Asymmetric Synthesis of All Stereoisomers of the Strigol Analogue GR24. Dependence of Absolute Configuration on Stimulatory Activity of Striga hermonthica and Orobanche crenata Seed Germination

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All four optically pure stereoisomers of the strigol analogue GR24 were prepared via two different routes. In the first approach enantiopure ABC-fragments 4 were used as the chiral source, whereas in the alternative route both antipodes of the latent D-ring 6 were employed. Bioassays revealed significant differences in activity between the four stereoisomers in the stimulation of germination of the parasitic weeds Striga hermonthica and Orobanche crenata.

Keywords: Striga; Orobanche germination, GR24

INTRODUCTION

Parasitic weeds belonging to the genera Striga, Orobanche, and Alectra cause severe damage to several graminaceous and leguminous crops in tropical and semitropical areas of the eastern hemisphere (Musselman, 1987; Parker and Riches, 1993). Seeds of these parasitic weeds usually remain dormant in the soil, but after exposure to a specific germination stimulant, which is exuded by the roots of a suitable host, the seeds germinate and attach to the root system of the host (Press et al., 1990). The stimulation of germination, in particular the compounds by which this process is triggered, has received much attention. The most prominent naturally occurring germination stimulant, viz. (+)-strigol (1) (Figure 1), has been isolated from the root exudate of the false host cotton (Gossypium hirsutum L.) (Cook et al., 1966), and its structure was elucidated (Cook et al., 1972). The absolute configuration was unambiguously determined several years later (Brooks et al., 1985). Recently, it has been shown to be the major Striga germination stimulant produced by the true hosts maize (Zea mays L.) and proso millet (Panicum miliaceum L.) (Siame et al., 1993). It was demonstrated that the absolute stereochemistry of strigol 1 is of prime importance with respect to seed germination activity (Hauck and Schildknecht, 1990; Bergmann et al., 1993).

In a study to design simpler analogues of (+)-strigol (1) with optimal bioactivity, Johnson et al. (1976, 1981) have prepared the rather readily accessible synthetic strigol analogue GR24 (2) (Figure 1). Its racemic preparation and the separation of its diastereomers was improved considerably (Mangnus et al., 1992a; Nefkens et al., 1996). The seed germination stimulatory activity toward several parasitic weed species is within the same order of magnitude as that of strigol (Pepperman et al., 1987; Hauck et al., 1992; Bergmann et al., 1993). For these reasons GR24 (2) has found widespread applications, including its use as a positive control in bioassays of Orobanche and Striga seed germination (Mangnus et al., 1992b). So far, only little attention has been paid to the influence of the absolute stereochemistry of synthetic strigol analogues on the biological activity. Recently, all four stereoisomers of GR7 (3) (Figure 1), a strigol analogue lacking the A-ring, have been prepared and tested on seed germination activity (Mangnus and Zwanenburg, 1992). It was concluded that the correct absolute stereochemistry at C-2` (R-configuration) is essential to exert maximal bioactivity.

The present paper deals with the asymmetric syntheses of all four stereoisomers of GR24 (2) and their germination activities on seeds of Striga hermonthica (Del.) Benth and Orobanche crenata Forsk.

MATERIALS AND METHODS

Nomenclature. The AUTONOM 1.0 program, provided by the Belstein Institute and Springer-Verlag (Weinheim, Germany) was used.

Syntheses. General Remarks. 1H-NMR (100 MHz and 400 MHz) spectra were recorded on Bruker AC 100 and Bruker AM-400 spectrometers, respectively (Me4Si as internal standard), both from Bruker (Wissembourg, France). All coupling constants are given as 3 in hertz, unless indicated otherwise. For mass spectra a double-focusing VG7070E mass spectrometer from VG Analytical (Manchester, U.K.) was used. Optical rotations were measured on a Perkin-Elmer 241 polarimeter. Elemental analyses were performed at the Department of Micro-analysis of this laboratory.

