successful in repairing urethral lesions in artificial urethral defects, and inclusion of GF has a profound effect on regenerative processes.

Introduction

Repair of severe hypospadias is often associated with multiple clinical interventional procedures and grafting. Current grafting techniques include the use of donor tissues such as preputial skin or buccal mucosa. In most instances, the initial results are satisfactory. But long-term follow-up studies show unsatisfactory results and complications, as fistula formations, recurrent strictures, and necrosis can occur in up to 54% of patients.¹,² Additionally, the amount of donor tissue or tissue that is suitable for grafting can be a limiting factor. These issues may be relieved by tissue engineering strategies for urethral reconstruction. However, the development of an optimal biomaterial for urethral reconstruction remains a significant challenge and suitable constructs are currently not available in clinical practice. Biocompatible and biodegradable biomatrices have already been successfully applied for urethral repair, mostly as an onlay.³–⁵ In studies with tubular biomatrices, cellular seeding of the scaffold was an absolute requirement to achieve successful repair of the defect.⁶–⁹ These findings appear to suggest that the use of acellular matrices for urethral reconstruction is limited. Previously, we tubularized small intestinal submucosa (SIS) (COOK) and molecularly defined biomatrices by suturing and compared the regenerative capacity in a rabbit urethral defect model. We reported equating results between these two scaffolds without noticeable complications.⁴ Construction of pretubularized type I collagen biomatrices and their supplementation with growth factors (GFs) may facilitate the surgical procedure.
and improve cellular ingrowth, respectively. For urethral regeneration, multiple GFs are involved because different cell populations are needed to restore the reconstructed site. Epidermal GF (EGF) is known to play a key role in urothelial regeneration, whereas fibroblast GF-2 (FGF-2) and vascular endothelial GF (VEGF) are involved in angiogenesis and blood vessel maturation. As all three GFs bind to heparin, we developed a tubular collagenous biomatrix containing heparin and supplemented this with heparin-binding EGF (HB-EGF), VEGF, and FGF-2. Biomatrices with and without heparin and GFs were used to replace rabbit urethra and were evaluated using retrograde urethrography and immunohistochemical examination.

Materials and Methods

Unless stated otherwise, all chemicals were purchased from Merck.

Preparation of tubular collagen biomatrices

For the construction of tubular biomatrices, insoluble type I collagen fibrils were purified from pulverized bovine Achilles tendon as previously described. A 0.67 wt% type I collagen suspension in 0.25 M acetic acid was prepared and incubated for 16 h at 4°C. Then the suspension was homogenized, deaerated by centrifugation at 2,500 g for 15 min, and poured into polystyrene tubular moulds (all at 4°C). Stainless-steel mandrels (diameter 3 mm) and spacers were placed. The molds were frozen at −20°C for 4 h followed by lyophilization (Fig. 1A). The tubular collagen biomatrices were crosslinked with 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (Fluka) and N-hydroxsuccinimide (Fluka). In brief, tubular biomatrices were incubated at a content of 0.4 mg collagen/mL in 50 mM 2-morpholinoethane sulphonate acid (USB Corporation) containing 40% (v/v) ethanol, 33 mM 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide, and 6 mM N-hydroxsuccinimide (pH 5.0), with or without 0.25% (w/v) heparin (Diosynth) for 4 h while shaking at 21°C. Biomatrices were washed with 0.1 M Na2HPO4, 1 M and 2 M NaCl, and demineralized water. Biomatrices were disinfected by 70% (v/v) ethanol washings and subsequently stored at −20°C. Before implantation, scaffolds were washed extensively with sterile phosphate-buffered saline (PBS, pH 7.4) under sterile conditions.

For correct GF supplementation, simultaneous incubation (3.5 mg/mL per GF), proportional incubation (5.5 μg FGF-2/mL, 3.5 μg VEGF/mL, 1.5 μg HB-EGF/mL), and preincubation of FGF-2 were investigated. In the latter case, disinfected heparinized biomatrices (4 mg matrix/mL) were incubated in sterile PBS containing 3.5 μg/mL human recombinant FGF-2 (R&D) for 1 h at 21°C. Next, human recombinant VEGF (R&D) and human recombinant HB-EGF (R&D) were added (final concentration of 3.5 μg/mL PBS per GF) and incubated for 16 h at 21°C. Subsequently, supplemented

FIG. 1. Overview of the construction, implantation, and explantation of tubular type I collagen matrices. Scheme of the tubular molding process (A), a cross-sectional scanning electron microscopical image of part of a crosslinked tubular matrix (B, bar represents 100 μm), surgical replacement by a tubular matrix (arrow) of a 1 cm urethral defect (C), and explant at 24 weeks (D). L: lumen. Color images available online at www.liebertonline.com/ten.
biomatrices were washed three times (each 15 min) with sterile PBS and used immediately.

