Composite Survival Index to Compare Virulence Changes in Azole-Resistant Aspergillus fumigatus Clinical Isolates

Eleftheria Mavridou1,2, Joseph Meletiadis3, Pavol Jancura5, Saiden Abbas5, Maiken C. Arendrup4, Willem J. G. Melchers1,2, Tom Heskes5, Johan W. Mouton1,2, Paul E. Verweij1,2

1 Department of Medical Microbiology, Nijmegen Institute for Infection, Inflammation and Immunity, Nijmegen, The Netherlands, 2 Nijmegen Institute for Infection, Inflammation and Immunity, Nijmegen, The Netherlands, 3 Laboratory for Clinical Microbiology, Attikon University General Hospital, Athens, Greece, 4 Department of Microbiological Surveillance and Research, Statens Serum Institut, Copenhagen, Denmark, 5 Institute for Computing and Information Sciences, Faculty of Science, Radboud University, Nijmegen, The Netherlands

Abstract

Understanding resistance to antifungal agents in Aspergillus fumigatus is of increasing importance for the treatment of invasive infections in immunocompromised patients. Although a number of molecular resistance mechanisms are described in detail, the potential accompanying virulence changes and impact on clinical outcome have had little attention. We developed a new measure of survival, the composite survival index (CSI) to use as a measure of the virulence properties of A. fumigatus. Using a novel mathematical model we found a strong correlation between the in vitro growth characteristics and virulence in vivo expressed as CSI. Our model elucidates how three critical parameters (the lag phase (t), decay constant (δ), and growth rate (ρ)) interact with each other resulting in a CSI that correlated with virulence. Hence, strains with a long lag phase and high decay constant were less virulent in a murine model of invasive aspergillosis, whereas high virulence for isolates with a high CSI was associated in vitro with rapid growth and short lag phases. Resistant isolates with cyp51A mutations, which account for the majority of azole resistant aspergillosis cases, did not show a lower virulence compared to azole-susceptible isolates. In contrast, the CSI index revealed that a non-cyp51A-mediated resistance mechanism was associated with a dramatic decrease in CSI. Because of its predictive value, the mathematical model developed may serve to explore strain characteristics in vitro to predict virulence in vivo and significantly reduce the number of experimental animals required in such studies. The proposed measure of survival, the CSI can be used more in a general form in survival studies to explore optimal treatment options.

Introduction

Aspergillus fumigatus is responsible for the majority of invasive fungal infections in immunocompromised patients. Timely treatment with antifungal drugs is essential for the management of this disease and numerous efficacy studies have been carried out both in animals and humans to support evidence-based treatment choices [1,2,3]. To date, most investigations into antifungal efficacy have concentrated upon growth inhibition (MIC) whereas the role of fungal virulence has been largely ignored. The role of virulence as a factor in the disease outcome of patients with aspergillosis has hardly been considered, despite data obtained for other microorganisms indicating that the factor virulence may affect treatment [4,5,6].

The recent emergence of acquired resistance of A. fumigatus to medical triazoles [7,8,9,10,11,12,13,14], has drawn attention to the question whether the evolution ofazole resistance has any impact on the ability of the fungus to cause infection in man, and subsequently, on the clinical outcome. A link between azole drug resistance and the virulence of A. fumigatus was first demonstrated by Willger and colleagues [15]. Loss of SrbA, a sterol regulatory element binding protein resulted in growth incapacity of A. fumigatus and inability to cause fatal infections in two murine models of invasive pulmonary aspergillosis [16]. Further examination of the SrbA null mutant revealed that SrbA played a critical role in resistance to the azoles. Moreover, we recently reported reduced virulence in clinical A. fumigatus isolates that had become resistant to azoles during azole therapy [17]. A reduction of virulence in the above-mentioned studies was observed in non-cyp51A gene associated resistance mechanisms, while cyp51A-associated short nucleotide polymorphisms (SNPs) are the most prevalent resistance mechanism in clinical A. fumigatus isolates. Numerous SNPs in the cyp51A gene have been reported in clinical A. fumigatus isolates [18,19,20,21], which confer increased minimal inhibitory concentrations (MIC) for azoles in vitro and reduced azole efficacy in vivo [22,23]. Although A. fumigatus isolates with cyp51A mutations cause invasive aspergillosis in humans, indicating their ability to cause infection, quantitative estimates of in vivo virulence of isolates harboring those mutations are lacking [24].


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* E-mail: MavridouE.Rita@gmail.com (EM); p.verweij@mmb.umcn.nl (PEV)
A standardized animal model to compare the virulence of different A. fumigatus isolates is absent. It has been shown previously that variation in virulence between A. fumigatus isolates exists in a murine infection model but this study did not report susceptibility data [25] and other models previously used for measuring virulence reported unsatisfactory results [26].

In the present study, we investigated whether cyp51A-associated azole resistance mechanisms favored a gain or loss of virulence in A. fumigatus. To that purpose, we used a simple in vitro non-neutropenic murine model of disseminated aspergillosis. In particular, we explored the effects of in vitro growth characteristics on survival and developed a novel mathematical model. We propose and describe a new composite survival index (CSI) that enabled the prediction of survival in the animal model. The CSI was subsequently used to determine the impact of resistance mechanisms on the virulence of A. fumigatus.

