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Dietary n-3 Fatty Acids Increase Spleen Size and Postendotoxin Circulating TNF in Mice; Role of Macrophages, Macrophage Precursors, and Colony-Stimulating Factor-1

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In experimental studies in mice, dietary supplementation with n-3 fatty acids (FA) alleviates inflammation and increases resistance to infection. Nevertheless, TNF production capacity was found to be increased in n-3 FA-fed mice. We previously found increased relative spleen weights in n-3 FA-fed mice. In this study, the nature of this increased spleen size was further investigated. Spleen cellularity was increased significantly in mice fed n-3 FA (fish oil 15% w/w), compared with controls fed corn oil (15%) or normal lab chow (p < 0.05). Experiments with T cell-deficient nude mice and experiments using macrophage depletion through liposomal dichloromethylene-biphosphonate revealed that the increase in spleen cellularity is T cell independent and largely due to macrophage accumulation in the spleen. Accumulation of marginal zone and red pulp macrophages was histologically and immunohistochemically confirmed. n-3 FA induced peripheral blood monocytes and an aspecific increase in bone marrow cellularity. Postendotoxin circulating TNF concentrations were increased significantly in n-3 FA-fed mice compared with controls. Splenectomy did not abolish this increase in circulating TNF. However, macrophage depletion through liposomal dichloromethylene-biphosphonate, circulating TNF was not detectable after endotoxin challenge. Circulating concentrations of CSF-1 did not differ between the various experimental groups. It is suggested that the cellular changes observed relate to increased constitutive production of TNF.


Fatty acids (FA) such as eicosapentaenoic acid and docosahexaenoic acid are polyunsaturated (n-3) FA, in which the last double bond is located between the third and fourth carbon atom from the methyl end of the FA chain. Dietary supplementation with n-3 FA results in modification of the FA composition of the cell membranes, thereby reducing the production and biologic activity of prostaglandins and leukotrienes (1). More recently, dietary supplementation with n-3 FA has been associated with modulations in the production of the proinflammatory cytokines IL-1 and TNF, the principal polypeptide mediators of inflammation (2, 3).

In experimental studies, dietary supplementation with n-3 FA has been shown to alleviate inflammation in various clinical disease states and in animal models (4–11). Preliminary evidence indicates that dietary n-3 FA supplementation may increase resistance to infection in murine trauma patients (12). n-3 FA increase survival following endotoxic shock or infections with live microorganisms in various animal models (13–18), and dietary n-3: FA increase IL-1α and TNF production capacity of mouse peritoneal cells (15, 19, 20). In the course of our experiments in mice, we noticed a 1.5-fold increase in relative spleen weight in n-3 FA-fed mice compared with controls. Similar observations have been reported by other groups studying mice or rats (21–23).

To elucidate the mechanism of the enhanced resistance to infections and possibly relate this to the increased spleen weight in mice, we studied the effects of dietary n-3 FA on the cellular composition of spleens, peripheral blood, and bone marrow. In addition, the effects of dietary n-3 FA on circulating concentrations of endotoxin-induced TNF and steady state CSF-1 were investigated.

Materials and Methods

Mice

In most experiments, 6-wk-old female Swiss mice, approximate weight 25 g, were used. The animals were housed in plastic cages (type II and type III; Macrolon, Beyer en Eggelaar, The Netherlands) under specific pathogen-free conditions at 23 ± 2°C and 7 ventilations/h. In some experiments, normal female BALB/c and T cell-deficient female BALB/c nude mice (nu/nu) were used. Nude mice were housed in cages covered with sterilized filter bonnets.

Diet

Mice were fed a fat-free standard reference diet a: a dry powder, supplemented with 14% fish oil (PO) concentrate and 1% corn oil (PO group), 15% corn oil (CO), or normal lab chow. The FC concentrate used was EPAX 3000 TG, a kind gift of Pronova (Bergen, Norway). The PO concentrate contained 37.1% n-3 FA, i.e., 17.7% eicosapentaenoic acid and 14.4% docosahexaenoic acid (w/w), α-linolenic acid (0.67 mg/g) and a natural mix of tocopherols (0.525 mg/g) were added to the PO diet to avoid essential FA deficiency. The PO concentrate was kept under strict anaerobic conditions until preparation of the diet to avoid auto-oxidation. Diets were prepared at least twice per week, and kept at 4°C until administration.
Table I. Effect of dietary n-3 FA on spleen, peripheral blood, and bone marrow cells

