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Improved Tumor Targeting of Radiolabeled RGD Peptides Using Rapid Dose Fractionation

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ABSTRACT

Arginine-glycine-aspartic acid (RGD) peptides preferentially bind to αvβ3 integrin, an integrin expressed on newly formed endothelial cells and on various tumor cells. When labeled with β-emitting radionuclides, these peptides can be used for peptide-receptor radionuclide therapy of malignant tumors. These studies aimed to investigate whether tumor targeting and tumor therapy could be optimized by dose fractionation.

The RGD-peptide DOTA-E-[c(RGDfK)]2 was labeled with 111In for biodistribution experiments and with 90Y for therapy experiments. In mice with NIH:OVCAR-3 ovarian carcinoma xenografts, optimal tumor uptake was obtained at peptide doses up to 1.0 µg (4.8 %ID/g). A peptide dose of 5 µg, required to administer the maximum tolerable dose (MTD) 90Y-DOTA-E-[c(RGDfK)]2, was administered as 5 portions of 1.0 µg. Tumor uptake of the fifth portion was significantly higher than that of the single 5.0 µg portion (3.3 %ID/g versus 2.1 %ID/g). The therapeutic efficacy of 37 MBq 90Y-DOTA-E-[c(RGDfK)]2 (1 × 5.0 µg) was compared with that of 37 MBq administered in five equal portions (5 × 1.0 µg). No difference in tumor growth between the fractionated and the nonfractionated therapy was observed. In conclusion, dose fractionation resulted in higher radiation doses. However, therapeutic efficacy of the radiolabeled peptide was not significantly improved by dose fractionation.

Key words: RGD peptides, αvβ3 integrin, tumor targeting, dose fractionation

INTRODUCTION

Integrins are a group of adhesion molecules consisting of two noncovalently bound transmembranous subunits (α and β). Integrin αvβ3 mediates cellular adhesion to extracellular matrix proteins (e.g., vitronectin, fibrinogen, laminin, collagen) through exposed tripeptide arginine-glycine-aspartic acid (RGD) amino-acid moieties.1 Integrin αvβ3 is preferentially expressed on proliferating endothelial cells.2 For their growth beyond a certain size, tumors are dependent on the formation of new blood vessels from preexisting capillaries (angiogenesis) and thus αvβ3 is expressed on the blood vessels of most growing tumors.3-5 In addition, αvβ3 is expressed on various tumor cell types (ovarian cancer, neuroblastoma, breast cancer, melanoma, and others). Due to the restricted expression of αvβ3 in tumors, αvβ3 is considered a suitable membrane structure for tumor targeting.6

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The potential of αβ3-binding peptides to serve as vehicles for targeting tumors with radionuclides has been investigated by several groups. Hauben et al. studied two radiolabeled cyclic pentapeptides (c(RGDyV) and c(RGDyD); IC50 = 2 nM) in nude mice with various tumors. Tumor uptake peaked 10 minutes after injection (3.0 ± 0.31 %ID/g versus 0.92 ± 0.16 %ID/g at 4 hours p.i.). Van Hagen et al. studied the cyclic peptide c(RGDyK) conjugated with DTPA. Uptake of the 111In-labeled peptide in CA20948 pancreatic tumors in rats amounted to 0.2 %ID/g at 24 hours p.i., while at this timepoint, the peptide was almost completely cleared from the blood (0.003 %ID/g).

In our previous studies, we have shown that the radiolabeled c(RGDyK) peptides specifically accumulated in αβ3-positive ovarian cancer xenografts in athymic mice. The dimeric RGD peptide was retained better in the tumor than the monomeric RGD peptide. The uptake of 111In-DOTA-E-[c(RGDyK)]2 in subcutaneous (s.c.) OVCAR-3 tumors peaked at 7.5 %ID/g 2 hours after intravenous injection. A single injection of this peptide labeled with 99mTc (37 MBq/mouse) caused a significant growth delay of the tumors. In the present study, the optimal peptide dose for peptide radionuclide therapy with the dimeric RGD peptides was determined. In addition, the effect of rapid dose fractionation on the radiation dose guided to the tumors and on the therapeutic efficacy was studied.

