Aspergillus Cell Wall Chitin Induces Anti- and Proinflammatory Cytokines in Human PBMCs via the Fc-γ Receptor/Syk/PI3K Pathway


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ABSTRACT Chitin is an important cell wall component of Aspergillus fumigatus conidia, of which hundreds are inhaled on a daily basis. Previous studies have shown that chitin has both anti- and proinflammatory properties; however the exact mechanisms determining the inflammatory signature of chitin are poorly understood, especially in human immune cells. Human peripheral blood mononuclear cells were isolated from healthy volunteers and stimulated with chitin from Aspergillus fumigatus. Transcription and production of the proinflammatory cytokine interleukin-1β (IL-1β) and the anti-inflammatory cytokine IL-1 receptor antagonist (IL-1Ra) were measured from the cell culture supernatant by quantitative PCR (qPCR) or enzyme-linked immunosorbent assay (ELISA), respectively. Chitin induced an anti-inflammatory signature characterized by the production of IL-1Ra in the presence of human serum, which was abrogated in immunoglobulin-depleted serum. Fc-γ-receptor-dependent recognition and phagocytosis of IgG-opsonized chitin was identified as a novel IL-1Ra-inducing mechanism by chitin. IL-1Ra production induced by chitin was dependent on Syk kinase and phosphatidylinositol 3-kinase (PI3K) activation. In contrast, costimulation of chitin with the pattern recognition receptor (PRR) ligands lipopolysaccharide, Pam3Cys, or muramyl dipeptide, but not β-glucan, had synergistic effects on the induction of proinflammatory cytokines by human peripheral blood mononuclear cells (PBMCs). In conclusion, chitin can have both pro- and anti-inflammatory properties, depending on the presence of pathogen-associated molecular patterns and immunoglobulins, thus explaining the various inflammatory signatures reported for chitin.

IMPORTANCE Invasive aspergillosis and allergic aspergillosis are increasing health care problems. Patients get infected by inhalation of the airborne spores of Aspergillus fumigatus. A profound knowledge of how Aspergillus and its cell wall components are recognized by the host cell and which type of immune response it induces is necessary to develop target-specific treatment options with less severe side effects than the treatment options to date. There is controversy in the literature about the receptor for chitin in human cells. We identified the Fc-γ receptor and Syk/PI3K pathway via which chitin can induce anti-inflammatory immune responses by inducing IL-1 receptor antagonist in the presence of human immunoglobulins but also proinflammatory responses in the presence of bacterial components. This explains why Aspergillus does not induce strong inflammation just by inhalation and rather fulfills an immune-dampening function. While in a lung coinfected with bacteria, Aspergillus augments immune responses by shifting toward a proinflammatory reaction.
associated molecular patterns (PAMPs), which are recognized by pattern recognition receptors (PRRs) to induce an innate immune response with consequent induction and shaping of an adaptive immune response (7). While \( (1,3) \)-glucan is known as the ligand for dectin-1 (8), recent studies propose different identities of the chitin recognition receptor and the induced immune response. Schlosser et al. identified FIBCD1 as a high-affinity chitin binding receptor of the intestine that controls immune responses against ingested parasites and fungi (9). Additionally, chitin was described to have proinflammatory properties by inducing IL-17 via the Toll-like receptor 2 (TLR2) pathway (10) in murine cells, and a recent study suggested recognition of chitin via the mannose receptor (MR) on the surface of the phagocytes and by the nucleotide-binding oligomerization domain 2 (NOD2) and TLR9 receptors in the cytoplasm, leading to the induction of the anti-inflammatory cytokine IL-10 in mouse macrophages (11). No pattern recognition receptor or signaling pathway triggered by chitin has been identified in human immune cells to date. In the present study, we aimed to elucidate the immunological properties of chitin isolated from the \( \text{A. fumigatus} \) cell wall and its receptor in human peripheral blood mononuclear cells (PBMCs) and the mechanism and circumstances under which chitin can switch between anti-inflammatory immune responses toward proinflammatory responses.