Materials and Methods

Isolates

Thirty clinical A. fumigatus isolates from different patients and hospitals were used in this study (Table S1). Microsatellite genotyping showed no genetic relationship among all isolates (Table S1). Ten isolates were defined as wild type based on the in vitro susceptibility profile and absence of mutations in the cyp51A gene (Table S1). Twenty isolates were defined as non-wild type based on the in vitro susceptibility profile and the presence of mutations in the cyp51A shown to be associated with azole resistance.

From the collection of the aforementioned 30 isolates, we used in total 15 clinical isolates for the in vivo studies; three WTs (V28–29, V52–76, AZN 8196), three isolates were used that harbored the TR44/L98H resistance mechanism, which are believed to be selected through exposure toazole fungicides in the environment [9]. This mechanism was found to be the dominant resistance mechanism in clinical isolates in the Netherlands, other European countries and in Asia [27,28,29] and has also been found in azole-resistant isolates recovered from the environment [8,30].

Five isolates harbored SNPs in cyp51A, which were selected during azole therapy, including substitutions at codon M220 (28–77 with M220V, v59–07 with M220K, codon G54 (G54W, isolate v59–73), and codon 138 (G138C, isolate v59–72). In addition, four isogenic A. fumigatus isolates used were cultured serially from a single patient (isolates S1, S2, R1, and R2) [17]. The patient had chronic granulomatous disease (CGD) and was treated with multiple regimens of antifungal azoles for a chronic pulmonary Aspergillus infection. On antifungal therapy, the azole susceptibility changed from a wild-type phenotype (S1 and S2) to a resistant phenotype (R1 and R2). Although the expression of the cyp51A gene in the isolates R1 and R2 was elevated compared with the S1 and S2 isolates, no SNPs were found in cyp51A [17]. All isolates were stored in 10% glycerol broth at −80°C and were revived by subculturing on Sabouraud dextrose agar (SAD) supplemented with 0.02% chloramphenicol for 5 to 7 days at 35°C.

Molecular Analysis

The morphological identification of the A. fumigatus isolates was confirmed by sequencing of the β-tubulin and calmodulin genes, as described previously [8]. Genetic relationships of all isolates were determined by microsatellite genotyping (Table S1) [31]. This previously described assay relied on the variability of STRs in the A. fumigatus genome. Three trinucleotide and three tetranucleotide repeats of six different loci were amplified by using fluorescently labeled primers. The amplified DNA fragments were determined by the addition of the GeneScan LIZ [500] marker and were analyzed with the Applied Biosystems 3730 DNA software system. The assignment of repeat numbers in each marker was determined from GeneScan data by using Peak Scanner version 1.0 software (Applied Biosystems). The cyp51A coding region and its promoter were sequenced as previously described [18,32].

In vitro Studies

Antifungal susceptibility testing was performed based on the M38-A2 method of the Clinical Laboratory Standards Institute (CLSI) by using a microbroth dilution format and MICs of voriconazole, posaconazole, and itraconazole were determined as the lowest drug concentration resulting in no visible growth after incubation for 48 h [33]. In vitro growth curves of the thirty isolates were determined by using a previously described microbroth kinetic system [34,35]. OD measurement can be used to quantify A. fumigatus growth and it describes changes in fungal biomass. Briefly, 96-well microtitration plates were inoculated with 200 μL of RPMI1640 with 0.165 M MOPS containing 2.5×104 conidia. To prevent evaporation, the vertical margins of the microtiter plates were sealed with autoclave sterilization tape. After agitation for 15 s the suspensions were incubated at 37°C inside a plate reader (Rosys Anthos ht3; Anthos Labtec Instruments GmbH, Salzburg, Austria) for 90 h. The optical density at 405 nm (OD) was automatically recorded for each well every 15 min. The changes in OD over time were used to generate growth curves for each isolate in triplicate (OD-growth). All studies were conducted twice. The mean value was used for statistical analyses.

In vivo Studies

To compare the in vivo virulence of each A. fumigatus isolate, a previously described experimental murine model of disseminated aspergillosis was used [17,22]. A subset of 15 out of the 30 A. fumigatus strains was chosen based on the azole resistance mechanism and the duration of the lag phase, which was recently implicated as an important marker to predict virulence [17]. A total of 660 outbred CD-1 female mice (20–25 g, 4–5 weeks old) were randomized into 60 groups (n = 11 per group). Each of these groups was infected intravenously by the tail vein with four different inocula of the aforementioned isolates: concentrations of 1×106, 5×106, 1×107, and 5×107 CFU per mouse. Post-infection viability counts of the injected inocula were determined to ensure that the correct inoculum had been injected. Mortality was monitored for 15 days. On day 15 post-infection, all remaining mice were humanely euthanized by cervical dislocation. To assess the variability of in vivo studies, A. fumigatus isolates with different azole resistance mechanisms (the wild type, G54W, TR44/L98H, M220I, and the M220I isolate) were tested in triplicate at a low (1×106 CFU per mouse) and a high (5×107 CFU per mouse) inoculum. The differences in percentage of survival and MST were <10% and <1 day among replicates, indicating very high reproducibility.

