Central and peripheral integration of interrenal and thyroid axes signals in common carp (Cyprinus carpio L.)

Edwin J W Geven, Gert Flik and Peter H M Klaren
Department of Animal Physiology, Faculty of Science, Institute for Water and Wetland Research, Radboud University Nijmegen, Toernooiveld 1, 6525 ED Nijmegen, The Netherlands

Abstract

In teleostean fishes the hypothalamic–pituitary–thyroid axis (HPT axis) and the hypothalamic–pituitary–interrenal axis (HPI axis) regulate the release of thyroid hormones (THs) and cortisol respectively. Since many actions of both hormones are involved in the regulation of metabolic processes, communication between both signal pathways can be anticipated. In this study, we describe central and peripheral sites for direct interaction between mediators of both neuroendocrine axes in the common carp (Cyprinus carpio). Despite suggestions in the literature that CRH is thyrotropic in some fish; we were not able to establish stimulatory effects of CRH on the expression of the pituitary TSHβ subunit gene. In preoptic area tissue incubated with 10^{-7} M thyroxine (T₄) a 2.9-fold increase in the expression of CRH-binding protein (CRHBP) was observed. Thus, T₄ could reduce the bioavailable hypothalamic crh via the up regulation of crhbp expression and hence down regulate the HPI axis. At the peripheral level, cortisol (10^{-6} M), ACTH (10^{-7} M), and α-MSH (10^{-7} M) stimulate the release of T₄ from kidney and head kidney fragments, which contain all functional thyroid follicles in carp, by two- to fourfold. The substantiation of three pituitary thyrotropic factors, viz. TSH, ACTH, and α-MSH, in common carp, allows for an integration of central thyro­tropic signals. Clearly, two sites for interaction between the HPT axis, the HPI axis, and α-MSH are present in common carp. These interactions may be key to the proper regulation of general metabolism in this fish.

Introduction

Thyroid hormones (THs) and corticosteroids are major endocrine signals that are involved in the regulation of fundamental and basic physiological processes in vertebrates. The action of THs is pleiotropic and often permissive, but generally contributes to the regulation of growth, metabolism, development, and metamorphosis (Blanton & Specker 2007). Glucocorticosteroids are considered to be prime stress hormones that govern the stress response through the redistribution of energy toward processes required for coping with the stressor (Wendelaar Bonga 1997). Because of the significance of THs and corticosteroids in the regulation of metabolic processes, we postulate that a bidirectional communication between these endocrine systems is a necessity for the integration and proper functioning of either system.

In teleostean fishes, the THs thyroxine (T₄, 3,5,3′,5′-tetraiodothyronine) and T₃ (3,5,3′-triiodothyronine) are the end products of the hypothalamic–pituitary–thyroid (HPT) axis where hypothalamic TRH stimulates the release of pituitary TSH, which stimulates the release of THs. Similarly, the hypothalamic–pituitary–interrenal (HPI) axis controls the release of cortisol from the interrenal cells in the head kidney, via hypothalamic CRH and pituitary ACTH. The hypothalamic and pituitary components of both axes are inhibited through negative feedback by their respective end products. The multilevel control of TH and cortisol release allows for potential multiple sites of interaction between both endocrine systems.

Interactions between the HPT and HPI axes have been described in teleostean fishes. Long-term and short-term exposure to cortisol or dexamethasone resulted in decreased levels of plasma THs in several fish species (Redding et al. 1984, 1986, Brown et al. 1991, Walpita et al. 2007). These decreased hormone levels were associated with either increased clearance of plasma THs (Redding et al. 1986) or changes in the activity and expression of deiodinases (Walpita et al. 2007). Stimulatory effects of cortisol on the HPT axis have also been suggested in teleost fish. In brook charr (Salvelinus fontinalis) long-term exposure to cortisol increased the hepatic conversion of T₄ to T₃ (Vijayan et al. 1988) and in Japanese flounder (Paralichthys olivaceus) cortisol augmented the effects of THs on the resorption of the dorsal fin ray (de Jesus et al. 1990). Experimental data on the effects of THs on the HPI axis in teleosts are scarce. In pre- and post-smolt coho salmon (Oncorhynchus kisutch), thyroxine treatment resulted in increased and decreased sensitivity of the head kidney to ACTH respectively (Young & Lin 1988).