<table>
<thead>
<tr>
<th></th>
<th>Fish oil</th>
<th>Corn oil</th>
<th>Normal</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Spleen</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Relative spleen weight (mg/g)</td>
<td>5.9 ± 0.5</td>
<td>4.8 ± 0.3</td>
<td>4.6 ± 0.2</td>
<td>0.01</td>
</tr>
<tr>
<td>Spleen cellularity (× 10⁶)</td>
<td>2.7 ± 0.2</td>
<td>2.0 ± 0.2</td>
<td>1.4 ± 0.5</td>
<td>0.02</td>
</tr>
<tr>
<td>B 220 positive spleen cells (%)</td>
<td>41 ± 4</td>
<td>36 ± 2</td>
<td>31 ± 6</td>
<td>0.04</td>
</tr>
<tr>
<td><strong>Peripheral blood</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hematocrit</td>
<td>43 ± 2.6</td>
<td>42 ± 3.2</td>
<td>43 ± 1.6</td>
<td>NS</td>
</tr>
<tr>
<td>White blood cell count (× 10⁶/ml)</td>
<td>12.2 ± 4.0</td>
<td>9.1 ± 1.9</td>
<td>7.9 ± 1.3</td>
<td>NS</td>
</tr>
<tr>
<td>Lymphocytes (%)</td>
<td>55 ± 8</td>
<td>69 ± 4</td>
<td>64 ± 5</td>
<td>NS</td>
</tr>
<tr>
<td>Granulocytes (%)</td>
<td>34 ± 8</td>
<td>25 ± 4</td>
<td>31 ± 6</td>
<td>NS</td>
</tr>
<tr>
<td>Monocytes (%)</td>
<td>11 ± 2</td>
<td>6 ± 1</td>
<td>5 ± 1</td>
<td>0.02</td>
</tr>
<tr>
<td><strong>Bone marrow</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cellularity (× 10⁶)</td>
<td>24 ± 5</td>
<td>21 ± 7</td>
<td>13 ± 5</td>
<td>0.02</td>
</tr>
<tr>
<td>M-CFC (No. macrophage precursors/10⁶ NBMPC)</td>
<td>58 ± 15</td>
<td>51 ± 17</td>
<td>77 ± 20</td>
<td>NS</td>
</tr>
<tr>
<td>ER-MP 21 high (erythroid cells) (%)</td>
<td>94 ± 22</td>
<td>80 ± 16</td>
<td>98 ± 27</td>
<td>NS</td>
</tr>
<tr>
<td>ER-MP 58 high (myeloid cells) (%)</td>
<td>22 ± 5</td>
<td>25 ± 3</td>
<td>22 ± 5</td>
<td>NS</td>
</tr>
<tr>
<td>ER-MP 12 high/20 neg (committed progenitors) (%)</td>
<td>50 ± 7</td>
<td>50 ± 2</td>
<td>54 ± 6</td>
<td>NS</td>
</tr>
<tr>
<td>ER-MP 12 pos/20 pos (immature myeloid precursors) (%)</td>
<td>1.2 ± 0.3</td>
<td>1.7 ± 0.2</td>
<td>1.6 ± 0.5</td>
<td>NS</td>
</tr>
<tr>
<td>ER-MP 12 neg/20 high (monocytes) (%)</td>
<td>5.4 ± 1.2</td>
<td>5.6 ± 0.5</td>
<td>5.0 ± 1.2</td>
<td>NS</td>
</tr>
</tbody>
</table>

Spleen analysis

The mice were anesthetized with ether, and blood was taken from the retrobulbar vessels after eye extraction. The animals were killed by neck dislocation, and spleens were removed and weighed. Total body weight was determined immediately before bleeding, and relative spleen weight was calculated afterward. A suspension of spleen cells was made over a nylon filter, and cells were counted with a Coulter counter (Coulter Corp., Hialeah, FL). The effect of dietary n-3 FA on spleen weight was determined in normal Swiss mice, normal BALB/c mice, and T cell-deficient nude BALB/c mice (nu/nu).

Spleen cells were phenotypically characterized with flow cytometry using a set of mAbs specific for mouse cells: 59-AD 2.2 detecting Thy-1 (T lymphocytes), RA3 6B2 detecting B220 (B lymphocytes), R129.19 detecting MT4 (CD4-positive T lymphocytes), 53-6.72 detecting Ly-2 (CD8-positive T lymphocytes), KT3 (CD3-positive T lymphocytes), and F4/80 (macrophages). Details concerning these Abs can be found in Leenen et al. (24).

