Role of Negatively Charged Residues in the Fifth and Sixth Transmembrane Domains of the Catalytic Subunit of Gastric H⁺,K⁺-ATPase*

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The role of six negatively charged residues located in or around the fifth and sixth transmembrane domains of the catalytic subunit of gastric H⁺,K⁺-ATPase, which are conserved in P-type ATPases, was investigated by site-directed mutagenesis of each of these residues. The acid residues were converted into their corresponding amides. S99 cells were used as the expression system using a baculovirus with coding sequences for the α- and β-subunits of H⁺,K⁺-ATPase behind two different promoters. Both subunits of all mutants were expressed as the wild-type enzyme in intracellular membranes of S99 cells as indicated by Western blotting experiments, an enzyme-linked immunosorbent assay, and confocal laser scan microscopy studies. The mutants D824N, E834Q, E837Q, and D839N showed no 3-(cyanomethyl)-2-methyl-8(phenylmethoxy)-imidazo[1,2a]pyridine (SCH 28080)-sensitive ATP dependent phosphorylation capacity. Mutants E795Q and E820Q formed a phosphorylated intermediate, which, like the wild type enzyme, was hydroxylamine-sensitive, indicating that an acylphosphate was formed. Formation of the phosphorylated intermediate from the E795Q mutant was similarly inhibited by K⁺ (I₅₀ = 0.4 mm) and SCH 28080 (I₅₀ = 10 nm) as the wild type enzyme, when the membranes were preincubated with these ligands before phosphorylation. The dephosphorylation reaction was K⁺-sensitive, whereas ADP had hardly any effect. Formation of the phosphorylated intermediate of mutant E820Q was much less sensitive toward K⁺ (I₅₀ = 4.5 mm) and SCH 28080 (I₅₀ = 1.7 μm) than the wild type enzyme. The dephosphorylation reaction of this intermediate was not stimulated by either K⁺ or ADP. In contrast to the wild type enzyme and mutant E795Q, mutant E820Q did not show any K⁺-stimulated ATPase activity. These findings indicate that residue Glu⁸²⁰ might be involved in K⁺ binding and transition to the E₂ form of gastric H⁺,K⁺-ATPase.

Transport ATPases are able to convert the energy from ATP into active ion transport. ATPases of the P-type class (1) form an acid-stable phosphorylated intermediate during the catalytic cycle. This phosphorylated intermediate contains an aspartyl phosphate residue present in a conserved domain in the large intracellular loop, which in mammalian P-type ATPases is located between the fourth and fifth transmembrane segments of the catalytic subunit (2).

Phosphorylation of this residue and ion transport are coupled in such a way that specific binding of the cation that has to be transported to the extracellular or intravesicular medium stimulates phosphorylation, whereas binding of the cation to be transported into the cytosol stimulates dephosphorylation. The latter process has only been demonstrated unequivocally in Na⁺,K⁺-ATPase and the gastric H⁺,K⁺-ATPase. The molecular mechanism of the coupling between the phosphorylation process on the one side and ion binding and transport on the other side is still far from being elucidated.

It is generally assumed that polar amino acid residues present in the transmembrane domains might play a key role in transmembrane ion transport. In particular, negatively charged residues like those originating from aspartate and glutamate are likely candidates for such a role (3, 4). The presence of four transmembrane segments in the N-terminal part of the catalytic subunits of these proteins is generally accepted. In these four transmembrane regions there is only one conserved negatively charged amino acid residue, which might be involved in transmembrane ion transport (3, 5–8). In the C-terminal part of the catalytic subunit, however, the secondary structure is still disputed. Most authors assume the presence of six transmembrane segments in this area, but several models with only four transmembrane segments have been proposed too (9, 10). In the last transmembrane segment there is a pair of negatively charged conserved amino acid residues, but mutational studies up to now give no indication for an important role in Na⁺,K⁺-ATPase (11, 12).

Most negatively charged residues are present in the fifth and sixth transmembrane segments (see Fig. 1) that are assumed to be immediately C-terminal of the large intracellular loop. This region of P-type ATPases, however, is rather peculiar. Because of the relatively large number of negatively charged and other polar residues, the hydrophobic index is rather low. Moreover, this region contains a number of proline residues, which generally give a break in an α-helix structure. In vitro translation studies with H⁺,K⁺-ATPase (13) did not show membrane insertion properties for the fifth and sixth transmembrane segments. For sarcoplasmic and endoplasmic reticulum (SERCA)¹-type Ca²⁺-ATPase, only a stop-transfer signal was found for the fifth but not for the sixth transmembrane segment. No signal anchor sequence was found for either of these.

