RESEARCH ARTICLE

Stress in Atlantic salmon: response to unpredictable chronic stress

Angelico Madaro1,*, Rolf E. Olsen1,2, Tore S. Kristiansen1, Lars O. E. Ebbesson3, Tom O. Nilsen3, Gert Flik4 and Marnix Gorissen4

ABSTRACT

Combinations of stressors occur regularly throughout an animal’s life, especially in agriculture and aquaculture settings. If an animal fails to acclimate to these stressors, stress becomes chronic, and a condition of allostatic overload arises with negative results for animal welfare. In the current study, we describe effects of exposing Atlantic salmon parr to an unpredictable chronic stressor (UCS) paradigm for 3 weeks. The paradigm involves exposure of fish to seven unpredictable stressors three times a day. At the end of the trial, experimental and control fish were challenged with yet another novel stressor and sampled before and 1 h after that challenge. Plasma cortisol decreased steadily over time in stressed fish, indicative of exhaustion of the endocrine stress axis. This was confirmed by a lower cortisol response to the novel stressor at the end of the stress period in chronically stressed fish compared with the control group. In the preoptic area (POA) and pituitary gland, chronic stress resulted in decreased gene expression of 11β-hsd2, gr1 and gr2 in the POA and increased expression of those genes in the pituitary gland. POA crf expression and pituitary expression of pomcs and mr increased, whereas interrenal gene expression was unaffected. Exposure to the novel stressor had no effect on POA and interrenal gene expression. In the pituitary, crfr1, pomcs, 11β-hsd2, grs and mr were down-regulated. In summary, our results provide a novel overview of the dynamic changes that occur at every level of the hypothalamic-pituitary-gland–interrenal gland (HPI) axis as a result of chronic stress in Atlantic salmon.

KEY WORDS: Salmo salar, Parr, Cortisol, HPI axis, crf, Gene study, Brain, Hypothalamus, Pituitary gland, Head kidney

INTRODUCTION

The intensity, duration, predictability and controllability of a stressor are important aspects to assess the severity of a stressor. In addition, stressors seldom come alone, adding a complex extra dimension to the definition of stress intensity. As opposed to wild fish, animals under farming conditions are confined and cannot escape from stressors. In aquaculture, fish are exposed to several simultaneous stressors. Examples include suboptimal/poor water quality, repeated handling, transport and crowding. The effects of many of these challenges have been studied, albeit mostly as a single stressor (Barton and Peter, 1982; Gorissen et al., 2012; Di Marco et al., 2008; Pottinger, 2010; Remen et al., 2012). Studies on mammals (Aguilera and Rabadan-Diehl, 2000; Dhabhar and McEwen, 1997; Grissom and Bhatnagar, 2009; Thorsell et al., 1999), as well as fish (Schreck, 2000), show they have great resilience to a single stressor given repeatedly over long periods of time; however, knowledge on how ectotherms respond to multiple persistent stressors is very scant. One study on Atlantic salmon (Salmo salar, L.) has shown that chronic stress followed by an additional maze challenge resulted in a suppressed cortisol response, and decreased neural plasticity and learning ability (Grassie et al., 2013). In this study, we aim to describe the effects of unpredictable chronic stress (UCS) on the stress axis of Atlantic salmon.

When a stressor is perceived, neuronal signals (visual, auditory and sensory) activate the hypothalamus and initiate a downstream activation of sympathetic fibres that in turn stimulate the chromaffin cells of the head kidney to release the catecholamines adrenaline and noradrenaline into the bloodstream as the initial stress response. The catecholamines prepare the animal for fight or flight by increasing gluconeogenesis and glycolysis, lipid degradation, etc. Secondly, the hypothalamic-pituitary gland–interrenal gland (HPI) axis (the equivalent of the hypothalamic-pituitary gland–adrenal axis in mammals) becomes activated (Arends et al., 1999; Wendelaar Bonga, 1997). In the hypothalamic preoptic area (POA), corticotropin-releasing factor (CRF) is released to activate the pituitary corticotropes (Huising et al., 2004). CRF, via its receptor CRF-R1, induces the synthesis of pro-opiomelanocortin (POMC) which is processed into adrenocorticotrophic hormone (ACTH) and released into the bloodstream (Sumpter et al., 1986). In the interrenal gland, ACTH induces the synthesis and release of cortisol via the melanocortin 2 receptor (MC2R) (Wendelaar Bonga, 1997), which is expressed exclusively on cortisol-producing cells therein. MC2R activation results in the activation of steroidogenic acute regulatory protein (STAR), which is responsible for the transport of cholesterol into the mitochondrial membrane, where it will be converted to corticosteroids including cortisol.

