Surface location and orientation of the lantibiotic nisin bound to membrane-mimicking micelles of dodecylphosphocholine and of sodium dodecylsulphate

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The interaction of nisin, a membrane-interacting cationic polypeptide, with membrane-mimicking micelles of zwitterionic dodecylphosphocholine and of anionic sodium dodecylsulphate was studied. Direct contacts have been established through the observation of NOEs between nisin and micelle protons. Spin-labeled DOXYL-stearic acids were incorporated into the two micellar systems. From the paramagnetic broadening effects induced in the 'H-NMR spectrum of nisin it is concluded that the molecule is localized at the surface of the micelles. The interactions of nisin with zwitterionic and with anionic micelles resemble each other as do the nisin conformations [van den Hooven, H. W., Doeland, C. C. M., van de Kamp, M., Konings, R. N. H., Hilbers, C. W. & van de Ven, F. J. M. (1995) Eur. J. Biochem. 235, 382-393]. The hydrophobic residues are immersed into the micelles and oriented towards the center, whereas the more polar or charged residues have an outward orientation. The micellar systems are considered to model the first step in the mechanism of antimicrobial action of nisin, this step is the binding of nisin to the cytoplasmic membrane of target bacteria. Detailed information on this initial binding step is obtained. Hydrophobic and electrostatic interactions appear to be involved in the nisin-micelle contacts. It is suggested that subtilin, a lantibiotic structurally related to nisin, has a comparable membrane interaction surface.

Keywords: bacteriocin; lanthionine-containing polypeptide; mode of action; NMR; spin label.
to the membrane. Here we report that in the presence of 

DODPC/cho and of SDS micelles it is located on their surface. 

These observations will be used to describe a model of the interaction between nisin and cellular membranes.

**MATERIALS AND METHODS**

**Chemicals.** Nisin was obtained as a gift of Aplin & Barrett. Prior to its use in NMR studies it was further purified by reverse-phase HPLC (Rollemo et al., 1991). [\(^{1}H_{15}\)]SDS and \([^{1}H_{16}]\)DODPC/cho were obtained from MSD Isotopes. [\(^{1}H_{15}\)]SDS was from Biorad and [\(^{1}H_{15}\)]DODPC/cho was a kind gift of Drs R. Dijkman and H. M. Verheij (University of Utrecht). 5-DOXYL- stearic and 16-DOXYL-stearic acid were purchased from Aldrich, while deuterated methanol was from Merck.

**ESR spectroscopy.** ESR spectra were recorded at 313 K on a Bruker ESP300 X-band spectrometer as described previously (Brown et al., 1981; Mendez et al., 1988, 1991). The pH of the micellar samples was 3.5 (pH meter reading).

**\(^{13}C\)-NMR spectroscopy of SDS and DODPC/cho.** Sample conditions were essentially as previously described (Brown et al., 1981). \(^{13}C\)-NMR spectra were measured at 100.62 MHz on a Bruker AM400 spectrometer, interfaced to a Spectr3000 computer. Standard composite-pulse proton decoupling was used. The experiments were conducted at 313 K. The number of datapoints was 32 k (1 k = 1024), the number of scans 512 and the sweep width 25 ppm or 55 ppm for SDS and DODPC/cho, respectively. The spectral region of 25 ppm for SDS was such that the resonance of the CH\(_2\) group next to the sulphate moiety of SDS was folded back in the spectrum. The data were processed using NMR1 software (New Methods Research, Inc., Syracuse, NY). No window function was used and the data were zero-filled to 64 k. The line width of the \(^{13}C\) resonances was determined by curve fitting.

**\(^{1}H\)-NMR spectroscopy of nisin/micelle complexes with spin-labels.** Samples for the experiments with spin-labels were prepared by first dissolving nisin in 0.5 ml H\(_2\)O/\(^{2}H\)H\(_2\)O (9:1) or in \(^{2}H\)H\(_2\)O to a concentration of 3 mM and then adding [\(^{1}H\)]SDS to a concentration of 100 mM or [\(^{1}H\)]DODPC/cho to a concentration between 100 mM and 120 mM, resulting in a 30–40 fold excess of detergent. The pH was adjusted to 3.5 (pH meter reading). A small aliquot (1–2 \(\mu\)l) of spin-labeled stearic acid, taken from 0.1 M stock solutions in deuterated methanol, was then added.