**RESULTS**

**Aspergillus fumigatus** cell wall chitin produces IL-1Ra in the presence of serum. The chitin samples purified from \( \text{A. fumigatus} \) cell wall contained 70% of the particles showing sizes of \(<0.5 \mu \text{m}\) (Fig. 1) and a degree of acetylation of \(~91\%\) (see Fig. S1 in the supplemental material). Purity of the chitin was ensured as follows. First, the 3% of \( (1,3) \)-glucan still remaining bound to chitin after the final chemical extraction was removed by an endo-\( (1,3) \)-glucanase treatment (see Materials and Methods). Second, the absence of endotoxin contaminations was ensured by the pretreatment of chitin suspension with polymyxin B. In order to identify immunological function of chitin, human PBMCs were stimulated \textit{in vitro} with chitin in the presence or absence of human serum. The proinflammatory cytokines IL-1\( \beta \), tumor necrosis factor alpha (TNF-\( \alpha \)), and IL-6 (Fig. 2A) were not detectable in the cell culture supernatant by enzyme-linked immunosorbent assay (ELISA) in the presence or absence of chitin. In addition, the anti-inflammatory cytokine IL-10 was not induced by chitin, neither on a protein level (Fig. 2A) nor on a transcription level, while lipopolysaccharide (LPS) and LPS with chitin induced high transcription of IL-10 (Fig. 2B). Although the level of IL-8 was high in the presence of chitin and serum, serum without chitin also induced a high level of IL-8, indicating that this high level of IL-8 is not due to the presence of chitin (Fig. 2C). However, chitin induced significant amounts of IL-1 receptor antagonist (IL-1Ra) compared to the medium control (Fig. 3A), in a dose-dependent manner (Fig. 3B). A kinetic analysis of IL-1Ra mRNA expression...
showed that IL-1Ra transcription peaked after 4 h of exposure of cells to chitin (Fig. 3C). The absence of transcription of IL-1β confirmed the lack of detection of this cytokine in the culture supernatant (Fig. 2A). In conclusion, serum was required for the induction of IL-1Ra transcription and release by PBMCs upon chitin stimulation.

Dectin-1, TLR2, TLR4, MR, and NOD2 are not involved in chitin-induced IL-1Ra induction. Subsequently, we aimed to identify the receptor that recognizes chitin that mediates IL-1Ra production by stimulation of PBMCs with chitin. Blocking dectin-1 with GE2 antibody did not affect chitin-induced IL-1Ra production (Fig. 4A). In addition, blocking TLR2 or TLR4 also did not influence IL-1Ra production by chitin (Fig. 4B and C). Since mannose receptor (MR) and NOD2 have been described as receptors for Candida albicans-derived chitin (11), we stimulated PBMCs with chitin and human serum in the presence of an MR neutralizing antibody as well as stimulated PBMCs of NOD2-deficient patients with chitin and the NOD2 ligand N-acetylmuramyl-ananyl-D-isoglutamine (MDP). Blocking MR did not result in significant differences of the production of IL-1Ra (Fig. 4D), and as depicted in Fig. 4E, MDP stimulation did not result in any IL-1Ra production in the NOD-deficient patients, while chitin induced IL-1Ra in equal amounts as in the healthy control. In conclusion, chitin-induced IL-1Ra is dectin-1, TLR2 and -4, MR, and NOD2 independent.

Immunoglobulin opsonization mediates chitin-induced IL-1Ra. Since none of the PRRs known to recognize fungi was identified as the receptor for the recognition of chitin to induce IL-1Ra, while human serum was crucial, we engaged in additional experiments to decipher the roles of different components present in human serum. In the presence of heat-inactivated serum, IL-1Ra production was not abolished but remained similar in concentration as in the presence of serum depleted of all immunoglobulins (Fig. 5E). This result suggested that IgGs are responsible for the induction of IL-1Ra by chitin. To demonstrate this, human immunoglobulins for intravenous (i.v.) administration (IVIGs) were added to the immunoglobulin-free serum, wherein IL-1Ra production could be restored in a dose-dependent manner, indicating that immunoglobulins are required for the induction of IL-1Ra by chitin (Fig. 5F). Fc-γ receptors recognize the Fc part of IgG and thus recognize IgG opsonized pathogens and particles. An Fc-γ receptor II (Fc-γRII) neutralizing antibody significantly reduced chitin-induced IL-1Ra production (Fig. 5G). To further elucidate whether chitin was directly bound and opsonized by IgG, we incubated 10% diluted serum or 1 mg/ml IVIGs in a polystyrene plate coated with chitin and subsequently detected bound IgG with an anti-Fc antibody. A significant amount of IgGs bound to chitin in both samples (Fig. 5H). To further elucidate whether the IL-1Ra induction was caused by immunoglobulins binding specifically to chitin, human serum was depleted from chitin-binding immunoglobulins. The efficacy of the pulldown experiment was tested by ELISA, resulting in a reduction of 73.5% of chitin-binding immunoglobulins. This result confirmed opsonization with IgG as the main mechanism for the IL-1Ra induction by chitin.