Solvents were dried using the following methods: Dichloromethane was distilled from P2O5. Diethyl ether was distilled...
from NaH. Hexane was distilled from CaH₂. Tetrahydrofuran was distilled from lithium aluminum hydride just before use. All other solvents were of analytical grade. Thin layer chromatography (TLC) was carried out on Merck precoated silica gel 60 F254 plates (0.25 mm) using the eluents indicated. Spots were visualized with UV or using a molybdate spray. “Flash” chromatography was carried out at a pressure of ca. 1.5 bar, using Merck Kieselgel 60H. Column chromatography at atmospheric pressure was carried out, using Merck Kieselgel 60.

Sodium hydride (60% in dispersion oil) was washed twice with hexane just before use. 3,3a,4,8b-Tetrahydrodienand1(b)uran-2-one (rac 4) and 5-bromo-3-methyl-2(5H)-furanone (5) were prepared following published methods (Magnus et al., 1992a). The synthesis of chlorolactones 6 and ent 6 was reported previously (Thuring et al., 1995).

For the resolution of rac 4 mixture crystalline cellulose triacetate-particle size 25 μm) and silica gel 60. 1H NMR (CDCl₃, 300 MHz) δ 8.9 Hz, H2 B-ring), 3.7 Hz, H2 A-ring), 7.9 Hz, H2 O-ring), 7.50 (d, 1H, 4J, 16.8 Hz, Ar H), 7.48 (d, 1H, 1J, 2.75 Hz, Ar H), 7.36 (3H, Ar H), 7.23–7.36 (3H, Ar H), 3.42 (4J, 1H, J = 9.3 Hz, CH₂), 3.94 (4J, 1H, J = 7.9 Hz, CH₂), 5.96 (d, 1J, J = 7.9 Hz, H₃b), 3.10 (dd, 1H, J = 16.9 Hz, J = 3.1 Hz, CH₃), 3.44 (dd, 1H, J = 16.9 Hz, J = 9.2 Hz, H₃a), 3.95 (m, 1H, H₃a), 5.96 (d, 1H, J = 7.9 Hz, H₃b), 6.17 (m, 1H, OCHO D-ring), 6.96 (m, 1H, =CH D-ring), 7.23–7.36 (3H, Ar H), 7.48 (d, 1H, 1J, J = 2.75 Hz, Ar H), 7.36 (3H, Ar H), 7.23–7.36 (3H, Ar H), 3.42 (d, 1H, J = 16.8 Hz, CH B-ring), 3.24 (m, 1H, H₃a), 3.41 (d, 1J, J = 16.8 Hz, CH B-ring), 3.24 (m, 1H, H₃a), 3.41 (d, 1J, J = 16.8 Hz, CH B-ring). 1H NMR (CDCl₃, 400 MHz) δ 2.05 (m, 3H, CH₃), 3.10 (dd, 1H, J = 16.9 Hz, J = 3.1 Hz, CH₃), 3.44 (dd, 1H, J = 16.9 Hz, J = 9.2 Hz, H₃a), 3.95 (m, 1H, H₃a), 5.96 (d, 1H, J = 7.9 Hz, H₃b), 6.17 (m, 1H, OCHO D-ring), 6.96 (m, 1H, =CH D-ring), 7.23–7.36 (3H, Ar H), 7.48 (d, 1H, 1J, J = 2.75 Hz, =CH O-ring), 7.36 (3H, Ar H), 7.23–7.36 (3H, Ar H), 3.42 (d, 1H, J = 16.8 Hz, CH B-ring), 3.24 (m, 1H, H₃a), 3.41 (d, 1J, J = 16.8 Hz, CH B-ring), 3.24 (m, 1H, H₃a), 3.41 (d, 1J, J = 16.8 Hz, CH B-ring).
Scheme 1. Resolution of rac 4 and Coupling with 5

![Diagram of Scheme 1](image)

a) cellulose triacetate chromatography, b) 1. KOtBu, HCO₂Me 2. 5

H₂O), 7.21-7.53 (m, 4H, Ar H), 7.39 (d, 1H, J = 2.8 Hz, -CHO); mass data were the same as for 7a. Anal. Calcd for C₂₂H₂₀O₅: C, 72.52; H, 5.53. Found: C, 71.91; H, 5.47.