Spin-label titrations at 298 and 313 K were performed with DOXYL- stearic acid concentrations in the range 0.1–0.6 mM to determine the optimum amount for large and specific effects. \(^{1}H\)-NMR experiments were performed at 400 MHz on a Bruker AM400 spectrometer, interfaced to an Aspect3000 computer. Two different types of 1D NMR spectra were recorded: (a) with a simple \(\pi/2\) pulse preceded by water irradiation, or (b) with a \(\pi/2\) pulse followed by a 25-ms spin-lock period as in a total correlated spectroscopy (TOCSY) sequence (Bax and Davis, 1985; Griesinger et al., 1988). The spin-label titrations were analyzed by subtracting the Fourier-transformed spectra recorded for samples with and without spin-label using standard Bruker software. 2D-MLEV17-TOCSY spectra (Bax and Davis, 1985; Griesinger et al., 1988) were recorded in the phase-sensitive mode using the time-proportional-phase-incrementation (TPPI) method (Redfield and Kunz, 1975; Marion and Wuthrich, 1983). The carrier was placed at the water resonance. The H\(_2\)O signal was suppressed by continuous irradiation during the relaxation delay of 1.5 s. The experiments were performed at 313 K. The spectral width was 12 ppm for nisin/SDS and 15 ppm for nisin/DODPC/cho samples in both dimensions; the number of data points amounted to 400 to 512 in \(t\), and 2048 in \(t\). The number of scans was typically 32. The mixing times were 25 ms and 22 ms for nisin/SDS and nisin/DODPC/cho, respectively. 2D-MLEV17-TOCSY experiments were performed with H\(_2\)O/\(^{2}H\)H\(_2\)O (9:1) and \(^{2}H\)H\(_2\)O samples. The spin-label concentrations of the samples were in the range of 0.2–0.4 mM. The data were processed on an Aspect3000 computer. A \(\pi/4\)-shifted sinebell apodization was used in both dimensions. 2D relative-difference spectra were obtained by subtracting TOCSY spectra recorded for samples with and without spin-label, using in-house-written software, on a Bruker Aspect3000 computer. Relative differences were obtained as \(I = (I_w - f I_s)/I_w\), where \(I\) is the peak height in the relative difference spectrum, \(I_w\) is the peak height in the spectrum recorded for a sample without and \(I_s\) with spin-label; \(f\) is the factor used to scale the latter TOCSY spectrum. To determine \(f\), rows and columns of the 2D spectra including peaks not affected by the spin-label were extracted. The optimal factor is determined using standard Bruker software by optimizing the nilling of the unaffected resonances in the difference spectrum. As noise signals can give rise to artifacts in relative difference spectra, peak heights below a chosen level were not taken into account.
'H-NMR spectroscopy of complexes of nisin and protonated micelles. Samples contained 4 mM nisin and a 25-fold excess of [1H]SDS or [1H]DodPCho in 0.5 ml H2O/HH2O (9:1) at pH 3.5. NOESY spectra (Jeener et al., 1979; Bodenhausen et al., 1984) with a mixing time of 500 ms were recorded at 298 K and 313 K on a Bruker AMX600, operating at 600 MHz and interfaced to a Bruker Aspect X32 computer. Acquisition was essentially the same as described above for the TOCSY spectra. The solvent was also irradiated during the mixing time. The number of scans was 64. Processing of the data was performed on a Bruker Aspect X32 computer. Prior to Fourier transformation a π/3-shifted sine bell apodization was applied in both dimensions and in the t1 direction the data were zero-filled to 1024. Baseline corrections were necessary and were performed after Fourier transformation.

RESULTS AND DISCUSSION

General remarks. The data obtained in this study are interpreted on the basis of the 3D structures of nisin when complexed to membrane-mimetic micelles of DodPCho or of SDS that are described in the accompanying report (van den Hooven et al., 1995). Two structured domains are found, a structured N-terminal domain encompassing the first three lanthionine rings A, B, and C, when nisin is bound to DodPCho, or the first two or three lanthionine rings when nisin is bound to SDS micelles. This domain is connected (via a hinge region) to a structured C-terminal domain encompassing the residues Lys22 to Ala28, involving three consecutive overlapping β-turns which include the two intertwined lanthionine rings D and E. Although the regions flanking the two domains show structural variability, a tendency for extended structures has been observed. The molecule is amphipathic in two ways. First, the charged and hydrophilic amino acids are mainly located in the C-terminal half of the molecule, whereas the majority of the residues in the N-terminal half is hydrophobic and only a single charged residue, Lys12, is present. Secondly, both of the aforementioned domains have a hydrophobic and a hydrophilic side.

