Heparan sulfate proteoglycans (HSPGs) are components of the basement membrane (BM) of various tissues. They consist of a core protein and negatively charged glycosaminoglycan side chains: the heparan sulfate proteoglycan (HSPG). This member of the heterogeneous family of the proteoglycans is composed of glycosaminoglycan side chains, the negatively charged heparan sulfate (HS) moieties, that are covalently bound to a core protein (13, 14). There are two classes of HSPGs: one class is present on the cell membrane of many cell types, the other class is expressed in the ECM and more specifically in BM (14). The BM-associated HSPGs bind to laminin, fibronectin, collagen type IV, and niadogen and therefore play an important role in BM assembly (15). Because of the negatively charged shield, they block passage of anionic macromolecules through the BM. The storage of growth factors is considered to be an important function of HSPGs (5, 16). This storage of growth factors, the important role in BM assembly and the barrier function of HSPGs, combined with the fact that abnormal proliferation, differentiation and increase in transepidermal water loss are important characteristics of a psoriatic lesion (17, 18), pointed towards a possible change of HSPGs in psoriasis.

Recently, three monoclonal antibodies directed against the HSPGs have become available: monoclonal antibody JM-13, directed against the highly sulfated domains of HS, mAb JM-403, directed against the unmodified/sulfated parts of HS, and mAb JM-72, an antibody directed against the core protein of BM HSPGs. These antibodies enabled us to study the distribution of HSPG in detail. The aim of the present study was to investigate immunohistochemically the localization and distribution of HSPG-core protein and HSPG side chains in psoriatic lesions and to compare them to normal skin and uninvolved psoriatic skin.

MATERIALS AND METHODS

Patients and biopsies

Twenty patients with psoriasis and 20 healthy volunteers participated in this study. The 20 patients suffering from psoriasis (7 women and 13 men, age range 24–72 years; mean 48.2 years) had used no topical treatment for at least 2 weeks, and no systemic anti-psoriatic therapy for at least 1 year. After local anaesthesia, a biopsy (44 mm) from a psoriatic lesion was taken in 20 patients. In 6 patients, an additional biopsy was taken from a clinically uninvolved port of the skin, at a distance of at least 20 cm from a lesion.

In addition, biopsies were taken from the normal skin of 20 healthy volunteers without history or sign of skin disease (19 men and one woman, age range 25–56 years; mean 29.9 years).

Biopsies were embedded in Tissue Tek OCT compound (Miles Scientific, Naperville, U.S.A.), snap frozen in liquid nitrogen and stored at −80°C. Tissues were sectioned at 7 μm and fixed in acetone (0°C) for 10 min and again stored at −80°C until use.

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Permission to conduct this study was obtained from the local Ethics Committee of the University Hospital Nijmegen. Informed consent was given by all subjects in this study.

**Antibodies**

For immunohistochemical staining, three monoclonal antibodies against HSPG were used: mAb JM-72 (19), which is directed against the BM-associated HSPG-core protein, and two mAbs, designated as JM-13 (20) and JM-403 (21) against the HS-side chains. The production, characterisation and demonstration of specificity of these monoclonal antibodies were carried out by van den Born et al., as previously published (19-21). The two mAbs JM-13 and JM-403 recognise different epitopes along the HS polysaccharide chain: JM-13 binds to N-sulfated epitopes in HS and heparin (which is closely related to HS but much more sulfated); JM-403 preferentially binds to low-sulfated HS preparations. Moreover, the epitope is fully dependent on the presence of an N-unsubstituted glucosamine residue, thus having a free amino group. These units are not found in hyaluronic acid, thereby excluding binding of JM-403 to hyaluronic acid (22). In conclusion: mAb JM-13 binds to an epitope in the highly sulfated domains of HS and heparin, whereas mAb JM-403 binds to unmodified/low sulfated regions of HS.

**Immunohistochemical staining**

For all three antibodies an indirect peroxidase technique was used. Control stainings were carried out using nonimmune mouse IgG. After thawing at room temperature for 30 min, slides were fixed again in acetone (0°C) for 10 min, washed in phosphate-buffered saline (PBS) and incubated for 1 h at room temperature with the primary antibodies [anti HSPG-core mAb JM-72 (ascites, 1:250 in PBS), anti HS mAb JM-403 (ascites, 1:250) and anti HS mAb JM-13 (undiluted culture supernatant)].