Chitin-induced IL-1Ra is dependent on actin polymerization and signals via spleen tyrosine kinase (Syk) and PI3K. Having identified opsonization with IgG as the main mechanism for the IL-1Ra induction by chitin, we further deciphered whether phagocytosis and the downstream signaling of Fc-γ receptor are needed for the induction of IL-1Ra by chitin. We stimulated PBMCs with chitin and human serum and blocked either the actin polymerization with cytochalasin D (Fig. 6A) or the downstream-signaling kinases of the Fc-γ receptor spleen tyrosine kinase (Syk) (Fig. 6B) or
PI3K (Fig. 6C) or the combination of Syk and PI3K. All resulted in a significant reduction of IL-1Ra, while IL-1Ra was completely abolished when Syk and PI3K were blocked simultaneously (Fig. 6D). Taken together, these findings indicate that recognition and phagocytosis via the Fc-γ receptor with subsequent Syk kinase and PI3K activation mediates the IL-1Ra induction by chitin. Chitin synergizes with Syk-independent PRR pathways, leading to IL-1β production that is dependent on immunoglobulins, Syk, and PI3K. Since the immune system is probably never confronted in nature with the highly purified chitin alone, but rather in the setting of different PRR ligands of the *A. fumigatus* conidia and hyphal cell wall, we stimulated PBMCs with chitin in the absence and presence of serum in combination with the NOD2, TLR2, TLR4, and dectin-1 ligands. In the absence of serum, no cytokines were induced. All non-Syk-dependent PRR pathways stimulated with ligands, such as MDP (NOD2 pathway), Pam3Cys (TLR2 pathway), and LPS (TLR4 pathway) synergized with chitin to induce higher production of IL-1β and TNF-α, while stimulation with β-glucan (dectin-1/Syk pathway) combined with chitin did not induce IL-1β (Fig. 7A), TNF-α (Fig. 7B), or IL-6 (see Fig. S1 in the supplemental material). In contrast, chitin did not have synergistic effects on the LPS-induced IL-1Ra production; instead, LPS combined with chitin-induced IL-1Ra additively (see Fig. S3 in the supplemental material). Next, we wanted to elucidate whether similar pathways responsible for IL-1Ra induction by chitin were mediating the synergistic response of chitin with other PAMPs. Using heat-inactivated serum, we observed a significantly higher ratio of LPS versus LPS/chitin synergistic induction of IL-1Ra, while this ratio was significantly lower in the presence of Ig-depleted serum. Further heat inactiva-