Miscellaneous observations suggesting a surface location of nisin. A number of initial observations indicated that nisin is localized at the surface of DodPCho and of SDS micelles. First, the line widths of 13C resonances of DodPCho and SDS near the polar head-group are (slightly) increased upon addition of nisin, whereas there is almost no effect on resonances from the micellar center. Second, amide proton exchange measured for nisin in both micellar systems, by dissolving a protonated sample in D2O and following the loss of intensity of the NH proton signals, is still quite fast. After an hour only a few NH resonances could be seen from H1′ protons of the αJ-unsaturated amino acids, Dha2, Dha5 and Dha33 (along the horizontal axis) to other nisin resonances and to the [1H]DodPCho resonances 15, 6-14 and 5 (numbering along the F2 axis, see Fig. 3). A comparison of NOESY spectra recorded for samples with protonated and with deuterated DodPCho shows that the cross-peaks from the resonance of the H6′ protons of Dha5 to the resonance of DodPCho and to the resonance of the H6′ proton of Lys16 are indeed two cross-peaks. Relatively strong NOEs between nisin and protonated DodPCho are observed for the H6′ resonances of Dha5 and the corresponding cross-peaks are boxed.

Nisin-detergent NOEs indicate complexation. A possible interaction of nisin and micelles might be detectable via direct NOE contacts between nisin and protonated detergent molecules. Therefore NOESY experiments were performed at 313 K, the temperature at which the data for the 3D-structure elucidation was obtained. To observe intermolecular NOEs it was necessary to change the nisin/detergent ratio from 1:30 to 40 to 1:25. Furthermore, the experiments were conducted at 600 MHz and a long mixing time of 0.5 s was chosen to increase NOE intensities. A distinction between intramolecular nisin and intermolecular nisin-detergent NOEs could be made by comparing NOESY spectra in the presence of protonated and of perdeuterated micelles. Additional information was obtained from a NOESY spectrum recorded at 298 K, where some overlap problems encountered at 313 K could be resolved. NOEs were observed between nisin and detergent resonances (Fig. 2) in the part of the spectrum downfield from the residual water signal, whereas this was precluded in the region upfield of the water signal due to overlap problems. The resonance positions of the 1H resonances of DodPCho and SDS are given in Fig. 3.

Quite a number of intermolecular NOEs were observed, which clearly demonstrate complexation between nisin and the
micelles. Of the detergent resonances those overlapping at about 1.3 ppm gave the strongest NOEs to resonances of nisin, but also cross-peaks from detergent resonances at the micellar surface to nisin resonances were observed; in two cases connectivities to the methyl group of the dodecyl chain were seen. The intensities of the nisin-detergent NOEs differ clearly. The micelles are dynamical aggregates. Individual detergent molecules can change places, can move along the diameter of the micelle and can swing up and down (Godici and Landsberger, 1974; Menger and Doll, 1984). Further, nisin is still flexible (van den Hooven et al., 1995) and its depth of immersion into the micelles may vary slightly in time. This explains, together with spin diffusion, that a number of detergent resonances give NOEs to nisin resonances, though to a different extent. As an example, in Fig. 2 it can be seen that the H\textsuperscript{2} protons of Da5 give relatively strong NOEs to the overlapping resonances of DodPCho, whereas those of Dhb2 and Dha33 are weak. Although the results obtained for both systems are not completely identical because of different overlap problems, it can be concluded that the relatively strong NOEs observed are from nisin protons distributed over the whole polypeptide sequence and located at the hydrophobic sides of the two structured domains in correspondence with the results obtained in the spin-label experiments.