1 The abbreviations used are: SERCA, sarcoplasmic and endoplasmic reticulum Ca²⁺-ATPase; E-P, phosphorylated intermediate; PAGE, polyacrylamide gel electrophoresis; PBS, phosphate-buffered saline; SCH 28080, 3-(cyanomethyl)-2-methyl-8(phenylmethoxy)imidazo[1,2α]pyridine.

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transmembrane segments (14). Lutsenko et al. (15) recently showed that extensive tryptic digestion of Na\(^+\),K\(^+\)-ATPase led to membrane release of a water-soluble fragment (Gln\(^{977}\)_Arg\(^{985}\)), which included the putative M5-M6 hairpin. Occlusion of K\(^+\), however, prevented the release of this fragment from the membrane. These findings suggest that (i) the putative transmembrane segments M5 and M6 might be present in the membrane in a form different from a classical \(\alpha\)-helix and (ii) this part of the \(\alpha\)-subunit plays a role in cation binding and transport.

The putative M5-M6 region in gastric H\(^+\),K\(^-\)-ATPase contains at least three conserved negatively charged residues (Glu\(^{794}\), Glu\(^{820}\), and Asp\(^{824}\)). Moreover, quite close to the C terminus of M6 there are three other negatively charged residues (Glu\(^{834}\), Glu\(^{837}\), and Asp\(^{839}\)), which are also conserved in other ATPases. Although most models for P-type ATPases (16, 17) do not consider these residues as being present in transmembrane segments, some models (see Fig. 1) do (18–21). One of the reasons for the discrepancies between the models for H\(^+\),K\(^-\)-ATPase and other mammalian P-type ATPases is the presence of two cysteine residues in the M5-M6 region, some models (see Fig. 1) do not consider these residues as being present in transmembrane segments, some models (see Fig. 1) do (18–21). One of the reasons for the discrepancies between the models for H\(^+\),K\(^-\)-ATPase and other mammalian P-type ATPases is the presence of two cysteine residues in the M5-M6 region, at least one of which is the target for extracellularly applied acid-activated omeprazole, an inhibitor of gastric acid secretion (19). Moreover, there is a cytosolically located tryptic digestion site at Lys\(^{793}\) (19, 20), of which the corresponding amino acid residue in most models is placed within M5. This results in a more C-terminally located position of the transmembrane segments M5 and M6 in H\(^+\),K\(^-\)-ATPase as compared with the original models for Ca\(^{2+}\)-ATPase and Na\(^+\),K\(^-\)-ATPase (16, 17).

Despite this uncertainty, a number of site-directed mutagenesis studies aimed at elucidating the function of these negatively charged residues in the M5-M6 region in Ca\(^{2+}\)-ATPase of both the SERCA (3, 5, 22–24) and the plasma membrane type (25) as well as in Na\(^+\),K\(^-\)-ATPase (26–29) have been performed recently. From these studies, several candidate amino acids for a role in transmembrane cation transport have been proposed, but a consistent model has not yet been obtained.

With the gastric H\(^+\),K\(^-\)-ATPase (18, 21, 30), only one study with mutants has been published until now (31), since functional expression of this enzyme system has only recently been successfully carried out (31–33). We report here mutational studies in which six negatively charged amino acid residues within or close to the fifth and sixth transmembrane segments of the catalytic subunit have been converted into the corresponding acid amides. The study shows that the mutation E795Q has no effect, whereas the mutation D824N, E834Q, E837Q, or D839N prevents the formation of a phosphorylated intermediate. The mutation E820Q results in a phosphorylated intermediate with a markedly reduced sensitivity toward both K\(^+\) and the specific H\(^+\),K\(^-\)-ATPase inhibitor 3-(cyanomethyl)-2-methyl-8{ phenyl methoxy}imidazol-1,2-ajpyridine (SCH 28080) in the phosphorylation reaction. The hydrolysis of the phosphointermediate is not stimulated by K\(^+\), and no K\(^+\)-stimulated ATPase activity can be determined. This emphasizes the importance of these negatively charged residues in the function of H\(^+\),K\(^-\)-ATPase.

**Site-directed Mutagenesis**

**Site-directed Mutagenesis of Gastric H\(^+\),K\(^-\)-ATPase**

The putative M5-M6 region in gastric H\(^+\),K\(^-\)-ATPase from Shull et al. (16), for Na\(^+\),K\(^-\)-ATPase from Shull et al. (17), for sarcoplasmic reticulum Ca\(^{2+}\)-ATPase from MacLennan et al. (16), and for plasma membrane Ca\(^{2+}\)-ATPase from Verma et al. (16). The amino acid residues mutated in this study in gastric H\(^+\),K\(^-\)-ATPase as well as the corresponding amino acid residues in the other ATPases are given in **boldface type**. The position of the M5 and M6 domains is given according to the original authors except for H\(^+\),K\(^-\)-ATPase and Na\(^+\),K\(^-\)-ATPase, where the models of Klaassen and De Punt (21) and Lutsenko and Kaplan (2), respectively, are used.