The animals were housed under standard conditions with water and food supplied ad libitum. Animal studies were carried out in strict accordance with the recommendations of the European Community (Directive 86/609/EEC, 24 November 1986). The protocol was approved by Animal Welfare Committee of the Radboud University (RU-DEC 2007-106). All efforts were made to minimize suffering. To prevent severe discomfort and substantial distress, throughout the whole study mice were monitored and
clinically inspected several times day and night. The experiment was immediately terminated if the humane endpoints were not fulfilled. Symptoms of discomfort and distress included loss of weight (15% loss within 72 h or 20% within 48 h); very high or very low body temperature; desiccation; lethargy; hyperactivity in terms of spinning around in circles constantly; eyes turning black; reduced mobility, and therefore, mice are unable to reach food or drink; or generally reduced food/water intake. All stressed animals were sacrificed by cervical dislocation. The murine model of disseminated aspergillosis is well established in our laboratory. Therefore, the veterinary experts and investigators are able to recognize the clinical symptoms in the animals at a very early stage and can ensure early termination before the humane endpoints are reached without affecting interpretation of the experimental results.

Modeling of the in vitro Growth in A. fumigatus

In vitro growth curves were analyzed with nonlinear regression analysis based on a novel mathematical model of fungal growth. Previous reports suggested Malthusian fitness as a possible measure model of fungal fitness [36]. This function assumes that the size of the population is limited by an asymptotic maximum (plateau). However, because the previous and our present growth kinetic experiments showed that a plateau phase was not reached, we set out to define a new model that better described the observed growth [34,35,37]. This model incorporated a piecewise function of 1) the lag phase \((\tau)\), which corresponded to no OD-growth changes observed until the hyphal length reached 70 \(\mu\)m; and 2) the growth phase of the fungus, which corresponded to OD-growth changes post \(\tau\). The lag phase was described as a constant function. The growth phase was modeled by the function composed of two main factors: linear OD growth and exponential decay. With these factors, the model reasonably captures the dynamics of the growth phase and the death phase which are constantly and simultaneously present during the life cycle of a cell population which is composed of a heterogeneous population of aging cells. The underlying assumptions of our model are currently utilized in prediction models for other cell types and organisms which take into account the variability in the aging process of the cells. To the best of our knowledge there has been no similar function model of fungal fitness [36]. This function assumes linear growth of the population (linear component), whereas, at the same time, cell death would occur due to ageing and nutrient consumption (exponential component). The proportion of cell death would increase over time.

\[
Y(t) = \begin{cases} 
Y_0 & ; t < \tau \\
Y_0 + v(t - \tau)\exp(-\lambda(t - \tau)) & ; t \geq \tau 
\end{cases}
\]

where \(Y(t)\) represents the fungal biomass assessed based on the optical density at time \(t\). For the mathematical growth function parameters, \(Y_0\) corresponded to the OD value for the period of the lag phase, \(t\) is the lag phase duration in hours, \(v\) is the growth rate expressed in OD \(\times h^{-1}\), and \(\lambda\) is the growth decay constant expressed in \(h^{-1}\). The use of linear components in a nonlinear equation is a common approach which increases the flexibility of the equation in order to capture complex phenomena like Aspergillus dynamic growth. In general, this growth model assumes linear growth of the population (linear component), whereas, at the same time, cell death would occur due to ageing and nutrient consumption (exponential component). The proportion of cell death would increase over time.

Clustering Analysis of Strains Based on the Fungal Growth

To identify whether there were differences in fungal growth between the strains, clustering analysis was performed on their growth parameters \((\tau, v, \lambda)\) as follows: Each parameter was standardized by computing its \(z\)-score. Then, since the number of clusters was unknown, we created an agglomerative hierarchical cluster tree by applying the centroid linkage method (UPGMC) with the Euclidean distance.

The quality of agglomerative clustering was assessed by the cophenetic correlation coefficient (CPCC). The significance of clustering was estimated by computing the probability of obtaining the CPCC by chance as follows: Given a group of \(N\) strains, we generated \(10^5\) random samples of \(N\) objects (“random” strains) with coordinates from the same range as the given \(N\) strains. Then, parameters of each random sample were \(z\)-score standardized and the objects were clustered by the same agglomerative technique as the original data. CPCC was computed for each random sample (CPCC\(_i\)). As a result, we accumulated the distribution of CPCC\(_i\) for these clusterings \(\{C^{i}\} = \{\text{CPCC}\_i \mid i \in \{1,2,\ldots,10^5\}\}\) and we counted how many times CPCC\(_i\) was equal to or greater than the cophenetic correlation CPCC\(_0\) of the clustering original strains \(C = \{\text{CPCC}\_i \in \mathbb{C}^{i} | \text{CPCC}\_i \geq \text{CPCC}\_0\}\). Then, the empirical \(p\) value of the original clustering was determined as

\[
P = \frac{|C|}{|C^{R}|}
\]

If clustering of the original strains was significantly better than clustering of random samples \((p < 0.05)\), the derived clustering structure had a low probability of being obtained by chance.

The CSI: a New Composite of Survival

For in vivo studies, survival data for each inoculum is analyzed based on the Kaplan Meier method (log-rank test), and \(SUR\%\) and \(MST\) is determined. \(SUR\%\) is calculated as the number of survivors over the total number of infected animals at the end of the experiment. \(MST\) is the time at which the fractional survival equals 50\% (GraphPad Prism, version 5.0). For inocula where 50\% mortality is not observed within the predefined time interval, \(MST\) is considered as maximum \(MST\) (\(MST_{\text{max}}\)) equal to the day of termination of the experiment.