**Biochemical characterization of tubular collagen matrices**

The degree of collagen crosslinking was determined by a trinitrobenzolsulfonic acid assay. The amount of bound heparin to the biomatrices was measured by hexosamine analysis. The bound GFs was determined using Western blot analysis. In short, biomatrix samples were run on sodium dodecyl sulfate–polyacrylamide gels, together with a series of known amounts of VEGF, FGF-2, and HB-EGF. Subsequently, gels were blotted, blocked using 1% bovine serum albumin (PAA), and incubated with either rabbit-anti-bovine FGF-2 (1:8000; Sigma-Aldrich), goat-anti-human HB-EGF (1:1000; R&D), or goat-anti-rabbit VEGF (1:400; R&D) containing 1% bovine serum albumin. After washing, blots were incubated with the appropriate secondary antibodies conjugated to horseradish peroxidase and washed. Peroxidase activity was detected by chemiluminescence using ECL Western Blotting Detection Reagents (GE Healthcare) followed by detection and digital quantification (Adobe Photoshop CS3).

**Scanning electron microscopy**

Scanning electron microscopy was performed to analyze the morphology of the biomatrices. The samples were fixed on a stub with double-sided carbon tape. Next, the samples were sputtered with an ultrathin gold layer and examined in a JEOL Scanning Electron Microscope 6310 at an accelerating voltage of 15 kV.

**Animals**

The animal experiment was approved by the Animal Ethics Committee of the Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands. A total of 32 male New Zealand white rabbits (2.5–3.5 kg) were housed individually in a temperature-controlled cage with 50%–55% humidity for a 12 h light–dark cycle with free access to standard laboratory chow and tap water. After a minimum of 7 days, the rabbits were acclimatized to ambient conditions. The surgical procedures have been described in detail elsewhere. In short, rabbits were placed on the operating table in the supine position and general anesthesia was achieved with 2% isoflurane by endotracheal intubation with spontaneous ventilation. Blood pressure, heart rate, and hemoglobin oxygen saturation were monitored continuously throughout the period of anesthesia. After shaving and washing of the ventral penis, the urogenital region was disinfected with povidone-iodine scrub. Next, a transurethral 8F catheter was placed in situ. Urethrography was performed preoperatively. Under a 2.5 optical magnification, a complete resection of a urethral segment of 1 cm in length was made at ∼1.5 cm below the distal urethral meatus with the penis in stretch. A tubular biomatrix of 1 cm (diameter 3 mm) was placed and secured with eight notched sutures (7/0 vicryl0; Ethicon). Two nonresolvable sutures (6/0 prolene0; Ethicon) were placed at the external ends of the tubular biomatrix for future reference. The subcutis and skin were closed in two layers with 7/0 vicryl suture. The transurethral catheter was removed directly after surgery.

**Expantation schedule**

All rabbits were euthanized with an overdose of pentobarbital. Then, urethrography and collection of the urethra was performed. The total penis was removed, fixed in 4% (v/v) formalin in PBS for 24 h, and embedded in paraffin. The rabbits were divided into two groups: a crosslinked collagen biomatrix group (COLX) and a group that received heparinized tubular biomatrices supplemented with VEGF, FGF-2, and HB-EGF (COLX-Hep-GF). Four rabbits were sacrificed in each group at 2, 4, 12, and 24 weeks after implantation (n = 32).

**Immunohistological examination**

Macroscopic inspection and evaluation of the urethra was performed. For histological examination, two samples of every group were cut into four equal transversal pieces. The other two samples were processed longitudinally. Then, sections were cut (4 μm) and stained with hematoxylin and eosin. Sections were scored independently by two panels of two investigators (J.N. and P.G., and M.K. and W.D.) for cellular infiltration, neovascularization, new extracellular matrix (ECM) formation, urothelium, and biomatrix.