Spin-labeled probes incorporated in micelles. The micelle-inserted parts of nisin can be probed with intramicellar spin-labels, for which the position in the micelle must be known. We determined the incorporation of the two spin-labeled 5-DOXYL-stearic and 16-DOXYL-stearic acids in micelles of DodPCho and of SDS through observation of a broadening of the ESR lines of the nitroxide spin-labels and a decrease in the hyperfine splitting. A similar approach has been used for DodPCho micelles (Brown et al., 1981). The micelles were found to consist of 30–50 detergent molecules as determined from the detergent concentration dependence of the widths of the ESR lines of the nitroxide radicals, as previously described (Brown et al., 1981). In the presence of nisin the spin-labels are incorporated into micelles, which consist of 30–50 detergent molecules (data not shown).

The paramagnetic spin-label probes induce in the NMR spectrum line broadening of the resonances of nuclei in their vicinity. This effect was used to determine the position of the spin-labels of the 5-DOXYL-stearic and 16-DOXYL-stearic acids in the micelles by recording \(^{13}C\)-NMR spectra at natural abundance. The resonance positions of the \(^{13}C\) resonances of DodPCho and SDS are given in Fig. 3. We note that the average number of spin-labels/micelle in our samples is smaller than one, so that interactions between spin-labels can be neglected. The line broadening caused by 5-DOXYL-stearic acid was most pronounced for the detergent resonances of \(^{13}C\) atoms located near the polar head-groups, whereas the effect of 16-DOXYL-stearic acid was the largest at the apolar end of the carbon chain (data not shown). Similar results have been obtained for other micellar systems (Brown et al., 1981; Mendz et al., 1988, 1991; Papavoine et al., 1994). These results indicate qualitatively that the nitroxide moiety of 16-DOXYL-stearic acid is most of the time near the center of the micelles, whereas the label of 5-DOXYL-stearic acid is, on average, close to the surface, i.e. near the phosphate groups of DodPCho and just underneath the sulphate of SDS.

The addition of nisin does not significantly alter the effect of the spin-labels on the \(^{13}C\) resonances of DodPCho and SDS, indicating that the relative orientation of the spin-labeled molecules within the micelles does not appreciably change. The nisin/detergent aggregate can be regarded as a complex with a typical micellar organization. These results formed the basis for the following studies on the interaction between nisin and the spin-labeled micelles.

Spin-labels demonstrate a location and orientation of nisin at the micellar surface. Using the two different spin-labels, intramicellar and extra-micellar locations can be determined. Using the same spin-label approach the polypeptide-hormone glucagon and the bee-venom constituent melittin were found to bind to the surface of DodPCho micelles (Brown et al., 1981, 1982). Studies of the myelin basic protein indicated that this molecule is also located primarily near the surface of DodPCho and of mixed DodPCho/palmitoyl/phosphatidic acid micelles (Mendz et al., 1984, 1988, 1990, 1991). A different situation was encountered for the major coat protein of bacteriophage M13; this protein is composed of a long hydrophobic helix that traverses the SDS micelle and a shorter amphipathic helix that is situated on the surface of the micelle (Papavoine et al., 1994). In the present study the homonuclear spectra with and without

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Fig. 3. Structures of the anionic sodium dodecylsulphate and the zwitterionic dodecylphosphocholine with numbering of carbon and hydrogen atoms. The \(^{13}C\)-NMR spectra of SDS and DodPCho have been partially assigned (Kragh-Hansen and Riisom, 1976; Brown et al., 1981). Based on chemical shift data, e.g. (Birdsall et al., 1972), multiplet structure and spin-decoupling experiments we assigned a few more \(^{13}C\) resonances.
spin-label were analyzed by relative-difference 2D spectra instead of 1D NMR, which allows for a larger number of resonances to be investigated unambiguously.