After immunohistochemical staining of spleen sections with RA3 6B2 (anti-B220), a mAb specific for mouse B cells*, the relative surface area of white pulp was quantitated using a Leitz Diaplan light microscope (Leitz, Wetzlar, Germany) and a Videoplan image-processing system (Kontron, FillErlands), as described (27). The following mAbs were used: ER-MP12 and ER-MP20, identifying subpopulations of macrophage precursors; ER-MP58, identifying myeloid cells; and ER-MP21, identifying transferrin receptor-expressing cells (28).

Postendotoxic circulating TNF

At a time period of 1.5 h after i.p. administration of 10 μg of LPS (Escherichia coli, serotype O55:B5; Sigma Chemical Co., St. Louis, MO), mice were anesthetized with ether, and blood was taken from the retrobulbar vessels after eye extraction. Blood samples (approximately 1 ml) were mixed with 100 μl of EDTA solution (21.4 mg/ml of EDTA in H₂O) and kept on ice until centrifugation at 1500 × g for 5 min. Resultant plasma was isolated and stored at −20°C until assay.

TNF-α was measured by ELISA using TN3, a hamster mAb specific for murine TNF-α, and lymphotoxin, as described (29, 30).

In separate experiments, the effect of splenectomy on postendotoxic circulating TNF was studied. Splenectomy was performed after ether anesthesia in 4-wk-old mice, 2 wk before the animals were started on the experimental diet.

Colon-stimulating factor-1

CSF-1 was determined in unstimulated serum samples by specific RIA, as described in detail elsewhere (31). This assay is based on the competition by CSF-1 for the interaction between ¹²⁵I-labeled, purified mouse L cell CSF-1 glycoprotein and a rabbit polyclonal Ab to purified L cell CSF-1 (32), and is more sensitive than the conventional CSF-1 bioassay based on bone marrow colony formation. Assays were conducted in duplicate on 20-μl samples. The concentration of CSF-1 in U/ml (1 U = 12 pg) was determined with reference to a standard curve prepared using a stable, partially purified L cell CSF-1 preparation.

Statistical analysis

Differences between groups were analyzed using the Kruskal-Wallis non-parametric ANOVA test, corrected for ties. Results were considered statistically significant at p < 0.05.

Results

Effects of FO supplementation on spleen cellularity

The effects of dietary n-3 FA on spleen, peripheral blood, and bone marrow cells are summarized in Table I. The following experiments were done to elucidate the mechanisms of the increased spleen weight in FO-fed mice. After 4 wk of dietary supplementation with FO or CO, experimental Swiss mice were compared with control animals on normal lab chow. FO-fed mice had significantly increased relative spleen weights (p = 0.01) and spleen...
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FIGURE 1. Time-effect relationships of dietary n-3 FA on relative spleen weight (A) and on postendotoxin circulating TNF (B). Relative spleen weight was increased significantly from 2 wk on FO diet, as was postendotoxin circulating TNF (n = 12).

cellularity (p = 0.02). In separate experiments, time-effect relationships in these responses were investigated. Mice were studied simultaneously after 0, 2, 4, and 6 wk of dietary FO supplementation. Increases in spleen weight were most prominent after 2 wk, and appeared somewhat blunted after 6 wk (Fig. 1A). Similar results were obtained in normal BALB/c mice. To investigate whether the increase in spleen size was accounted for or mediated by T cells, T cell-deficient female BALB/c mice were fed FO for 0, 2, or 4 wk. Relative spleen weight was increased significantly from 2 wk on, indicating that the increase in spleen weight was independent of the presence of T cells (results not shown).

Phenotypic characterization of spleen cells using a set of mAbs revealed no percentual differences in T lymphocytes or macrophages between the various dietary groups. However, FO-fed mice had a small but significant increase in B220-positive B lymphocytes (Table I).

FIGURE 2. BALB/c mice were fed FO for 0, 2, or 4 wk. At the end of the diet period, subsets of mice were injected i.v. with liposomal C12MDP 24 h before LPS challenge. In contrast to mice injected with saline, animals injected with liposomal C12MDP did not show a significant increase in relative spleen weight following 2 or 4 wk of FO supplementation (A). Postendotoxin circulating TNF was not detectable in the liposomal C12MDP-treated mice (B). *, Indicates a statistically significant difference compared with mice that had no FO diet.

