Human Golgi Antiapoptotic Protein Modulates Intracellular Calcium Fluxes

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Golgi antiapoptotic protein (GAAP) is a novel regulator of cell death that is highly conserved in eukaryotes and present in some poxviruses, but its molecular mechanism is unknown. Given that alterations in intracellular Ca\(^{2+}\) homeostasis play an important role in determining cell sensitivity to apoptosis, we investigated if GAAP affected Ca\(^{2+}\) signaling. Overexpression of human (h)-GAAP suppressed staurosporine-induced, capacitative Ca\(^{2+}\) influx from the extracellular space. In addition, it reduced histamine-induced Ca\(^{2+}\) release from intracellular stores through inositol trisphosphate receptors. h-GAAP not only decreased the magnitude of the histamine-induced Ca\(^{2+}\) fluxes from stores to cytosol and mitochondrial matrices, but it also reduced the induction and frequency of oscillatory changes in cytosolic Ca\(^{2+}\). Overexpression of h-GAAP lowered the Ca\(^{2+}\) content of the intracellular stores and decreased the efficacy of IP\(_3\) providing possible explanations for the observed results. Opposite effects were obtained when h-GAAP was knocked down by siRNA. Thus, our data demonstrate that h-GAAP modulates intracellular Ca\(^{2+}\) fluxes induced by both physiological and apoptotic stimuli.

INTRODUCTION

Recently, a novel regulator of cell death was identified (Gubser et al., 2007). This protein was named Golgi antiapoptotic protein (GAAP) because of its predominant localization in the Golgi and its ability to suppress apoptosis. GAAP is a predicted seven-transmembrane protein and was identified initially in certain poxviruses (vaccinia virus and camelpox virus) where it affects virus virulence. GAAPs are highly conserved in a broad range of organisms including human, orangutan, dog, mouse, rat, *Xenopus laevis*, and zebrafish, and related proteins are present in *Drosophila* and *Arabidopsis*. Human (h)-GAAP is expressed ubiquitously in human tissue and shares 73% aa identity with viral (v)-GAAP. Stable expression of either v-GAAP or h-GAAP suppressed cell death induced by a broad variety of intrinsic and extrinsic apoptotic stimuli. Conversely, knockdown of h-GAAP in tissue culture cells by siRNA resulted in cell death.

Ca\(^{2+}\) functions as a ubiquitous intracellular signal to many different biological processes. Ca\(^{2+}\)-induced signaling arises from Ca\(^{2+}\) entry across the plasma membrane and/or release from intracellular stores, predominantly the endoplasmic reticulum (ER) and Golgi. Ca\(^{2+}\) is released from intracellular stores by inositol-1,4,5-trisphosphate (IP\(_3\)), which interacts with IP\(_3\) receptors (IP\(_3\)Rs) that are Ca\(^{2+}\) release channels present in the ER and Golgi (Pinton et al., 1998). Furthermore, IP\(_3\)R activity is modulated by Ca\(^{2+}\) itself, ATP, phosphorylation, and interacting proteins (Foskett et al., 2007). Ca\(^{2+}\) that enters the cytosol activates cytosolic enzymes and is taken up by mitochondria, which play an important role in decoding Ca\(^{2+}\) signals during normal cell physiology (Berridge et al., 2003). Mitochondrial Ca\(^{2+}\) uptake is mediated by a low-affinity Ca\(^{2+}\) uniporter that senses the high Ca\(^{2+}\) microdomains that are established at the tight junctions between the ER and mitochondria (Rizzuto et al., 1998). Recently, tight junctions with a putative role in Ca\(^{2+}\) signaling were also observed between Golgi and mitochondria (Dolman et al., 2005).