**Transmembrane Segments**

Transmembrane segments (14). Lutsenko et al. (15) recently showed that extensive tryptic digestion of Na\(^+\),K\(^-\)-ATPase led to membrane release of a water-soluble fragment (Gln\(^{977}\)_Arg\(^{985}\)), which included the putative M5-M6 hairpin. Occlusion of K\(^+\), however, prevented the release of this fragment from the membrane. These findings suggest that (i) the putative transmembrane segments M5 and M6 might be present in the membrane in a form different from a classical \(\alpha\)-helix and (ii) this part of the \(\alpha\)-subunit plays a role in cation binding and transport.

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Production of Recombinant H+,K+-ATPase—Sf9 cells were grown at 27 °C either in 100-mI spinner flask cultures or as monolayer cultures in 175-cm² culture flasks as described by Klausen et al. (32). For production of H+,K+-ATPase, the cells were infected at a multiplicity of infection of 3 in the presence of 1° ethanol (5%) with the DLZJ1 virus. Big Blue positive recombinant viruses and infected Sf9 cells were harvested by centrifugation for 3 days. Occasionally the multiplicity of infection was varied from 0.01 to 10, and the Sf9 cells were incubated up to 5 days.

Confocal Laser Scan Microscopy—Sf9 cells were grown on sterile microscope coverslides in complete growth medium and infected with a multiplicity of infection of 3 at 27 °C. After infection, cells were incubated at 27 °C for 4 h in complete growth medium with additions as indicated. Cells were washed three times for 5 min with phosphate-buffered saline (pH 7.4) followed by a short wash with 0.1% (w/v) Mowiol (Hoechst, Amsterdam, The Netherlands), 2.5% (w/v) gelatin, and 2% fetal bovine serum. From this wash on, the coverslides were kept in the dark as much as possible, and specific binding sites for antibodies were blocked by incubation for 30–60 min in PBS, 0.05% polyoxymethylene sorbitan monolaurate (Tween 20), 1% (w/v) gelatin, and 2% fetal calf serum with gentle rocking. After washing with PBS, 0.05% Tween 20 three times for 5 min each, the cells were incubated with the polyclonal antibody HH-K (60 mg/ml) in PBS–5% (v/v) fetal bovine serum. Free antibodies were removed by washing the cells as above. Next, the cells were incubated with a fluorescently labeled secondary antibody (Dako, Glostrup, Denmark) for 1 h in PBS, 0.05% Tween 20, 1% (w/v) gelatin, and 2% fetal bovine serum. From this moment on, the coverslides were kept in the dark as much as possible. After washing as above, the cells were desalted by a short wash with water, dehydrated with 100% methanol, dried, and mounted in 10% glycerol, and 0.012% bromphenol blue. The protein was solubilized by addition of 100 mM NaOH in 0.1 M TrisCl (pH 8.5). They were examined on a Bio-Rad MRC1000 confocal microscope. Images were averaged over eight scans.

Preparation of Sf9 Membranes—The Sf9 cells were harvested by centrifugation at 2,000 × g for 5 min. After resuspension at 0 °C in 0.25 M sucrose, 2 mM EDTA and 25 mM Hepes/Tris (pH 7.0), the membranes were sonicated three times for 15 s at 60 W (Branson Power Company, Danbury, CT). After centrifugation for 30 min at 10,000 × g, the supernatant was recentrifuged for 60 min at 100,000 × g at 4 °C. The pellets were resuspended in the same mentioned buffer and stored at –20 °C.

Protein Determination—Protein was determined with the modified Lowry method described by Peterson (39) using bovine serum albumin as a standard.

Quantification of the Expression Level—The H+,K+-ATPase α-subunit content of the membrane fraction was determined by a quantitative enzyme-linked immunosorbent assay (40), using the monoclonal antibody 5D6 (41, 42) and using purified pig H+,K+-ATPase as reference.