**FIG 4** Dectin-1, TLR2, TLR4, MR, and NOD2 are not involved in the chitin-induced IL-1Ra. (A to E) Chitin-induced IL-1Ra in culture supernatants of PBMCs of healthy volunteers stimulated in the absence or presence of a neutralizing antibody or molecule like (A) GE2 for blocking dectin-1 (*n* = 5), (B) anti-TLR2 for blocking TLR2 (*n* = 6), (C) *Bartonella* (Bart) LPS for blocking TLR4 (*n* = 8), and (D) anti-MR for blocking the mannose receptor (*n* = 12), or (E) two NOD2-deficient patients. IL-1Ra was measured in the cell culture supernatant by ELISA. Statistical analysis was performed with the Wilcoxon signed-rank test. ns, not significant.
**FIG 5** Complement and immunoglobulin opsonization mediate the chitin-induced IL-1Ra. PBMCs from healthy volunteers were stimulated with RPMI or chitin in the presence of (A) pooled serum (PS) or heat-inactivated pooled serum (HI PS) \((n = 11)\), (B) MBL-deficient serum with and without additional purified MBL \((n = 3)\), or (C) isotype control or CR3 neutralizing antibody \((n = 8)\). (D) PBMCs from a kindlin-3-deficient patient or two controls were stimulated with chitin. (E) PBMCs from healthy volunteers were stimulated with RPMI or chitin in the presence of pooled serum or immunoglobulin-depleted serum (Ig) \((n = 11)\) or (F) Ig-depleted serum supplemented with decreasing concentrations of human immunoglobulins (IVIGs) \((1 \text{ mg/ml, } 100 \mu\text{g/ml, and } 10 \mu\text{g/ml})\) \((n = 6)\). (H and I) Binding of IVIGs to chitin was determined by coating an ELISA plate with chitin and a secondary incubation with (H) 10% pooled serum or 1 mg/ml IVIGs or (I) 10% pooled serum after mock treatment or after pulldown of chitin-binding immunoglobulins, and the OD was measured \((n = 6)\). (G and J) PBMCs

(Continued)
tion of the Ig-depleted serum did not change the ratio (Fig. 7C). In addition, blocking Syk and PI3K in PBMCs reduced the synergistic effect of chitin on LPS stimulation significantly, and the combination of both blockers completely inhibited the effect (Fig. 7D). Collectively, these data suggest synergism induced by chitin is also immunoglobulin/FcR dependent and provide evidence for cross talk between chitin-induced FcR signaling and other PRR pathways, with the exception of dectin-1.

DISCUSSION

No pattern recognition receptor or signaling pathway activated by chitin has been identified in human immune cells to date, and immunostimulatory capacities range from absence of effects to anti-inflammatory and even proinflammatory properties (16). In the present study, we describe a mechanism by which chitin can induce immune responses in human immune cells and identify that chitin has both anti-inflammatory properties by inducing IL-1Ra and proinflammatory effects by inducing IL-1β/TNF in synergy with Syk-independent PRR pathways. Interestingly, none of the previously proposed chitin receptors, such as TLR2, MR, NOD2, or dectin-1, were involved in the induction of IL-1Ra in human PBMCs, but phagocytosis and recognition via the Fc-γRII with subsequent Syk and PI3K signaling of IgG-opsonized chitin was identified as the signaling pathway. Strikingly, the effect was only observed with chitin particles in the presence of immunoglobulins. Therefore, we propose that chitin has a dual immunological function by dampening and controlling immune responses on one hand and orchestrating proinflammatory responses when chitin is presented in combination with PAMPs on the other.

Purified Aspergillus chitin does not induce any proinflammatory cytokines in PBMCs both in the absence and in the presence of human serum. This is in line with former studies by Mora-Montes et al. and Bueter et al. (16, 17), which described chitin as an immunologically inert particle, not inducing TNF-α, IL-1β, IL-6, or IL-10 production (17) or inflammasome activation (18). This was observed in several cell types, such as murine bone marrow-derived macrophages, M1 and M2 macrophages, as well as

Figure Legend Continued

from healthy volunteers were stimulated with RPMI or with chitin in the presence of (G) an anti-Fc-γRII antibody or the isotype control (n = 3) or (J) in the presence of mock-treated pooled serum or with pooled serum depleted from chitin-binding immunoglobulins. (A to F, G, and J) IL-1Ra was measured in the cell culture supernatant by ELISA. Statistical analysis was performed with the Wilcoxon signed-rank test (**, P < 0.01; ***. P < 0.001).
In addition, the use of *Aspergillus* to isolate chitin and the different chitin purification method could account for the differences observed in cytokine responses. The presence of a contaminant even at low dose in the chitin sample can have a significant effect on the response to chitin since we have shown that polysaccharides can have an opposite effect from chitin on the immune response when tested in combination with chitin.