Alterations in the finely tuned intracellular Ca\(^{2+}\) homeostasis and compartmentalization contribute to the induction of apoptosis. The switch from the control of physiological functions to the involvement in this death program most likely entails changes in the tightly regulated spatiotemporal Ca\(^{2+}\) signaling pattern affecting cytosolic effector proteins and effector organelles (Orrenius et al., 2003). Ca\(^{2+}\)
signaling between storage organelles and mitochondria plays an important role in sensitizing cells to apoptosis (Pinton and Rizzuto, 2006). Molecular and pharmacological approaches that lowered Ca\(^{2+}\) levels in the stores and thereby reduced Ca\(^{2+}\) signaling to the mitochondria, protected cells from apoptosis, whereas conditions that increased Ca\(^{2+}\) levels in the stores had the opposite effect (Ma et al., 1999; Nakamura et al., 2000; Pinton et al., 2001; Pinton and Rizzuto, 2006). Moreover, both antiapoptotic (e.g., Bcl-2 and Bcl-X\(_L\)) and proapoptotic (e.g., Bak and Bak) partially localize at the ER to regulate Ca\(^{2+}\) signaling (Oakes et al., 2003; Chen et al., 2004; White et al., 2005).

On the basis of the localization of h-GAAP at intracellular Ca\(^{2+}\) stores and the established importance of intracellular Ca\(^{2+}\) signaling in sensitizing cells to apoptosis induction, we hypothesized that the antiapoptotic role of h-GAAP may be mediated by modulating the Ca\(^{2+}\) content of these stores and/or the flux of Ca\(^{2+}\) between these stores and the closely opposed mitochondria. Here, evidence is presented that h-GAAP alters intracellular Ca\(^{2+}\) fluxes induced by both a physiological stimulus (histamine) and an apoptotic stimulus (staurosporine).

**MATERIALS AND METHODS**

**Cells and Medium**

U2OS-neo and U2OS-h-GAAP cell lines were described previously (Gubser et al., 2007). Cells were grown in minimal essential medium (Invitrogen, Carlsbad, CA) supplemented with 10% (vol/vol) fetal bovine serum and 10 \(\mu\)g/ml Ciproxin (Bayer, Newbury, Berks, United Kingdom) at 37°C in a 5% CO\(_2\) atmosphere.

**Antibodies, Conjugates, and Reagents**

Mouse monoclonal antibodies against IP\(_3\), R3, calnexin, Bcl-2, and caspase were obtained from BD Transduction Laboratories (Lexington, KY). Mouse mAb against protein disulfide isomerase (PDI) was from StressGen (San Diego, CA), against Bcl-2 (Upstate Laboratories, Lake Placid, NY). Rabbit polyclonal antibodies against Bax and Bak were from Cell Signaling Technology (Beverly, MA). Rabbit polyclonal antibody (Rbt 476) against IP3R (all isoforms) was described previously (Ma et al., 2002). Coelenterazine-W, coelenterazine-N, fura-2 acetoxyethyl ester (fura-2/AM), and Rhod-2/AM were from Molecular Probes (Eugene, OR), histamine and ionomycin from Sigma-Aldrich (Poole, Dorset, United Kingdom), 2-APB from Calbiochem (La Jolla, CA), and STS from Roche (East Sussex, United Kingdom).

**RNA Interference**

Sequences of small interfering RNA1 (siRNA1) and siRNA2 (Ambion, Austin, TX) were described previously (Gubser et al., 2007). Cells were grown to 50% confluence in six-well plates and transfected with 1 \(\mu\)g of each of the above siRNAs using siFECTamine (IC-Vec; www.icvec.com) according to the manufacturer’s instructions.

**Immunoprecipitation**

Coimmunoprecipitation was performed as described for Bcl-2 interaction with IP\(_3\)R (Chen et al., 2004). Abs used were anti-IP\(_3\)R Ab (BD Biosciences, Poole, United Kingdom; 1:200), anti-HA mAb (1:200) and the control Ab used was a mouse IgG2a Ab-1 (Stratec Scientific, Bedfordshire, United Kingdom; 1:150). Proteins were resolved by SDS-PAGE and transferred onto Hybond-P PVDF membranes (Amersham, Bucks, United Kingdom).