Although, \(SUR\%\) is considered to be a measure for estimating virulence in general, it does not take into account \(MST\), which is also an outcome of Kaplan Meier analysis. We therefore propose a new composite index of these two parameters of survival, the CSI. CSI is defined to include both terms as follows:

\[
CSI = 1 - \frac{SUR\% \times MST}{100\% \times MST_{\text{max}}}
\]

where \(MST_{\text{max}}\) is the length of the in-vitro experiment, which equals to the day of termination of the experiment. The denominator of the second term is a normalizing constant that restricts CSI between 0 and 1. CSI corresponds to the degree and rate of mortality, for which high mortality corresponds to high CSI. The CSI itself is used here as a direct measure of virulence, but can be applied as an index value for any survival curve.

Correlation between Fungal Growth and CSI

A multiple logistic regression analysis was employed to determine the relationship between virulence markers and fungal growth-curve parameters. Logistic functions were used to fit three
different virulence markers as follows

\[ MST \sim \frac{MST_{\text{max}}}{1 + e^{-g(t,\nu,\lambda)}} \]

\[ SUR\% \sim \frac{100}{1 + e^{-g(t,\nu,\lambda)}} \]

\[ CSI \sim \frac{1}{1 + e^{-g(t,\nu,\lambda)}} \]

in which \( g(t,\nu,\lambda) \) is the following function of growth parameters:

\[ g(t,\nu,\lambda) = at + bv + c\lambda. \]  (3)

\( a, b, \) and \( c \) are regression coefficients. Virulence was quantified by using \( MST, SUR\% \), and \( CSI \). Thus, for all virulence measurements (response-dependent variables), the regression coefficients were estimated independently. In addition, the regression for each response model was cross-validated by using Stein formula [38] of adjusted \( R^2 \) to assess the loss of predictive power of each response model.

**Comparison of the Virulence-marker Models**

The two virulence markers with the best goodness of fit were compared to assess which was explained better by the growth parameters. To this end, we computed standardized residuals for each of the two response models using leave-one-out cross validation.

Given a model, one response value was omitted and the multiple logistic regression analysis was performed on the remaining response values to estimate the regression coefficients. Afterwards the removed response was predicted by the logistic function by using the estimated coefficients. The standardized residual \( R \) of the predicted value \( (V_{\text{pred}}) \) and the observed value \( (V_{\text{obs}}) \) of the omitted response was computed by using the formula

\[ R = \frac{|V_{\text{pred}} - V_{\text{obs}}|}{\text{std}(V)}, \]

in which \( \text{std}(V) \) was the sample standard deviation of all response values. The procedure was repeated for each response value. For each strain, we compared the standardized residuals of response values of both models: the lower the standardized residuals, the better the fit.

**Construction of a General Model**

The logistic function-based model given by Equation 4 was used to incorporate the four-dimension dataset as a general model in which \( CSI \) was associated with growth-curve parameters \((\nu,\lambda)\) and inoculum size, \( \Phi \):

\[ CSI \sim \frac{1}{1 + \exp \left( \Phi(EC_{50} - (at + bv + c\lambda + \log(\Phi)) \right)} \]  (4)

in which \( EC_{50} \) is a parameter associated with a \( CSI \) value of 0.5.

**Results**

**Phenotype and in vitro Growth Characteristics**

The in vitro susceptibility phenotype to itraconazole, posaconazole, and voriconazole, the underlying resistance mechanism, and the short tandem repeat (STR) profile of the 30 clinical \( A. fumigatus \) isolates are shown in Table S1. The recently proposed interpretative breakpoints were used to categorize the phenotypes of the isolates [39]. For each isolate, the growth curve was characterized using a microbroth kinetic system [34,37]. In this system, microscopic examination has confirmed that an increase in the optical density corresponds to the multicellular development of \( A. fumigatus \) [37]. Previously multiple phases (lag phase, log phase, and two transition phases) were identified to describe the growth of \( A. fumigatus \) in vitro [37], however our novel mathematical model identified only two phases; the lag phase and the decaying linear OD growth phase, the latter including the log and two transition phases (Fig. 1, Equation 1). The proposed growth function describes the first phase through the duration of the lag phase \((t)\), and the second phase by the linear OD growth rate \((\nu)\) and the decay constant \((\lambda)\). The stationary phase was not observed during the incubation period of 90 h. The mathematical model of growth fitted well to the observed growth curves (Fig. S1) with \( R^2 \) values ranging from 0.98 to 0.99 with no zero value within the 95% CI of the model parameters for all isolates (Table S2). The correlation study of the growth parameters \((\nu,\lambda)\) revealed that two parameters, \( \nu \) and \( \lambda \), exhibit a high and statistically significant linear correlation (Table 1). Despite this correlation, the \( CSI \) measure using both growth parameters can be still applied as a predictive tool which is verified by F-test on nested models of \( CSI \) (Table 4).