The two different spin-labels induce specific line-broadening effects in the ^H-NMR spectrum of nisin in the presence of DodPCho and of SDS. These labels do not change the chemical shifts, indicating that they have little or no effect on the conformation of nisin. A spin-label concentration of 0.2 mM to 0.4 mM was found to be most appropriate for optimal, strong and specific, broadening effects in the spectrum of 3 mM nisin samples. The line broadening was estimated from the peak height in relative-difference spectra (Fig. 4) obtained by subtracting 2D-MLEV17-TOCSY spectra recorded for samples with and without spin-label. The resonances of the α,β-unsaturated amino acids did not give rise to cross-peaks in the TOCSY spectra, but were well resolved in the 1D NMR spectrum. For these resonances the spin-label-induced line broadening was evaluated from a 1D difference spectrum. In the 2D TOCSY difference spectra the influenced protons are traced back via the observed cross-peaks. If a majority of the cross-peaks, stemming from a given proton, showed up in the relative difference spectra a line broadening effect was attributed to that particular proton.
Schematic representation of the interaction of nisin with a SDS micelle. Nine SDS molecules with extended conformation were placed to form a circular array to represent a micelle. One representative structure of nisin (van den Hooven et al., 1995) was positioned at the surface of the micelle. The N-terminus of nisin is at the upper left corner, whereas the C-terminus is at the upper right corner. For the N-terminal structured domain the side chains of Leu6 and Leu16 are the deepest immersed into the micelle, whereas the side chain of Lys12 is protruding outwards.

The effects of both spin-labels on nisin/DodPCho and on nisin/SDS were the same. The 5-DOXYL-stearic and 16-DOXYL-stearic acids essentially affect the same nisin resonances, although 5-DOXYL-stearic acid is more effective in the sense that at a given concentration for several resonances the broadening is stronger than for 16-DOXYL-stearic acid and 10–15% of the resonances affected by 5-DOXYL-stearic acid are not influenced by 16-DOXYL-stearic acid. The resonances of the following residues are broadened: Ile1, Dha5, Leu6, Ala15, Leu16, Met17, Met21, Ala24, Ala25 [Ala3, 3-methylalanly moiety of (2S, 3S, 6R)-3-methyllanthionine], sAla28 [sAla, t-alanyl moiety of meso-lanthionine], Ser29, Ile30, His31, Val32, Dha33 and Lys34. For Gly10 only a few cross-peaks with low intensity were observed in the TOCSY spectra hampering the evaluation of the data for this residue. The strongest effects, i.e. the highest extent of line broadening, are observed for the residues Ile1 and Ile30–Lys34. The protons giving rise to broadened resonances are distributed over the whole, more or less extended, molecule and are not confined to a limited region of the molecule. These and other observations (see above) are interpreted in terms of a model in which nisin is localized at the surface of both DodPCho and SDS micelles.

From the distribution of the residues which exhibit broadened resonances information about the orientation of the molecule on the surface of the micelles is obtained. The positions of the protons of which the resonances are broadened by 5-DOXYL-stearic acid are indicated (Fig. 5) in the 3D structure of nisin complexed to DodPCho micelles (van den Hooven et al., 1995). In the structured N-terminal domain comprising the residues Ala3 to sAla19 all hydrophobic residues except Ile4 are influenced by the spin-labels. The affected protons reside on one side of this amphipathic domain (Fig. 5b), meaning that the hydrophobic residues are immersed below the charged surface of the micelles. In the structured C-terminal domain encompassing the residues Met21 to sAla28 the amino acids Lys22, Ala23, sAla26 and His27 are on one side and Met21, Ala24, Ala25 and sAla28 on the other side. The latter are much more influenced by the spin-labels than those on the other side (Fig. 5c). This suggests that as for the N-terminal domain, the hydrophobic side of the C-terminal domain is immersed in the hydrophobic interior of the micelle, whereas the more hydrophilic side of this part of the molecule resides at the charged surface. The global picture that emerges from these observations is that nisin is localized at the surface of the micelles and oriented such that the hydrophobic residues are immersed below the surface, whereas the more polar or charged residues have an outward orientation (Fig. 6).

The results obtained from the NOESY experiments are in accord with those of the spin-label experiments, which suggests that the DOXYL-stearic acids have no major influence on the interaction between nisin and the micelles, justifying the use of these paramagnetic probes. Overlap problems are much less severe in the spin-label approach, where also the aliphatic spectral region can be utilized to derive information. This renders it much more suitable for the study of the interaction between polypeptides and micelles than the polypeptide-detergent NOEs.
Temperature coefficients as indicators of solvent shielding.