**Digital Imaging Microscopy of Cytosolic and Mitochondrial Ca\(^{2+}\) Concentrations**

Cells (3 \(\times\) 10\(^4\)) seeded on 24-mm glass coverslips were coloaded with 3 \(\mu\)M fura-2/AM and 5 \(\mu\)M rhod-2/AM for 25 min at 37°C and used for monitoring simultaneous changes in mitochondrial and cytosolic Ca\(^{2+}\) concentration as described by Visch et al., 2004). The fura-2 and rhod-2 dyes were excited at 380 and 540 nm, respectively. The fura-2 fluorescence emission ratio at 492 nm was monitored as a measure of the free cytosolic Ca\(^{2+}\) concentration after alternating excitation at 340 and 380 nm. In all experiments, the fluorescence emission signal was normalized to its prestimulatory value, which was set at 1.

**Luminescence Monitoring of Ca\(^{2+}\)**

For luminescence measurement of Ca\(^{2+}\), 5 \(\times\) 10\(^5\) cells were seeded on 13-mm glass coverslips, transfected with targeted aqueorin (Pinton et al., 1998) using FuGENE 6 reagent (Roche), and analyzed as described (Visch et al., 2004; Visch et al., 2006).

**45Ca\(^{2+}\) Fluxes**

45Ca\(^{2+}\) fluxes were performed as described (Kasri et al., 2006). Briefly, cells were grown to confluency, permeabilized with saponin, loaded with 4Ca\(^{2+}\), and washed with efflux medium containing 4 \(\mu\)M thapsigargin (TG) to block ATP-dependent Ca\(^{2+}\) uptake. IP\(_3\)-stimulated Ca\(^{2+}\) release was initiated by the addition of efflux medium containing the indicated concentration of IP\(_3\), and 2 min later the efflux medium was collected and counted for radioactivity. After correction for the passive leak induced by TG alone, the amount of radioactivity released by IP\(_3\) was expressed as a percentage of the total amount of radioactivity present in the stores as determined by addition of the Ca\(^{2+}\) ionophore A23187. To assess the rate of passive Ca\(^{2+}\) leakage induced by TG alone, the efflux medium was replaced every 2 min during 18 min. At the end of the experiment, all residual radioactivity was released by incubation with 1 ml of 2% SDS for 30 min. For each data point, the amount of radioactivity that was still present in the stores was calculated, expressed as a percentage of the total amount present at the onset of the experiment, and plotted as a function of time.

**Calculations**

Data are presented as mean values ± SEM. Differences were tested for significance using the Student’s t test.

**RESULTS**

**h-GAAP Decreases Cytosolic and Mitochondrial Ca\(^{2+}\) Rises Triggered by an Apoptotic Stimulus**

In this study, possible effects of h-GAAP on intracellular Ca\(^{2+}\) on intracellular Ca\(^{2+}\) signaling were investigated using U2OS cells that stably expressed hemagglutinin (HA)-tagged h-GAAP predominantly at the Golgi but also at the ER (hereafter referred to as h-GAAP cells; Gubser et al., 2007). Essentially the same results were obtained with two independently constructed h-GAAP cell lines. Except in experiments in which parental U2OS cells were transfected with siRNA, U2OS cells containing the empty plasmid vector were used as a control (hereafter referred to as U2OS-neo cells). Western blot analysis was performed to exclude the possibility that h-GAAP overexpression affected the expression levels of ER chaperones or Bcl-2 family members. The data show that the expression levels of calnexin, PDI, Bcl-2, Bcl-X\(_L\), BAX, and Bak in h-GAAP cells was not altered compared with U2OS-neo cells (Supplementary Figure S1).