In several species from all non-mammalian vertebrate classes, CRH does not only stimulate the release of ACTH,
but also that of tsh (De Groef et al. 2006). Indeed, in common carp (Cyprinus carpio), CRH has been suggested to exhibit thyrotropic activity. In this species, TRH does not stimulate the release of TSH from cultured pituitary cells (Kagabu et al. 1998). Moreover, experimental treatment with thyroxine resulted in a marked hypocortisolemia in carp, which was accompanied by an increased mRNA expression of CRH-binding protein (CRHBP) in the preoptic area and unchanged levels of CRH and prepro-TRH mRNA (Geven et al. 2006). It appears that, in common carp, not TRH but CRH is controlling the activity of the thyroid gland. Because of its corticotropin and putative thyrotropic activity, CRH neurons may constitute a central site for the communication between the HPT and the HPI axis in common carp.

An investigation on the location of the thyroid gland in common carp revealed another putative site for the integration of the HPT and the HPI axes signals. Whereas in most fishes the thyroid follicles are located in the subpharyngeal region, surrounding the ventral aorta, in common carp all functional thyroid follicles, as characterized by iodine uptake and TSH-mediated T₄ release, are scattered throughout the kidney and head kidney (Geven et al. 2007). The close juxtaposition in the head kidney of TH-producing follicles to cortisol-producing interrenal cells strongly hints at a paracrine interaction between both endocrine tissues.

Besides the apparent communication between HPT and HPI axes signals in common carp, a third endocrine signal appears to be involved in these axes. Plasma levels of α-MSH are increased in hyperthyroid and stressed carp (Metz et al. 2005, Geven et al. 2006), the exact physiological role of which still is unclear. We hypothesize that in common carp the preoptic area and the head kidney represent a central and a peripheral site respectively, for the integration of signals of the HPT and the HPI axis. In this study, we investigated this hypothesis by performing in vitro incubations of preoptic area tissues, pituitary glands, and renal tissues with several mediators of both neuroendocrine systems, including α-MSH.

Materials and Methods

**Animals**

Common carp (C. carpio), hereafter called carp, of the all male E4XR3R8 isogenic strain (Bongers et al. 1998) were obtained from the Department of Fish Culture and Fisheries of Wageningen University (The Netherlands). Fish were kept in 140-L tanks with aerated, circulating, city of Nijmegen tap water, at a photoperiod of 16h light:8h darkness at 23 °C. Carp were fed Trouvix dry food pellets (Trouv Nutrition International, Putten, The Netherlands) once daily at a ration of 1-5% of the estimated body weight. Before collection of tissues, fish were deeply anesthetized with 0-1% (v/v) 2-phenoxyethanol and killed by spinal transection. Animal handling followed approved university guidelines.

Static incubation of preoptic area and pituitary gland

The pituitary gland and the preoptic area containing the nucleus preopticus (NPO) were dissected as described by Metz et al. (2006b). The preoptic areas were diced in ~2 mm³ sized fragments, while the pituitary glands were kept intact. The quality of dissection was assured by stereomicroscopic analysis. The preoptic area fragments and pituitary glands were immediately transferred to 1 ml ice-cold Leibovitz's L-15 medium (Invitrogen) containing 100 μg/ml kanamycin (Invitrogen), and antibiotic/antimycotic (1 X) (Invitrogen). After 1 h, the preoptic area fragments were carefully transferred to 1 ml fresh L-15 medium supplemented with T4 (at concentrations of 10⁻⁸ and 10⁻⁷ M respectively), while the pituitary glands were transferred to 1 ml fresh L-15 medium supplemented with T₄ (10⁻⁸ and 10⁻⁷ M), ovine (O) CRH (10⁻⁷ M), or human TRH (10⁻⁷ M). L-15 medium of controls did not receive any supplement. Thyroxine, oCRH, and hTRH were obtained from Sigma Chemical Co.

Preoptic area fragments and the pituitary glands were incubated for periods as indicated in the legends to the figures. All tissues were incubated at 22 °C, while continuously shaking (200 r.p.m). Each medium was replaced at 1, 6, 12, and 24 h after the start of incubation. After the incubation, the preoptic area fragments and pituitary glands were immediately stored at −80 °C until further processing.

**RNA extraction and cDNA synthesis**

To extract total RNA, the preoptic area fragments and the pituitary glands were homogenized in 500 μl TRIzol reagent (Invitrogen) by ultrasonification. Following treatment with DNase, 1 μg RNA was reverse transcribed to cDNA in a 20 μl reaction mixture containing 300 ng random primers, 0.5 mM dNTPs, 10 mM dithiothreitol, 10 U RNase DNase, 1 mg RNA was reverse transcribed to cDNA in a 25 ml reaction mixture consisting of 300 ng random primers, and hTRH were obtained from Sigma Chemical Co.

