Acute disseminated candidiasis is a life-threatening condition that occurs predominantly in immunocompromised hosts. The mortality rate associated with disseminated candidiasis is high (31), and the incidence of this disease has increased in recent years (1, 3). Candida albicans ranks fourth among the organisms most frequently isolated from blood cultures in the United States (1). Clinically, systemic candidiasis sometimes mimics gram-negative sepsis. Viable Candida cells and cell wall constituents are able to induce the synthesis of proinflammatory cytokines in vitro (13, 14, 18), similar to gram-negative bacteria and their lipopolysaccharide (LPS) component (23). However, the role of these cytokines in systemic candidiasis is probably beneficial rather than deleterious (20, 26). In contrast, the induction of proinflammatory cytokines, such as interleukin-1α (IL-1α) and IL-1β and tumor necrosis factor alpha (TNF-α), is merely a deleterious event in gram-negative sepsis (4) and treatment aimed at blocking cytokine action has proved beneficial in various experimental models (2, 27, 30). In vivo, Candida grew better in plasma samples of LDLR−/− mice than in control plasma samples and peritoneal macrophages of LDLR−/− mice challenged with heat-killed C. albicans produced more cytokines than did those of controls. This latter phenomenon was probably due to increased binding of yeast cells to macrophages of LDLR−/− mice. These data suggest that hyperlipoproteinemia is deleterious in systemic candidiasis.

MATERIALS AND METHODS

Animals. Heterozygous C57BL/6 LDLR−/− mice and wild-type littermates were obtained from Jackson Laboratory (Bar Harbor, Maine) as mating pairs and bred in our local facility. For experiments, 6- to 8-week-old mice, weighing 20 to 25 g, were used. The animals were fed standard laboratory chow (Hope Farms, Woerden, The Netherlands) and housed under specific-pathogen-free conditions. The experiments were approved by the ethical committee for animal experiments at the Catholic University Nijmegen.

C. albicans infection. C. albicans (strain UCS20), maintained on agar slants at 5°C, was inoculated into 100 ml of Sabouraud broth and cultured for 24 h at 37°C. After three washes with pyrogen-free saline by centrifugation at 1,500 × g, the viability of yeast cells was at least 98%, as confirmed by plating serial dilutions on Sabouraud dextrose agar plates. Mice were injected intravenously (i.v.) in the retro-orbital plexus with 105 or 107 CFU of C. albicans. Survival was assessed daily for 14 days in groups of at least 15 animals. In separate groups, after 4 h and 1 and 3 days, subgroups of five mice were killed by cervical dislocation and blood samples were collected for the measurement of plasma cytokine concentrations.
The outgrowth of microorganisms from the livers, spleens, and kidneys of animals was quantified on days 1 and 3 after infection with C. albicans. For this purpose, the organs were removed aseptically, weighed, and homogenized in sterile saline in a tissue homogenizer. The number of viable C. albicans cells in tissue samples was determined by plating serial dilutions on Sabouraud dextrose agar plates as described previously (15), and CFU were counted after overnight incubation at 37°C. The results were expressed as the log CFU per gram of tissue.

Cytokine production. Resident peritoneal macrophages were harvested by rinsing the peritoneal cavity aseptically with cold phosphate-buffered saline containing 0.38% (wt/vol) sodium citrate. After centrifugation, cells were resuspended in RPMI 1640 (Dutch modification: Flow Laboratories, Irvine, United Kingdom) containing 1 mM pyruvate, 2 mM l-glutamine, and 100 μg of gentamicin per ml and incubated for 1 h at 37°C (10^6/well) in 96-well microtiter plates (Costar Corporation, Cambridge, Mass.). Nonadherent cells were discarded, and the remaining cell population consisted of more than 90% macrophages, as assessed by light microscopy. Heat-killed (30 min, 100°C) C. albicans cells were radioiodinated by the iodogen method (8) with minor modifications. Briefly, 10^9 C. albicans cells in 50 mM sodium phosphate buffer (pH 7.4) were iodinated in an iodogen-coated vial (20 μg iodine/50 μl) in the presence of 300 μCi of Na^125I (specific activity, 15 mCi/μg; Amersham) in a total volume of 100 μl. After 15 min of incubation, the suspension was washed three times with phosphate-buffered saline and resuspended in RPMI 1640 containing 1 mM pyruvate, 2 mM l-glutamine, and 100 μg of gentamicin per ml (labelling efficiency, 45%). Peritoneal macrophages from LDLR_/_ and control mice were collected as described above. Macrophages (2 x 10^5) were incubated with 10^6 CFU of radiiodinated C. albicans in a total volume of 200 μl of RPMI 1640 at 37°C in 96-well microtiter plates. After 5, 15, and 30 min, the supernatant was discarded and the radioactivity associated with adherent cells was measured with a gamma counter. Nonspecific binding was determined in wells in which C. albicans cells were incubated in the absence of macrophages. Total binding of C. albicans cells to macrophages (including both specific binding to receptors and nonspecific binding through membrane hydrophobicity or other nonspecific interactions) was determined by subtracting the binding of C. albicans cells to plastic from total counts.