Cortisol acts as glucocorticoid and mineralocorticoid in teleosts as these animals do not produce aldosterone synthase (Wendelaar Bonga, 1997). Specific actions of cortisol are effected by receptor specificity, i.e. the mineralocorticoid (MR) and glucocorticoid receptor (GR) profile of the target cell. MR and GR are transcription factors, mediating activation or inhibition of target gene expression. They are also involved in the negative feedback regulation of the HPI axis at the level of the hypothalamus and pituitary gland (Atkinson et al., 2008; Bury et al., 2003; Cole et al., 2000). Cortisol also exerts negative feedback on hypothalamic CRF synthesis (Bernier and Peter, 2001; Bernier et al., 1999) and ACTH secretion by the pituitary gland (Fryer et al., 1984), whereas 11β-hydroxysteroid dehydrogenase 2 (11β-HSD2) counteracts cortisol actions by converting cortisol into the inactive cortisone (Mommsen et al., 1999). CRF binding protein (CRF-BP) provides yet another way of controlling the HPI axis. It modulates the effect of CRF and CRF-related peptides by binding these peptides and reducing their bioavailability (Geven et al., 2006; Huising et al., 2008; Emanuel et al., 2014; Seasholtz et al., 2002) with a decreased release of ACTH as a result.
In the short term, the stress response works to the benefit of the animal: faced with a danger or challenge, the consorted action of the catecholamines and cortisol increases heart rate and blood flow to the muscles and increases processes aimed at providing more energy. Less vital bodily functions including growth, the immune system, reproduction and digestion are put on hold. If this situation persists, and the balance is not restored, the animal's condition changes from eustress to distress. Distress reflects the situation in which the response is inadequate and maladaptive (Kupriyanov and Zhdanov, 2014; Wendelaar Bonga, 1997).

Allostasis, or maintenance of stability through change (Sterling and Eyer, 1988), involves resetting of physiological and behavioural set points of regulatory mechanisms to optimize performance in accordance with predicted environmental demands at minimal costs (McEwen and Lasley, 2002). Low allostatic load, or eustress, can have a positive effect on the animal’s performance (Kupriyanov and Zhdanov, 2014). In contrast, long-lasting and/or repeated stressors result in a chronic stress condition in which a lasting initial stress response proves to be inadequate and the alarm signals become deleterious (allostatic overload/distress) (McEwen and Seeman, 1999).

Much stress research has addressed isolated hypothalamicus tissue, pituitary gland or interrenal gland, and, to the best of our knowledge, only Fuzzen and colleagues (2010) have conducted a study on zebrafish that covers all levels of the stress axis. Although effects of single, acute stressors have been studied in Atlantic salmon (POMC, POMC1, POMC2, MC2R, CRF1, CRF2), long-lasting and/or repeated stressors have a positive effect on the animal's performance (Kupriyanov and Zhdanov, 2014; Wendelaar Bonga, 1997).

The main objective of this study was to determine how freshwater Atlantic salmon parr respond to unpredictable chronic stress. Stressors were selected based on what salmon may experience in aquaculture. We measured food intake and growth throughout the experiment. Plasma cortisol levels were analysed at regular intervals throughout the experiment. After 3 weeks of UCS, all fish were subjected to yet another, novel stressor and sampled before and 1 h after this novel stressor. Total mRNA levels of crf, crfhp, 11βhsd2, gr1, gr2 and mr in the POA, crfr1, pomca1 and pomcb1, 11βhsd2, gr1, gr2 and mr in the pituitary gland and mc2r, star, 11βhsd2, gr1, gr2 and mr in interrenal tissue were analysed at both these time points.

### RESULTS

#### Effect of UCS on feeding and growth

After 23 days, control fish had grown by 41% from 63.6±1.2 g at the start of the study to 89.5±2.4 g; the chronically stressed (hereafter, UCS) fish had grown by 8.5% to 69.0±1.4 g (F_{2,173}=54.2, P=0.001; Fig. 1). This represents a daily growth rate of 1.42% and 0.53%, respectively (t=5.433, df.=4, P=0.0056; Fig. 2). The reduced growth rate in the UCS group corresponds with a reduced appetite throughout the study (Fig. 3). Fish were starved for 12 h before sampling days (indicated by arrows in Fig. 3). On the following day, a compensatory food intake was observed in all groups.