SDS-PAGE of the Phosphorylated Proteins—After phosphorylation of 50–100 µg of membranes with [γ-32P]ATP, the reaction was terminated upon the addition of 0.5 M of stopping solution. After a 30-s centrifugation at 16,000 × g, the pellets were washed with 0.5 M of 30% (w/v) sucrose and resuspended in 40 µl of 2 × concentrated sample buffer, containing 100 mM Tris-Cl (pH 6.8), 5% dithiothreitol, 4% SDS, 20% glycerol, and 0.01% bromphenol blue. The protein was solubilized during a 30-min incubation at room temperature. Samples of 5 µl (2–10 pCi) of [γ-32P]-P-protein, were applied on the SDS-gels (Mini-Prutein II; Bio-Rad). Running gels were prepared with 6% polyacrylamide/N,N’-bismethacrylamide (ratio 40/1.07) in 100 mM Na+-phosphate buffer (pH 6.5) and 0.2% SDS, while the stacking gel contained 4% polyacrylamide. In the same 0.1 M Na+-phosphate buffer (pH 6.5), the electrophoresis was performed at 0 °C, first during 20 min at 30 mA and next at 60 mA. The upper electrophoresis was followed by another 0.1% SDS. After fixation in 5% (w/v) methanol, 7.5% (w/v) acetic acid for 15–30 min at room temperature, the gels were autoradiographed at –20 °C on a Kodak film (X-Omat AR, Eastman Kodak Co., Rochester, NY), for 3–16 h.

Western Blotting—Protein samples from the membrane fraction were solubilized in SDS-PAGE sample buffer and separated on SDS-gels containing 10% acrylamide according to Laemmli (43). For immunoblotting, the separated proteins were transferred to Immobilon polyvinylidene difluoride membranes. The α- and β-subunit of H+,K+-ATPase were detected as described earlier (32), with the polyclonal antibody HKB (38) recognizing the 565–585 region of the α-subunit of H+,K+-ATPase and the monoclonal antibody 2G11 (44) evoked against the β-subunit of H+,K+-ATPase, respectively.

K+-ATPase Activity Assay—The K+-activated ATPase activity was determined with a radiochemical method (45). For this purpose, 0.6–5 µg of Sf9 membranes were added to 100 µl of medium, which contained 10 µM γ32P-ATP (specific activity 100,000 Ci/mmol), 1.0 mM MgCl2, 0.2 mM EDTA, 0.1 mM ouabain, 1 mM NaN3, 25 mM Tris-HCl (pH 7.0), andvarying concentrations of KCl. After incubation for 30 min at 37 °C, the reaction was stopped by adding 500 µl of ice-cold 10% (w/v) charcoal in 6% (w/v) trichloroacetic acid, and after 10 min at 0 °C, the mixture was centrifuged for 10 s (10,000 × g). To 0.2 ml of the clear supernatant, containing the liberated inorganic phosphate (γ32P), 3 ml of OptiFluor (Camlab Packard, Tilburg, The Netherlands) was added, and the mixture was analyzed by liquid scintillation analysis. In general, two or three samples were run in each experiment. K+-ATPase activity is presented as the percentage of the activity in the absence of added K+ (5 µM), which is 70–150 mP of P, liberated per mg per h. The latter activity is endogenously present in membranes of Sf9 cells.

ATP Phosphorylation Capacity—ATP phosphorylation was determined as described before (37). Sf9 membranes (40–100 µg) were incubated at 0 °C in 25 mM Tris-acetic acid (pH 6.0), 1 mM MgCl2 with and without 0.1 mM SCH 28080 in 0.2% (w/v) ethanol in a volume of 50 µl. After 30–60 min of preincubation, 10 µl of 0.6 µM γ32P-ATP was added, and the mixture was incubated for another 10 s at 0 °C. The reaction was stopped by adding 5% trichloroacetic acid in 0.1 M phosphoric acid, and the phosphorylated protein was collected by filtration over a 0.8-µm membrane filter (Schleicher and Schuell, Dassel, Germany). After repeated washing, the filters were analyzed by liquid scintillation analysis. The SCH 28080-sensitive part is defined as the H+,K+-ATPase-dependent phosphorylation; the activity in the presence of SCH 28080 is the background phosphorylation.

Dephosphorylation Studies—After ATP phosphorylation as described above, the reaction mixture was diluted from 60 to 200 µl with nonradioactive ATP (final concentration 1 mM) in order to prevent rephosphorylation with radioactive ATP and the ligands to be tested (5 mM ADP, 10 mM K+ (46)). The mixture was further incubated for 5 or 10 s at 0 °C. Thereafter, the reaction was stopped as described above, and the residual phosphorylation level was determined.

Hydroxylamine Sensitivity of the Phosphorylated Intermediate—After ATP phosphorylation, the acid-denaturated membranes present on the membrane filters were washed with 0.5 mM imidazole-HCl (pH 7.5). After exposure of the filter to either ice-cold 0.5 M hydroxylamine-imidazole (pH 7.5) or 0.5 M imidazole-HCl (pH 7.5) for 10 min, the membranes were washed with 5% trichloroacetic acid in 0.1 M phosphoric acid and analyzed by liquid scintillation analysis.

