Substantial Contribution of Submicroscopical Plasmodium falciparum Gametocyte Carriage to the Infectious Reservoir in an Area of Seasonal Transmission

André Lin Ouédraogo1,2*, Teun Bousema3, Petra Schneider4, Sake J. de Vlas5, Edith Ilboudo-Sanogo1, Nadine Cuzin-Ouattara1, Issa Nébié1, Will Roeffen2, Jan Peter Verhave2, Adrian J. F. Luty2, Robert Sauerwein2

1 Department of Biomedical Sciences, Centre National de Recherche et de Formation sur le Paludisme, Ouagadougou, Burkina Faso, 2 Department of Medical Microbiology, Radboud University Medical Centre, Nijmegen, The Netherlands, 3 Department of Infectious & Tropical Diseases, London School of Hygiene & Tropical Medicine, London, United Kingdom, 4 Centre for Immunity, Infection and Evolution, University of Edinburgh, Edinburgh, United Kingdom, 5 Department of Public Health, Erasmus MC, University Medical Centre Rotterdam, Rotterdam, The Netherlands

Abstract

Background: Man to mosquito transmission of malaria depends on the presence of the sexual stage parasites, gametocytes, that often circulate at low densities. Gametocyte densities below the microscopical threshold of detection may be sufficient to infect mosquitoes but the importance of submicroscopical gametocyte carriage in different transmission settings is unknown.

Methodology/Principal Findings: Membrane feeding experiments were carried out on 80 children below 14 years of age at the end of the wet season in an area of seasonal malaria transmission in Burkina Faso. Gametocytes were quantified by microscopy and by PfS25-based quantitative nucleic acid sequence-based amplification assay (QT-NASBA). The children’s infectiousness was determined by membrane feeding experiments in which a venous blood sample was offered to locally reared Anopheles mosquitoes. Gametocytes were detected in 30.0% (24/80) of the children by microscopy compared to 91.6% (65/71) by QT-NASBA (p < 0.001). We observed a strong association between QT-NASBA gametocyte density and infection rates (p = 0.007). Children with microscopically detectable gametocytes were more likely to be infectious (68.2% compared to 31.7% of carriers of submicroscopical gametocytes, p = 0.001), and on average infected more mosquitoes (13.2% compared to 2.3%, p < 0.001). However, because of the high prevalence of submicroscopical gametocyte carriage in the study population, carriers of sub-microscopical gametocytes were responsible for 24.2% of the malaria transmission in this population.

Conclusions/Significance: Submicroscopical gametocyte carriage is common in an area of seasonal transmission in Burkina Faso and contributes substantially to the human infectious reservoir. Submicroscopical gametocyte carriage should therefore be considered when implementing interventions that aim to reduce malaria transmission.


Editor: David Joseph Diemert, Sabin Vaccine Institute, United States of America

Received September 19, 2009; Accepted November 22, 2009; Published December 22, 2009

Copyright: © 2009 Ouédraogo et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was funded by the Netherlands Foundation for the Advancement of Tropical Research (WOTRO WM 93-350). Teun Bousema is supported by a Rubicon fellowship of the Netherlands Organization for Scientific Research (NWO; Rubicon 825.08.025). Petra Schneider is funded by the Wellcome Trust. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: andrelin.cnrfp@fasonet.bf

Introduction

The transmission of malaria depends on the presence of mature sexual stage parasites, gametocytes, in the human peripheral blood. Once ingested by a mosquito taking a blood meal, gametocytes develop through different mosquito-specific stages and ultimately result in infection of the mosquito salivary glands with sporozoites. This renders the mosquito infectious to humans. There is growing awareness that targeting gametocytes, either alone or as part of integrated control programmes, is essential for malaria control and elimination efforts [1,2,3,4]. The identification of the human infectious reservoir is therefore important for successful malaria control.

Gametocytes frequently occur at low densities, making microscopical detection complicated [5]. In the last decade, molecular tools have become available to detect and quantify gametocytes at densities well below the microscopical threshold, in the order of 0.02–10 gametocytes/μL of blood [4]. Using these techniques, it has become evident that the proportion of gametocyte carriers in the population has been grossly underestimated and that the gametocyte reservoir may be 2–5 fold larger than assumed based on microscopy [6,7]. Carriers of gametocytes at submicroscopical levels are capable of infecting mosquitoes [7,8,9,10,11,12,13,14,15,16,17,18,19] although at a lower degree than those with gametocytes detectable microscopically in whom gametocytes are present at higher densities [12,18].
The importance of submicroscopical gametocyte carriage for malaria epidemiology and malaria control is the subject of some debate. While carriers of gametocytes at submicroscopical densities were concluded to be as important for the human infectious reservoir as carriers of microscopically-detectable gametocytes in areas of perennial transmission in Kenya [18] and Thailand [16], data from the Gambia suggest that submicroscopical gametocyte carriers only form a very small fraction of the infectious reservoir in this area of seasonal transmission [20]. This suggests that the relevance of submicroscopical gametocyte carriage may depend on transmission settings.