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**Table 1. Biochemical Characteristics of Biomatrices**

<table>
<thead>
<tr>
<th>Tubular Scaffolds</th>
<th>Crosslinked using EDC/NHS</th>
<th>Amine group content (nmol/mg biomatrix)</th>
<th>Heparin content (μg/mg biomatrix)</th>
<th>VEGF content (ng/mg biomatrix)</th>
<th>FGF-2 content (ng/mg biomatrix)</th>
<th>HB-EGF content (ng/mg biomatrix)</th>
</tr>
</thead>
<tbody>
<tr>
<td>COL</td>
<td>No</td>
<td>306 ± 16</td>
<td>0*</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>COLX</td>
<td>Yes</td>
<td>189 ± 10</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>COLX-Hep-GF</td>
<td>Yes</td>
<td>135 ± 4</td>
<td>178 ± 15</td>
<td>116 ± 63</td>
<td>104 ± 6</td>
<td>162 ± 27</td>
</tr>
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</table>

The amounts of amine groups (indicative for crosslinking), heparin, and various growth factors are given. COL were used as reference. *Set to 0.

EDC, 1-ethyl-3-(3-dimethyl aminopropyl) carbodiimide; NHS, N-hydroxysuccinimide; s.d., standard deviation; VEGF, vascular endothelial growth factor; FGF-2, fibroblast growth factor-2; HB-EGF, heparin-binding epidermal growth factor; COL, noncrosslinked collagen biomatrices; COLX, crosslinked collagen biomatrices; COLX-Hep-GF, heparinized collagen biomatrices supplemented with growth factors; —, not determined.
FIG. 2. Retrograde urethrograms. The urethrograms of the COLX group showed normal caliber of the urethra at 2, 4, 12, and 24 weeks after implantation (A, C, E, G). In the COLX-Hep-GF group (B, D, F, H), rabbits had relative narrowing of the urethra at 2 weeks and diverticula after 4 weeks. Arrows indicate the position of the biomatrix. COLX, crosslinked collagen biomatrix; COLX-Hep-GF, heparinized collagen biomatrices supplemented with growth factors.
remnants. In case of dissimilar scores, slides were re-evaluated until agreement was reached.

Immunohistochemistry was performed for CD31, pancytokeratin, desmin, and smooth muscle actin. In brief, paraffin sections were deparaffinized in xylene and hydrated in a degrading series of ethanol. After blocking of endogenous peroxidase activity, sections were incubated with monoclonal mouse anti-human CD31 (1:20; Dako), monoclonal mouse anti-human cyto-keratin (1:800; LabVision), monoclonal mouse anti-human desmin (1:200; Biogenex), or monoclonal mouse anti-human smooth muscle actin (1:15,000; Sigma). After PBS washings, sections were incubated with the appropriate peroxidase-conjugated secondary antibodies. After PBS washings, sections were developed with power diaminobenzidine (DAB) (Immunologic) as a chromogen and dehydrated and then slides were mounted with permount.

Results

Biomatrix characterization

Table 1 reviews the biochemical characteristics of the biomatrices. The biomatrices were highly porous (pores ranging 100–150 μm; Fig. 1B) and successfully crosslinked as evidenced by a decrease of amine group content of 38% and 56% for the crosslinked and heparinized biomatrices, respectively. Heparinized biomatrices contained 18 wt% of heparin. Estimates from Western blot analyses revealed a total amount of 0.4 μg bound GFs per milligram of scaffold (VEGF 30%, FGF-2 27%, HB-EGF 42%).

The method used to bind GFs involved incubation of FGF-2 after which VEGF and HB-EGF were added. This method was found to be the most optimal method resulting in binding approximately equal amounts of each GF, as HB-EGF was highly competitive in binding heparin, especially in binding approximately equal amounts of each GF, as HB-EGF was highly competitive in binding heparin, especially with respect to FGF-2. Other incubation methods, namely simultaneous or proportional incubation, were not adequate. The GF incubation concentrations were selected based upon previous work.11

Clinical observations

All animals recovered satisfactory from the initial surgery (Fig. 1C) and survived without any sign of difficulty in urinating or urinary retention. No strictures or urethrocutoaneous fistula were observed in the group without GFs (COLX) (Fig. 1D). The retrograde urethrogram of the group without GFs showed normal caliber of the urethra at all time points (Fig. 2A, C, E, F). Only two rabbits developed small diverticula at 12 weeks. In the group with heparin and GF-supplemented tubular biomatrices (COLX-Hep-GF) (Fig. 2B, D, F, H), all four rabbits showed relative narrowing of the urethra at 2 weeks (Fig. 2B). In 11 of 12 rabbits receiving COLX-Hep-GF tubular biomatrices (4, 12, and 24 weeks postrepair groups), a diverticula of the urethra was observed. One rabbit in this group showed a urethrocutoaneous fistula at 24 weeks (Fig. 2D). Macroscopic inspection confirmed these findings on urethrography.