Since homologous antibodies against the peptides we wished to quantify are not available for carp, and heterologous antibodies were validated for qualitative purposes only, we measured the expression of αh, prepro-trh (pp-trh), urotensin I (utsI), and αhbp mRNA in the preoptic area and of TSHβ (tshb) subunit, proopiomelanocortin (pomc), proproconvertase subtilisin/kexin type 1 (pksk1), and prolactin (prl) mRNA in pituitary gland by real-time quantitative PCR (RQ-PCR). In general, cDNA was diluted ten times, but a portion of pituitary gland cDNA was diluted 500 times to measure pomc gene expression. Totally 5 μl cDNA was used in a 25 μl reaction mixture consisting of 12.5 μl Sybr Green Master Mix (PE Applied Biosystems Benelux, Nieuwerkerk aan den IJssel, The Netherlands), and 3 μl of each primer (600 nM final concentration). The primer sets used for PCR are shown in Table 1. The RQ-PCR was performed on a

Table 1 Primer sequences with corresponding GenBank accession numbers. Open reading frame positions are relative to the start codon.

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GeneAmp 5700 Sequence Detection System (PE Applied Biosystems). The reaction mixture was incubated for 10 min at 95 °C, followed by 40 cycles of 15 s denaturation at 95 °C and 1 min annealing and extension at 60 °C. Analysis of dissociation plots confirmed the specificity of the PCRs. Cycle threshold values were determined from amplification curves. The expression of genes of interest was calculated relative to 40S ribosomal protein S11 mRNA expression.

Static incubation of kidney and head kidney

Head kidneys and kidney tissue were removed from the animal, diced into ~ 2 mm3 sized fragments and immediately placed in appropriate volume of ice-cold Leibovitz's L-15 medium (Invitrogen) containing 100 μg/ml kanamycin (Invitrogen) and antibiotic/antimycotic (1X) (Invitrogen). After 1 h, the head kidney and kidney fragments were carefully transferred to 2 ml fresh L-15 medium supplemented with cortisol (at concentrations of 10-7 and 10-6 M respectively) human ACTH (10-7 M) or monoacetyl α-MSH (10-7 M), while control incubations did not receive any supplement. Cortisol, hACTH, and α-MSH were from Sigma Chemical Co.

Tissues were incubated for 24 h at 22 °C, while continuously shaking (200 r.p.m.), after which the 2 ml incubation medium was separated from the tissue by centrifugation (4 °C, 1000 g, 15 min) and reduced to 0.5 ml by vacuum drying. The incubation medium was applied to a Sephadex LH-20 column to isolate the THs and to remove salts (Mol & Visser 1985). In short, glass pipettes were filled with 1 ml Sephadex LH-20 (Amersham Biosciences) suspension in water (10% w/v) and equilibrated with 3 volumes of 1 ml 0.1 M HCl. Samples were acidified with an equal volume of 1 M HCl and loaded on to the column. The samples were then eluted from the column with 5 volumes of 1 ml 0.1 M HCl for the removal of ions, 4 volumes of 1 ml H2O to neutralize the column, and 3 volumes of 1 ml 0.1 M NH4/ETOH to collect THs. The fractions containing THs were vacuum dried and reconstituted in 60 μl 50 mM sodium barbitone/0.1% BSA buffer (pH 8.6). Total thyroxine was measured in duplicate with a total T4 ELISA (Human Gesellschaft für Biochemica und Diagnostica GmbH, Wiesbaden, Germany) according to the manufacturer’s instruction. Standards were prepared in the same barbitone buffer as the samples were. The intra-assay and inter-assay coefficients of variation for the T4 ELISA reported by the manufacturer are 4-2 and 3-3% respectively. The reported cross reactivity of the ovine anti-T4 antibody to D-T4 is 98% (the reactivity to L-T4 is set at 100% as a reference), and to L-T3 and D-T3 is 3 and 1.5% respectively. Cross-reactivities of the antibody to diiodothyronine, diiodothyrosine, and iodothyrosine are <0.01%. The reported sensitivity of the T4 ELISA is 4 ng/ml T4. Protein content of the incubated tissues was determined by Bio-Rad protein assay (Bio-Rad) using BSA as a reference.