While former studies focused on IL-10 as a potent anti-inflammatory cytokine, less attention has been paid to another potent endogenous anti-inflammatory cytokine, namely, IL-1Ra. In this study, stimulation of PBMCs with chitin in the presence of human serum resulted in IL-1Ra transcription and production. The fine-tuned balance of pro- versus anti-inflammatory cytokines of the IL-1 family is important during systemic inflammation, and several reports point to a crucial role for a balanced IL-1 response to prevent *Aspergillus*-induced pathology (19–21).

Since the MR has previously been reported to be involved in chitin-induced immune responses in mice (11), we investigated mannose-binding lectin (MBL), which shares homology with the MR, as well as blocking the MR on PBMCs. However, MBL and MR were not involved in IL-1Ra induction by chitin. Although CR3 plays a crucial role in mediating adaptive T cell responses against *Aspergillus* conidia (22, 23), this receptor was also not involved in chitin-induced IL-1Ra. Moreover, stimulation of PBMCs of a kindlin-3-deficient patient (defective CR3 signaling [15]) with chitin revealed an intact IL-1Ra pathway. These data suggest that CR3 and MBL are not involved in chitin-induced IL-1Ra production. However, the fact that IL-1Ra induction by chitin was partly dependent on heat inactivation suggests a role for the complement system in the induction of IL-1Ra by chitin. Since Agarwal et al. had shown in an earlier study that chitin is a poor inducer of the alternative complement pathway (24), the role of complement activation seen in this study could be caused by activation of the classical pathway. Elucidation of this should be part of future studies.

In contrast to complement, immunoglobulins were essential for mediating the induction of IL-1Ra induced by chitin. Chitin showed direct interaction with immunoglobulins, and the replenishment of IVIGs in Ig-depleted serum restored IL-1Ra induction. In addition, IL-1Ra was significantly lower when serum was depleted from chitin-binding immunoglobulins, suggesting that the presence of anti-chitin-specific IgGs mediates the IL-1Ra induction of opsonized chitin. Blocking Fc-receptor signaling, subsequently leading to IL-1Ra. Moreover, we identified that IL-1Ra induction of opsonized chitin was dependent on phagocytosis, Syk, and PI3K activation.

Other polysaccharides present in the cell wall of *Aspergillus*, such as galactosaminogalactan and β-glucan, can also induce IL-1Ra (19, 28). However, both polysaccharides do not require human serum for the induction of IL-1Ra. Interestingly the induction of IL-1Ra by β-glucan was shown to be independent of dectin-1 but dependent on PI3K (29). Therefore, it might be that the PI3K-Akt pathway is the final common pathway essential for IL-1Ra induction by polysaccharides, but that this pathway can be induced via different mechanisms, depending on the ligand. Furthermore, it has been shown that insoluble fibrillar polysaccharides can induce potent inflammatory responses by engaging multimerization of pattern recognition receptors (PRRs) and formation of supramolecular PRR complexes, which is known as the fibril hypothesis (30). We observed striking effects when cells

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**FIG 7** Chitin synergizes with PRR ligands not activating Syk, leading to high IL-1β production dependent on immunoglobulins, Syk, and PI3K. PBMCs from healthy volunteers were stimulated with (A and B) chitin, MDP, Pam3Cys, LPS, or β-glucan alone or with the combination of PRR ligands with chitin in the absence or presence of human pooled serum (n = 5 to 18) or (C and D) with LPS and LPS together with chitin, either (C) in the presence of heat-inactivated (HI) pooled serum, Ig-depleted serum (Ig- ), or heat-inactivated Ig-depleted serum (HI Ig- ) (n = 9) or (D) after blocking Syk kinase or PI3K alone or in combination (n = 5 to 7) (C and D). The ratio between LPS-induced IL-1β and chitin plus LPS-induced IL-1β was calculated. (A to D) IL-1β or TNF-α (B) was measured in the cell culture supernatant by ELISA. Statistical analysis was performed with the Wilcoxon signed-rank test (*, P < 0.05; **, P < 0.01; ***, P < 0.001; ns, not significant).
were stimulated with opsonized chitin in the presence of other PRR ligands. Ligands for TLR2, TLR4, and NOD2 mediated potent synergistic effects on proinflammatory cytokines such as IL-1β. Interestingly, the combination of chitin with β-glucan, the ligand for dectin-1, did not induce IL-1β production, not even in the presence of human serum. Recently, cross talk between Fc-γ receptors and other TLRs has been described (26). Vogelpoel et al. observed synergism with TLR2 and TLR4 ligands, but not with dectin-1 ligands, which is in line with the data from our study. Whether this is because Fc-γ receptors and dectin-1 both signal via Syk-PI3K-Akt needs to be further elucidated (31, 32). Since the synergism was dependent on IgGs and PI3K signaling, it is tempting to speculate that independent of a specific receptor, opsonized polysaccharides can induce potent synergistic proinflammatory responses via Fc-γ receptor activation when other proinflammatory ligands are present.