Histological examination

The microscopical evaluation of urethral regeneration was semiquantified and is summarized in Table 2. Representative images are shown in Figures 3 and 4.

<table>
<thead>
<tr>
<th>COLX-Hep-GF</th>
<th>Weeks of implantation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Granulocytes</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Giant cells</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Macrophages</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Neovascularization</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Biomatrix remnants</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>ECM production</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Fibroblast-like cells</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Urothelium</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Number of glands</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Glandular hyperplasia</td>
<td>++ ++ ++ ++</td>
</tr>
<tr>
<td>Budding</td>
<td>++ ++ ++ ++</td>
</tr>
</tbody>
</table>

Scoring was done by two independent couples. Events were scored ranging from sporadic (sp) to abundant (+). COLX, crosslinked collagen biomatrix; ECM, extracellular matrix; COLX-Hep-GF, heparinized collagen biomatrices supplemented with growth factors; –, not present.

Two weeks evaluation. Granulocytes infiltrated the biomatrix, regardless of the biomatrix modification procedure with GFs (Fig. 3A, B). More giant cells were observed in the COLX-Hep-GF biomatrix. Neovascularization (CD31) was observed at the rim of the biomatrix (Fig. 4A, B). The COLX-Hep-GF biomatrix revealed purple-pink-colored matrix structures on hematoxylin and eosin staining, indicative of the presence of immobilized heparin. Despite precautions to completely remove the native urothelium, some remnants with smooth muscle cells underneath could be localized at the dorsal site of the urethra in almost all rabbits. This was confirmed by staining for desmin. Tissue regeneration was observed in the biomatrix as well as below the biomatrix. In this manner, the tubular construct also facilitated urethral reconstruction by functioning as an onlay.

Four weeks evaluation. The amount of granulocytes decreased in the COLX biomatrix but not in the COLX-Hep-GF biomatrix (Fig. 3C, D). More fibroblast-like cells and newly formed ECM were present in the COLX-Hep-GF biomatrix compared with COLX. As in the latter, still empty porous biomatrix structures could be found. The number of giant cells, present at the periphery of the biomatrix, was higher in COLX-Hep-GF compared with COLX. Also, neovascularization was increased in the COLX-Hep-GF group,
based upon more CD31-positive vessels. Newly formed and
organized urothelium was cytokeratin positive and found to
be increased in the COLX-Hep-GF biomatrix, though only
sporadically present in the COLX biomatrix (Fig. 4C, D).

Twelve weeks evaluation. The cellular infiltration of
granulocytes was reduced in all biomatrices (Fig. 3E, F).
More lymphocytes and macrophages were present in the
COLX-Hep-GF biomatrix in comparison to the COLX bio-
natrix. In both types of biomatrices, the level of interstitial
ECM production and fibroblasts was increased in compari-
sion to the situation at 4 weeks after implantation. In
the COLX group, the scaffold was completely replaced by
urethra-like tissue. In contrast, biomatrix remnants were still
observed in the COLX-Hep-GF biomatrix. In both bio-
matrices, urothelium and capillaries became more evident.
Neovascularization (CD31 staining) was comparable be-
tween both groups. A profound increase of glands, which
were also positive for cytokeratin, and fusion of these was
present in the COLX-HEP-GF in comparison to the COLX
biomatrix (Fig. 4E, F).

Twenty-four weeks evaluation. The inflammatory re-
response was additionally decreased. Both types of biomatrices
were replaced by urethra-like structures; a normal transitional
layer of epithelium was surrounded by vascularized tissue
(Fig. 3G, H). COLX-Hep-GF biomatrices were almost fully
degraded. A decrease in the number of glands was seen in
both groups in comparison with 12 weeks after implantation
(Table 2). Fusion of these, or budding, was increased in COLX
and decreased in COLX-Hep-GF. No muscle cells were visible
in the new tissue of both groups. A clear transition could be
made between old and new tissue, as the latter was recog-
nizable by underlying desmin-positive muscle (Fig. 4G, H).
Both old and new tissues had cytokeratin-positive epithelium
that was comparable to one another.