**Differential Survival Outcomes Reflect a Strain-dependent Virulence Distinction**

To explore whether there was a variation in virulence of our strains in vivo, we performed survival experiments with 15 \( A. fumigatus \) strains using 4 different inocula for each strain. Table 2 depicts differential survival rates for each strain of mice groups inoculated with the same lethal or sublethal inoculum. Susceptibility to infection was most prominent in animals infected with an inoculum of \( 5 \times 10^7 \) CFU and no clear differences in percentage of survival \((SUR\%)\) and median survival time \((MST)\) were observed, with exception of two strains. In mice infected with azole-resistant isolates R1 and R2, 100% mortality occurred in a time period that...
was significantly longer than the other groups. Lower mortality and longer MST was also observed at the inoculum of $10^7$ CFU. In contrast, mice infected with the parental S2 rapidly succumbed at a dose of $10^6$ conidia and $5 \times 10^3$, while these concentrations showed a sublethal outcome in the case of infection with wild type or any of the other mutants. Moreover, both highest inocula concentrations resulted in lowest survival in shortest time compared to all other groups (Table 2). Similarly, the second lowest SUR% with short MST was found for the groups infected with the G54W mutant, however, for the main two lowest inocula.

As our survival data suggested a wide variation in the virulent traits of some strains, we next addressed the question if the observed variation in SUR% and MST could be correlated with characteristics of the in vitro growth curves. Furthermore, to improve the sensitivity to detect in vivo virulence differences, we developed a dynamic marker, the Composite Survival Index (CSI), which describes the rate (MST) and extent of killing (SUR%) (Equation 2).

Cluster Analysis Reveals Heterogeneous Growth within the Wild Type and Azole-resistant Populations

A clustering approach was applied to the growth-curve parameters ($\tau$, $\nu$, $\lambda$) estimates of all strains to explore primary differences in growth between the wild type and the non-wild type isolates more accurately (Fig. S2 & Fig. 2). Statistical analysis revealed that clustering was significantly better than that found for a random population (CPCC = 0.922, $p<10^{-5}$). Interestingly, there was no single-cluster formation of the wild type population, indicating that the wild type strains did not share similar growth characteristics. Similarly, no single cluster was observed for any of the strains harboring cyp51A mutations (Fig. 2).

**Virulence in A. fumigatus is Related to Growth**

To determine whether growth of *A. fumigatus* was associated with virulence, we investigated the relationship between $\text{MST}$, $\text{SUR}^\%$, the new proposed Composite Survival Index (CSI) and the growth properties ($\tau$, $\nu$, $\lambda$) of each strain. This index incorporates both SUR% and MST and may be viewed as an indicator of virulence of a strain (Equation 2). Multiple logistic regression analysis was used to fit in vitro growth characteristics $\tau$, $\nu$, and $\lambda$ to all three virulence markers (Fig. 3). For each virulence marker, the fitting procedures estimated different regression coefficients ($a$, $b$, $c$) (Table 3).

While regression analysis revealed a weak correlation between $\text{MST}$ and the growth properties ($R^2 = 0.36$; Fig. 3A), a strong correlation was found for the other two responses (for $\text{SUR}^\%$, $R^2 = 0.84$; for $\text{CSI}$ $R^2 = 0.92$; Fig. 3B & 3C).

Regression coefficients $a$, $b$, and $c$ for the growth parameters $\tau$, $\nu$ and $\lambda$ (Equation 3), respectively, associated with CSI, were as follows: $a = -0.51$ (95%CI: $-0.72$, $-0.29$), $b = 1835.78$ (95%CI: $1811.09$, $1860.46$) and $c = 183.578$ (95%CI: $181.109$, $186.046$).

### Table 1. Sample correlation coefficients between the growth parameters ($\tau$, $\nu$, $\lambda$).

<table>
<thead>
<tr>
<th>Correlation Coefficients</th>
<th>$\tau$</th>
<th>$\nu$</th>
<th>$\lambda$</th>
<th>$p$-values</th>
<th>$\tau$</th>
<th>$\nu$</th>
<th>$\lambda$</th>
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<td>$p&lt;0.05$</td>
<td>1</td>
<td>0.9938</td>
<td>0.5851</td>
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<tr>
<td>$\nu$</td>
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<td>0.8651</td>
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<td>0.9938</td>
<td>1</td>
<td>0.000031</td>
</tr>
<tr>
<td>$\lambda$</td>
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<td>0.8651</td>
<td>1</td>
<td>$p&lt;0.05$</td>
<td>0.5851</td>
<td>0.000031</td>
<td>1</td>
</tr>
</tbody>
</table>

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### Table 2. Differential survival outcome reflects strain-dependent virulence distinction.