Growth of C. albicans in vitro. To investigate whether the growth of C. albicans in the plasma of LDLR_/_ mice is similar to that in plasma of control mice, 2 x 10^5 CFU of C. albicans were incubated in plasma samples obtained from LDLR_/_ and control mice diluted 50% with Sabouraud broth in 10-ml glass tubes (Hospidel, Nieuwkoop, The Netherlands) in a final volume of 2 ml. After 12 h of incubation at 37°C, aliquots of 0.1 ml were removed, serial dilutions were plated on Sabouraud agar, and CFU were counted after overnight incubation at 37°C. The adherence of C. albicans cells to tubes was checked and found to be less than 1% of total counts.

Determination of plasma lipids. Cholesterol and triglycerides were determined by enzymatic methods with a Hitachi 747 analyzer.

Cytokine measurements. TNF-α, IL-1α, and IL-1β concentrations were determined by specific radiommunosassays developed in our laboratory, as previously described (21).

C. albicans binding studies. C. albicans cells were radiiodinated by the iodogen method (8) with minor modifications. Briefly, 10^9 C. albicans cells in 50 mM phosphate buffer (pH 7.4) were iodinated in an iodogen-coated vial (20 μg iodine/50 μl) in the presence of 300 μCi of Na^125I (specific activity, 15 mCi/μg; Amersham) in a total volume of 100 μl. After 15 min of incubation, the suspension was washed three times with phosphate-buffered saline and resuspended in RPMI 1640 containing 1 mM pyruvate, 2 mM l-glutamine, and 100 μg of gentamicin per ml (labelling efficiency, 45%). Peritoneal macrophages from LDLR_/_ and control mice were collected as described above. Macrophages (2 x 10^5) were incubated with 10^6 CFU of radiiodinated C. albicans in a total volume of 200 μl of RPMI 1640 at 37°C in 96-well microtiter plates. After 5, 15, and 30 min, the supernatant was discarded and the radioactivity associated with adherent cells was measured with a gamma counter. Nonspecific binding was determined in wells in which C. albicans cells were incubated in the absence of macrophages. Total binding of C. albicans cells to macrophages (including both specific binding to receptors and nonspecific binding through membrane hydrophobicity or other nonspecific interactions) was determined by subtracting the binding of C. albicans cells to plastic from total counts.

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Determination of plasma lipids. Cholesterol and triglycerides were determined by enzymatic methods with a Hitachi 747 analyzer.

Statistical analysis. The survival curves for control and LDLR_/_ mice were compared by the Kaplan-Meyer log rank test. Differences in concentrations of cytokines and in organ counts of microorganisms were analyzed by the Mann-Whitney U test. Differences were considered significant at P < 0.05. All experiments were performed at least twice.

**RESULTS**

**C. albicans infection.** Plasma cholesterol concentrations decreased significantly during C. albicans infection in both mouse strains (P < 0.05) but remained three to four times higher in LDLR_/_ mice than in control mice (P < 0.01) (Table 1). The initial plasma triglyceride levels in LDLR_/_ mice were two times higher than those of controls (P < 0.01). Four hours after infection, triglyceride levels decreased significantly in LDLR_/_ mice (P < 0.05) and only marginally in control animals (P > 0.05) (Table 1). The triglyceride levels 4 h after C. albicans infection did not differ significantly between the two mouse strains (P > 0.05) (Table 1). After i.v. injection of either 10^6 or 10^7 CFU of C. albicans, LDLR_/_ mice died significantly earlier than did control animals (P < 0.05) (Fig. 1). One day after infection with 10^6 CFU of C. albicans, yeast outgrowth in the kidneys of LDLR_/_ mice was significantly higher compared with that of controls; 3 days after infection, outgrowth of C. albicans was increased in both the kidneys and livers of LDLR_/_ mice (Table 2). No difference in the outgrowth of C. albicans in the spleen was detected between the two mouse strains (Table 2).