Discussion

Literature data suggest that cell seeding is critical to re-
generate urethral tissue and avoid stricture formation when
collagen grafts are used for urogenital reconstructions.6–9
Cell seeding necessitates tissue harvesting for the cellular
components, causing an increase in morbidity, and—more
importantly—comes with a complex, labor-intensive, and
time-consuming procedure. It would be advantageous to de-
velop an acellular biomatrix with a high capacity to regenerate
urethral tissues. A tubular biomatrix may be a suitable alter-
native for patients in need for a total urethral reconstruction,
such as patients with severe hypospadias or long urethral
strictures. In a previous study,7 we investigated an onlay
urethral procedure in a rabbit model with an unmodified,
acellular collagen-based biomatrix, compared it with an SIS,
and observed comparable regeneration of the urethra. En-
couraged by the results of that study, we decided to develop
novel, molecularly defined tubular biomatrices (containing
porous of about 100–150 µm substituted with three GFs (VEGF,
FGF-2, and HB-EGF), by coupling with covalently attached
heparin. These GFs are known to have a stimulatory effect on
various cellular processes including cell influx, angiogenesis,
and proliferation/differentiation, thus improving the regen-
orative capacity of the construct. It is known from literature
that epidermal GF plays a key role in urothelial regeneration,10
whereas FGF-2 and VEGF are involved in angiogenesis and
blood vessel maturation.11 In an in vitro pilot study, optimal
proliferation of urothelial cells (SCaBER) was seen by the sole
use of EGF (unpublished results).

We evaluated the regenerative capacity of these bio-
matrices without and with GFs (COLX and COLX-Hep-GF) in a
rabbit model by replacing 1 cm urethra, a critical-sized de-
fect,12 and compared the outcome at 2, 4, 12, and 24 weeks
after implantation. Successful regeneration without urethral
occlusion was achieved with the tubular COLX biomatrix.
Retrograde urethrogram analysis indicated normal caliber of
the urethra at all time points, underscored its effectiveness.
Histology of the COLX biomatrix indicated a mild cellular
response, with the formation of normal urothelium and
capillaries within 24 weeks. For the COLX-Hep-GF biomatrix,
a profound effect on tissue regeneration was observed, in-
cluding an increase in ECM, urothelium, capillaries, glands,
and the number of granulocytes, lymphocytes, and macro-
phages. Sievert et al. already showed the importance of the
time-dependent changes in GF expression during regenera-
tion. Through the addition of GF to the biomatrix we mimic a
depot of GFs in a slow release system, leading to faster ure-
thal regeneration.18 Additionally, genetically engineered
urethral grafts expressing human VEGF significantly induced
neovascularization in the urethral wall.19 VEGF attracts en-
dothelial cells and the combination with FGF-2 has proven to
further enhance an early and well-developed vasculature.11
Nevertheless, although we assumed that the addition of GF
would facilitate and quicken urethral regeneration, urethro-
gram analysis of the COLX-Hep-GF group indicated a relative
narrowing of the replaced urethral segment at 2 weeks and
dilatation (diverticula) at 12 and 24 weeks. The histological
analyses suggest that dilatation is caused by an increase in
urothelium ingrowth in comparison to the unmodified bio-
matrix. The fact that FGF-2 and HB-EGF promote epithelial

![Figure 3](https://www.liebertonline.com/ten)
FIG. 4. Immunohistochemical analysis of blood vessels, urothelium, glands, and muscle cells in biomatrices. Neovascularization (CD31 staining, blue arrowheads) in the biomatrix at 2 weeks after implantation (A, B). Newly formed urothelium at 4 weeks after implantation (cytokeratin staining, black arrowheads), indicating a more organized urothelial layer in the COLX-Hep-GF biomatrix in comparison to the COLX biomatrix (C, D). This new urothelium also resulted in an increase and fusion of glands (black arrows) in the COLX-Hep-GF group in comparison to the normal COLX biomatrix (E, F, H). Both types of biomatrices were replaced by urethra-like structures and only distinguishable from normal urethra by the lack of smooth muscle cells in the newly formed tissue compared with original tissue (desmin staining, red arrowhead) (G). Bars represent 1 mm and 200 μm within the inset. L: lumen; red arrow: biomatrix remnants; white arrow: newly formed extracellular matrix. Color images available online at www.liebertonline.com/ten.
migration and reepithelialization in wound healing\textsuperscript{20,21} may explain the urothelium migration and proliferation into the wall of the biomatrix. This may be prevented by binding HB-EGF only to the luminal side of the biomatrix wall. Epithelial glands that were formed inside the construct seemed to fuse into lacunae. This may explain the decrease in the absolute number of glands after 24 weeks. In a human setting, periurethral glands are of eminent importance; for example, glands of Littré are needed for a normal secretion of the seminal fluid.\textsuperscript{22,23} Additionally, these glands are known to form, at times, small diverticula known as lacunae of Morgagni.\textsuperscript{22,23}