**MATERIALS AND METHODS**

**Radiolabeling of the RGD Peptide**

The synthesis and characterization of the dimeric cyclic RGD peptide E-[c(RGDyK)]2; conjugated with the chelator 1,4,7,10-tetraazadodecane- N,N,N,N-tetracetic acid (DOTA) was described previously. The peptide has a high affinity for the αβ3 integrin (IC50 = 0.9 nM). DOTA-E-[c(RGDyK)]2 was radiolabeled with 111In to obtain 111In-DOTA-E-[c(RGDyK)]2.

Briefly, 250 µg DOTA-E-[c(RGDyK)]2 was dissolved in 4.5 mL of 0.25 M ammoniumacetate buffer, pH 7.0 and 160 MBq 111InCl3 (Mallinckrodt, Petten, The Netherlands) was added. The mixture was heated for 15 minutes at 100°C. The radiochemical purity was determined by reversed-phase high-performance liquid chromatography (RP-HPLC) (HP 1100 series, Hewlett Packard, Palo Alto, CA) using a C18 column (RX-C18, 4.6 × 250 mm, Zorbax) eluted with an isocratic eluent (33% acetonitrile in 25 mM phosphate buffer, pH 6.0) at 1.0 mL/minute. The radioactivity of the eluate was monitored using an in-line radiodetector (Radiomatic Flo-One Beta series A-500, Packard BioScience, Zellik, Belgium).

**Animal Model**

In our animal model, 0.2 mL of a cell suspension of NIH:OVCAR-3 human ovarian carcinoma cells (5 × 10⁷ cells/mL) was injected s.c. in the right flank of 6–8 week-old female nude BALB/c mice. The NIH:OVCAR-3 tumor cells express the αβ3 integrin. Two (2) weeks after inoculation of the tumor cells, when the diameter of the tumors was 6–8 mm, the mice were i.v. injected with the radiolabeled peptides via the tail vein.

**Biodistribution Studies**

In these experiments, the effect of the peptide dose escalation and the effect of peptide dose fractionation on the biodistribution of the radio-labeled peptide was determined. Five groups of five mice received 0.5, 1.0, 2.0, 5.0, or 10 µg of the DOTA-E-[c(RGDyK)]2 peptide labeled with 0.4 MBq 111In (dose escalation). Two (2) hours after injection of the radiolabeled peptide, the mice were euthanized with CO2/O2. Blood was collected by cardiac puncture, and tissues (tumor, muscle, lung, spleen, kidney, liver, small intestine) were dissected, counted, and weighed.
The effect of dose fractionation on the biodistribution of $^{111}$In-DOTA-E-[c(RGDfK)]$_2$ was studied in two groups of mice which received four injections with 1 $\mu$g of nonradioactive DOTA-E-[c(RGDfK)]$_2$ with intervals of 1 hour or 2 hour. One (1) or 2 hours after the fourth injection, the mice in these groups received 1 $\mu$g of DOTA-E-[c(RGDfK)]$_2$ labeled with 0.4 MBq $^{111}$In. Two (2) hours after injection of the radiolabeled peptide, the mice were euthanized with CO$_2$/O$_2$. Blood was collected by cardiac puncture, and tissues (tumor, muscle, lung, spleen, kidney, liver, small intestine) were dissected and weighed.

The activity in tissues and injection standards was measured in a shielded well-type scintillation gamma counter (Wizard, Pharmacia, Turku, Finland) and expressed as %ID/g.

Peptide-Receptor Radionuclide Therapy

The therapeutic efficacy of 37 MBq $^{99m}$Tc-DOTA-E-[c(RGDfK)]$_2$ (specific activity: 7.4 MBq/$\mu$g) administered as a bolus injection was compared to the therapeutic efficacy of the same dose of $^{99m}$Tc-DOTA-E-[c(RGDfK)]$_2$ administered in five equal portions. Four groups of 10 mice with s.c. NIH:OVCAR-3 tumors were used in this experiment (Groups A–D). Group A received 37 MBq $^{99m}$Tc-DOTA-E-[c(RGDfK)]$_2$ (peptide dose: 5.0 $\mu$g) as a single bolus injection (0.2 mL). Group B received the same dose of $^{99m}$Tc-DOTA-E-[c(RGDfK)]$_2$ administered in 5 equal portions ($5 \times 7.4$ MBq: $5 \times 1.0 \mu$g) with a 1-hour interval between the five injections. The mice in group C received 5 equal doses of the scrambled sequence control peptide $^{99m}$Tc-DOTA-E-[c(RGKfD)]$_2$ ($5 \times 7.4$ MBq, $5 \times 1.0 \mu$g, 1-hour interval). The mice in group D received 5 subsequent injections with nonradioactive DOTA-E-[c(RGKfD)]$_2$ peptide ($5 \times 1.0 \mu$g, 1-hour interval) and also served as a control group.