| Isolates | Inoculum $10^6$ CFU | | | | Inoculum $5 \times 10^9$ CFU | | | | Inoculum $10^7$ CFU | | | | Inoculum $5 \times 10^7$ CFU | |
|----------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|---------------------|
|          | SUR% | MST | SUR% | MST | SUR% | MST | SUR% | MST | SUR% | MST | SUR% | MST |
| WT (AZN-8196) | 100 | 15 | 72.72 | 15 | 18.18 | 8 | 0 | 3 |
| WT (V52–76) | 63.63 | 15 | 36.36 | 12 | 9.09 | 4 | 0 | 2 |
| WT (V28–28) | 100 | 15 | 63.63 | 15 | 0 | 6 | 0 | 2 |
| TR54/L98H (V52–35) | 90.90 | 15 | 45.45 | 8 | 18.18 | 8 | 0 | 3 |
| TR54/L98H (V45–07) | 100 | 15 | 27.27 | 11 | 45.45 | 11 | 0 | 2 |
| TR54/L98H (V51–76) | 100 | 15 | 18.18 | 8 | 0 | 4 | 0 | 2 |
| M220I (V28–77) | 81.81 | 15 | 63.63 | 15 | 0 | 7 | 18.18 | 5 |
| M220K (V59–27) | 100 | 15 | 63.63 | 15 | 45.45 | 12 | 0 | 2 |
| M220V (V13–09) | 81.81 | 15 | 9.90 | 10 | 9.09 | 9 | 0 | 2 |
| G54W (V59–73) | 45.45 | 8 | 0 | 8 | 0 | 4 | 0 | 3 |
| G138C (V59–72) | 90.09 | 15 | 54.54 | 15 | 0 | 6 | 0 | 2 |
| S1 (V67–38) | 90.09 | 15 | 63.63 | 15 | 36.36 | 15 | 0 | 2 |
| S2 (V67–37) | 36.36 | 5 | 0 | 2 | 0 | 2 | 0 | 1 |
| R1 (V67–36) | 100 | 15 | 90.09 | 15 | 90.09 | 15 | 0 | 6 |
| R2 (V67–35) | 100 | 15 | 100 | 15 | 63.63 | 15 | 0 | 6 |

**Table 1.** Sample correlation coefficients between the growth parameters ($\tau$, $\nu$, $\lambda$).

**Table 2.** Differential survival outcome reflects strain-dependent virulence distinction.

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between the respective growth characteristics negative coefficients a and c and the CSI, which means that strains with a high growth rate have high CSI values.

The linear combination \( g(\tau, v, \lambda) \) of the growth parameters (Equation 3) combined with the estimated regression coefficients for CSI provides a good measure of the virulence of any given A. fumigatus isolate. Accordingly, an isolate with \( g(\tau, v, \lambda) = 0 \), will have a CSI of 0.5, which corresponds to 50% SUR% and an MST of 15 days. This is also the case for any other combination of SUR% and MST that results in a CSI of 0.5 based on Equation 2. By definition, CSI is always less than or equal to 1-SUR%. Thus, a CSI of up to 0.5 is obtained only with a SUR% of at least 50%.

Figure 3B and 3C show, for the SUR% and CSI models, that the virulence of the two azole-resistant isogenic isolates, R1 and R2, was reduced relative to the other isogenic azole susceptible isolates and all other strains. Specifically, the SUR% in mice exposed to infection by R1 and R2 was greater than 60% and \( g(\tau, v, \lambda) \) was greater than 0. Two other isolates with mutations in cyp51A (TR34/L98H_45-07 and M220K) were less virulent than all WT isolates and the other cyp51A mutants. Eleven out of 15 isolates displayed a high CSI of 0.9, which corresponded with an increased probability of death when the value of \( g(\tau, v, \lambda) \) exceeded 2.2.

To verify whether all growth characteristics were significant for the CSI model, we additionally performed multiple regressions on models with reduced numbers of growth parameters. The F-test comparison of the original and nested models revealed that a reduction in each parameter resulted in a significant decrease in the performance of the model (Table 4). Table 4 also shows that extending the original model by \( Y_0 \) parameter did not lead to statistically significant improvement of the fit.

The Novel Composite Survival Index is the Superior Virulence Marker

We performed a comparative leave-one-out cross-validation study between the SUR% and the CSI models to verify the superiority of CSI over SUR%. Figure 4 demonstrates that the CSI model describes the relationship between fungal growth and virulence better than the SUR% model because 12 of the 15 isolates had less standardized residuals for CSI than for SUR% (Fig. 4, markers below the diagonal). To test the significance of the improvement in the residual values, we applied a binomial statistical test, which showed that the CSI model was significantly better than the SUR% model \( (p<0.02) \).

Prediction of Virulence via CSI

Mice survival for the aforementioned studies was assessed following an infection of \( 10^6 \) CFU per mouse. To further confirm the validity of CSI, we carried out the infection in groups with three additional doses of conidia. Figure 5 depicts the resulting CSI versus the function \( g(\tau, v, \lambda) \) relationships for four different inocula. There was no clear sigmoidal relationship between CSI and \( g(\tau, v, \lambda) \) for the groups infected with \( 1 \times 10^3, 5 \times 10^3, \) and \( 5 \times 10^5 \) CFU per mouse. In panel with an infection of \( 10^6 \) CFU, only the initial, shallow part of the sigmoidal shape of the CSI curve is apparent, whereas increasing the concentration to \( 5 \times 10^6 \) results in a steeper gradient of the curve. Lastly, the highest infection concentration of \( 5 \times 10^6 \) was associated with reaching the plateau of the sigmoidal curve and results in no differentiation between the different strains.

As expected, different inocula affected the virulence of individual isolates, indicating that CSI was also dependent on the inoculum size. Therefore, we defined a general model for

1043.1, 2628.5), \( a = -526.92 \) (95% CI: -1009.6, -44.23). The negative coefficients a and c indicate a negative relationship between the respective growth characteristics \( \tau \) and \( \lambda \) and the CSI.
predicting the virulence of a strain based on the initial inoculum ($\Phi$) and the growth of the strains (Fig. S3; Equation 4).