After having been washed twice in PBS the slides were incubated for 30 min with a solution of rabbit-anti-mouse-immunoglobulins (DAKOPATTS, Copenhagen, Denmark) conjugated with peroxidase, diluted 1:50 in phosphate buffer containing 5% human AB-serum. After two further washes in PBS and a preincubation with sodium acetate buffer (pH 4.9), slides were stained in a solution of 3-amino-9-ethyl-carbazole (AEC) in sodium acetate buffer containing 0.01% H2O2 for 10 min. Slides were counterstained with Mayer’s haematoxylin (Sigma, St Louis, U.S.A.) and mounted in glycerin gelatin (Sigma, St Louis, U.S.A.).

**Histological examination**

The distribution of the staining in dermis, epidermis and basal membrane was assessed by two independent investigators using a 3-point scale: 0 = staining not present, 1 = discontinuously distributed staining, 2 = continuously distributed staining. In addition, staining of BM of blood vessels in the dermis was scored as being either present or absent and the presence of cell membrane-associated staining was recorded as well.

**RESULTS**

All control stainings using nonimmune mouse IgG were negative.

**JM-13**

All 20 specimens of psoriatic skin showed a striking staining pattern, completely different from the staining pattern in normal and uninvolved skin. JM-13 staining was consistently absent in the tips of the dermal papillae. A clear, linear staining was present invariably in the lower parts of the dermal papillae (Fig. 1b).

The JM-13 staining in all 20 specimens of normal skin showed a continuous staining of the BM at the dermo-epidermal junction (Fig. 1a). The clinically uninvolved skin of psoriatic patients showed a pattern that was similar to that observed in healthy volunteers. In contrast to the antibodies JM-403 and JM-72, mAb JM-13 only stained the BM of the dermo-epidermal junction and not the BM of the bloodvessels.

**JM-403**

In all specimens from normal skin, JM-403 staining, directed against the unmodified/low sulfated parts of HS, displayed a strong honeycomb-like staining in the epidermis (Fig. 2a). The staining pattern suggested staining of membrane-bound structures or intercellular substances. This staining pattern was most pronounced in the stratum spinosum and stratum granulosum and was less prominent in the stratum corneum and basal layers. In uninvolved epidermis of psoriatic patients the cell membrane associated staining was seen more focally and was less conspicuous, whereas this staining pattern was invariably absent in the lesional skin of all psoriatic patients (Fig. 2b).

In the BM of skin and bloodvessels the JM-403 staining was bright, continuous and present in all specimens.

**JM-72**

Staining with the antibody JM-72, directed against the core protein of the HSPG, was positive in all specimens. In normal
Fig. 2. Immunohistochemical staining of HS-side chains by mAb JM-403. (a) Normal skin. Note the honeycomb-like staining in the epidermis. (b) Psoriasis. Note the absence of honeycomb-like staining.

Fig. 3. Immunohistochemical detection of HSPG-core protein by mAb JM-72. (a) Normal skin. (b) Psoriasis.

skin, uninvolved psoriasis and psoriatic lesions the staining was restricted to the BM of the dermal-epidermal junction and the BMs of the blood vessels in the dermis. There was a strong and continuous linear staining (Fig. 3). No additional structures were stained.

**DISCUSSION**

At present little is known about the distribution of HSPGs in human skin. In normal human epidermis, Tammi et al. localised keratinocyte surface-associated HSPGs by means of electron microscopy (23), and recently, van den Born et al. described the expression of BM-associated HSPGs in normal human skin (19).

In this study we analysed the distribution of HSPGs in psoriasis, uninvolved psoriasis and normal skin by means of immunohistochemical staining. Three new monoclonal antibodies were used, two directed against the HS-side chains (JM-13 and JM-403) and one against the HSPG-core protein (JM-72) (19–21). mAb JM-72 gave a continuous, similar staining in all specimens, indicating that the core protein of HSPG is present in normal skin, uninvolved psoriasis and psoriatic skin without a difference in distribution. This also applies to the staining of the BM by the mAb JM-403, directed against the HS-side chains, which also gave a bright, continuous staining in all specimens.