Analysis of Data—The Ip values for K+ and SCH 28080 were iteratively determined by fitting the concentration relationship to the logistic equation Y = A + (B – A) / (1 + (X/Xo)α), where A represents the lower plateau, B is the top plateau, C is Ip, and D is the Hill coefficient; the values of X and C were entered as the logarithm of concentration) using the nonlinear regression computer program InPlot (GraphPAD Software for Science, San Diego, CA). All data are presented as mean values with standard error of the mean, and differences of average were tested for significance by means of Student’s t test.

Chemicals—[γ-32P]ATP (3000 Ci/mmol, 1, Amersham, Buckinghamshire, UK) was diluted with nonradioactive Tris-ATP (pH 6.0) to a specific radioactivity of 20–100 Ci/mmol. SCH 28080, kindly provided by Drs. J. Forte (Chugai Pharmaceuticals, Tokyo, Japan) and M. Pfeuffer (Schering, Berlin, Germany), was dissolved in ethanol and diluted to its final concentration of 0.1 mM in 0.2% ethanol. The antibodies 2G11, HKB, and HKAc2 were gifts of Drs. J. Forte (University of California, Berkeley), M. Caplan (Yale University), and A. Smolka (University of South Carolina), respectively.

RESULTS

Six baculoviruses were produced, each of which contains coding sequences for the β-subunit as well as for a mutated α-subunit of gastric H+,K+-ATPase. In each of the mutants, one of the negatively charged carboxyl residues located in or around the fifth and sixth transmembrane segments of the α-subunit had been converted into an acid amide residue. These viruses were used to infect Sf9 cells. Fig. 2 shows that the α-subunit present in the membrane fractions of Sf9 cells infected with these mutated viruses has the same apparent molecular mass as the α-subunit of the enzyme of pig gastric mucosa (47). The antibody used to detect the α-subunit on the
Site-directed Mutagenesis of Gastric H⁺,K⁺-ATPase

The amount of phosphorylated intermediate was measured both after preincubation with 0.1 mM SCH 28090 and in its absence and was expressed as pmol of E-P/mg of protein. In the presence of SCH 28080, the amount of phosphorylated intermediate was in most cases not different between the mutants and the wild type enzyme (Table I; only with mutant E820Q this background phosphorylation was significantly higher than with the wild type enzyme (see below). The difference between the ATP phosphorylation level in the absence of SCH 28080 and in its presence was taken as the H⁺,K⁺-ATPase-specific phosphorylated intermediate. Table I shows that in the four mutants in which no 100-kDa phosphoprotein could be seen in the SDS gels (Fig. 5), the amount of specific phosphorylated intermediate was not significantly different from zero. The amount of phosphorylated intermediate of the two mutants of which a 100-kDa phosphoprotein could be visualized on the SDS gel (E795Q and E820Q) indeed showed a significant phosphorylation capacity.

The amount of H⁺,K⁺-ATPase protein expressed was measured using an enzyme-linked immunosorbent assay (40), based on specific binding of the produced protein to the monoclonal antibody 5B6 (41), which recognizes an epitope in the intracellular loop between the fourth and fifth transmembrane domain of the catalytic subunit of pig gastric H⁺,K⁺-ATPase (42). By comparing the amount of immunoreactive protein with that of the pig enzyme, the amount of baculovirus-produced (mutated) H⁺,K⁺-ATPase could be quantitated. Table I shows that the amount of immunoreactive protein in the particulate membrane fractions varied from 0.84 to 4.13% of the total amount of protein. Although no explanation can be given for this variation, the table clearly shows that all viruses produced a considerable amount of mutated H⁺,K⁺-ATPase protein, indicating that the lack of activity of the mutants D824N, E834Q, E837Q, and D839N is not due to lack of biosynthesis. The amount of phosphorylated intermediate produced by the mutants E795Q and E820Q either expressed per mg of protein or per mg of H⁺,K⁺-ATPase was significantly reduced compared with the wild type enzyme.