The size of the tumors was measured twice weekly in 3 dimensions using a caliper. The volume was estimated assuming the tumors were ellipsoids, using the formula: volume = $4/3\pi r_1 r_2 r_3$, where $r_1$, $r_2$, and $r_3$ are the length, width, and height, respectively. At the time of the injection the volume of the s.c. tumors ranged from 27 to 150 mm$^3$. When the volume of the tumors exceeded 2 cm$^3$, the mice were taken out of the experiment and euthanized.

Statistical Analysis

Statistical analysis was performed using a GraphPad Prism 3.0 (GraphPad Software, San Diego, CA). The biodistribution data were analyzed using the one-way analysis of variance ANOVA test. The level of significance was set at $p = 0.05$. If $p < 0.05$, Tukey-Kramer multiple comparisons post-tests were performed.

RESULTS

Radionuclide Peptides

RP-HPLC analysis indicated that the radiochemical purity of DOTA-E-[c(RGDfK)]$_2$, labeled with $^{111}$In and $^{99m}$Tc, was 96% and 95%, respectively. The $\alpha\beta$ binding capacity of the $^{111}$In-labeled peptide was confirmed in an in vitro cell binding assay on IGROV-1 human ovarian carcinoma cells as described previously.

Peptide-Dose Escalation Study

The biodistribution of $^{111}$In-DOTA-E-[c(RGDfK)]$_2$ was studied in athymic mice with s.c. OVCAR-3 tumors at a peptide dose of 1.0 $\mu$g per mouse. The biodistribution data were analyzed using the one-way analysis of variance ANOVA test. The level of significance was set at $p = 0.05$. If $p < 0.05$, Tukey-Kramer multiple comparisons post-tests were performed.
%ID/g, p < 0.01). The uptake of the fractionated dose was not affected by the time interval between the injections (1 hour or two h, p > 0.05).

**Peptide-Receptor Radionuclide Therapy**

The MTD of \(^{90}\text{Y}-\text{DOTA-E-}[\text{c(RGDfK)}]\) in this mouse model is 37 MBq per mouse as determined previously. The maximum specific activity of \(^{90}\text{Y}\)-labeled DOTA-E-[c(RGDfK)]\(_2\) is 7.4 MBq/µg. Therefore, the MTD of the peptide has to be administered with a peptide dose of at least 5.0 µg. The biodistribution experiments indicated that 5.0 µg of peptide can be targeted more efficiently to the OVCAR-3 tumor in this mouse model when administered as a fractionated dose (5 × 1.0 µg). Therefore, the therapeutic efficacy of a single bolus injection of 37 MBq \(^{90}\text{Y}\)-labeled DOTA-E-[c(RGDfK)]\(_2\) was compared to that of the same-activity dose administered as five subsequent injections with a 1-hour time interval in a peptide-receptor radionuclide therapy experiment (Fig. 3).

Dose fractionation of the activity dose of 37 MBq of \(^{90}\text{Y}\)-DOTA-E-[c(RGDfK)]\(_2\) in five equal portions did not improve the growth-inhibitory effect of the radionuclide therapy compared to therapy with 37 MBq of \(^{90}\text{Y}\)-DOTA-E-[c(RGDfK)]\(_2\) administered in one injection of 5.0 µg of peptide. The growth rate of the tumors in both of these groups was significantly delayed compared to the growth of the tumors in the mice that received \(^{90}\text{Y}\)-labeled scrambled control peptide or nonradioactive peptide.

**DISCUSSION**

The present study aimed to optimize peptide-receptor radionuclide therapy with a \(^{90}\text{Y}\)-labeled dimeric cyclic RGD peptide using rapid-dose fractionation. The maximum specific activity of the \(^{90}\text{Y}\)-labeled DOTA-E-[c(RGDfK)]\(_2\) peptide that we use for peptide receptor radionuclide therapy is 7.4 MBq/µg, while a total activity dose of 37 MBq can be administered safely to mice. Therefore, a 37 MBq dose contains at least 5.0 µg of the DOTA-E-[c(RGDfK)]\(_2\) peptide. However, the peptide dose escalation study indicated that, at peptide doses exceeding 1.0 µg/mouse, the uptake in the tumor decreased, most likely due to saturation of the αβ₃ binding sites in the tumor.