Studying the role of chitin-specific antibodies during aspergillosis might lead to new insights that could be relevant for the clinical setting. Indeed, an increase in antichitin antibodies has been shown in immunocompetent patients with chronic pulmonary aspergillosis and allergic aspergillosis (S. Raj and J.-P. Latgé, unpublished data). The role of the level of antibodies on the immune response toward chitin should be investigated in the future.

Another important question is how the opsonization of chitin with specific antibodies could influence disease severity. Only few data exist; however, Kin et al. have shown that antibodies against bacterial polysaccharides can bind to chitin and dampen the overall immune response to chitin (33). In addition, since we provide evidence that chitin induces a different immune response when tested alone or with other PAMPs and since we know that the different polysaccharides of *A. fumigatus* will activate differently inflammatory responses, it is now of interest to test the effect of combinations of the cell wall polysaccharides to see if these combinations are neutral, antagonistic, or synergistic.

In conclusion, we have identified a novel mechanism through which chitin has strong anti-inflammatory effects and in the setting of other PAMPs can boost proinflammatory responses (Fig. 8). These responses were critically dependent on immunoglobulins, Fc-γ receptor signaling, and phagocytosis. These data might explain why the responses reported by chitin are so diverse. Moreover, since immunoglobulins might also influence other polysaccharide-induced immune responses, future studies need to focus on immunoglobulins and FcR signaling when investigating fungal or fungal-cell wall component-induced immune responses.

**MATERIALS AND METHODS**

**Volunteers and patients.** Blood was collected from healthy volunteers or patients by venous blood puncture. Two NOD2-deficient individuals had the homozygous NOD2 insertion of a cysteine at position 1007 (rs2066847) that results in a frameshift of the coding sequence, which was analyzed as described previously (34). Serum was used from two patients who had a deficiency in mannose-binding lectin (MBL) with serum MBL levels below 0.04 mg/liter. One patient had the autosomal recessive mutation in *FERMT3*, the gene encoding kindlin-3, leading to the leukocyte adhesion deficiency type III (35, 36).

**Ethics statement.** All experiments were performed and conducted in accordance with good clinical practices, the Declaration of Helsinki, and the approval of the Arnhem-Nijmegen Ethical Committee (no. 2010/104). Blood from volunteers and patients was taken after written informed consent was obtained.

**PRR ligands, blockers, and other stimuli.** *E. coli* lipopolysaccharide (LPS [1 ng/ml]) (TLR4 ligand [E. coli serotype O55:B5]) was obtained from Sigma-Aldrich, St. Louis, MO), Pam3Cys (5 μg/ml) (TLR2 ligand) was obtained from EMC Microcollections, Tübingen, Germany, and N-acetylglucosaminyl-ananyl-β-D-glucopyranoside (MDP [5 μg/ml]) (NOD2 ligand) was obtained from Sigma-Aldrich. Fungal cell wall β-(1,3)-glucan (10 μg/ml) (dectin-1 ligand) was kindly provided by David L. Williams (East Tennessee State University, Johnson City, TN). This polysaccharide was isolated from *C. albicans* yeast cells and was verified to be exempt of
proteins. The suspension contained water-insoluble microparticles (1 to 5 μm) (37).

