Biphasic Effect of MCH on α-MSH Release From the Tilapia (Oreochromis mossambicus) Pituitary

DIET GRÔNEVELD,1 PAUL H. M. BALM AND SJOERD E. WENDELAAR BONGA

Department of Animal Physiology, Faculty of Science, University of Nijmegen, Toernooiveld, 6525 ED Nijmegen, The Netherlands

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GRÔNEVELD, D., P. H. M. BALM AND S. E. WENDELAAR BONGA. Biphasic effect of MCH on α-MSH release from the tilapia (Oreochromis mossambicus) pituitary. PEPTIDES 16(5) 945–949, 1995.—The effect of melanin-concentrating hormone (MCH) on the release of α-melanocyte stimulating hormone (α-MSH) from the tilapia pituitary gland was studied in vitro. In a superfusion set up, 10 nM to 1 pM synthetic salmon MCH caused a concentration-dependent inhibition of α-MSH release from tilapia neurointermediate lobes (NILs). Immunoneutralization of MCH in tilapia NILs further indicated that endogenous MCH has an inhibitory effect on the melanotropes. The release of monoacetylated α-MSH release was more strongly inhibited by MCH than that of des-, and diacetylated α-MSH, indicating that MCH modulates the secretory signal of the melanotropes in a quantitative and qualitative manner. A high concentration of MCH (10 pM) substantially increased the release of α-MSH. Further evidence in support of a stimulatory action of high concentrations of MCH was provided by the observation that the MCH analogue MCH(2–17) at 10 and 35 pM enhanced α-MSH release as well. Therefore, we conclude that the response of pituitary melanotropes to MCH is biphasic, as was reported previously for the effects of MCH on other targets in fish and mammals. Under physiological conditions the inhibitory action of MCH on fish melanotropes most likely dominates.

Requests for reprints should be addressed to Diet Grôneveld.

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The purpose of the present study was to gain more insight into the action of MCH at the pituitary level. We therefore quantitatively and qualitatively analyzed the effects of MCH and MCH(2–17), a MCH analogue lacking the N-terminal amino acid, which was helpful in elucidating the mechanisms of action of MCH on the teleost skin (11,26), on the release of α-MSH in vitro from the tilapia pituitary gland.

METHOD

Animals

Male and female tilapia (Oreochromis mossambicus), average weight 80 g, were bred in the aquarium facility of the Dept. of Animal Physiology of the University of Nijmegen. They were kept in fresh water at 28°C and were fed a commercial dried fish food (Tetramin). The fish were kept in glass aquaria, illuminated by overhead TL tubes, with a day–night rhythm of 12 h light and 12 h darkness. Immediately after removal from the tank, the animals were sacrificed by spinal transection and pituitary glands were dissected from the brain.

Superfusion of Neurointermediate Lobe Tissue

One or two freshly dissected pituitaries or NILs were placed on a nylon gauze in a 10-µl superfusion chamber. A multichannel peristaltic pump (Whatson Marlow) was used to pump carbogen-aerated incubation medium (IM) [142 mM NaCl, 2 mM KCl, 2 mM CaCl₂, 15 mM HEPES (pH 7.38), 0.3 mg/ml bovine serum albumin (Sigma), 2.5 mg/ml glucose] through each chamber at a rate of 30 µl/min. The superfusion chambers as well as the IM were thermostat controlled at 28°C. The effluent from each chamber was collected in fractions over varying periods with an Isco rate of 30 µl/min. The superfusion chambers as well as the IM were thermostat controlled at 28°C. The effluent from each chamber was collected in fractions over varying periods with an Isco (Retriever II model) fraction collector. During at least 3 h superfusion fractions were collected before switching to pulse medium (Retriever II model) fraction collector. During at least 3 h superfusion fractions were collected before switching to pulse medium containing MCH or MCH(2–17) (both synthesized by Dr. T.O. Matsunaga at the lab of Dr. V. J. Hruby and kindly provided by Dr. M. E. Hadley, Tucson, AZ) (32), for a 25–30-min period. After the pulse the superfusion was continued with IM. The collected fractions were stored immediately at −20°C until α-MSH radioimmunoassay.