Previously, staurosporine (STS)-induced apoptosis was demonstrated to be partially, but significantly, inhibited in h-GAAP cells (Gubser et al., 2007). The exact mechanism by which STS induces cell death is unknown, but STS-induced cell death is at least partially Ca\(^{2+}\)-dependent (Oakes et al., 2003; Chen et al., 2004; White et al., 2005). Therefore, we first addressed a possible role of h-GAAP in the STS-induced changes in intracellular Ca\(^{2+}\) homeostasis (Boehning et al., 2003). To this end, cells were coloaded with the cytosolic Ca\(^{2+}\) indicator fura-2 and the mitochondrial Ca\(^{2+}\) indicator rhod-2, treated with STS, and analyzed by digital imaging microscopy. Initial measurements of the resting cytosolic Ca\(^{2+}\) concentration using fura-2 revealed no detectable differences between U2OS-neo and h-GAAP cells (resting fura-2 ratio in U2OS-neo cells was 0.35 ± 0.02, \(n = 85\) cells, measured on 5 d; resting fura-2 ratio in h-GAAP was 0.35 ± 0.01, \(n = 85\) cells, measured on 5 d). On addition of 2 \(\mu\)M STS, U2OS-neo cells displayed a gradual increase in both mitochondrial and cytosolic Ca\(^{2+}\) concentration (Figure 1, A and B). Both increases were virtually abolished by 2-aminocythoxy-diphenylborate (2-APB), a drug that suppresses extracellular Ca\(^{2+}\) entry by inhibiting store-operated Ca\(^{2+}\) channels and indicating that STS acts to stimulate the capac-
Ca²⁺ its effects on the increase in cytosolic and mitochondrial 

h-GAAP Decreases Histamine-induced Rises in Cytosolic and Mitochondrial Ca²⁺

To gain more insight into a possible role of h-GAAP in the regulation of intracellular Ca²⁺ signaling, we investigated its effects on the increase in cytosolic and mitochondrial Ca²⁺ concentration evoked by the IP₃-generating hormone histamine. To prevent capacitative Ca²⁺ entry, stimulation with histamine was performed in the absence of extracellular Ca²⁺ (Peppiatt et al., 2003). Similarly, no increases were observed when using Ca²⁺-free medium (data not shown). Importantly, expression of h-GAAP strongly reduced the STS-induced increase in both mitochondrial and cytosolic Ca²⁺ concentration.

To gain further support for a role of h-GAAP in regulating STS-induced Ca²⁺ fluxes, we next assessed the effect of h-GAAP down-regulation. Parental U2OS cells were transfected with siRNA1 (hereafter referred to as h-GAAP siRNA) or siRNA2 (hereafter referred to as control siRNA), shown before to decrease h-GAAP or be without effect on h-GAAP expression, respectively (Gubser et al., 2007). Cells were tested for their response to STS at 2 d after transfection, at which time they were shown before to be still alive (Gubser et al., 2007). Cells transfected with h-GAAP siRNA showed a much larger STS-induced increase in mitochondrial and cytosolic Ca²⁺ concentration than untreated U2OS-neo cells (Figures 1, A and B) or parental U2OS cells treated with either control siRNA or transfection reagent alone (data not shown). Also in h-GAAP siRNA-treated cells, the STS-induced increase in mitochondrial and cytosolic Ca²⁺ concentration was virtually completely inhibited by 2-APB. These results show that h-GAAP reduces the STS-induced increase in mitochondrial and cytosolic Ca²⁺ concentration, which depends on the influx of Ca²⁺ across the plasma membrane.

h-GAAP Decreases Histamine-induced Rises in Cytosolic and Mitochondrial Ca²⁺

h-GAAP Immunoprecipitates with the IP₃R

The above results suggested that h-GAAP might interact with IP₃Rs thus providing a potential explanation for its inhibitory effect on the flux of Ca²⁺ through these receptors. Because currently there exists no good antibody against h-GAAP for immunoprecipitation purposes, we made use of
the HA-tag that was fused to h-GAAP. The results obtained show that the anti-HA mAb coprecipitated IP₃R subtype 3 (IP3R3; Figure 2A), which is the most abundant subtype in U2OS cells (data not shown). The specificity of this reaction was confirmed by the failure of the antibody to precipitate any sarcoendoplasmic reticulum Ca²⁺/H⁺-ATPase isoenzyme 2b (SERCA2b), the predominant SERCA protein in non-muscle cells. In the reciprocal experiment, immunoprecipitation of IP3R3 brought down h-GAAP (Figure 2B). These results demonstrate that h-GAAP interacts, either directly or indirectly, with the IP₃R.