Statistical analysis

All data are represented as mean values ± s.d. The number of different preparations (n) is given in parentheses. Differences between groups were assessed with Student’s parametric t-test for unpaired observations, or Mann–Whitney’s non-parametric U-test, where appropriate. Statistical significance was accepted at P<0.05 (two-tailed), probabilities are indicated by asterisks (*, P<0.05; **, P<0.01; ***, P<0.001).

Results

Effects of CRH and TRH on pituitary tshb gene expression

Thyroxine, at concentrations of 10-8 and 10-7 M, significantly down regulated the expression of tshb subunit mRNA.
by 35% (P=0.01) and 45% (P=0.02) compared with control incubations respectively (Fig. 1A), which demonstrated the viability of the pituitary gland preparation. Neither TRH (10^{-7} M) nor CRH (10^{-7} M) altered *tshb* subunit gene expression (Fig. 1B and C). Incubation of pituitary glands with TRH (10^{-7} M) increased the expression of *pomc* (P=0.04) and *pcsk1* (P=0.002) 1.7- and 2.3-fold respectively (Fig. 2D), which confirmed the bioactivity of TRH. The integrity and bioactivity of the CRH preparation used was confirmed in our laboratory by mass spectrometry and the stimulatory action on the release of ACTH and α-MSH from carp pituitary glands *in vitro* (Metz et al. 2004, van den Burg et al. 2005).

**Effects of T\(_4\) on gene expression in the preoptic area**

Thyroxine, at 10^{-7} M, significantly increased the expression of *crh*, *prepro-trh*, and *crhbp* in the preoptic area 4.6-(P=0.0001), 2.9-(P=0.002), and 2.1-fold (P=0.04) respectively (Fig. 2). The expression of *uts1* remained unchanged (Fig. 2D). Incubation with 10^{-8} M T\(_4\) had no statistically significant effects on the expression of any of the genes tested (Fig. 2).

**Effects of cortisol, ACTH and α-MSH on the release of T\(_4\) from renal tissues**

Cortisol at 10^{-6} M, but not at 10^{-7} M, stimulated the release of T\(_4\) from head kidney (P=0.04) and kidney tissue (P=0.01) 3.5-fold (Fig. 3). ACTH and α-MSH (both at 10^{-7} M) increased the release of T\(_4\) from head kidney and kidney tissue two- to four-fold (Fig. 3). Basal and stimulated T\(_4\) secretion from the head kidney, overall, was ten times lower than that of the kidney (Fig. 3).

**Discussion**

We demonstrate here that the NPO and renal tissues are putative sites for interaction between mediators of the HPT- and the HPI axis in carp. Thyroxine affects the CRH system in the preoptic area. Peripherally, cortisol and ACTH both stimulate the release of THs from renal tissues, viz. the head kidney and kidney.

Despite indications that preoptic CRH may be involved in the regulation of the HPT axis in some fish (Kagabu et al. 1998, Geven et al. 2006, De Groef et al. 2006), no effect of CRH on the expression of carp pituitary *tshb* subunit could be demonstrated *in vitro*. The absence of a thyrotropic action of preoptic CRH can also be inferred from experimental results obtained *in vivo*. When carp were exposed to a 24 h confinement stressor, the HPI axis was markedly activated, as exemplified by an increased expression of *crh* mRNA in the preoptic area (Huising et al. 2004). However, this increase was not accompanied by increased expression of pituitary *tshb*.

Figure 1 Relative mRNA expression levels of pituitary *tshb* subunit upon (A) 36 h incubation with T\(_4\) (10^{-8} M, n=6 and 10^{-7} M, n=5) and (B) 6, 12, 24, and 36 h incubation with TRH (10^{-7} M, n=4) and (C) CRH (10^{-7} M, n=6). Relative mRNA expression of pituitary *pomc* and *pcsk1* upon (D) 6 h incubation with TRH (10^{-7} M, n=4).
In the thyroid interaction in carp, the CRH and TRH systems play a crucial role. These neuropeptides are synthesized in the hypothalamus and released into the bloodstream to stimulate the release of thyroid-stimulating hormones (TSH) from the pituitary gland. This interaction is essential for the regulation of thyroid function.

The expression of CRH and TRH is regulated by a variety of factors, including thyroid hormones (THs). THs can modulate the activity of the hypothalamic-pituitary-thyroid (HPT) axis at different levels. In vitro studies have shown that exposure to THs can alter the expression of CRH and TRH genes. For instance, T4 (thyroxine) has been found to increase the mRNA expression of crh (CRH) and trh (TRH) genes in hypothalamic tissue (Geven et al., 2006).