Here, we determine the contribution of submicroscopical densities of gametocytes to the human infectious reservoir in an area of seasonal malaria transmission in Burkina Faso.

Methods

This study was conducted in September-November 2005 in the village of Laye, 30 km northwest of Ouagadougou, Burkina Faso. The area is characterised by Sudanese savannah with a marked wet season from June to October and an estimated entomological inoculation rate of 300–500 infective bites per person per year [21]. Asexual parasite carriage in the population shows seasonal fluctuations and was recently estimated at 60–90% in children below 15 years of age and 20–50% in adults [22]. Clearance was received by the Ministry of Health of Burkina Faso. Children below 14 years of age were randomly selected from village census lists and written informed consent was obtained from parents/guardians after the purpose of the study was explained. Children were accompanied to the Centre National de Recherche et de Formation sur le Paludisme (CNRFP) 1 to 2 days after the consenting procedure. At CNRFP, children underwent a clinical examination and their axillary temperature was measured. Children were enrolled in membrane feeding experiments regardless of symptoms or the presence of asexual malaria parasites or gametocytes. Venous blood samples (3 mL) were drawn into heparin-containing tubes for membrane feeding and for gametocyte detection both by microscopy and by real-time Pfs25 quantitative nucleic acid sequence based amplification (QT-NASBA). For all membrane-feeding assays, 3 mL venous blood samples were obtained and fed to ~50 locally colony-reared 4–5-day-old female A. gambiae sensu stricto mosquitoes. The mosquito colony was established three years prior to the current experiments. Blood was offered via an artificial membrane attached to a water-jacketed glass feeder maintained at 37°C. After 10–15 min, fully fed mosquitoes were selected and kept on glucose at 29°C [7]. Unfed and partially fed mosquitoes were removed by aspiration and discarded. Mosquito midguts were examined after 7 days for the presence of oocysts following dissection in 2% mercurochrome. A second microscopist confirmed the presence of oocysts in midguts that were scored positive.

Only the total number of oocysts per batch of fed mosquitoes was recorded; not the number of oocysts per individual mosquito. Experiments in which a minimum of 10 mosquitoes were examined on day 7 after feeding were included in the analyses. After membrane feedings, children with fever (axillary temperature ≥37.5°C) and malaria parasites were treated with artesinisin-based combination therapy according to the national guidelines. Individuals for whom infections other than malaria were suspected were accompanied to the nearest health facility for appropriate clinical care. The study protocol was viewed and approved by the Ministry of Health of Burkina Faso on August 8th 2000 (Research’s Authorization number 2000/3174/MS/SG/DEP).

Microscopic Detection of P. falciparum Parasites

Samples were considered negative if no parasites were detected in 100 fields. Both asexual stage and gametocyte densities were simultaneously assessed by counting against 1000 leucocytes in the thick smear. The lower limit of microscopy for gametocyte quantification was therefore estimated at 8 gametocytes/μL of blood. Parasite counts were converted to numbers of parasites per μL by assuming a standard count of 8000 leucocytes/μL of blood. Each sample was read independently by two microscopists and the mean density was used. A third reader was involved when the first two readers disagreed about the prevalence of gametocytes or their estimated densities differed ≥30%. In these cases the mean density of the two closest readings was used.

Gametocyte Detection by Real Time Pfs25 QT-NASBA

Gametocyte detection by Pfs25 QT-NASBA was performed as described elsewhere using a NuchiSens EasyQ analyzer (BioMérieux) [23,24]. Nucleic acid was extracted from 50-μL blood samples as described by Boom et al. [25]. The first part of the RNA extraction was done in the field following the original guanidinium isothiocyanate (GuanSCN) RNA extraction method [25] until the nucleic acids were bound to silica dioxide particles. At this point, samples were stored at −20°C and transferred to the laboratory for completion of the extraction and QT-NASBA analysis. The number of gametocytes was calculated in relation to a standard gametocyte stage V dilution series [26], using the time point of amplification at which the fluorescence detecting target amplicons exceeded the mean fluorescence of three negative controls + 20 standard deviations. The Pfs25 QT-NASBA technique is gametocyte-specific and has a detection limit of 20–100 gametocytes/ml [24]. Samples with Pfs25 QT-NASBA gametocyte concentrations <20 gametocytes/mL were considered gametocyte negative.