Calcd for C₂₂H₂₀O₅: C, 72.52; H, 5.53. Found: C, 72.17; H, 5.53. 1H-NMR and mass data were the same as for compound 7a and 7b. Starting from rac 4 (302 mg, 1.73 mmol) and chloralactone ent 6 (379 mg, 1,91 mmol). The fast-moving diastereomer ent 7a (202 mg, 32%) was obtained as a white solid, and crystallization from diisopropyl ether/ethyl acetate afforded analytically pure ent 7a. The slow-moving diastereomer ent 7b (215 mg, 34%) was obtained as a white solid, which gave an analytically pure sample after crystallization from hexane/ethyl acetate.

ent 7a: mp 171-173.5 °C; [α]D = -278° (c 0.1, CHCl₃). Anal. Calcd for C₂₂H₂₀O₅: C, 72.52; H, 5.53. Found: C, 72.58; H, 5.45. 1H-NMR and mass data were the same as for compound 7a.

ent 7b: mp 193-193.5 °C; [α]D +340° (c 0.1, CHCl₃). Anal. Calcd for C₂₂H₂₀O₅: C, 72.52; H, 5.53. Found: C, 72.17; H, 5.53. 1H-NMR and mass data were the same as for compound 7b.

Cycloreversion of Adducts 7a,b and ent 7a,b. A solution of the enantiopure cycloadduct 7 (93 mg, 0.26 mmol) in o-dichlorobenzene (40 mL) was heated at 180 °C for 14 h. The solvent was removed in vacuo. The residue was purified by flash chromatography (SiO₂, hexaneeethyl acetate 2:1) to give the corresponding enantiomer of 2 (44 mg, 58%) as a solid. All enantiomers of 2 prepared via this procedure showed identical physical and chiroptical properties as described above.

Biological Activity. Seeds. Seeds of Striga hermonthica [from Sorghum bicolor (L.) Moench] and Orobanche crenata (from Vicia faba L.) were harvested in Burkina Faso in 1994 and in Egypt in 1991, respectively, and were stored in the dark at room temperature until use in germination tests. Bioassays were carried out essentially following the procedure of Mangnus et al. (1992b) with minor modifications. Only analytically pure samples as judged from correct elemental analyses were used in these experiments.

Preparation of Test Solutions. A compound to be tested was weighed out very accurately to the amount of 2.5 mg, dissolved in 5 mL of acetone p.a., and diluted with demineralized water to 25 mL. Aliquots of this stock solution were further diluted with water to obtain test solutions containing 1, 0.5, 0.1, 0.05, and 0.001 mg/L test compound and 0.2, 0.1, 0.02, 0.01, and 0.0002% (v/v) acetone, respectively.

Bioassays. For surface sterilization seeds of S. hermonthica were subsequently exposed to 70% (v/v) ethanol for 5 min and sodium hypochlorite (2% active chlorine) for 2 min with agitation. Seeds of O. crenata were exposed to an aqueous solution of sodium hypochlorite (2% active chlorine) for 5 min with agitation. The seeds were then thoroughly rinsed with water and dried.

For conditioning the sterilized seeds were spread on glass fiber filter paper disks (8-mm diameter; approximately 30–70 seeds per disk) in Petri dishes, each containing two disks (Striga) or four disks (Orobanche), wetted with water, and stored in the dark for 14 days at 20 °C for Orobanche seeds and at 30 °C for Striga seeds. Then the conditioning water was removed and replaced by 100 μL of test solution per disk (Orobanche) or 3 mL per Petri dish (Striga). After incubation for 24 h (Striga) and 5 days (Orobanche) in the dark at the indicated temperatures, the germination percentage was determined under a microscope. Seeds were considered to be germinated if the radical protruded through the seed coat.