Now that both the conformation (van den Hooven et al., 1995) and the location of nisin complexed to DodPCho and to SDS micelles is known, we can attempt to interpret the temperature coefficients. Amide proton temperature coefficients are indicative of hydrogen bonding and/or of shielding from the solvent. These coefficients have been calculated from the assignments at different temperatures for nisin in aqueous solution (Slipper et al., 1989) and in micellar systems (van den Hooven et al., 1993) and are shown in Fig 7. The magnitudes of the amide-proton temperature coefficients are nearly the same for nisin complexed to DodPCho and to SDS micelles. Two large differences of about −8 ppb/K are observed between nisin in water and in the micellar systems, strikingly for both dehydroalanines. For Dha5 this can be explained by a different conformation of the flanking (trans) peptide bonds which flipped by almost 180° when transferred from water to a membrane-like environment (van den Hooven et al., 1995). There are no indications that the conformation of the C-terminus, including Dha33, in water differs from that in the micellar systems, and in all three systems no hydrogen bonds were observed for the residues Ser29–Lys34. The NH resonances of the five consecutive residues Ile30–Lys34 all have a reduced temperature coefficient in the micellar systems compared to nisin in aqueous solution. These coefficients are medium-range for nisin in the micellar systems and the corresponding amide protons are not the slowest exchanging NH protons. Our explanation of this difference in temperature coefficients is that the nisin amide protons of the residues Ile30–Lys34 are in the micellar systems more shielded from the solvent than in aqueous solution, which is caused by an interaction with the micelles. This is in correspondence with the spin-label experiments, where the largest induced line-broadening effects were observed for resonances of the C-terminal residues Ser29–Lys34, proving that this sequence is in close contact with the micelles.

Model of pore-forming activity. In current models on the mechanism of action of nisin it is assumed that it forms transient multistate pores in the cytoplasmic membrane of the target cell (Sahl, 1991; García Garcerá et al., 1993; Driessen et al., 1995). The first step in the formation of these pores is the binding of nisin to the membrane, where nisin is supposed to adopt an amphipathic conformation upon contact with the membrane, the contacts of which are primarily based on ionic forces (Sahl, 1991). The binding step is followed by aggregation and membrane-potential-dependent insertion. The order of the latter two events is not known. In the pores the nisin molecule is thought to be amphipathic and oriented with its hydrophobic side towards the membrane and with its hydrophilic side towards the water-filled channel. The amphipathicity has been demonstrated for nisin in aqueous solution (van de Ven et al., 1991) and for nisin complexed to membrane-mimetic micelles (van den Hooven et al., 1995).

Opinions differ with regard to the orientation of nisin with respect to the membrane in the binding step. It has been suggested that the charged and polar residues contact the membrane surface, while the hydrophobic side remains exposed to water (Sahl, 1991). If we take the binding to the micellar systems studied here as a model for membrane binding we prefer the opposite view with the polar residues at the surface and the hydrophobic residues immersed in the membrane.

The N-terminal half of the nisin molecule contains mainly hydrophobic residues, the most hydrophobic ones are immersed in the micelles. For this half of the molecule hydrophobic interactions are likely to dominate the nisin-micelle contacts. The C-terminal half is much more hydrophilic and again the hydrophobic residues of the structured domain are immersed in the micelles, whereas the polar and charged residues reside at the charged surface. For this part of the nisin molecule both hydrophobic and electrostatic interactions are likely to be involved in the nisin-micelle contacts. The interactions with the micelles are strongest for the C-terminal residues Ser29–Lys34 as determined from the spin-label experiments as the strongest spin-label-induced line-broadening effects. This is corroborated by the temperature coefficients, which suggested that the amide protons of the residues Ile30–Lys34 are shielded from the solvent by the micelles (see above).

The antimicrobial activity of nisin has been found to be influenced by the phospholipid composition (Gao et al., 1991; García Garcerá et al., 1993; Driessen et al., 1995). Different activities were observed for liposomes composed of phosphatidylcholine (PtdCho) or phosphatidylglycerol (PtdGro; Driessen et al., 1995). It has been suggested that nisin induces the release of non-bulky anionic fluorescent indicators from phosphatidylcholine liposomes possibly as an anion carrier, an activity
which can be enhanced by a trans-negative membrane potential ($\Delta \Psi$). This activity is not observed for PtdGro liposomes. Nisin associates with the anionic surface of PtdGro liposomes, and disturbs the lipid dynamics near the phospholipid polar headgroup-water interface in the absence of a $\Delta \Psi$. This process is not directly responsible for pore formation since there is no general loss of the permeability barrier or integrity of the membrane. Nisin requires a threshold $\Delta \Psi$ for pore formation in PtdGro liposomes. The activity of nisin with PtdGro liposomes has been proposed to resemble the in vivo activity more closely than that of nisin with PtdCho liposomes (Driessen et al., 1995).