**h-GAAP Alters the IP₃-induced Ca²⁺ Release Response from Intracellular Ca²⁺ Stores in Permeabilized Cells**

Because h-GAAP can be coprecipitated with IP₃Rs, we next assessed the possibility that it might affect the characteristics of the IP₃-induced Ca²⁺ release response. To gain access to the IP₃R, we made use of a permeabilized cell system. Cells were permeabilized with saponin, loaded to steady state with ⁴⁵Ca²⁺, washed to remove excess ATP using an efflux medium containing the SERCA inhibitor TG to prevent Ca²⁺-reuptake, and challenged with either the Ca²⁺ ionophore A23187, to determine total releasable ⁴⁵Ca²⁺ or the indicated concentration of IP₃. After 2 min, the time required for completion of the rapid phase of the IP₃-induced Ca²⁺ release response, the medium was removed, and the amount of ⁴⁵Ca²⁺ released was determined, corrected for passive ⁴⁵Ca²⁺ leakage, and expressed as percentage of total releasable ⁴⁵Ca²⁺. Maximum stimulation with IP₃ released a significantly smaller fraction of total releasable Ca²⁺ in h-GAAP cells (p < 0.005, Figure 2C). The latter finding was not likely to be due to a decrease in the number of IP₃Rs because immunoblot analysis of cell lysates revealed no detectable difference in expression of the most abundant subtype 3 (Figure 2A).

Calculation of the EC₅₀ value showed that h-GAAP did not affect the sensitivity for IP₃ (1.35 ± 0.33 and 1.28 ± 0.14 μM IP₃ for U2OS-neo and h-GAAP cells, respectively). We observed a small effect of h-GAAP on the cooperativity of the IP₃-induced Ca²⁺ release response, as measured by calculation of the Hill coefficient (1.22 ± 0.22 and 1.02 ± 0.15 for U2OS-neo and h-GAAP cells, respectively), but this difference was not statistically significant. These results indicate that h-GAAP decreases the efficacy of IP₃ without altering its potency. It remains to be established whether this involves a direct or indirect interaction between h-GAAP and IP₃Rs.

**h-GAAP Lowers the Amount of Stored Ca²⁺ in Intact Cells**

Next, we measured the steady-state Ca²⁺ content of IP₃R-regulated Ca²⁺ stores in intact cells expressing h-GAAP. To this end, cells were loaded with the cytosolic Ca²⁺ indicator fura-2, transferred to a Ca²⁺ free medium, and treated either with the Ca²⁺ ionophore ionomycin (1 μM) or the SERCA inhibitor BHQ (20 μM). Because the Ca²⁺ content of these stores is maintained by a pump-leak system, SERCA inhibi-
tion will lead to passive release of stored Ca\textsuperscript{2+} into the cytosol. The results show that both BHQ (Figure 3A) and ionomycin (Figure 3B) evoked a transient rise in cytosolic Ca\textsuperscript{2+} concentration, the amplitude of which was significantly reduced in h-GAAP cells. Importantly, and in contrast to what was observed with histamine, the upstroke of the Ca\textsuperscript{2+} transients induced by BHQ and ionomycin was unaltered in h-GAAP cells.

To investigate the effects of h-GAAP on the Ca\textsuperscript{2+} concentration in ER and Golgi separately, cells were transfected with organelle-targeted aequorins, permeabilized with saponin at 20 h after transfection, and assayed for ATP-dependent Ca\textsuperscript{2+} uptake under "cytosolic" conditions at a free Ca\textsuperscript{2+} concentration of 0.1 \mu M. In both organelles the steady-state Ca\textsuperscript{2+} concentration appeared lower (~20%) in h-GAAP cells (p < 0.01, Figure 3, C and D). Importantly, no major differences in the initial rate of Ca\textsuperscript{2+} uptake were observed, indicating that h-GAAP does not alter the SERCA pump capacity of the intracellular stores.