**Table 1.** Biodistribution (%ID/g ± SD) of \(^{111}\text{In}-\text{DOTA-E-}[\text{c(RGDfK)}]\) at Various Peptide Doses in Athymic Mice with Subcutaneous OVCAR-3 Tumors 2 H after Injection

<table>
<thead>
<tr>
<th>Dose</th>
<th>0.5 µg</th>
<th>1.0 µg</th>
<th>2.0 µg</th>
<th>5.0 µg</th>
<th>10 µg</th>
<th>5 × 1.0 µg 1-h interval</th>
<th>5 × 1.0 µg 2-h interval</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood</td>
<td>0.12 ± 0.02</td>
<td>0.08 ± 0.01</td>
<td>0.08 ± 0.01</td>
<td>0.06 ± 0.01</td>
<td>0.06 ± 0.01</td>
<td>0.08 ± 0.01</td>
<td>0.04 ± 0.01</td>
</tr>
<tr>
<td>Muscle</td>
<td>0.42 ± 0.02</td>
<td>0.28 ± 0.03</td>
<td>0.27 ± 0.07</td>
<td>0.13 ± 0.01</td>
<td>0.16 ± 0.08</td>
<td>0.29 ± 0.11</td>
<td>0.28 ± 0.06</td>
</tr>
<tr>
<td>Tumor</td>
<td>4.87 ± 0.60</td>
<td>4.73 ± 0.65</td>
<td>3.20 ± 0.26</td>
<td>2.07 ± 0.46</td>
<td>1.54 ± 0.15</td>
<td>3.28 ± 0.60</td>
<td>3.18 ± 0.56</td>
</tr>
<tr>
<td>Lung</td>
<td>0.97 ± 0.07</td>
<td>1.11 ± 0.09</td>
<td>0.72 ± 0.08</td>
<td>0.47 ± 0.09</td>
<td>0.44 ± 0.06</td>
<td>0.70 ± 0.10</td>
<td>0.64 ± 0.01</td>
</tr>
<tr>
<td>Spleen</td>
<td>2.03 ± 0.08</td>
<td>1.99 ± 0.11</td>
<td>1.50 ± 0.20</td>
<td>1.14 ± 0.15</td>
<td>0.79 ± 0.03</td>
<td>1.58 ± 0.12</td>
<td>1.23 ± 0.10</td>
</tr>
<tr>
<td>Kidney</td>
<td>2.91 ± 0.11</td>
<td>2.93 ± 0.28</td>
<td>2.37 ± 0.36</td>
<td>2.04 ± 0.18</td>
<td>2.26 ± 0.12</td>
<td>2.08 ± 0.14</td>
<td>1.93 ± 0.25</td>
</tr>
<tr>
<td>Liver</td>
<td>2.18 ± 0.08</td>
<td>2.03 ± 0.26</td>
<td>1.55 ± 0.24</td>
<td>0.93 ± 0.10</td>
<td>0.70 ± 0.08</td>
<td>1.50 ± 0.19</td>
<td>1.17 ± 0.14</td>
</tr>
<tr>
<td>Intestine</td>
<td>3.46 ± 0.37</td>
<td>3.08 ± 0.92</td>
<td>2.51 ± 0.39</td>
<td>1.61 ± 0.30</td>
<td>1.06 ± 0.14</td>
<td>2.25 ± 0.30</td>
<td>1.93 ± 0.29</td>
</tr>
</tbody>
</table>

In the two groups of mice that received a fractionated dose (5 × 1.0 µg), the biodistribution of the fifth fraction of the dose is shown.
tumor. Similarly, the uptake in most normal tissues tested (lung, spleen, kidney, liver, and intestine) was significantly lower at the higher peptide doses. This observation was in line with earlier studies in which we showed that part of the uptake of the peptide in normal tissues can be blocked with an excess of cold peptide, indicating that the uptake in these tissues is saturable. Whether this is due to expression of αvβ3 integrin in these tissues remains to be investigated.