In contrast, the conformation (van den Hooven et al., 1995), location and orientation of nisin relative to the micelles are similar for both zwitterionic and anionic detergents, suggesting that binding to bilayers composed of different lipids will not affect the 3D structure of nisin nor its orientation relative to the membrane. A tighter interaction between nisin and anionic lipids than between nisin and zwitterionic lipids may be the explanation. It has been suggested that nisin interacts in a different manner with PtdCho and PtdGro liposomes (Driessen et al., 1995). Evidence for a tight interaction between nisin and PtdGro liposomes has been obtained (Driessen et al., 1995).

To be able to form pores the cationic nisin requires a trans-negative membrane potential (Sahl et al., 1987; Driessen et al., 1995), which forces the molecule into a state that renders the membrane leaky. Evidence was obtained that nisin can traverse the membrane of liposomes composed of zwitterionic PtdCho (Driessen et al., 1995). It is conceivable that anionic phospholipid head-groups interact with the surface-bound cationic nisin and prevent a deep penetration of nisin in the membrane in absence of a $\Delta \Psi$. It is tempting to speculate that a trans-negative membrane potential above a certain threshold pulls the nisin molecule with surrounding anionic lipids in a trans-membrane orientation (Fig. 8 and Driessen et al., 1995) with the hydrophobic side of nisin and the attached lipid head-groups facing the center of the water-filled pore and the hydrophobic surface of nisin and the fatty acid chains of the lipids pointing to the lipid bilayer. Both the polar sides of the cationic nisin and the anionic lipids face the lumen of the pore, which explains the observed non-selective efflux (Ruhr and Sahl, 1985; Kordel and Sahl, 1986; Sahl, 1991). In such a (tentative) pore nisin is located on a highly curved surface, which resembles the micellar situation. It is clear from the present study that nisin can be situated on such a curved surface. Within this model the presence of a flexible hinge region between the two structured domains proposed for nisin in aqueous solution (van de Ven et al., 1991) and complexed to micelles (van den Hooven et al., 1995) can be rationalized, since in the membrane-bound state the nisin molecule is probably elongated, whereas it has to be more bent in the inserted state. During the proposed insertion process the orientation towards the lipids does not change, whereas in other models (Sahl, 1991; García Gascón et al., 1993) the interactions between cationic nisin and the lipid head-groups have to be broken to let the nisin molecule slide into the hydrophobic core of the membrane.

**Related lantibiotics and nisin mutants.** A comparison with related polypeptide sequences has been performed in order to investigate whether the effects that amino acid substitutions have on the bactericidal activity are compatible with our model. So far up to about 25 lantibiotics, subdivided in types A and B, are known (Jung, 1991; Bierbaum and Sahl, 1993; Sahl et al., 1995). Lantibiotics of the first type, including nisin, act primarily by membrane perturbation, while those of the other type appear to inhibit enzyme function. The type-A lantibiotics nisin, subtilin, epidermin and galliderinin are considered because they act according to a similar mechanism and because of the structural homology.

The type-A lantibiotic subtilin (Gross et al., 1973) bears a strong similarity to nisin: the sequence identity between the two proteins amounts to 60%, the number of lanthionine rings is the same and they are at identical positions in the proteins (Fig. 9). The structure of subtilin in aqueous solution has been determined using NMR spectroscopy and has been suggested to be similar to that of nisin (Chan et al., 1992). In view of the results obtained above for the location and orientation of nisin with respect to the micelles it is interesting to consider possible differences in hydrophathy (Kyte and Doolittle, 1982) for the substituted residues. Large changes occur for the residues at positions 2, 4, 12, 17, 20 and 29, indicated by the thick double-headed arrows in Fig. 9. Apart from residue 17 and 29, these residues are, as evidenced by the spin-label approach, not in contact with
the micelles but are exposed to water. Resonances of Met17 were influenced by the spin-labels. However, for nisin/DodPC 
the side chain of Met17 is pointing in the direction of the 
residue Ala13, which has an outward orientation. Since the lengths 
of the side chains of Met and Glu are the same it is likely that 
the hydrophilic group of Glu in subtilin resides at the surface. 
Residues His31 and Val32 in nisin have no equivalent in subtilin 
and we assume that the presence of a lysine residue at position 
29 in subtilin compensates for the loss of the charged His31. 
Conservative substitutions of hydrophobic by hydrophobic resi-
dues or of hydrophilic by hydrophilic residues are indicated by 
thin double-headed arrows in Fig. 9 and are thought to have no 
major effect on the process of membrane interaction. Thus, the 
differences are such that the membrane interaction surface as 
found for nisin is conserved for subtilin.