#### Plasma cortisol

Throughout the experiment, plasma cortisol levels were generally elevated in the stressed groups, but the amplitude of the response decreased towards the end of the study (interaction: F_{3,160}=3.496, P=0.005; Fig. 4). On day 5, cortisol production in the UCS group following chosing was not significantly different compared with controls.

There was a significant interaction effect (UCS×acute stressor: F_{1,1111}=15.61, P=0.0001) on cortisol levels after 23 days. Cortisol increased in both groups after the stress test, but less so for UCS fish (91.26±25.05 ng ml\(^{-1}\)) than for control fish (163.3±21.98 ng ml\(^{-1}\); Fig. 5).

#### Gene study

POA

Fig. 6 shows the transcript abundance of selected genes in the POA of control and UCS fish before and 1 h following exposure to a novel stressor on day 23. A significant interaction (UCS×acute stressor) was observed in crf expression (F_{1,103}=4.830, P=0.03).

---

**List of symbols and abbreviations**

- **11βHSD2**: 11-β-hydroxysteroid dehydrogenase 2
- **20S**: 20S ribosomal protein
- **ACTH**: adrenocorticotropic hormone
- **CRF**: corticotropin-releasing factor
- **CRFBP**: corticotropin-releasing factor binding protein
- **E**: entopenduncular nucleus
- **EF1α**: elongation factor 1α
- **GR**: glucocorticoid receptor
- **HPA**: hypothalamic-pituitary gland–interrenal gland
- **MC2R**: melanocortin 2 receptor
- **MR**: mineralocorticoid receptor
- **MSH**: melanocyte-stimulating hormone
- **POA**: preoptic area
- **PM**: preopticus magnocellularis
- **POMC**: pro-opiomelanocortin
- **PPα**: preopticus parvocellularis pars anterior
- **SGR**: specific growth rate
- **STAR**: steroidogenic acute regulatory protein
- **UCS**: unpredictable chronic stress

---

**Fig. 1.** Body mass of Atlantic salmon parr fed *ad libitum*, at the beginning and at the end of the experiment. Bars represent means±s.e.m.; different letters indicate a significant difference (one-way ANOVA followed by Sidak’s *post hoc* test; P<0.025). Number of fish: N=45, 60 and 71 for control day 0, after 23 days in optimal breeding conditions and after 23 days under the effect of unpredictable chronic stress (UCS), respectively.

**Fig. 2.** Specific growth rate of salmon. Specific growth rate (SGR) is expressed as a percentage of body mass gain per day (g day\(^{-1}\)) of salmon parr fed *ad libitum* (control) and of fish exposed to UCS for 23 days. Values represent means±s.e.m. (N=3; Mann–Whitney *U*-test, P<0.05).
Fish subjected to 23 days of chronic stress showed elevated basal levels of crf mRNA (Fig. 6A); 1 h after exposure to the novel stressor, expression levels of crf in control and UCS fish had not changed. A significant interaction effect was also observed for crfbp (UCS×acute stressor: F1,105=4.723, P=0.032). There were no differences in basal transcript abundance for crfbp between control and UCS fish. However, 1 h after exposure to the acute stress, crfbp expression was significantly higher in the control group compared with the chronically stressed group (Fig. 6B). mRNA for H1βhsd2 (Fig. 6C) and the glucocorticoid receptors gr1 and gr2 (Fig. 6D, E) showed similar patterns: a lower amount of transcript in UCS fish compared with controls but no effect of the novel stressor. An interaction was seen in mineralocorticoid receptor (mr) mRNA levels (UCS×acute stressor: F1,103=17.9, P<0.001): 23 days of chronic stress had no effect on basal mr transcript abundance, whereas the acute challenge increased mr mRNA in control fish but not in UCS fish (Fig. 6F).

In situ hybridisation
Transcript abundance of crf mRNA in the nucleus preopticus parvocellularis pars anterior (PPa) and the nucleus preopticus magnocellularis (PM) was visibly more pronounced in UCS fish (Fig. 7C, D) compared with control (Fig. 7A, B). The images shown are from a single fish per treatment.