Pretreatment with liposomal C12MDP 24 h before endotoxin challenge did not significantly affect spleen weight. However, in mice that had been on FO diet for 2 or 4 wk, liposomal C12MDP resulted in a decrease in spleen weight (Fig. 2A). In these FO-fed mice, treatment with liposomal C12MDP resulted in spleen weights that were not significantly different from mice that had been on normal diet only. Depletion of marginal zone and red pulp macrophages was histologically confirmed, but accurate quantitation of F4/80-positive macrophages was not possible in cell suspensions, in which their numbers were equal in control and FO-fed mice. However, for other reasons (see below), the results indicate that the increase in spleen weight in FO-fed mice is mainly attributable to an increase in the splenic macrophage population that is depleted by pretreatment with liposomal C12MDP.

To a lesser extent, B lymphocytes seem to contribute to the increased spleen cellularity in FO-fed mice. Quantification of the relative surface area of red and white pulp in spleen sections indicated a decrease in T and B cell-containing white pulp (Fig. 3).

The effect of FO supplementation on peripheral blood cells

To investigate a possible relation between circulating blood cells and changes in spleen cellularity, peripheral blood cells were quantitated. Hematocrit did not differ between the various treatment
groups. White blood cell count was higher in FO-fed mice, but this difference did not reach statistical significance. Analysis of the light-scatter profile of peripheral blood cells showed a significant increase in percentage of monocytes in FO-fed mice ($p = 0.02, n = 6$).

The effect of FO supplementation on bone marrow cells

Possible relations between changes in spleen and peripheral blood cellularity and the cellular composition of bone marrow were investigated. Bone marrow cellularity was increased significantly in FO-fed mice compared with mice fed CO or normal lab chow. This increase in bone marrow cellularity was equally distributed over the various hemopoietic lineages, since differential flow-cytometric analysis of bone marrow cells, using ER-MP12 and ER-MP20 mAbs, showed no significant differences between the various experimental groups (Table I). Moreover, soft agar cultures of bone marrow cells did not reveal a relative increase in macrophage precursors, measured as macrophage colony-forming cells or macrophage cluster-forming cells.

The effect of FO supplementation on postendotoxin TNF

To investigate the suggested increase in mononuclear phagocyte functionality, we measured circulating TNF concentrations after endotoxin challenge. Postendotoxin concentrations of circulating TNF were significantly higher in FO-fed mice than in control mice. Postendotoxin TNF was 10.4 ± 6.6 ng/ml in FO-fed mice, 2.2 ± 1.4 in CO-fed mice, and 2.7 ± 1.3 in mice fed normal diet (mean ± SD, $p < 0.05$). In separate experiments, time-effect relationships in TNF production and FO diet were investigated. Similar to the increase in spleen weight, increases in postendotoxin circulating TNF were most prominent after 2 wk of FO diet, and appeared somewhat blunted after 6 wk (Fig. 1B). Postendotoxin TNF in T cell-deficient nude BALB/c mice was increased significantly from 2 wk on, indicating that the increase in postendotoxin circulating TNF was independent of the presence of T cells (results not shown).

Splenectomy, performed 2 wk before starting on the FO-supplemented diet, did not influence the increase in postendotoxin circulating TNF concentrations after 2 and 4 wk of diet: at both time points, postendotoxin circulating TNF concentrations were increased significantly compared with baseline (results not shown). At baseline and at 2 and 4 wk of dietary FO supplementation, postendotoxin circulating TNF was not detectable in liposomal Cl$_2$MDP-treated mice (Fig. 2B). These observations suggest that splenic macrophages, despite their accumulation after dietary FO supplementation, are not the most important contributors to the production of circulating TNF. On the other hand, the macrophage population that is depleted by liposomal C12MDP suggest that the observed increase in spleen size following dietary FO supplementation is largely due to accumulation of macrophages in the spleen. Since TNF is an important growth factor for lymphoid organs, increased endogenous TNF production in FO-fed mice may contribute to this effect (44). The phenotypic characterization of spleen cell suspensions using a set of mAbs is not suitable for the assessment of the number of macrophages in the spleen, since especially tissue-fixed macrophages are lost in the process of making a cell suspension over a nylon filter. We therefore regard the concept that dietary FO supplementation leads to macrophage accumulation in the spleen as valid.