To establish more firmly that h-GAAP has an effect on the steady-state Ca\textsuperscript{2+} content of the intracellular stores, we next determined this content at different times after transfection of parental U2OS cells with h-GAAP siRNA. Comparison with parental U2OS cells transfected with control siRNA revealed no detectable difference at 20 h after transfection (Figure 3E). At 40 h after transfection, however, the amplitude of the ionomycin-induced increase in cytosolic Ca\textsuperscript{2+} concentration was significantly higher (~35%) in h-GAAP down-regulated cells (p < 0.001). Organelle-targeted aequorins revealed that this increase in Ca\textsuperscript{2+} concentration occurred in both ER and Golgi (p < 0.05, Figure 3F).

**hGAAP Decreases the Sensitivity to Histamine Induction of Oscillatory Cytosolic Ca\textsuperscript{2+} Changes**

In the experiments described thus far, cells were stimulated with a "pharmacological" concentration of histamine (100 \mu M). In the remainder of this study, we assessed the possible consequences of these findings on Ca\textsuperscript{2+} signaling in intact cells under more "physiological" conditions. Cells were loaded with fura-2 and superfused with medium containing (sub)micromolar concentrations of histamine. Digital imaging microscopy of individual cells revealed that 0.3 and 1.0 \mu M histamine increased the cytosolic Ca\textsuperscript{2+} concentration in ~70 and ~95% of the U2OS-neo cells, respectively. For h-GAAP cells, these values were ~20 and ~70%, respectively, indicating a reduced sensitivity to hormonal induction of an increase in cytosolic Ca\textsuperscript{2+} concentration. In a small percentage of the responding U2OS-neo cells (~5–10% at 1.0 \mu M histamine), the initial large Ca\textsuperscript{2+} transient was followed by Ca\textsuperscript{2+} oscillations (Figure 4, A and B). These Ca\textsuperscript{2+} oscil-
DISCUSSION

In this study, evidence is presented that h-GAAP, a novel regulator of cell death, reduces both extracellular Ca\(^{2+}\) influx evoked by staurosporine, a widely used apoptosis inducer, and intracellular Ca\(^{2+}\) release evoked by histamine, known to exert its effect on intracellular Ca\(^{2+}\) through IP\(_3\).

Using a permeabilized cell system, which allows experimental control of the cytosolic compartment, h-GAAP overexpression was demonstrated to lower the efficacy of IP\(_3\), as demonstrated by a reduction of the maximum amount of total (A23187-) releasable Ca\(^{2+}\) that could be released by IP\(_3\). Because neither the amount of IP\(_3\)R nor their affinity for IP\(_3\) were detectably altered in h-GAAP-overexpressing cells, this result suggests that h-GAAP either decreases the IP\(_3\)-sensitive part of the (A23187)-releasable Ca\(^{2+}\) store or, alternatively, decreases the Ca\(^{2+}\) release properties of the IP\(_3\)-channels. In intact cells, h-GAAP overexpression was shown to reduce the amount of total (ionomycin- or BHQ-) releasable Ca\(^{2+}\), consistent with a reduced filling state of the intracellular stores. Consistent with the above results, the cytosolic and mitochondrial Ca\(^{2+}\) increases in response to a pharmacological histamine concentration (100 μM) were down-regulated in intact cells overexpressing h-GAAP and up-regulated when h-GAAP was knocked down. Furthermore, stimulation with a more close to physiological concentration of histamine (1 μM) revealed that h-GAAP rendered cells less sensitive to the induction of cytosolic Ca\(^{2+}\) oscillations, characteristic for these low concentrations of histamine. Together, these data suggest that h-GAAP reduces both the total amount of releasable Ca\(^{2+}\) and its maximum amount that can be released by IP\(_3\), thereby attenuating IP\(_3\)-induced cytosolic and mitochondrial Ca\(^{2+}\) signaling.