In a series of experiments, the incubation conditions for the wild type virus have been optimized. The period of infection and the multiplicity of infection were varied, and in these experiments both the amount of SCH 28080-sensitive phosphorylated intermediate and the amount of immunoreactive H⁺,K⁺-ATPase produced by the insect cells were determined. Fig. 6 shows that for the wild type enzyme there is a positive correlation between these two parameters, although individual experiments do deviate considerably from the calculated regression line. From this kind of experiment the conditions used in the present study (3 days of infection and a multiplicity of infection of 3) were chosen. The figure clearly shows that in addition to the mutants D824N, E834Q, E837Q, and D839N, which have no measurable phosphorylation capacity, mutant E795Q has a relatively low phosphorylation capacity, compared with its expression level. The phosphorylation capacity of mutant E820Q was slightly lower than that of the wild type enzyme.

The two mutants E795Q and E820Q, which show the presence of a SCH 28080-sensitive phosphorylated intermediate, have been studied in more detail. The membranes containing the (mutated) enzyme were preincubated for 60 min at 0 °C (pH 6.0) with varying concentrations of either K⁺ or SCH 28080. Fig. 7A shows that the K⁺ sensitivity of mutant E795Q (I₅₀ = 0.45 ± 0.10 mM; n = 3), is similar to that of the wild type enzyme (I₅₀ = 0.38 ± 0.04 mM; n = 5). The I₅₀ value of mutant E820Q, however, is 10 times higher (I₅₀ = 4.5 ± 1.2 mM; n = 4), and complete inhibition is not reached at 30 mM. Some reduction was also found with either 100 mM Na⁺ or choline⁺ (not shown), indicating that the inhibitory effect of high [K⁺] on the level of the phosphorylated intermediate of E820Q might even be in part nonspecific. This mutant has additionally a 100 times lower sensitivity toward SCH 28080 (Fig. 7B). An I₅₀ value of 1.7 ± 0.6 μM (n = 4) was found, whereas the wild type enzyme has an I₅₀ value of 14 ± 3 nM (n = 4) and mutant E795Q has an I₅₀ value of 8 ± 3 nM (n = 4).

The effects of K⁺, SCH 28080, ouabain, and vanadate, if present during the preincubation step, on the ATP phosphorylation level are also reflected in the autoradiograms of the SDS-PAGE gels (Fig. 4, middle and lower panels) of these two mutants. The figure clearly shows that these reagents have

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**Fig. 5. Autoradiogram of SDS-polyacrylamide gel of the ATP-phosphorylated Sf9 membranes infected with mutated recombinant baculoviruses.** Wild type enzyme and the H⁺,K⁺-ATPase mutants E795Q, E820Q, D824N, E834Q, E837Q, and D839N were phosphorylated at 0 °C with 0.1 mM [γ⁻³²P]ATP in the presence of 1 mM MgCl₂ and 20 mM Tris-acetic acid (pH 6.0). The acid-quinched samples were solubilized and subjected to SDS-PAGE at pH 6.5 as described under "Experimental Procedures." Uninfected Sf9 membranes as well as gastric H⁺,K⁺-ATPase (pig) were used as control.

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**Table I**

*Properties of Sf9 membranes after infection with recombinant baculovirus*

<table>
<thead>
<tr>
<th>Mutant</th>
<th>Uninfected</th>
<th>Wild type enzyme</th>
<th>E-P</th>
<th>H⁺,K⁺-ATPase</th>
<th>E-P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>pmol/mg protein</td>
<td>pmol/mg protein</td>
<td>% of pig enzyme</td>
<td>pmol/mg H⁺,K⁺-ATPase</td>
<td></td>
</tr>
<tr>
<td>Uninfected</td>
<td>0.87 ± 0.18</td>
<td>-0.03 ± 0.04</td>
<td>0.00 ± 0.00</td>
<td>1.97 ± 0.40</td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>0.69 ± 0.09</td>
<td>1.15 ± 0.11</td>
<td>1.50 ± 0.17</td>
<td>78.1 ± 10.2</td>
<td></td>
</tr>
<tr>
<td>E795Q</td>
<td>0.76 ± 0.16</td>
<td>0.65 ± 0.12**</td>
<td>4.13 ± 0.86***</td>
<td>19.7 ± 6.0</td>
<td></td>
</tr>
<tr>
<td>E820Q</td>
<td>1.05 ± 0.15**</td>
<td>0.52 ± 0.04***</td>
<td>1.86 ± 0.45</td>
<td>32.2 ± 7.1</td>
<td></td>
</tr>
<tr>
<td>D824N</td>
<td>0.77 ± 0.16</td>
<td>0.04 ± 0.02***</td>
<td>3.13 ± 0.82***</td>
<td>2.4 ± 1.3</td>
<td></td>
</tr>
<tr>
<td>E834Q</td>
<td>0.76 ± 0.17</td>
<td>0.02 ± 0.03***</td>
<td>1.36 ± 0.23</td>
<td>-4.5 ± 4.0</td>
<td></td>
</tr>
<tr>
<td>E837Q</td>
<td>0.53 ± 0.13</td>
<td>-0.03 ± 0.05***</td>
<td>0.84 ± 0.28**</td>
<td>-9.3 ± 8.6</td>
<td></td>
</tr>
<tr>
<td>D839N</td>
<td>0.67 ± 0.08</td>
<td>0.07 ± 0.04***</td>
<td>1.97 ± 0.40</td>
<td>4.3 ± 1.8</td>
<td></td>
</tr>
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</table>
Surprisingly, the ADP, suggesting that this mutation did not lead to a blockade is responsible for the band of 140 kDa and is also present in SCH 28080-insensitive phosphorylation, which in the SDS gel is responsible for the band of 140 kDa and is also present in uninfected cells, also decreased with time. The rate of dephosphorylation of this SCH 28080-insensitive phosphoprotein was increased by ADP but not by the presence of K⁺ (not shown).