In each test series aqueous solutions with 0.2% (v/v) acetone were used as negative control. Test solutions of a 1:1 diastereomeric mixture of GR24 at the same concentrations were used as positive controls. All tests were performed in duplicate, and in each test the germination percentages were determined on 6 disks (Striga) or 12 disks (Orobanche) per treatment.

RESULTS AND DISCUSSION

Synthesis. To achieve the resolution of GR24 (2), two synthetic approaches were considered. The first approach (Scheme 1) involves the separation of tricyclic lactone rac 4 into its enantiomers, followed by formylation and coupling of the individual enantiomers with racemic 5-bromofuranone (5) in an analogous manner as described previously for the racemic preparation of GR24 (Mangnus et al., 1992a).

Tricyclic lactone rac 4 was chromatographically resolved using cellulose triacetate (CTA) as the chiral stationary phase. It has been demonstrated that a wide variety of racemic γ- and δ-lactones can be separated by column chromatography on the chiral phase CTA, on both analytic and preparative scales (Francotte and Lohmann, 1987). In the present case 500 mg of rac 4 was resolved by “flash” chromatography at a pressure of ca. 1.2 bar, using 50 g of CTA and 95% ethanol as the eluent, in a total recovery of enantiopure tricyclic lactones 4 and ent 4 of 68%. The enantiopurity of 4 and ent 4 was assessed by HPLC, using cellulose carbbamate as the chiral stationary phase. The diastereomeric mixtures obtained after formylation and coupling of 4 and ent 4 with 5 (Scheme 1) were both separated by flash chromatography (SiO₂) to give 2a, ent 2b and 2b, and ent 2a (ee values > 98%) in total isolated yields of 59% and 60%, respectively. The ee values were determined by 1H NMR using the chiral shift reagent Eu(hfc)₃.
In the second approach the complete resolution of GR24 (2) was accomplished by formylation of rac 4, followed by coupling with the homochiral latent D-rings 6 and ent 6, respectively (Scheme 2).

The asymmetric syntheses of 6 and ent 6, together with their application in the preparation of all four homochiral diastereomers of GR7 (3), have recently been described (Thuring et al., 1995). The sequence as outlined in Scheme 2 gave the cycloadducts 7a and 7b and their corresponding enantiomers in diastereomeric ratios of approximately 1:1. These reactions proceeded with complete stereocontrol as no signals arising from the respective C₂-epimers could be detected in the NMR spectra. Cycloreversion by heating the homochiral adducts of 7 in o-dichlorobenzene at 180 °C afforded the corresponding enantiopure stereoisomers of GR24 (2), which have the same [α]D values as those prepared by the route depicted in Scheme 1. It was essential to control the reaction temperature and time carefully to avoid concomitant epimerization at C₂ of 2.

**Determination of Absolute Configuration.** It was attempted to establish the absolute configuration of the GR24 (2) stereoisomers by comparison of their circular dichroism (CD) spectra with those of the corresponding stereoisomers of strigol 1, which have been reported (Heather et al., 1976; Frischmuth et al., 1993). However, such a correlation is not reliable, because GR24 (2) has a different ABC-chromophore. Therefore, an X-ray diffraction analysis of 2a was undertaken (Moers et al., 1995) to establish its absolute configuration. A stereoview of 2a is depicted in Figure 2. With the absolute configuration of 2a known, the configuration of the remaining stereoisomers could be assigned on the basis of the synthetic sequence.