**Discussion**
We characterised the stress response of Atlantic salmon parr after exposing them to UCS for 3 weeks. We made several major observations: (1) the UCS resulted in suppression of food intake and growth; (2) the cortisol response abated over the 3 weeks of UCS.
and the cortisol response to a novel stressor was more pronounced in control fish than in UCS fish; (3) in the POA, basal transcript abundance of the genes involved in negative feedback of the stress-axis decreased in UCS fish, whereas the novel stressor resulted in limited effects in both control and UCS fish; (4) genes involved in negative feedback in the pituitary gland increased in UCS fish compared with controls – a novel stressor increased expression of *pomc* paralogues in the control group and decreased transcript abundance in the UCS fish; and (5) the novel stressor increased the transcript abundance of *mc2r* and *11βhsd2* transcripts in the UCS fish, while no differences were found for the other genes studied.

As a general remark, we would like to stress that we do not – as yet – know how changes in the expression of genes, as measured by mRNA, are representative of the levels of proteins for which they encode. That means that our data should be interpreted with care (Maier et al., 2009; Schwanhäusser et al., 2011).

**Growth and plasma cortisol**

The UCS approach appeared to induce chronic stress as both growth and food intake were reduced over the 3 weeks of the study. To the best of our knowledge, there are no similar studies on the effect of UCS on fish appetite and growth, although our results are in line with other studies where repeated stress reduced appetite and growth (McCormick et al., 1998).

We also observed a chronic down-regulation of the cortisol response during the trial. However, on day 5 of the experiment, the UCS group displayed a remarkable low cortisol production post-stress. Perhaps the UCS together with the high frequency of samplings during the first days of the experiment resulted in an exhaustion of the HPI axis and thus a failure to mount a proper cortisol response following stress. Further, a lowered cortisol response was measured at the end of the trial when the fish were exposed to a novel stressor. These observations are in line with the general down-regulation of the HPI axis that is a common feature during chronic stress and stress adaptation in fish and mammals (Barton, 2002; Barton et al., 1987; Vijayan and Leatherland, 1990).

**POA**

Exposing Atlantic salmon to UCS resulted in a general down-regulation of the examined genes in the POA except for *crf*, which was up-regulated. The *crf* up-regulation measured by qPCR was also validated by *in situ* hybridisation with more pronounced staining of hypothalamic *crf* mRNA in the PPa and the PM in UCS fish compared with controls. This increase in *crf* seems at odds with
the abated cortisol response, but the high levels of crf, together with the high cortisol levels compared with baseline cortisol, may explain the reduction of appetite that we observed in the UCS fish, as CRF and cortisol are known suppressors of appetite in fish (Bernier and Peter, 2001; Gregory and Wood, 1999).

POA crfbp transcript abundance was not affected by chronic stress. Studies on trout (Alderman et al., 2008; Doyon et al., 2005) reported that CRFBP may play a role in the inhibition of CRF signalling at its receptor in the acute stress response and this mechanism was confirmed by in vitro studies (Manuel et al., 2014). The effect of chronic stress in our study is in line with the findings of another experiment on rainbow trout (Jeffrey et al., 2012) where crfbp mRNA was unaffected by chronic stress caused by social interactions.

During chronic stress, HPI axis down-regulation (Barton, 2002; Barton et al., 1987; Vijayan and Leatherland, 1990) in salmonids and in other teleostean fish results from negative feedback mediated by the two glucocorticoid receptors, GR1 and GR2, of which GR2 has an affinity for cortisol that is similar to/higher than that of MR (Bury et al., 2003; Greenwood et al., 2003; Prunet et al., 2006; Stolte et al., 2008). In the present study, UCS induced a down-regulation of both gr mRNAs in the POA while mr transcript amount was unaffected. A reduced expression of gr1 and gr2 and an increase in crf mRNA is in line with previous observations in zebrafish (Chakravarty et al., 2013; Piato et al., 2011) and trout (Doyon et al., 2005). Low levels of gr1 and gr2 may protect neurones from high levels of cortisol during chronic stress. GR-containing cells are more prone to apoptosis by elevated levels of glucocorticoids (Aluru and Vijayan, 2006; Piato et al., 2011; Sapolsky et al., 2000). In contrast, 11βhsd2 was down-regulated as a result of UCS. This enzyme catalyses the degradation of cortisol to cortisone, thus regulating cortisol availability to corticoid receptors (Funder et al., 1988). The lower 11βhsd2 transcript level may result in higher cortisol availability; we interpret these results combined as a search for a new balance in cortisol action under chronic stress conditions. Promotor analysis in zebrafish revealed putative sites for cortisol-mediated regulation of transcription of 11βhsd2 and (chemical) inhibition of 11βhsd2 activity resulted in increased POA crf expression and elevated cortisol levels (Alderman and Vijayan, 2012). Taken together, these findings indicate a pivotal role for 11βhsd2 in negative feedback control of the stress axis.