Immunoneutralization of MCH From Tilapia NILs

NILs from tilapia kept in dark glass aquaria were dissected and individually preincubated in 100 µl IM, containing 5 mg/ml (w/v) glucose, during 1 h at 28°C. Subsequently, the NILs were transferred into 30 µl IM containing either 0.1% (v/v) MCH antiserum (a kind gift of Dr. B.I. Baker and Dr. H. Kawauchi) (28) or 0.1% (v/v) normal rabbit serum. The vials were aerated with carbogen, sealed, and shaken gently at 28°C for a 15-h incubation period. After this static incubation, media were collected and stored at −20°C until α-MSH radioimmunoassay; subsequently, the NILs were superfused as described above.

Reversed-Phase HPLC Analysis

To determine the effect of MCH on the profile of the secretory signal, superfusate was submitted to HPLC analysis. Different forms of α-MSH were separated on a Spherisorb 10 ODS column (Bisschof) as described previously (25). In short, the primary solvent was buffer B (0.5 M formic acid, 0.14 M pyridine, pH 3.0) and elution was accomplished with a gradient of 1-propanol at a flow rate of 2 ml/min. Fractions of 0.6 ml were collected. The fractions were dried in a Speedvac concentrator (Savant), diluted into 0.05 N HCl, 50% methanol, and submitted to α-MSH radioimmunoassay.

α-MSH Radioimmunoassay

The α-MSH radioimmunoassay (RIA) with L9 α-MSH antiserum has been described previously (24). The antiserum is equally sensitive to des-, mono- and diacetyl α-MSH. Cross-reactivity of the α-MSH antiserum with MCH [preliminary results have been reported by De Koning et al. (13)] and MCH(2–17) was assessed (see the Results section).

Processing of Data and Statistics

The results of the superfusions are either expressed as pg α-MSH per min per NIL, or as a percentage of the basal release (=100%). Basal release was defined as the average α-MSH release during the secretagogue pulse were expressed as a percentage of the basal release in 20–30 min prior to addition of secretagogue, and basal release values ranged from 30 to 200 pg/min/NIL. For the concentration–response relationships of MCH- or MCH(2–17)-induced effects, values for maximal inhibition or stimulation of α-MSH release during the secretagogue pulse were expressed as a percentage of the prepulse values. Correction of α-MSH values for cross-reactivity after a 10-µM MCH pulse was performed as follows: proceeding from 0.0032% cross-reactivity (see the Re-
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RESULTS

Quantitative Effects of MCH on α-MSH Release

At concentrations below 1 μM, MCH caused a concentration-dependent inhibition of α-MSH release during in vitro superfusion of tilapia NILs [Fig. 1(A,C)], and at 10 μM α-MSH release was stimulated [Fig. 1(B)]. The effect of MCH on the α-MSH release lasted for the duration of the pulse; after the pulse α-MSH release did restore to basal levels. The MCH concentration inducing 50% of maximum inhibition of α-MSH release (EC50), using the data points up to and including 250 nM, was estimated to be 19 nM with a maximum inhibition of 42%. At 10 μM MCH the α-MSH release was stimulated with a maximum stimulation of 96 ± 17% (p < 0.001) [Fig. 1(C)]. Taking into account the slight cross-reactivity of MCH with the α-MSH antiserum (0.0032%) (Fig. 2), this percentage decreased marginally to 87 ± 22% (p < 0.001 compared with basal release). However, because the dilution curve for MCH only approximately parallelled the α-MSH dilution curve in the α-MSH radioimmunoassay (Fig. 2; the slope of log transformed curves was significantly different, p < 0.01) an additional control was performed. To mimic pulse conditions, MCH was added to prepulse superfusates to a final concentration of 10 μM. This yielded a nonsignificant increase (p = 0.88) of immunoreactive α-MSH of 2 ± 10 pg (and 16 ± 16%; n = 6) compared with untreated prepulse fractions. During the 10 μM MCH pulse the increase of immunoreactive α-MSH was 108 ± 33 pg (95 ± 28%; n = 5), which was significantly higher (p < 0.01) than the increase measured when MCH was added afterwards to superfusates. MCH(2–17), a MCH analogue that, in contrast to MCH, did not cross-react with the α-MSH antiserum (Fig. 2), induced a significant stimulation of the α-MSH release at concentrations of 10 and 35 μM (Fig. 3). The stimulation of α-MSH release by 10 μM MCH(2–17) was comparable with that evoked by 10 μM MCH.

Effect of Endogenous MCH on α-MSH Secretion

After incubation of tilapia NILs with 0.1% MCH antiserum, to immunoneutralize endogenous MCH, α-MSH release significantly increased compared with NILs incubated in medium containing normal rabbit serum [Fig. 4(A)]. After 15 h of static incubation, the antiserum was washed out by superfusion of the NILs with fresh incubation medium. During this superfusion the α-MSH release of the NILs incubated with the MCH antiserum returned to control levels within 2 h [Fig. 4(B)].