Sample Size Considerations

Based on a previous study in the area, we expected a gametocyte prevalence of 10–20% by microscopy and 70–80% by Pfs25 QT-NASBA [23]. Including 80 individuals in the membrane feed experiments would allow us to detect a threefold lower infectiousness of submicroscopical carriers compared to microscopical gametocyte carriers [12,20] when we assumed that 60% of the microscopical gametocyte carriers infected at least one mosquito [18] (Za = 1.645; Zb = 0.84).

Data Analysis

Data analyses were performed using SPSS version 16.0 (SPSS Inc., Chicago, IL, USA) and Stata 10 (Statcorp, Texas US). Densities of gametocytes were analysed on a log10-scale. The prevalence of mosquito infection (i.e. whether an individual infected at least one mosquito) and the proportion of infected mosquitoes were used as outcomes of the membrane feeding experiments and were related to Pfs25 QT-NASBA gametocyte density, age and fever in logistic and linear regression models. Individual oocyst densities in mosquitoes were not recorded.

Results

We enrolled 80 children in our study who were aged 2.9–13.6 years. In line with previous studies from the study area, microscopy indicated an asexual parasite prevalence of 82.5% (66/80) and a gametocyte prevalence of 30.0% (24/80); Table 1 [22,23]. When the Pfs25 QT-NASBA was used for gametocyte detection, 91.6% (65/71) individuals were shown to be carrying gametocytes. There was a strong correlation between gametocyte densities detected by QT-NASBA and microscopy for microscop-
Figure 1. The relationship between 500 parasites/μL and the proportion of infected mosquitoes in membrane feeding experiments. The solid line indicates the best fitted line \( Y = 0.0176\ln(X) + 0.0187 \) (\( R^2 = 0.153 \)). The dashed line indicates the estimated microscopic threshold for gametocyte detection, 8 gametocytes/μL, when screening 100 high power fields (i.e. \(~1000\) white blood cells).

doi:10.1371/journal.pone.0008410.g001

Table 1. Baseline characteristics.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, median (IQR)</td>
<td>6.2 (4.8–9.5)</td>
</tr>
<tr>
<td>Sex, % male (n/N)</td>
<td>55.0 (44/80)</td>
</tr>
<tr>
<td>Fever, % (n/N)</td>
<td>22.5 (18/80)</td>
</tr>
<tr>
<td>Asexual parasite prevalence, % (n/N)</td>
<td>82.5 (66/80)</td>
</tr>
<tr>
<td>Asexual parasite density/μL, median (IQR)(^a)</td>
<td>1028.5 (585–3662)</td>
</tr>
<tr>
<td>Symptomatic malaria(^a)</td>
<td>15.0 (12/80)</td>
</tr>
<tr>
<td>Microscopic gametocyte prevalence, % (n/N)</td>
<td>30.0 (24/80)</td>
</tr>
<tr>
<td>Microscopic gametocyte density/μL, median (IQR)(^a)</td>
<td>4.00 (16–45)</td>
</tr>
<tr>
<td>QT-NASBA gametocyte prevalence, % (n/N)(^a)</td>
<td>91.6 (65/71)</td>
</tr>
<tr>
<td>QT-NASBA gametocyte density/μL, median (IQR)(^a)</td>
<td>7.9 (1.4–48.9)</td>
</tr>
</tbody>
</table>

\( ^a \)Fever = temperature \( \geq 37.5^\circ C \);
\( ^a \)Fevers were considered if \( \geq 500 \) parasites/μL;
\( ^a \)Only gametocyte densities \( \geq 20 \) gametocytes/μL were considered 500 parasites/μL positive.

doi:10.1371/journal.pone.0008410.t001

Discussion

In this study, we observed that submicroscopic gametocyte carriage is common in children in an area of seasonal malaria in Burkina Faso. Although, on average, carriers of gametocytes at submicroscopic densities infected significantly fewer mosquitoes than individuals with microscopically-detectable densities of gametocytes. The relative contribution to transmission of microscopic and submicroscopic gametocyte carriage was determined based on their prevalence in the studied population and the proportion of mosquitoes infected by each group. This resulted in a relative contribution to malaria transmission of 24.2% for submicroscopic gametocyte carriage, compared to 73.8% for microscopic gametocyte carriage.