However, staining with JM-13, directed against a different epitope on the HS-side chain, showed a clearly different pattern in all specimens. It was absent in the tips of the dermal papillae of all psoriasis specimens. The heterogeneity in staining of the HS-side chains by the two antibodies JM-13 and JM-403 can be explained by the different epitopes recognised. The epitope of JM-403 is located in the unmodified/low sulfated domains of HS. This is in contrast with the epitope of JM-13, which is located in the N-sulfated parts of HS-side chain.

Several mechanisms can explain the absence of JM-13 staining in the tips of the dermal papillae. This might be the result of decreased accessibility of HS due to epitope masking by cationic molecules or immune complexes, or alternatively the epitopes are lost due to enzymatic cleavage of HS. A third possibility could be a steric hindrance of BM components caused by changed molecular interactions. These three possibilities are discussed below.

In psoriasis, neutrophil polymorphonuclear leukocytes (PMNs) and plasma proteins move from the tip of the capillary loops in the dermal papillae into the epidermis (24, 25). It is striking to observe that in this part of the basal membrane, where the PMNs and plasma proteins pass, the JM-13 staining is absent. It is tempting to assume a connection between these events. Maybe the JM-13 epitope is blocked by plasma and/or PMN-derived cationic proteins, remaining the unmodified/low sulfated regions in HS accessible for JM-403 binding. This mechanism has been described in previous studies on SLE-nephritis, showing a binding of cross-reactive anti-nuclear antibodies to HS (26, 27). Recently, the adherence of leukocyte...
integron Mac-1 to heparan sulfate has been described (28). The adhesion to heparan sulfate of this integrin, which co-ordinates adhesive functions including leukocyte migration, strengthens this explanation for the absence of JM-13 staining.

A second explanation might be the lytic action of inflammatory cells, such as PMNs, that may destroy the JM-13 epitope. Thirdly, many interactions between the HS-side chain and the BM components fibronectin, laminin and collagen type IV are known. Battaglia et al. describe the formation of ternary complexes between laminin, nidogen and proteoglycan, suggesting a key role for nidogen in basement membrane assembly (15, 29). It could thus be that one of these interactions is changed in the tips of the dermal papillae in such a way that the accessibility of the JM-13 epitope is altered by steric hindrance of BM components. A previous study already found a general loss of BM integrity in psoriatic lesions (30) and by electron microscopy, Cox (31) found foci of discontinuity in the basal lamina, especially in active lesions.

A second interesting finding was the honeycomb-like staining of the epidermis with mAb JM-403 in normal skin and non-lesional psoriatic skin and the remarkable absence of this staining in psoriasis. Most likely this is due to binding of the antibody to the keratinocyte plasma membrane-associated HSPGs. In literature, however, such crossreactivity has never been described (32, 33). Two possible candidates of cell membrane-associated HSPGs in skin which share homologies with BM-associated HSPGs are syndecan and CD44v3. The possibility of staining of hyaluronic acid by mAb JM-403 was ruled out, as the epitope of mAb JM-403 is fully dependent on the presence of an N-unsubstituted glucosamine residue, thus having a free amino group. These units are not found in hyaluronic acid (22).

Both CD44v3 and syndecan-1 are known to play an important role in cell-cell interactions, cell-ECM interactions and the binding of growth factors (34–36). These biological functions are important in the regulation of cell growth and migration. Absence of either syndecan-1 or CD44v3 in the epidermis of a psoriatic lesion could contribute to changes in proliferation, differentiation and migration in psoriasis.

In conclusion, an altered HSPG expression in psoriasis was described compared to normal skin, namely the absence of high sulfated HS-side chains (JM-13) in the tips of the dermal papillae in psoriatic patients and the different staining pattern with mAb JM-403 against low sulfated HS-side chains in psoriasis compared to uninvolved and normal skin.

To the best of our knowledge, these changes in HSPG expression in psoriasis have never been described before. These data may provide insight into the role of HSPGs in psoriasis, which remains further to be elucidated.

REFERENCES