Four hours after infection, plasma TNF-α concentrations were below the detection limit and IL-1α concentrations were higher in LDLR_/_ mice than in controls (140 ± 22 versus 78 ± 48 pg/ml, respectively; P < 0.05). Circulating IL-1β concentrations were similar in both strains (56 ± 29 versus 37 ± 28 pg/ml, respectively; P > 0.05). Plasma TNF-α concentrations were significantly higher in LDLR_/_ mice compared with those of control animals at both 24 (42 ± 5 versus 18 ± 4 pg/ml, respectively; P < 0.01) and 72 (302 ± 237 versus 98 ± 88 pg/ml, respectively; P < 0.02) h after infection (Fig. 2). No differences in plasma IL-1α and IL-1β concentrations were observed at these time points (Fig. 2).

**In vitro cytokine production.** We investigated the capacity of peritoneal macrophages of both mouse strains to produce cytokines when stimulated in vitro with heat-killed C. albicans. Compared with those of controls, the TNF-α concentrations in supernatants from macrophages of LDLR_/_ mice were significantly higher (Fig. 3a). The IL-1α and IL-1β concentrations were only marginally increased (Fig. 3a). The cell-associated IL-1α concentration was significantly increased in macrophages of LDLR_/_ mice, but the cell-associated TNF-α and

**TABLE 1. Plasma cholesterol and triglyceride concentrations before and 4 h after C. albicans infection in LDLR_/_ and C57BL/6j mice**

| Mouse strain | Time       | Mean concn (mmol/liter) ± SD
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Cholesterol</td>
</tr>
<tr>
<td>LDLR_/_</td>
<td>Before</td>
<td>9.55 ± 1.11*</td>
</tr>
<tr>
<td></td>
<td>After</td>
<td>6.76 ± 0.92**,</td>
</tr>
<tr>
<td>C57BL/6j</td>
<td>Before</td>
<td>2.25 ± 0.45</td>
</tr>
<tr>
<td></td>
<td>After</td>
<td>1.84 ± 0.90**,</td>
</tr>
</tbody>
</table>

* Mice were infected i.v. with 10^6 CFU of C. albicans. Each group consisted of five mice.

**FIG. 1. Survival during C. albicans infection. LDLR_/_ mice (closed symbols) infected with either 10^6 (triangles) or 10^7 (circles) CFU of C. albicans died significantly earlier than did C57BL/6j mice (open symbols) (P < 0.05; Kaplan-Meyer log rank test). The data are pooled results of two experiments with at least 15 animals per group.**
CANDIDIASIS AND HYPERLIPOPROTEINEMIA

TABLE 2. Outgrowth of C. albicans in the organs of LDLR\(^{-/-}\) and C57BL/6J mice after infection with 10\(^6\) CFU of C. albicans

<table>
<thead>
<tr>
<th>Mouse strain</th>
<th>Organ</th>
<th>Day</th>
<th>Log CFU/g of tissue(^a)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LDLR(^{-/-})</td>
<td>Kidney</td>
<td>1</td>
<td>6.3 ± 0.5(^*)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>7.4 ± 0.3**</td>
</tr>
<tr>
<td></td>
<td>Liver</td>
<td>1</td>
<td>4.8 ± 0.3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>5.2 ± 0.2**</td>
</tr>
<tr>
<td></td>
<td>Spleen</td>
<td>1</td>
<td>4.9 ± 0.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>5.1 ± 0.5</td>
</tr>
<tr>
<td>C57BL/6J</td>
<td>Kidney</td>
<td>1</td>
<td>5.9 ± 0.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>6.8 ± 0.3</td>
</tr>
<tr>
<td></td>
<td>Liver</td>
<td>1</td>
<td>4.8 ± 0.1</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>4.9 ± 0.1</td>
</tr>
<tr>
<td></td>
<td>Spleen</td>
<td>1</td>
<td>5.1 ± 0.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>5.0 ± 0.2</td>
</tr>
</tbody>
</table>

\(^a\) Data are means ± standard deviations of pooled data from two experiments with 10 animals per group. *, \(P < 0.01\); **, \(P < 0.05\).

Discussion

The main conclusion from the present study is that hyperlipoproteinemia has deleterious effects on the outcome of severe C. albicans infection, in contrast to gram-negative bacterial infections. We have shown that LDLR\(^{-/-}\) mice, with seven- to nine-times-higher LDL levels, are more susceptible to C. albicans infection than are their wild-type littermates. The earlier mortality of LDLR\(^{-/-}\) mice was associated with increased outgrowth of C. albicans in their organs, and these mice produced significantly more proinflammatory cytokines than did control mice.