The hypothalamic CRHBP (CRH-binding protein) plays a significant role in regulating the bioavailability of CRH. CRHBP binds to CRH and reduces its bioactivity, thereby controlling the release of ACTH (adrenocorticotropic hormone) from the pituitary gland. The expression of crhbp mRNA is increased in response to stress and thyroid hormones (Geven et al., 2006).

The regulation of CRH and TRH expression by THs is not only limited to the hypothalamus but also affects other tissues, such as the kidney. Exposure to THs can alter the expression of these genes in kidney tissue as well (Geven et al., 2006).

In conclusion, the interaction between the thyroid and hypothalamic systems is complex, and various factors, including THs, play a crucial role in regulating the expression of CRH and TRH genes. Understanding this interaction is essential for the proper functioning of the thyroid gland and maintaining hormonal balance.
functional protein concentrations in the hypothalamus in situ is difficult to estimate and awaits the development of a quantitative assay for CRHBP protein. Still, our data suggests that CRHBP from the preoptic area, where expression is T4-sensitive in vitro as well as in vivo, may fulfill a role as a central messenger that allows for the interaction of the HPT axis with the HPI axis in carp.

We also identified a peripheral site for the interaction between the HPT and the HPI axes: the head kidney and kidney. We already established that short- and long-term incubation with THs have no effect on the release of cortisol from head kidney fragments (Geven et al. 2006). Conversely, exposure of head kidney and kidney fragments to cortisol and ACTH stimulated the release of T4 from these tissues. The TH-releasing properties of cortisol and ACTH are consistent with the expression of their specific receptors, i.e. the glucocorticoid receptor and the type 2 melanocortin receptor respectively, in head kidney as well as kidney (Metz et al. 2005, Stolte et al. 2008).

Although cortisol has been shown to stimulate iodide uptake and thyroglobulin synthesis in synergy with TSH in several mammalian thyroid cell cultures (Roger & Dumont 1983, Gérard et al. 1989, Becks et al. 1992, Takiyama et al. 1994), we describe here a direct and independent effect of cortisol on the thyroid gland of a teleost. Since the kidney is devoid of interrenal cells and a local cortisol-mediated effect therefore is not possible, we conclude that ACTH has a direct effect on the release of T4 in the kidney. Such a direct effect of ACTH could also apply to the head kidney. However, the ACTH-induced release of T4 may also represent a paracrine effect of endogenous cortisol released upon stimulation by ACTH, as exogenous cortisol mimicked the effect of ACTH on the release of T4 from head kidney tissue. Studies into the cellular localization of the type 2 melanocortin receptor in head kidney tissue may reveal the presence of this receptor in thyrocytes, and thus can provide evidence for a direct mode of action of ACTH.

Another novel finding of this study is that the pituitary POMC-derived hormone α-MSH also has thyroid-stimulating properties in carp. The effect of α-MSH in the kidney is consistent with the expression of type 5 melanocortin receptor (Metz et al. 2005). Although the MC5R is not expressed in the head kidney of carp, the release of pituitary α-MSH is mainly attributed to TRH (Lamers et al. 1994, van den Burg et al. 2003, 2005). The in vitro stimulation of αMSH expression by T4, and the in vivo stimulation of pomc and pcsk1 expression by TRH observed in this study, is commensurate to the endocrine cascade by which THs control the release of α-MSH in carp.

The widespread distribution of melanocortin receptors in fish illustrates the pleiotropic functions for α-MSH, which includes the regulation of food intake and metabolism (Metz et al. 2006a). The intracerebroventricular injection of [Nle4, α-Phe7] α-MSH, an α-MSH agonist, inhibited food intake in goldfish (Carassius auratus; Cerdá-Reverter et al. 2003) and peripherally, α-MSH exhibited lipolytic effects in hepatocytes of rainbow trout (Oncorhynchus mykiss; Yada et al. 2000, 2002). The concerted actions of THs, cortisol, and α-MSH on the regulation of metabolic processes may form the basis for the integration of these signals in carp.

In conclusion, this study identifies a central and a peripheral site in carp for the communication between the interrenal axis and the thyroid axis and, additionally, α-MSH as a thyroid-stimulating factor. Centrally, T4 can inhibit the HPI axis via CRHBP in the preoptic area. Peripherally, cortisol and ACTH stimulate the release of T4 from renal tissues, as does α-MSH. The intimate interrelationships between these neuroendocrine systems are pivotal for the regulation of general metabolism.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References
