Selective replication of ribosomal DNA repeats after loss of the abnormal oocyte phenotype in *Drosophila melanogaster*

(Southern blotting technique/nontranscribed spacer)

F. Graziani*, L. Vicari*, E. Boncinielli*, C. Malva†, A. Manzi*, and C. Mariani‡*

*Istituto Internazionale di Genetica e Biofisica, Consiglio Nazionale delle Ricerche, and ‡Istituto di Biologia e Genetica, Università di Napoli, Naples, Italy

Communicated by M. S. Meselson, July 17, 1981

**Abstract**

*Drosophila melanogaster* females homozygous for the abnormal oocyte mutation produce a large excess of female offspring when crossed with *XY/0* males. After several generations in *abo* homozygous stock, this maternal effect is no longer observed. The disappearance of the *abo* phenotype is coupled with an increase in the amount of DNA coding for rRNA (rDNA). We have used restriction endonuclease analysis of total DNA extracted from adult females and from single female larval brains to investigate the molecular organization of rDNA before and after the loss of the *abo* phenotype. The rDNA increase is associated with variations of the restriction pattern of the nontranscribed spacer, probably due to a selective increase of rDNA repeats.

---

**Materials and Methods**

*Drosophila* Stocks. *XY¹ Y² y w f 1/0* and *Cy* was obtained in the University of Chicago collection and carries an extra *Y* chromosome. Wild-type Canton S with the second chromosome *abo/ Cy* was isolated by M. S. Meselson. This fragment had been obtained after HindIII/Hae III digestion of plasmid DM256 and is constituted by the 4.0-kilobase (kb) entire nontranscribed spacer and a region of 900 base pairs that is transcribed into the external spacer (11). Cloned DNA was prepared as described in David et al. (12). Nick-translations were performed according to Rigby et al. (13). Restriction endonuclease digests were performed according to the manufacturer’s instructions (Bethesda Research Laboratories, Rockville, MD), New England Biolabs, and Miles. Some HindIII digests were performed with enzyme kindly supplied by R. Di Lauro. DNA fragments were separated by gel electrophoresis on 0.8% agarose gel in Tris/borate/EDTA buffer with HindIII-digested DNA as marker. Blotting was according to Southern (14) on Schleicher & Schuell nitrocellulose filters. Preincubation and hybridization were according to Endow and Glover (15). Filters were exposed on flash-activated autoradiographic film (Kodak 5R), using an intensifying screen.

**Results**

Homozygous *abo* (first generation, *G₀*) and heterozygous females were crossed to *XY¹/0* males. Table I presents the results of these crosses. As the data illustrate, *abo* is a recessive allele. We performed the same crosses with *abo/abo* homozygous females after 19 generations in homozygous condition (*G₁₉*). In this last cross a practically normal sex ratio—that is, the loss of the *abo* phenotype—was observed.

---

**References**

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U. S. C. §1734 solely to indicate this fact.

Abbreviations: rDNA, structural cistrons for rRNA; kb, kilobase(s).

†Present address: Dept. of Genetics, Katholieke Universiteit, Nijmegen, The Netherlands.
Table 1. rDNA content of X/X females with the indicated chromosome 2 constitution, expressing or not the abo phenotype when crossed with XY/Y<sup>0</sup> w//0 males

<table>
<thead>
<tr>
<th>Genotype*</th>
<th>Phenotype</th>
<th>Sex ratio in the progeny, δ/β</th>
<th>% rDNA</th>
</tr>
</thead>
<tbody>
<tr>
<td>abo/Cy</td>
<td>Wild type</td>
<td>4587/4312 = 1.06</td>
<td>0.286 ± 0.023</td>
</tr>
<tr>
<td>abo/abo G&lt;sub&gt;0&lt;/sub&gt;</td>
<td>Mutant</td>
<td>591/4328 = 0.09</td>
<td>0.290 ± 0.024</td>
</tr>
<tr>
<td>abo/Cy</td>
<td>Wild type</td>
<td>2981/3428 = 0.89</td>
<td>0.658 ± 0.038</td>
</tr>
<tr>
<td>abo/abo G&lt;sub&gt;0&lt;/sub&gt;</td>
<td>Mutant</td>
<td>3150/3206 = 0.98</td>
<td>0.287 ± 0.018</td>
</tr>
</tbody>
</table>

The last three hybridizations were performed with DNA extracted from females with an equal number of generations in the respective stocks. rDNA is expressed as percent of total DNA, mean ± SEM.