How h-GAAP exerts these effects remains to be established. In this study, we showed that h-GAAP inhibits the influx of extracellular Ca\(^{2+}\) influx and decreases the IP\(_3\)-mediated release of Ca\(^{2+}\) from the stores. In addition, we showed that h-GAAP co-precipitated with IP\(_3\)Rs, suggesting an interaction. Such an interaction, which may be either direct or indirect, could be involved in the ability of h-GAAP

Figure 4. h-GAAP reduces the sensitivity to hormonal induction of cytosolic Ca\(^{2+}\) oscillations and reduces their frequency. (A and B) Fura-2-loaded cells were stimulated with different concentrations of histamine at the indicated time and monitored for their Ca\(^{2+}\) response. (A) Average percentage of the responding cells (±SEM) that produce Ca\(^{2+}\) oscillations. ** p < 0.01. (B) Representative traces of three oscillating neo and three h-GAAP cells. (C and D) Fura-2–loaded cells transfected with either control siRNA or h-GAAP siRNA were stimulated with different concentrations of histamine at the indicated time and monitored for their Ca\(^{2+}\) response. (C) Representative histamine-induced Ca\(^{2+}\) oscillations of three cells transfected with either siRNA. (D) Average percentage of cells (±SEM) that respond with Ca\(^{2+}\) oscillations to the indicated histamine concentrations. * p < 0.05.
to suppress Ca\(^{2+}\) fluxes. However, the observed effects of h-GAAP may equally well be explained by its ability to reduce the filling state of the Ca\(^{2+}\) stores. Therefore, the importance of this interaction for the observed function of h-GAAP requires further investigation.

A reduction in Ca\(^{2+}\) filling state of the intracellular stores is usually associated with an increase in capacitative Ca\(^{2+}\) entry across the plasma membrane, resulting in an increase in cytosolic Ca\(^{2+}\) concentration. However, under resting conditions no such increase in cytosolic Ca\(^{2+}\) concentration was observed in cells overexpressing h-GAAP, despite a decrease in the amount of total releasable Ca\(^{2+}\). This result suggests that h-GAAP exerts an inhibitory effect on the process of capacitative Ca\(^{2+}\) entry. In agreement with this idea, the STS-induced increase in cytosolic and mitochondrial Ca\(^{2+}\) concentration, which depended completely on the presence of extracellular Ca\(^{2+}\) and was abolished by 2-APB, an inhibitor of capacitative Ca\(^{2+}\) entry channels, was decreased in cells overexpressing h-GAAP and was increased in cells in which this protein was down-regulated.

STS and histamine increased the cytosolic Ca\(^{2+}\) concentration with different kinetics, a relatively slow increase after addition of STS versus a relatively fast increase after stimulation with histamine. In contrast to STS, histamine readily increased the cytosolic Ca\(^{2+}\) concentration in the absence of extracellular Ca\(^{2+}\), reflecting the IP\(_3\)-induced release of Ca\(^{2+}\) from intracellular stores. The present finding that h-GAAP lowers the histamine-induced increase in cytosolic Ca\(^{2+}\) concentration in the absence of external Ca\(^{2+}\) strongly supports the idea that GAAP exerts its effect by reducing the IP\(_3\)R-mediated release of Ca\(^{2+}\) from intracellular stores. In doing so, h-GAAP likely also reduces the capacitative entry of Ca\(^{2+}\). It remains to be established whether a similar mechanism underlies the inhibitory effect of h-GAAP on the STS-induced entry of extracellular Ca\(^{2+}\).

The ability of h-GAAP to interfere with intracellular Ca\(^{2+}\) signaling poses a plausible explanation for its ability to suppress apoptosis. This idea is supported by observations that modulation of IP\(_3\)R activity (by antisense knockdown, genetic deletion, or using a cell-permeable inhibitory peptide that interferes with the IP\(_3\)R-cytochrome c interaction) rendered cells less sensitive to apoptosis triggered by both intrinsic and extrinsic pathways (Joseph and Hajnoczy, 2007). Moreover, Bcl-2 and Bcl-X\(_{L}\), two major antiapoptotic proteins, interact with the IP\(_3\)R and alter its activity, though in opposite ways: Bcl-2 decreases the IP\(_3\)R opening probability, whereas Bcl-X\(_{L}\) increases it (Oakes et al., 2003; Kim, H. E., Du, F., Fang, M., and Wang, X. (2005). Formation of apoptosome complexes and formation of Golgi Ca\(^{2+}\)/H\(_{11001}\) gradients in pancreatic acinar cells. J. Biol. Chem. 280, 15794–15799.


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