Effects of MCH on the Release of α-MSH Isforms

To study the qualitative effect of MCH on α-MSH release, the isoforms of α-MSH released before and during pulses with 100 and 250 nM MCH were analyzed by HPLC. Des-, mono-, and diacetylated α-MSH were released during MCH pulses as well as under control conditions (Fig. 5). During the 100 and 250 nM MCH pulses the amounts of monoacetylated α-MSH decreased relatively more than the amounts of des- and diacetylated α-MSH, when compared to control. Taking the area under the curve of diacetylated α-MSH (di), monoacetylated α-MSH (mono), and desacetylated α-MSH (des), we calculated di/mono and des/mono ratios of 0.45 and 0.38 for control, 0.73 and 0.50 during a 100 nM MCH pulse, and 0.95 and 0.61 during a 250 nM MCH pulse, respectively. The highest increase of the ratios was found at the concentration (250 nM) where the overall inhibitory effect was the strongest (Figs. 1 and 5).

DISCUSSION

The present results demonstrate that the MCH concentration–response curve for α-MSH release from the tilapia NIL is biphasic. At concentrations below 1 μM, synthetic MCH caused a concentration-dependent inhibition of α-MSH release. Concentrations above 1 μM induced a concentration-independent release of α-MSH.

FIG. 3. Stimulatory effect of MCH(2–17) on α-MSH release during in vitro superfusion. Concentration–response relationship is given for the effect of high concentrations of MCH(2–17) on α-MSH release. Numbers of incubations are given in parentheses. ***p < 0.001, **p < 0.01, *p < 0.05.
The different mode of action of these two inhibitors might differ from dopamine, which indiscriminately inhibited the release of mono- and diacetylated α-MSH from tilapia NILs (25). However, with respect to the effect on differential α-MSH release MCH resembles TRH, which also enhances the di/mono ratio of released α-MSH (25).

Immunoneutralization of MCH in NILs in vitro showed that in tilapia endogenous MCH inhibits α-MSH release from the melanotropes and that this inhibitory effect persists in vitro as long as 18 h, because after washout of the MCH antisera α-MSH release restored to the low control levels. The inhibitory effect of MCH on tilapia melanotropes is in line with observations in trout and eel, that immunoabsorbance of endogenously released MCH enhanced the ratio of released over stored amounts of α-MSH (9). These findings indicate that under physiological conditions the effect of MCH on α-MSH is inhibitory.

High concentrations of MCH (10 μM) and MCH(2–17) (10 and 35 μM) significantly increased α-MSH release. The physiological importance of this stimulation of α-MSH release is as yet unclear. It is doubtful whether concentrations up to 10 μM MCH, which are very high compared with MCH plasma values of trout (10 to 300 pM) (14–16,21), occur in the teleost pituitary. However, it cannot be excluded that at the MCH nerve terminals in the vicinity of the melanotropes MCH occasionally reach micromolar concentrations. We demonstrated that the stimulation of α-MSH release by MCH could not be attributed to the slight cross-reactivity of MCH with the antibody used in the α-MSH RIA. The occurrence of this cross-reactivity supports the notion that the tertiary structures of α-MSH and MCH are related (11). Apparently, this structural similarity is lost when the N-terminal amino acid (Asp) of MCH is removed, because MCH(2–17) appeared not to cross-react with the α-MSH antibody (this study) and MCH(2–17) exhibited almost no α-MSH-like activity on skin melanophores (11,26).

The stimulatory effect of MCH on the α-MSH release most probably is transduced by a mechanism different from that of the skin melanophores, because the effect of MCH(2–17) on α-MSH release from tilapia NILs was similar to that of MCH. Possibly, two MCH receptor subtypes, an inhibitory and a stimulatory one, occur in the teleost pituitary. However, we cannot exclude that MCH, at a high concentration, binds to receptors for other α-MSH release stimulating agents, such as TRH or CRH (25,31), although no structural similarity with these peptides has been described.
The finding that MCH interacts with the \( \alpha \)-MSH cell not only has implications for our understanding of the roles of both cell types in background adaptation, but probably also for their functions in the response to stressors. In tilapia, both cell types are responsive to acidification as a stressor (17,24), and in trout MCH secretion and biosynthesis are affected during injection and disturbance stress (6,15).

REFERENCES