Nonlinear multiple regression analysis showed the ability of the model to predict virulence, as shown by the good overall statistics of the fit ($R^2 = 0.82$, $p<0.001$; Fig. S3). The mean (95%CI) estimates of the regression coefficients $a$, $b$, $c$, $d$ and the EC$_{50}$ was $-0.06$ ($-0.10$, $-0.01$), $207.03$ ($111.45$, $307.24$), $-156.42$ ($-266.68$, $-58.31$), $3.81$ ($2.74$, $5.96$) and $6.60$ ($6.01$, $7.13$), respectively.

Figure 3. Relationship between the virulence markers MST, and SUR % and CSI and the function $g(t, v, \lambda)$. The logistic function was used to fit three different virulence markers: the median survival time (MST, panel A), the survival percentage (SUR %, panel B) and the composite survival index (CSI, panel C). No in vitro-in vivo correlation was found between MST and $g(t, v, \lambda)$, whereas a strong correlation was found for the other two virulence markers. The symbols correspond to the observed CSI values, while the solid line is the outcome of the prediction model.

doi:10.1371/journal.pone.0072280.g003

Discussion

The primary aim of our study was to determine the impact of acquisition of azole resistance by A. fumigatus on virulence and to determine which in vitro growth characteristics are critical for in vivo survival. Our in vivo experiments showed that virulence, expressed as SUR% and MST, is variable, even between wild type isolates, and that azole-resistant clinical A. fumigatus isolates with cyp51A mutations are not less virulent. However, development of azole resistance may be associated with loss of virulence as was
apparent with the set of four isogenic isolates, where the transition from an azole-susceptible phenotype to an azole-resistant phenotype was associated with higher SUR% and MST. A novel non-cyp51A mediated resistance mechanism was recently reported in these isolates, which consisted of a mutation in the CCAAT-binding transcription factor complex subunit HapE [40]. Although the ergosterol biosynthesis pathway is critical for growth and proliferation of the fungus (and thus is an important drug target), it has previously been shown that SNPs in the cyp51A gene of Candida occurred without major perturbation of the haem environment or activity, and as a consequence, allowed resistant mutants to produce ergosterol and retain fitness [41]. Our study suggests that this may also be the case in A. fumigatus that harbor SNPs in cyp51A. Although the exact role of HapE is not yet understood, isolates with the HapE mutation exhibited altered growth characteristics, such as impaired growth, suggesting that this mutation has implications for a broad range of processes in the fungal cell.

From a clinical perspective our observations are of importance as they indicate that A. fumigatus isolates with an azole-resistant phenotype due to a cyp51A-mediated resistance mechanism are capable of causing a similar spectrum of azole diseases as the wild-type isolates [10, 11, 12, 21, 28, 42].

This supports the clinical experience of increasing reports of cases of non-invasive and invasive aspergillosis due to azole-resistant isolates, and the high probability of failure during azole therapy [12, 43]. Furthermore, for TR34/L98H mutations, which are considered to have developed through exposure to 14β-demethylase inhibitors (DMIs), any fitness loss would be an important disadvantage when competing with wild-type field isolates. Our findings indeed indicate that the virulence of TR34/

<p>| Table 3. Descriptive statistics of fitting the $g(t, v, \lambda)$ function against the composite survival index (CSI), the percentage of survival (SUR%) and the median survival time (MST) for the inoculum $10^7$ CFU. |
|---------------------------------|----------|----------|----------|----------|</p>
<table>
<thead>
<tr>
<th>Regression coefficients (95% CI ±)</th>
<th>a</th>
<th>b</th>
<th>c</th>
<th>R²</th>
<th>R²adj</th>
<th>MSE</th>
</tr>
</thead>
<tbody>
<tr>
<td>MST</td>
<td>0.162 (−0.071, 0.395)</td>
<td>−501.294 (−1054.08, 51.497)</td>
<td>367.81 (−195.07, 930.7)</td>
<td>0.37</td>
<td>−0.12</td>
<td>11.85</td>
</tr>
<tr>
<td>SUR%</td>
<td>0.354 (0.122, 0.586)</td>
<td>−1562.43 (−2472.47, −652.43)</td>
<td>968.64 (80.79, 1856.48)</td>
<td>0.82</td>
<td>0.68</td>
<td>162.4</td>
</tr>
<tr>
<td>CSI</td>
<td>−0.511 (−0.741, −0.281)</td>
<td>1887.502 (983.5, 2791.51)</td>
<td>−988.59 (−1782.6, −194.6)</td>
<td>0.91</td>
<td>0.84</td>
<td>0.007</td>
</tr>
</tbody>
</table>

MSE, mean squared error; doi:10.1371/journal.pone.0072280.t004

Figure 4. Leave one out-cross validation analysis. The CSI model describes the relationship between fungal growth and virulence better than the SUR% model because 12 of the 15 isolates had less standardized residuals for CSI than for SUR% (markers below the diagonal) (p<0.02). The shapes and colors of the symbols used for the observed data represent the same isolates as those defined in Figure 3. doi:10.1371/journal.pone.0072280.g004

Table 4. The F-test statistics of nested CSI models compared to the original CSI model.