The dectin-1 inhibitor GE2 was a kind gift from Gordon Brown (University of Aberdeen, Scotland). Bartonella quintana LPS was prepared and purified as described elsewhere (38) and used as a TL4 inhibitor (100 ng/ml) (22). Isotype control mouse IgG1 (10 μg/ml) and anti-TLR2 (10 μg/ml) were obtained from eBioscience, Halle-Zoerles, Belgium. Isotype control goat IgG (10 μg/ml), anti-human β2-integrin (anti-CR3 [10 μg/ml]), and isotype control mouse IgG2b (10 μg/ml) were obtained from R&D Systems Minneapolis, MN. Isotype control mouse IgG1κ (10 μg/ml) and anti-human mannose receptor (MR) (anti-human CD206 [10 μg/ml]) were obtained from BioLegend, San Diego, CA. Anti-human CD32 (anti-Fc-γ receptor II [10 μg/ml]) was obtained from StemCell Technologies SARL, Grenoble, France. Wortmannin (100 ng/ml, dissolved in dimethyl sulfoxide [DMSO]) was obtained from Sigma. R406 [10 μl] and anti-human mannose receptor (MR) (anti-human CD206 using 4-hydroxy-benzhydrazide (43). This was repeated until no more supernatant for reducing sugar released by protein) at 37°C overnight followed by centrifugation and checking the degree of acetylation in the chitin preparation was determined by three different methods. (i) The first method was the Cibacron brilliant red G330 solution (1 mg/ml) was prepared by ultrasonication using a Soniprep 150 probe sonicator (MSE, London, United Kingdom). The sonicator was set to 50% power and tuned to 50%, reaching an amplitude of 16 μm for 30 s. The sonication reduced the molecular weight while not affecting the degree of acetylation (45, 46). (iii) The third method was a Fourier transform infrared spectroscopy (FT-IR) method performed using the absorption ratio A1635/A1650 (47, 48). For FT-IR, dry chitin was suspended uniformly between IR windows, and the spectrum (1,400 to 4,000 cm⁻¹) was recorded in a Jasco FT/IR-6100 apparatus, subtracting the spectra of air.

Experiments were performed with four different batches of chitin purified from the A. fumigatus mycelial cell wall, with a final concentration of 10 μg/ml.

To determine the size of the chitin particles, the chitin suspension was subjected to flow cytometry (FC500 flow cytometer [Beckman Coulter]) and compared with reference beads of 0.1 and 0.25 μm from Thermofluidic or 3, 6, and 10 μm (Flow Cytomark Fluorospheres) from Beckman Coulter. The data were analyzed using Kaluza Analysis version 1.3 (Beckman Coulter).

PBMC isolation. Venous blood was drawn in 10 ml EDTA tubes. The blood was diluted 1:1 with phosphate-buffered saline (PBS). Subsequently PBMCs were isolated using Ficoll-Paque (GE Healthcare, Zeist, The Netherlands) density gradient centrifugation. The PBMC layer was collected and washed twice in cold PBS. Cells were reconstituted in RPMI+, consisting of RPMI 1640 culture medium (Dutch modification [Gibco, Invitrogen, Breda, The Netherlands]) supplemented with 10 μg/ml gentamicin, 10 mM l-glutamine, and 10 mM pyruvate (Gibco).

The cells were counted with a particle counter (Beckmann Coulter, Woerden, The Netherlands), and the concentration was adjusted to 1 × 10⁷ cells/ml.

PBMC stimulation. PBMCs were plated in a 96-well plate (Corning, NY) at a final concentration of 2.5 × 10⁶/ml in an end volume of 200 μl per well. Stimulations were performed in the presence of 10% human serum, either not depleted or depleted of all immunoglobulins (BBI Solutions, Cardiff, United Kingdom). Serum was either complement active, if not otherwise indicated, or heat inactivated by incubation for 30 min at 56°C in a water bath according to a commonly used protocol (49). After 1 h of preincubation with inhibitor or medium, stimuli or medium was added. Cells were incubated at 37°C with 5% CO₂, and after 24 h, supernatants were collected and stored at −20°C.

Cytokine measurements. IL-1β, tumor necrosis factor (TNF-α), IL-6, IL-8, IL-10, and IL-1Ra were measured in the cell culture supernatants using commercial ELISA kits (IL-1β, TNF-α, and IL-1Ra from R&D Systems, and IL-6, IL-8, and IL-10 from Sanquin) according to the instructions supplied by the manufacturer.