There are numerous reports that suggest that some individuals can infect mosquitoes despite the absence of microscopically detectable gametocytes [7,8,10,12,16,18,19]. However, it is unclear (i) how important this phenomenon is in areas of seasonal malaria [20] and (ii) how important submicroscopic gametocyte densities are for malaria transmission in the general, typically asymptomatic, population. Several detailed studies on the infectiousness of carriers of submicroscopic gametocytes have been carried out, but only after chemotherapeutic treatment of symptomatic malaria cases [7,18,20]. Because children in those studies all had high densities of asexual parasites in the weeks prior to the mosquito feeding experiments, they were more likely to have gametocytes at the time of the membrane feedings [5]. Although it was previously shown that gametocyte carriage may be common in asymptomatic individuals as well [27], findings from clinical trials cannot be extrapolated to the general population. To the best of our knowledge, the infectious reservoir has never been determined in the general population by means of the combination of molecular gametocyte detection tools with membrane feeding experiments used in the study described here. Although our experiments were restricted to children, our findings provide valuable information about the infectiousness of a cross-section of the population living in an area of seasonal malaria transmission. At the end of the wet season, 90% of the children in our study area carried gametocytes [23], two-thirds of them at densities below the microscopic threshold of detection. The infectiousness of individuals with submicroscopic gametocyte densities was lower than that of children with microscopically-detectable densities in terms of (i) prevalence of infection, (ii) proportion of infected mosquitoes and (iii) oocyst burden in mosquitoes. These observations are largely in agreement with previous studies [12,18] although a study in symptomatic children in Kenya reported that submicroscopic gametocyte carriers were as likely as microscopically-detectable gametocyte carriers to be infectious and only the average number of infected mosquitoes and the oocyst burden were lower for submicroscopic gametocyte carriers [18]. This difference may be due to the different populations: we enrolled asymptomatic children compared to the Kenyan study where children were
sampled 14 days after a clinical malaria episode [18]. We nevertheless consider the current findings biologically more plausible since the chance of a submicroscopical gametocyte carrier being classified as ‘infectious’ (i.e. infecting at least one mosquito) is likely to be lower if submicroscopical gametocyte carriers on average infect a lower proportion of mosquitoes [12,18]. Contrary to a recently published hypothesis that infection of mosquitoes by submicroscopical gametocytemia may be rare in areas of seasonal malaria transmission [20], our findings suggest that carriers of submicroscopical gametocyte densities may be common in these circumstances. The relative contribution to transmission per gametocyte carrier may be lower for submicroscopical gametocyte carriers but their relative abundance in a population appears to counterbalance this and makes them important contributors to malaria transmission. To reliably determine the influence of transmission intensity and seasonality on the occurrence and infectiousness of submicroscopical gametocyte densities, a direct comparison is needed where the infectivity of different populations is assessed at several time-points in the season. For this, a series of membrane feeding experiments are needed throughout the year. We have previously reported that submicroscopical gametocyte carriage is common in these circumstances. They were unable to infect mosquitoes [8,12,18]. This could be partly due to transmission reducing immune responses [8,29]. Alternatively, we cannot rule out that artefacts resulting from RNA degradation have resulted in unrealistically low estimates of gametocyte densities in occasional samples. We also observed that some carriers of gametocytes at high density were unable to infect mosquitoes [8,12,18]. This could be partly due to transmission reducing immune responses [8,29]. Further studies should therefore be conducted at different time-points, include all age groups and preferably incorporate transmission reducing immune responses and sexing of gametocytes to further elucidate the detailed processes that determine the composition of the human infectious reservoir of malaria in a given transmission setting.

Acknowledgments

We are grateful to all the inhabitants of the study village for their cooperation, the Ministry of health of Burkina Faso and the staff of CNRFP for their assistance in the implementation of this study. We thank Marcel de Kruijf for his assistance in the QT-NASBA analyses.

Author Contributions

Conceived and designed the experiments: ALO SJdV EIS NCO IN JPV. Analyzed the data: ALO TB PS SjdV AJFL. Wrote the paper: ALO TB AJFL RWS.

References

area of low and seasonal transmission in Tanzania. TropMedIntHealth 12: 547–553.