When the 5.0-mg dose was administered in five equal portions with a 1- or 2-hour time interval, the uptake in the tumor of the last 1.0-mg portion (3.2 ± 0.2 %ID/g) was significantly higher than that of the 5.0-mg dose injected as a bolus (2.1 ± 0.5 %ID/g). These data indicate that the uptake of the 5 × 1.0-μg dose was even higher, because the uptake of the first 1.0-μg fraction in the tumor was 4.7 ± 0.6 %ID/g. These results indicate that the mean uptake of the 5 × 1.0-μg dose is approximately 4.0 %ID/g, being the mean tumor uptake of the first and the fifth dose. Thus, the biodistribution study showed that rapid-dose fractionation can increase the uptake of the radiolabeled peptide almost twofold. These data suggest that within 1 hour after targeting the αvβ3 binding sites in the tumor with the RGD-peptide, new αvβ3 binding sites are available for binding newly administered RGD-peptide.

Despite the fact that peak tumor uptake of the radiolabeled peptide was higher when the fractionated dose scheme was applied, the therapeutic effect of 37 MBq ⁹⁰Y-DOTA-E-[c(RGDfK)]₂ was not enhanced significantly when administered as 5 equal fractions of 7.4 MBq. This could be due to the fact that the therapeutic effect of ⁹⁰Y-DOTA-E-[c(RGDfK)]₂ administered as a single bolus was considerable.

Various studies have investigated the potential of dose fractionation in radionuclide therapy.

![Figure 2](image2.png)

**Figure. 2.** Tumor uptake of ¹¹¹In-DOTA-E-[c(RGDfK)]₂ peptide in athymic mice with subcutaneous OVCAR-3 tumors 2 hours after injection at various peptide doses. Note that optimal tumor uptake is obtained with a peptide dose of 0.5 and 1.0 μg. At higher peptide doses, the uptake in the tumor was lower due to saturation of the αvβ3 binding sites in the tumor. In addition, the uptake in the tumor of the fifth fraction of five 1.0-μg doses of DOTA-E-[c(RGDfK)]₂ (with 1- or 2-hour time intervals) is shown.

![Figure 3](image3.png)

**Figure. 3.** Growth curves of the s.c. OVCAR-3 tumors in the four groups of mice (7–10 mice/group) after intravenous injection of 37 MBq ⁹⁰Y-DOTA-E-[c(RGDfK)]₂ (5.0 μg) (●), 5 × 7.4 MBq ⁹⁰Y-DOTA-E-[c(RGDfK)]₂ (5 × 1.0 μg) (○), 5 × 7.4 MBq control peptide ⁹⁰Y-DOTA-E-[c(RGKfK)]₂ (5 × 1.0 μg) (■), or nonradioactive DOTA-E-[c(RGDfK)]₂ (5 × 1.0 μg) (□).
rationale and the design of these studies varied largely. Several studies aimed to overcome heterogeneity of intratumoral distribution of the tumor radiation doses. Other studies aimed to reduce marrow toxicity and thus to increase the total radioactivity dose that can be administered. Only a few studies aimed to enhance radioactivity dose to the tumor and thus to enhance the therapeutic effect of the same activity dose antibody administered as multiple injections. In mice with HeLa xenografts, the tumor targeting with 22 MBq 125I-labeled anti-cytokeratin monoclonal antibody administered as one, three, or ten injections was investigated. In contrast with our findings, these investigators found reduced uptake in the tumors when the dose was administered in multiple injections. However, in that study the time interval between the injections was much longer and the antibody protein dose did not saturate the target antigen in the tumor. Goel et al. described a study in mice with LS174T tumors in which the researchers compared the therapeutic efficacy of an activity dose of 37 MBq 131I-labeled CC49 antibody construct either as a single bolus or as 4 injections (4 × 0.25 MBq) with a 1-day time interval. They found that the therapeutic efficacy of the fractionated dose was significantly higher than that of the single injection.

In conclusion, the present study shows that rapid-dose fractionation can improve the targeting of αβ3-expressing tumors with radiolabeled dimeric cyclic RGD peptide. Our studies suggest that 1 hour after targeting the αβ3 binding sites in the tumor with the RGD-peptide, new αβ3 binding sites are available again for binding RGD peptides.

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