**Biological Activity.** The germination stimulatory activity of all stereoisomers of GR24 2a,b and ent 2a,b was assayed using seeds of S. hermonthica and O. crenata. In each bioassay a diastereomeric mixture of GR24 was included as a positive control. In preliminary experiments the concentration-dependent activity range (GR24) of seeds of S. hermonthica has been established. Maximal germination percentages were obtained within the concentration range 0.01–1 mg/L. Half-maximal activity was observed at approximately 0.001 mg/L (data not shown). The relative bioactivity of the individual stereoisomers of GR24 was therefore assayed at an optimal concentration (0.1 mg/L) and at a sensitive concentration (0.001 mg/L). It was anticipated that the latter should exhibit more profound differences. Relevant data are collected in Table 1.

The same stereoisomers were also tested for stimulant activity on seeds of O. crenata, using four concentrations. The results are shown in Figure 3. These data (Table 1; Figure 3) reveal that there is a significant difference in stimulatory activity among the four stereoisomers. For both parasitic species, the enantiomer of GR24 possessing the “natural” absolute stereochemistry, viz. 2a, is considerably more active than its optical antipode ent 2a. The difference in activity amounts to at least a factor of 100. The relative importance of the different stereoergic centers can be addressed by comparison of the bioactivities of the

**Table 1. Germination Percentages for Seeds of S. hermonthica after Exposure to Solutions (0.1 and 0.001 mg/L) of GR24 Enantiomers 2 and the Corresponding Racemic Mixture of Diastereomers rac 2**

<table>
<thead>
<tr>
<th>entry</th>
<th>compound</th>
<th>configuration at C-2</th>
<th>at 10⁻¹ mg/L</th>
<th>at 10⁻³ mg/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2a</td>
<td>R</td>
<td>56.2 ± 4.6</td>
<td>32.5 ± 4.5</td>
</tr>
<tr>
<td>2</td>
<td>ent 2b</td>
<td>S</td>
<td>40.8 ± 2.5</td>
<td>0.7 ± 0.4p</td>
</tr>
<tr>
<td>3</td>
<td>ent 2a</td>
<td>S</td>
<td>4.0 ± 0.6</td>
<td>0.5 ± 0.5p</td>
</tr>
<tr>
<td>4</td>
<td>2b</td>
<td>R</td>
<td>54.0 ± 2.6</td>
<td>0.4 ± 0.4p</td>
</tr>
<tr>
<td>5</td>
<td>rac 2c</td>
<td>R/S</td>
<td>47.1 ± 3.9</td>
<td>33.2 ± 2.2</td>
</tr>
</tbody>
</table>

* The data presented ± SE are from one representative experiment. p Not significantly different from aqueous control (without stimulant). c Equimolar mixture of two racemic diastereomers.
Figure 3. Germination percentages for seeds of O. crenata after exposure to different concentrations of GR24 enantiomers. The data presented ± SE are from one single representative experiment.

C₃₅C₈₆-epimer 2b and the C₂⁻epimer ent 2b. The data in Table 1 (entries 2 and 4) reveal in the case of S. hermonthica a slight preference for the correct stereocchemistry at C₂. In contrast, the data obtained for O. crenata (Figure 3) suggest that for this species the absolute configuration at C₃₅-C₈₆ is more important for the stimulatory activity than the configuration at C₂. This result is in contrast with data obtained from a comparative study of GR7 stereoisomers, which revealed a more profound role of the C₂⁻configuration (Mangnus and Zwanenburg, 1992). This discrepancy indicates that one cannot attribute the configuration of a particular stereogenic center as solely relevant for the biological activity by disregarding the entire three-dimensional structure. Because GR7 lacks the A-ring, its smaller BC-part is apparently more flexible with respect to interactions within the receptor cavity. This implies a less critical requirement for its absolute configuration in comparison with the ABC-fragment of GR24.

Concluding Remarks. Two routes for the asymmetric synthesis of all four stereoisomers of GR24 have been presented. The route that requires the homochiral latent D-rings 6 and ent 6 is generally applicable in the enantioselective synthesis of strigol analogues. The relative importance of the absolute configuration at the different stereogenic centers in GR24 for the germination stimulatory activity has been assessed, indicating that the molecular shape is determined by the combination of all stereocenters.

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LITERATURE CITED


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