The increase in spleen size after dietary n-3 FA has not been described before, but may be induced through a feedback mechanism following the accumulation of macrophages in the red pulp of the spleen. It is apparently not due to increased concentrations of CSF-1. This is in accordance with the observation that the number of red pulp macrophages is reduced only slightly in CSF-1-deficient op/op mice (43). Other factors, including IL-3, granulocyte-macrophage CSF, and the proinflammatory cytokines, may be considered. The results of our experiments with macrophage depletion using liposomal Cl$_2$MDP suggest that the observed increase in spleen size following dietary FO supplementation is largely due to accumulation of macrophages in the spleen. The immunohistochemical staining of spleen sections with B220 showed a decrease in the relative surface area of white pulp in the FO-fed mice, thereby supporting the concept of accumulation of red pulp macrophages in FO-fed mice. As regulator cells, macrophages are responsible for the growth of lymphoid organs such as the spleen. Since TNF is an important growth factor for lymphoid organs, increased endogenous TNF production in FO-fed mice may contribute to this effect (44). The phenotypic characterization of spleen cell suspensions using a set of mAbs is not suitable for the assessment of the number of macrophages in the spleen, since especially tissue-fixed macrophages are lost in the process of making a cell suspension over a nylon filter. We therefore regard the concept that dietary FO supplementation leads to macrophage accumulation in the spleen as valid.

Discussion

The present study shows that dietary n-3 FA supplementation has a significant effect on the generation and distribution of mononuclear phagocytes in mice: n-3 FA induce a generalized increase in bone marrow cellularity, peripheral blood monocytopsis, and accumulation of macrophages in the spleen, leading to an increase in spleen size. What could be the mechanism of these n-3 FA-induced changes?

The aspecific increase in bone marrow cellularity without preferential stimulation of monocytopsis is in accordance with our observation on CSF-1: this specific monocyte growth factor was not increased in the n-3 FA-fed mice. The generalized increase in bone marrow cellularity suggests that earlier, broad spectrum hemopoietic growth factors such as IL-3 and granulocyte-macrophage CSF are involved. The production of these factors is largely regulated by proinflammatory cytokines including IL-1 (36, 37). We have shown previously that IL-1α production capacity is increased in mice after dietary n-3 FA (15). Increased constitutive production of this cytokine may be involved in the increased bone marrow cellularity observed in the present study. Moreover, Pelus and others have shown that PGE$_2$ has an important modulatory effect on hemopoiesis in mice: exogenous administration of PGE$_2$ reduced nucleated bone marrow and splenic cellularity, while blockade of PGE$_2$ biosynthesis increased bone marrow and splenic cellularity, especially in the presence of IL-1α (38–42). These observations may partly explain the results obtained in the present study, since dietary supplementation with n-3 FA in mice basically leads to increased IL-1α and a decreased PGE$_2$ production capacity.

Peripheral blood monocytopsis induced by dietary n-3 FA has not been described before, but may be induced through a feedback mechanism following the accumulation of macrophages in the red pulp of the spleen. It is apparently not due to increased concentrations of CSF-1. This is in accordance with the observation that the number of red pulp macrophages is reduced only slightly in CSF-1-deficient op/op mice (43). Other factors, including IL-3, granulocyte-macrophage CSF, and the proinflammatory cytokines, may be considered. The results of our experiments with macrophage depletion using liposomal Cl$_2$MDP suggest that the observed increase in spleen size following dietary FO supplementation is largely due to accumulation of macrophages in the spleen.
postendotoxin circulating TNF. It might be expected that Kupffer cells of the liver, due to their sinusoidal location, might be a major target for the uptake of liposomal Cl2MDP. In fact, recent evidence indicates that macrophage depletion in mice results in a 50 to 70% reduction in TNF mRNA in the liver following endotoxin challenge (46). Therefore, the Kupffer cells may be the most important contributors to the production of postendotoxin TNF. Similar to peritoneal macrophages, Kupffer cells may have increased TNF production capacity at the single cell level following dietary n-3 FA. We did not assess the number of Kupffer cells. The small increase in relative liver weight in FO-fed mice does not rule out a substantial increase in the number of Kupffer cells.

In conclusion, dietary n-3 FA supplementation in mice induces an aspecific increase in bone marrow cellularity, peripheral blood monocytecytosis, accumulation of macrophages in the spleen, and increased postendotoxin circulating concentrations of TNF. The mechanism of these changes remains to be elucidated.

Acknowledgments

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References