In order to test whether the phosphorylated intermediate of the wild type enzyme and of the mutants E795Q and E820Q was an acylphosphate, the intermediate was treated with hydroxylamine, which converts the acylphosphate into a hydroxy-acylphosphate too. However, a further identification of the nature of this phosphorylated product (not shown) both SERCA-type phosphorylation of this SCH 28080-insensitive phosphoprotein was increased by ADP but not by the presence of K⁺ (not shown).
Fig. 9. Effect of hydroxylamine on the residual amount of the phosphorylated intermediate of H⁺,K⁺-ATPase obtained from Sf9 membranes infected with recombinant baculoviruses. Membranes obtained from Sf9 cells either uninfected (triangles) or infected with wild type virus (circles), mutant E795Q (diamonds), or mutant E820Q (squares) were phosphorylated at 0 °C with 0.1 μM [γ-32P]ATP in the presence of 1 mM MgCl₂ and 20 mM Tris-acetic acid (pH 6.0). The phosphorylated membranes were treated at pH 7.5 with hydroxylamine (closed symbols) or imidazole (open symbols) as described under "Experimental Procedures," and the residual phosphorylation level (E·P) was expressed as a percentage of the control and plotted as a function of time.

Ca-ATPases and Na⁺,K⁺-ATPase can be excluded as candidates for this 140-kDa protein.

Phosphorylation and dephosphorylation are key steps in the catalytic cycle of H⁺,K⁺-ATPase. Fig. 10 shows the K⁺ dependence of the overall ATPase activity of membranes of uninfected Sf9 cells and cells infected with viruses expressing the wild type enzyme and the mutants E795Q and E820Q. In this assay a relatively low ATP concentration (10 μM) had to be used in order to obtain significant stimulation by K⁺. The wild type enzyme showed a biphasic activation curve with a maximum at 1 mM K⁺. This activation could be completely blocked by 100 μM SCH 28080 (not shown). A similar biphasic activation curve was found with the pig enzyme using a comparable low (5 versus 10 μM) ATP concentration (45). The E820Q mutant showed, like the uninfected cells, no K⁺-dependent ATPase activity. Low K⁺ activated the ATPase activity of mutant E795Q, although the maximal level reached was less than that of the wild type enzyme, as is also the case for the phosphorylation capacity of this mutant (Fig. 6). At high K⁺, however, less inhibition was found with this mutant than with the wild type enzyme. The maximal ATPase activity with the wild type enzyme was only 60% above the background activity, whereas the steady-state phosphorylation level was 170% above the control level (Table I). The relatively minor increase in the ATPase assay is due to the high basal ATPase activity of the membranes of Sf9 cells. It is tempting to speculate that the high basal ATPase activity is related to the relatively high dephosphorylation rate of uninfected cells in the absence of K⁺ (Fig. 9).

**DISCUSSION**

In the present study we converted six negatively charged amino acid residues present in the M5-M6 region of the α-subunit of gastric H⁺,K⁺-ATPase in their acid amide counterpart as a first approach to establish the importance of these glutamate and aspartate residues for the function of this transport enzyme. The studies were carried out in the baculovirus system, in which we were able to express the enzyme functionally, by constructing viruses with the (mutated) α-subunit and the β-subunit behind two different promoters (32, 37). As a main functional parameter, we measured the presence of a phosphorylated intermediate both qualitatively by autoradiography of the 100-kDa phosphorylated intermediate and quantitatively by measuring the SCH 28080-sensitive ATP-phosphorylation level. We also determined the biosynthesis of immunoreactive α-subunit using a specific enzyme-linked immunosorbent assay (40). Although the amount of immunoreactive α-subunit varied considerably from experiment to experiment, we have no indication that proteolytic breakdown of one of the mutants was enhanced. Moreover, we have no indication of routing problems with any of the mutants.