Pulldown assay of chitin-binding immunoglobulins in human serum. To deplete serum from chitin-binding immunoglobulins, 1 ml of RPMI supplemented with 10% serum containing either 1 mg chitin from a stock concentration of 10 mg/ml or an equal amount of distilled water (for mock treatment) was incubated overnight at 37°C and shaken at 130 rpm. On the next day, the chitin-containing or mock-treated suspension was centrifuged (20 min, 14,000 rpm, room temperature), and the supernatant without beads was recovered. The depletion efficacy was checked by a chitin-binding IgG ELISA.

Chitin-binding IgG ELISA. A polystyrene 96-well plate was coated with 10 μg/ml chitin (100 μl per well) overnight at room temperature. After being washed and blocked with PBS with 1% bovine serum albumin (BSA) for 1 h, wells were incubated with 10% serum in RPMI+, mock-treated serum, or depleted serum for 1 h. IgGs bound to chitin were detected by using an anti-human IgG (whole particle) labeled with peroxidase in a concentration of 1:1,000 (Sigma-Aldrich). The enzymatic reaction was started by adding the TMB ([3,3′,5,5′-tetramethylbenzidine]) and H₂O₂-containing substrate, which was stopped after 10 min by 10% sulfuric acid solution. The optic density was measured by a photometric ELISA reader.

qPCR. RNA was isolated from 1 × 10⁶ PBMCs after stimulation for 4 h and 24 h with Aspergillus chitin in the presence of either medium or 10% human serum using Trizol reagent (Invitrogen) according to a protocol supplied by the manufacturer. RNA (500 ng) was reverse transcribed into cDNA using the iScript cDNA synthesis kit (Hercules, Bio-Rad Laboratories, CA). Quantitative PCR (qPCR) analysis was performed using SYBR green master mix (Applied Biosystems, Carlsbad, CA) and the Applied Biosystems 7300 real-time PCR system. As in the PCR protocol, the following conditions were used: 2 min at 50°C and 10 min at 95°C, followed by 40 cycles at 95°C for 15 s and 60°C for 1 min. For the amplification of human IL-1Ra (hIL-1Ra), the primers 5′ GCCCTCCGACGATCCACTAAC 3′ and 5′ TCCGAGATTCTGAAGGCTTG 3′ were used, and for the amplification of hIL-1β, the primers 5′ GCAACTGTTCCTGAACTCAACT 3′ and 5′ ATCTTTTGCGTCTGAACCTTACT 3′ were used in each of loading concentrations of RNA between the different conditions, qPCR results were corrected with the β2 microglobulin (β2m) housekeeping gene amplified using the primers 5′ ATGATGTAGTCGCGTGTG 3′ and 5′ CCAAATGCGCGGATCTTCAAAC 3′. Primer efficiency was evaluated using a standard curve. IL-1Ra threshold cycle (Ct) values were compared with the β2m Ct, by calculating the ΔCt, and the fold difference in loading concentrations of RNA between the different conditions, qPCR results were corrected with the β2 microglobulin (β2m) housekeeping gene amplified using the primers 5′ ATGATGTAGTCGCGTGTG 3′ and 5′ CCAAATGCGCGGATCTTCAAAC 3′. Primer efficiency was evaluated using a standard curve. IL-1Ra threshold cycle (Ct) values were compared with the β2m Ct, by calculating the ΔCt, and the fold
change was calculated relative to the RPMI-stimulated level to determine the effect of stimulation with.chitin on IL-1α expression.

**Statistical analysis.** The Wilcoxon signed-rank test was used to determine differences between stimulation with and without inhibitors or between different sera. A P value of <0.05 was considered statistically significant as shown by asterisks in the figures (*, P < 0.05; **, P < 0.01; ***$, P < 0.001). Graphs represent cumulative results from all performed experiments and are presented as means ± standard errors of the means (SEM). Data were analyzed with GraphPad Prism v 5.0.

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at http://mbio.asm.orglookup/suppl/doi:10.1128/mBio.01823-15/-/DCSupplemental.

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**REFERENCES**