These results implicate that the tubular collagen COLX biomatrix is capable of and succeeds in the regeneration of functional urethra-like tissue. The tubular COLX biomatrix had better functional outcome in comparison to tubular COLX-hep-GF biomatrix. GFs substantially improved “molecular” features of healing but failed to be superior in functional outcome (narrowed urethras, urethras with diverticula, and one fistula). The present study shows that caution should be regarded with respect to the use of GFs, and that biomatrices without GFs do not have to be inferior to those with GFs.

In binding GFs, the biomatrices were first bound with the glycosaminoglycan heparin. After 12 weeks, COLX-Hep-GF biomatrix remnants were still present, whereas the plain collagen biomatrix was fully degraded. This may be explained by the fact that the incorporated heparin is highly negatively charged because of the presence of sulfate and carboxylic groups, which may protect collagen from proteolysis, and has an inhibitory effect on the macrophage adhesion, thus delaying matrix degradation.\textsuperscript{24}

Even though every precaution was taken to completely remove the native urothelium during surgery, this was extraordinarily difficult without causing excessive bleeding from the corpus cavernosa. As a result, some native urothelium was left at the dorsal site of the urethra. These urothelial remnants tend to induce tissue regeneration at the outside of the tubular construct, probably creating a new lumen through which urine was voided. As a result, the tubular construct most likely functioned as an onlay, sometimes, implying that at least small parts of the biomatrix could be lost because of voiding during (bio)matrix remodeling. The use of running sutures at the anastomoses or a temporary transurethral catheter may solve this problem.

Several investigators studied onlay urethroplasty in a rabbit model.\textsuperscript{3-5} Single-layer SIS appears to be inferior to spontaneous urethral regeneration, possibly because SIS impedes smooth muscle cell regeneration, although results have been variable.\textsuperscript{3,7} The conflicting results might be the consequence of dissimilarities in different SIS batches.\textsuperscript{25}

Similar to SIS, our constructs did not support smooth muscle cell ingrowth (smooth muscle actin). This is in contrast with results obtained with a decellularized tubularized construct created from SIS where complete regeneration of all urethral layers was reported as early as 12 weeks after urethra repair.\textsuperscript{5} However, the clinical importance and relevance of muscular cell ingrowth in a urethra is not clear. Urethroplasty with SIS resulted in fibrotic tissue covered by urethral mucosa without muscle cells at the site of SIS implantation.\textsuperscript{26} Animal studies have indicated that the urethra is capable of complete regeneration over a 15 mm defect area when only the ventral wall of the urethra is excised.\textsuperscript{7} However, a circumferential defect as used here does not heal spontaneously and this would imply that severe scarring may occur. Previous studies have shown that with an acellular tubularized construct the maximum defect distance that can be supported by normal tissue regeneration is 0.5 cm, albeit that the longest time period studied was only 4 weeks.\textsuperscript{17}

Collectively, in the present study, the repair of a 1 cm defect with collagen-based tubular biomatrices resulted in excellent regeneration of the urethra without urethral occlusion. In the future, long-term outcome of such matrices has to be studied. The inclusion of a support, for example, a polymer, may help to prevent voiding of urine alongside instead of through the construct.

Conclusion

Tubular, molecularly defined collagen biomatrices are successful in repairing urethral lesions of 1 cm in a rabbit model. The results with these developed biomatrices are promising, but further studies aimed at fine-tuning the effect of bound GFs must be performed, particularly to prevent (initial) excessive cell proliferation and to induce ingrowth of smooth muscle. Additionally, a composite construct may facilitate future use of longer tubular segments in urethral reconstruction.

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Disclosure Statement

No competing financial interests exist.

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