In general, mortality after infection may be due to lethal cytokinemia or to functional impairment by the growth of microorganisms in the organs of an animal. The increased susceptibility of LDLR\(^{-/-}\) mice to C. albicans is likely to be due to the latter mechanism, because C. albicans did not induce an impressive cytokinemia. The hypothesis that cytokines are not responsible for the deaths of animals during C. albicans infection is in agreement with studies showing that the stimulation of neutrophils and macrophages by TNF and IL-1 enhanced their capability to kill C. albicans cells (6, 29), whereas anti-TNF antibodies (26) or pharmacologic inhibition of proinflammatory cytokines proved to be deleterious during severe C. albicans infection (20). Thus, proinflammatory cytokines seem to play a beneficial rather than a deleterious role in the defense against C. albicans. It should be noted that despite the greater cytokine response, LDLR\(^{-/-}\) mice were not protected against C. albicans infection, probably due to the overwhelming outgrowth of yeast cells in their organs.

It may be hypothesized that the enhanced outgrowth of C. albicans in the organs of LDLR\(^{-/-}\) mice is due to elevated lipoprotein concentrations. Normal serum has a candidicidal effect (22), and as shown by in vitro growth experiments, this property was significantly decreased in plasma samples from LDLR\(^{-/-}\) mice. This effect may be due to the use of lipoproteins as a nutrition factor by C. albicans, as has been suggested by earlier studies showing increased growth of C. albicans in lipid-containing parenteral solutions compared with that in formulations without lipid contents (5, 9, 16). Another possible mechanism by which lipoproteins could influence C. albicans growth is interaction with other plasma factors. We cannot

![FIG. 2. In vivo cytokine concentrations during C. albicans infection. Shown are circulating concentrations of proinflammatory cytokines in control (open bars) and LDLR\(^{-/-}\) (shaded bars) mice at 4, 24, and 72 h after the injection of 10\(^6\) CFU of C. albicans. *, \(P < 0.05\).](image)
exclude the possibility that plasma candidicidal factors, such as platelet microbicidal protein (32) and the calprotectin complex (19), are bound and inactivated by lipoproteins.

The higher cytokine concentrations during infection in LDLR−/− mice, compared with those of controls, were probably at least in part a response of the host against enhanced C. albicans outgrowth in the organs of LDLR−/− mice. However, surprisingly, stimulated in vitro with heat-killed C. albicans, macrophages of LDLR−/− mice produced significantly more TNF-α and IL-1α than macrophages of control mice did. Most likely, this was due to the observed increased binding of C. albicans to macrophages of LDLR−/− mice compared with that of control macrophages. This phenomenon may be explained by the influence of constitutively increased lipoprotein concentrations in LDLR−/− mice on the Candida-binding proteins on macrophage. It has been shown previously that hypercholesterolemia is able to modify the number and clustering of other receptors, such as the LPS receptor CD14 (7, 25). Earlier, we observed similar higher binding of radiolabelled LPS to macrophages of LDLR−/− mice, followed by higher cytokine production (21). Thus, similar changes in the number and/or clustering of Candida-binding proteins may facilitate the binding of C. albicans to macrophages of LDLR−/− mice, with a subsequent increase in cytokine production. Hyperlipoproteinemia could also modify the hydrophobicity of cells, which may also influence the adherence of C. albicans to macrophages, as has been shown for endothelial cells (10). Which of these mechanisms is responsible for the observed increase in cytokine production by macrophages of LDLR−/− mice is under study.

An alternative desirable experiment to our model in order to investigate the influence of hyperlipoproteinemia in C. albicans infection would have been to infuse lipoproteins into animals before and during infection. However, an infusion of lipoproteins into mice is not possible and other models of lipoprotein infusion in rabbits (11) and rats (24) are short-term models that are not suitable for sustained lipid infusion during systemic candidiasis. Therefore, the genetically modified mouse model is a good alternative for studying the in vivo effects of hyperlipoproteinemia in models of sustained infection.

In conclusion, hyperlipoproteinemia has deleterious effects on the course of acute disseminated C. albicans infection, in contrast to its beneficial effect in gram-negative infection. Although no epidemiological studies have been done to show a relationship between hyperlipoproteinemia and increased susceptibility to C. albicans, an infusion of lipoproteins into a patient with disseminated candidiasis under the presumptive diagnosis of gram-negative sepsis may prove deleterious. These divergent effects of hyperlipoproteinemia should be taken into account when the use of lipoproteins as an adjuvant treatment of sepsis is considered.

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