The contribution of individual amino acids to the biological 
activity can be studied with the aid of nisin mutants. A natural 
nisin variant with an Asn at position 27 instead of a His (Mulders 
et al., 1991) and two nisin double mutants Met7Gln/Gly18Dhb 
and Met7Gln/Gly18Thr (Kuipers et al., 1992) have almost the 
same activity as wild-type nisin (De Vos et al., 1993; Kuipers 
et al., 1992). This is in line with the preceding discussion, the 
involved residues are not expected to interact with the micelles 
except for residue Met17 (but see discussion above and the leg-
end to Fig. 9). In this respect it is also noted that mutagenesis 
of Dha5 to Ala in subtilin had no effect on the ability to inhibit 
and lyse vegetative cells, though an intact Dha5 residue ap-
ppeared critical for spore-outgrowth inhibition, which apparently 
occurs by a different mechanism (Liu and Hansen, 1993).

Degradation of nisin at Dha5 can result in an opening of the 
first ring leaving the hydrophobic residues 4 and 6 much more 
polar (Chan et al., 1989a; Rollema et al., 1991). The observed 
drop in activity may be ascribed to hampered membrane interac-
tions.

The first 11 residues of the type-A lantibiotics gallidermin 
and epidermin, including the rings A and B, resemble those in 
nisin and subtilin. The sequence for the first 11 residues of gal-
lidermin is

\[
\text{Ile1-Ala2-Ala3-Lys4-Phe5-Leu6-Ala7-Ala8-Pro9-Gly10-Ala11;}
\]

the sequence of epidermin is identical to gallidermin with the 
exception of an Ile at position 6. Compared to nisin, in this part 
of the sequence of gallidermin three residues are different: Dhb2 
versus Ala, Ile4 versus Lys and Dha5 versus Phe (the first men-
tioned residues are in nisin). An extra difference is encountered 
between nisin and epidermin, namely a Leu or an Ile at position 
6. Only the lysine residue at position 4 causes a dramatic change 
in hydrophathy. However, in nisin this residue was shown not to 
be in contact with the micelles. It is therefore very likely that 
this marked difference does not influence the binding of the lan-
tibiotic to the membrane; as far as the first 11 residues are con-
cerned the membrane interaction surfaces of gallidermin and 
epidermin are thus highly similar to that of nisin.

Conclusions. In combination with our work on the conformation 
of nisin in aqueous solution (van de Ven et al., 1991) and when 
complexed to micelles (van den Hooven et al., 1995) the binding 
to micelles can be described in detail. The micellar systems are 
considered to model the first step in the mechanism of action of 
nisin, this step is the binding of nisin to the cytoplasmic mem-
brane of target bacteria. Detailed information on this initial bind-
ing step is obtained and an extension of the model for pore for-
mation (Driessen et al., 1995) is proposed. In aqueous solution, 
the nisin molecule is amphipathic and consists of two structured 
domains of the residues Ala3−aza19 and Lys22−aza28, con-

nected via a peptide segment which shows structural variability 
that is most likely caused by flexibility. The molecule binds par-
allel to the membrane surface, where its amphipathicity is re-
tained. Upon binding, the conformation is changed significantly 
in the first ring around the important residue Dha5, where two 
(trans) peptide bonds flip. Structural variability is observed for 
the region connecting the two structured domains. The molecule 
is partially immersed, with its hydrophilic residues in the bi-
layer, whereas the hydrophilic residues are at the surface or just 
outside the membrane in contact with lipid head-groups.

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