Table 1 also illustrates the rDNA content measured by hybridization of the labeled rRNA to DNA. DNA was extracted from adult females of the following genotype: abo/Cy heterozygous, abo/abo G<sub>0</sub> homozygous females, and abo/abo G<sub>19</sub> females that no longer showed the abo phenotype.

abo/Cy and abo/abo G<sub>0</sub> females have 0.39% rDNA. As expected, because abo is recessive, after these additional 19 generations in heterozygous condition, the rDNA content remains unchanged (Table 1; compare lines 1 and 2 with 4 and 5). On the contrary, a 2-fold increase in rDNA per X chromosome (0.65% rDNA) is observed in adult G<sub>19</sub> abo/abo females that had a normal sex ratio when crossed to X0 males. This last value, even though lower than previously reported (4, 5), confirms the increase of rDNA after the loss of the abo phenotype.

To investigate the molecular organization of the rDNA, total DNA extracted from abo/Cy adult females and from G<sub>0</sub> abo/abo females was analyzed by means of double digestion with HindIII and Hae III restriction endonucleases. Fragments were separated on agarose gels, then blotted and hybridized with labeled probes from cloned nontranscribed spacer. Results (Fig. 1, lanes a and b) show that there is a wide length heterogeneity of nontranscribed spacers of abo/Cy and abo/abo G<sub>0</sub> females, ranging between 15 and 2.5 kb, with discrete length classes, 4.5 kb being the predominant class; however the two types of female have the same pattern of length heterogeneity. Furthermore, the rDNA extracted from single female larval brain has exactly the same length pattern as the rDNA from the stock, thus indicating that the observed pattern is not the average of different individual patterns (Fig. 2, lanes a-e).

The restriction pattern of total DNA extracted from adult G<sub>19</sub> females is shown in Fig. 1, lane c. The lengths of nontranscribed spacers of these females range between 18 and 2.4 kb. In addition to the predominant class of 4.5 kb present in the original stock, new bands are observed. The most evident band has a length of 3.8 kb, but new bands of 18, 4.5, 3.2, and 2.8 kb, and smaller, are detectable. Experiments performed on DNA extracted from individual larval brains of the same abo/abo G<sub>19</sub> females shows (Fig. 2, lanes f-k) that in this case also the patterns of single fly brains correspond to those observed for DNA extracted from population samples.

The presence of classes of nontranscribed spacers overrepresented in the G<sub>19</sub> phenotypically reverted females suggests that differential replication of the rDNA genes occurs during this process and that, in the ribosomal gene cluster, only some classes of rDNA repeats are augmented with respect to the others.

**DISCUSSION**

The main purpose of our study was to analyze the molecular organization of ribosomal genes before and after the loss of the abo phenotype by means of restriction analysis, using the nontranscribed spacer as a molecular marker of rDNA organization.

The results indicate that the patterns of nontranscribed spacers in heterozygous abo/Cy females and in first generation abo/abo females are identical. On the contrary, new fragments, especially one at 3.8 kb, are found in HindIII/Hae III double digests of DNA extracted from abo/abo females after 19 generations, when the abo phenotype is no longer observed. The same bands are also found in brains of single female larvae.

It is difficult to explain this appearance of a new pattern. The abo/abo females used to "found" the homozygous stock had the same pattern as that of the heterozygous females, and no variations have been found among individuals of the stocks. These observations rule out the possibility of strong selective pressure in favor of a particular rDNA organization. The results obtained with DNA extracted from the diploid and near-diploid tissue of single female larval brains indicate that the new pattern is not the average of different individual patterns and, in addition, that this is a general phenomenon rather than a particular event occurring in tissues with polytene chromosomes. Our data suggest that the new pattern may be a result of a selective increase of particular ribosomal genetic units present in undetectable amounts in first generation abo/abo females.

Other examples of selective amplification of particular rDNA repeats are known, even though only at a somatic level. In recent studies it was demonstrated that during polytenization (17) the genes from only one nucleolus organizer are replicated and that, among these genes, some repeats are preferentially replicated. This last observation implies that the entire ribosomal block is not uniformly replicated in polytene cells, suggesting that polytenization of rDNA may occur by an extrachromosomal mechanism.
Because it appears that the selective rDNA increase observed during the reversion of \textit{abo/abo} homozygous females is not necessary (18) for the suppression of the \textit{abo} phenotype, this phenomenon could be concomitant with other more important events occurring in the heterochromatic region of the \textit{X} chromosome.