None of the mutants E834Q, E837Q, and D839N showed any ATP phosphorylation capacity, suggesting that each of these residues is essential for the enzyme to become phosphorylated. It might be that these residues are involved in H⁺ binding, which is essential for ATP phosphorylation. Similar residues in other P-type ATPases have met less attention until now, due to the fact that in nearly all models these residues are located in the intracellular loop between M6 and M7 and not in transmembrane segments (see Fig. 1). Further studies on the precise location of these residues and mutational studies in other P-type ATPases are needed to reach more definite conclusions.

The H⁺,K⁺-ATPase mutant D824N was also not active in terms of phosphorylation capacity by ATP. This Asp residue is completely conserved in Na⁺,K⁺-ATPase and Ca²⁺-ATPases from both sarcoplasmic reticulum and plasma membrane. Mutation of the similar Asp residue in either Na⁺,K⁺-ATPase (26, 28) or plasma membrane Ca²⁺-ATPase (25) did not result in active enzyme either. In SERCA1a Ca²⁺-ATPase, the similar mutant D800N did not show Ca²⁺ occlusion or Ca²⁺-induced phosphorylation by ATP (52). Thus, it is possible that this residue is involved in the binding of cations by all P-type ATPases, and thus amino acid substitutions affect phosphorylation from ATP.

Mutation of Glu⁷⁰⁵ into Gln results in formation of a phos-
phosphorylated intermediate with apparently normal behavior toward K* and SCH 28080. The amount of this intermediate formed is smaller than that for the wild type enzyme, as might be expected from the measurement of immunoreactive α-subunit. This might be due to the production of more inactive α-subunit. Mutational studies of the similar residue in SERCA Ca2+-ATPase and Na+,K+-ATPase suggest a more important role of this residue in these two P-type ATPases. The obtained results, however, markedly depend on the type of the amino acid residue chosen to replace the glutamate present in these two enzymes and possibly on the expression system used. Replacement of Glu779 into Asp or Leu in Na+,K*-ATPase expressed in HeLa cells did not result in active enzyme, whereas mutation of this Glu into Gln or Ala gave an active enzyme (27). Mutation of the same Glu residue into an Asp, using the baculovirus system, resulted in an active enzyme with only reduced cation affinity (29). Replacement of Glu779 by a Lys even resulted in an increase in cation affinity in the latter system. Moreover, Glu779 in Na+,K*-ATPase is the target for the carboxyl-specific reagent 4-(diazomethyl)-7-(diethylamino)-coumarin (53, 54), which inactivates the enzyme in a cation-protective way.

In SERCA Ca2+-ATPase, mutation of the corresponding residue, Glu779, into Gln results in inhibition of Ca2+ transport (53) and Ca2+ occlusion (55). Phosphorylation from ATP at 2.5 mM Ca2+ and Ca2+-induced inhibition of phosphorylation by inorganic phosphate still occurs (22). The dephosphorylation reaction of the ADP-insensitive intermediate was blocked in this mutant. Mutation of this Glu residue by either Gly or Ala resulted in similar effects. Replacement of Glu by Lys resulted in a mutant in which Ca2+ had no effect on phosphorylation from either ATP or inorganic phosphate (56). Moreover the dephosphorylation step was not inhibited by Ca2+ in this mutant. These experiments led Andersen to the suggestion that Glu779 might participate in countercountransport of two protons/Ca2+-ATPase cycle (56).

The most interesting mutant made in the present study is E820Q. This mutant yields a phosphorylated intermediate from ATP, but preincubation with either K* or SCH 28080 had, in contrast to the wild type enzyme, hardly any effect on the steady-state ATP phosphorylation level. The hydrolysis of this phosphorylated intermediate was insensitive to both ADP and K*, and no K*-activated ATPase activity could be detected in this mutant. In the pig enzyme, K* lowers the steady-state phosphorylation level both by shifting the E2 equilibrium to the right, which is assumed to occur through a cytosolic K* site. This is ex­plained from the measurement of immunoreactive α-sub­unit. Mutation of the same Glu residue into an Asp, using the baculovirus system, resulted in an active enzyme in a cation-protective way. There are indications that the similar residue in other P-type ATPases the amino acids present on the site similar to Glu820 are involved in cation binding.

In summary, the present study emphasizes the importance of Glu820 for coupling between ATP phosphorylation and K* transport in gastric H+,K*-ATPase. Further studies with this enzyme are necessary to understand the structural basis for the specificity of and the kinetic differences between the various P-type ATPases.

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REFERENCES