<table>
<thead>
<tr>
<th>Growth parameters kept</th>
<th>F</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>t</td>
<td>27.76</td>
<td>0.0002</td>
</tr>
<tr>
<td>r</td>
<td>134.14</td>
<td>7.17x10^-08</td>
</tr>
<tr>
<td>k</td>
<td>43.26</td>
<td>2.59x10^-05</td>
</tr>
<tr>
<td>σ</td>
<td>78.33</td>
<td>1.30x10^-07</td>
</tr>
<tr>
<td>v</td>
<td>54.11</td>
<td>9.89x10^-07</td>
</tr>
<tr>
<td>d</td>
<td>67.87</td>
<td>2.87x10^-07</td>
</tr>
<tr>
<td>tσxY₀</td>
<td>4.61</td>
<td>0.055</td>
</tr>
</tbody>
</table>

The last row demonstrate F statistics when the original model is nested with respect to the extended model by adding Y₀ into the linear term g(c, r, λ). doi:10.1371/journal.pone.0072280.t004
that other factors than growth may play role in virulence of this strain related with host and/or fungus. For example, immunogenic molecules on the fungal cell wall surface that affect docking and pathogen recognition result in reduced virulence [44], whereas spore production, spore decay, spore settlement, spore germination, and mycelia growth rate are important fitness components for fungi [36]. Interestingly, a change in virulence was also observed and correctly predicted by the model for the M220K isolate. The CSI was approximately 0.5 times lower than that of the wild type isolates and the other cyp51A mutants, and around 0.5 times greater than the least virulent isolates R1 and R2, indicating reduced virulence. If as stated above cyp51A-mutations have little or no fitness costs, differences in virulence as observed in the M220K isolate are probably due to changes in other genes or pathways, similar to those that may occur in wild type isolates. As we used clinical isolates and the resistance mechanism as selection criterion, the genetic background of our isolates will vary significantly. The use of recombinants would be an interesting next approach to further explore the contribution of individual SNPs on the virulence of *A. fumigatus*.

The CSI may be a useful tool to determine the impact of mutations on virulence. Moreover, the use of a general model (Equation 4) allows prediction of virulence *in vivo*, thereby reducing the number of animals required in such experiments. We have applied this tool in our laboratory to allow us to calculate the inoculum required for studies; this has led to substantial savings in time and animals.

In our study, we employed a relative simple and conservative murine model of aspergillosis. We used a nonneutropenic model, although patients with invasive aspergillosis often suffer from neutropenia and the opportunistic fungus is believed to be only infective when patients carry immunodeficiencies in one way or another. Once the fungal growth overcomes the immune defense the progress of the disease starts. This bears similarity to the model used. Mice are susceptible to conidia in high amounts and by increasing the infection-dose mortality increased. We could therefore avoid the use of immunosupressors that has been
described to alter A. fumigatus growth and thereby affect the interpretation of our results [25,45]. Moreover, the long-term effects of immunosuppressors on the fungus or host are almost completely unknown. Regarding the route of infection, there are two reasons for having chosen the disseminated model as opposed to an inhalation model. First, this infection model is well established and allows excellent control of infection parameters (such as inoculum), whereas the pulmonary model comes in a variety of forms with several technical problems including less well controlled infection. Secondly, Thomas and Elkinton reported that virulence and infectivity can primarily be measured when experimental infections occur through the intravenous portal because it involves establishment and spread within the host [46].

Multiple factors require investigation in the context of comparative virulence. We here focused on one aspect, the impact on survival and virulence in the process of disseminated growth characteristics and conclude that these have a major comparative virulence. We here focused on one aspect, the because it involves establishment and spread within the host [46].

Supporting Information

Figure S1 Fitting of the in vitro growth curves of fifteen A. fumigatus isolates. The proposed model (Equation 1) for simulating the growth of A. fumigatus in vitro (blue line) fitted well to the observed growth curves (red line) of fifteen clinical isolates with $R^2$ values ranging from 0.98 to 0.99. (TIF)

Figure S2 Distribution of the wild-type population and populations with diverse cyp51A mutations. Azole susceptible isolates (wild-type population) are black colored; red color, TR34/L98H mutants; blue, isogenic resistant strains R1 and R2; cyan, isogenic susceptible strains S1 and S2; green, M220I, M220K, M220V, TR46/L98, G138C, G54W. (TIF)

Figure S3 Prediction of CSI based on the inoculum size and the growth characteristics. Graphs depicting the goodness of fit ($R^2=0.82$, $p<0.001$) of the full general model predicting CSI based on the growth-curve parameters $\tau$, $\nu$ and $\lambda$ and the inoculum size $\Phi$ (the four inocula are depicted with different colors) for the fifteen A. fumigatus strains. (TIF)

Table S1 Characteristics of thirty clinical A. fumigatus isolates used in our studies. (DOC)

Table S2 In vitro growth characteristics and growth phase fit of thirty clinical A. fumigatus isolates. (DOC)

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Author Contributions

Conceived and designed the experiments: EM JM JWM PEV. Performed the experiments: EM. Analyzed the data: EM JM JWM PJ SA TH. Contributed reagents/materials/analysis tools: WJGM MCA. Wrote the paper: EM JM JWM PEV. Provided molecular data: WJGM. Designed and supervised the mathematical analysis: TH. Critical revision for important intellectual content: MCA.

References

Prediction of Virulence in Aspergillus fumigatus