The final stress test had no major influences on gene expression in the POA; only in control fish was a rapid up-regulation of mr seen. Possibly, MR is the main corticoid receptor responsible for the mediation of cortisol action in the POA in Atlantic salmon after acute stress. In mammals, corticosteroid effects mediated by MRs are involved in suppression of basal and stress-induced ACTH secretion (de Kloet et al., 2005). Studies in rats support the idea that even when MRs are occupied in resting conditions by basal levels of corticosteroids, they still have important roles in the sensitivity and feedback responses when high levels of corticosteroids are present (Ratka et al., 1989; Dallman, 1993; Oitzl et al., 1994). In contrast,
the UCS group displayed no effects on mr transcript abundance despite a persistent gr downregulation. De Kloet (1991) hypothesised that an increased amount of MRs relative to GRs may be predictive for a reduced responsiveness of the HPI axis to stress. If this holds for fish, POA should be tested by measurement of the number of receptor proteins. Furthermore, additional regulatory mechanisms may be present in other tissues. For example, Stolte and colleagues (2008) showed that carp, when repeatedly given a temperature shock, displayed a down-regulation of the GRs in the brain but not in the POA and pituitary gland, suggesting a central initiation by telencephalic pallial areas homologous to the mammalian hippocampus, amygdala and prefrontal cortex (Mueller, 2012; Mueller et al., 2011) of stress axis control, rather than direct feedback via the POA or pituitary gland.

**Pituitary gland**

We analysed the transcript abundance of pomca1 and pomcb1, paralogues that originate from the salmonid genome duplication (Arends et al., 1998; Leder and Silverstein, 2006; Macqueen and Johnston, 2014; Salbert et al., 1992). No changes in pomca1 and pomcb1 mRNA transcripts were observed when fish were subjected to either UCS or the novel stressor. In the pituitary gland, pomc is expressed in both the corticotrope cells (pars distalis) and the melanotrope cells (pars intermedia). In the pars distalis, POMC is processed by prohormone convertase 1 to ACTH and β-endorphin.
whereas in the pars intermedia POMC, through an additional action of prohormone convertase 2, POMC is processed into α-MSH (melanocyte-stimulating hormone) and β-endorphin (Takei and Loretz, 2006). We isolated the mRNA of the entire pituitary gland. Therefore, we are unable to discern where in the pituitary gland the measured pomer was produced. Both pomer paralogues possess the required dibasic cleavage sites to produce ACTH and α-MSH.

Plasma concentrations of α-MSH (data not shown) were not affected by UCS treatment or by exposure to the novel stressor. Unfortunately, ACTH was not analysed as a reliable assay for plasma ACTH in salmon is lacking. In our study, we observed an increase of poma1 and pomb1 mRNA in UCS fish compared with controls: in line with these results, in a trout study, Winberg and Lepage (1998) found increased pomer expression levels as a result of chronic stress induced by social interactions. In our study, the additional novel stressor resulted in increased poma1 and pomb1 expression in control fish but not in the UCS fish, which is corroborated by the elevated plasma cortisol levels.

In contrast to what we observed in POA tissue, all examined genes involved in negative feedback (gr1, gr2, mr and 11βhsd2) were up-regulated in the pituitary gland following chronic stress. However, the capacity to metabolise cortisol into the inactive cortisone may have increased as indicated by the enhanced expression of 11βhsd2 and taken together this could indicate an enhanced turnover of cortisol under stress conditions. Studies on transgenic mice homozygous for a targeted disruption of the gr gene (Cole et al., 1995) and for a mutation of the dimerisation loop domain in GR (Reichardt et al., 1998) suggest that the down-regulation of Crf expression by negative feedback triggered by glucocorticoids is GR–DNA binding independent, whereas Pomc transcription is under negative control by GR–DNA binding (Newton, 2000). In our study, we observed a decrease in POA and an increase in pituitary gland gr transcripts of chronically stressed fish, pointing to a stronger feedback control over the pituitary gland.

The effects of the acute novel stressor on the pituitary gland were significantly different in control and chronically stressed fish. In control fish, the novel stressor did not affect crfr1, while pomer mRNA increased, in agreement with studies in zebrafish (Fuzzen et al., 2010) and trout (Gilchrist et al., 2000). We underline that in
In the present study leads to two different lack of effect following the acute and chronic stress in both the groups, respectively, has been observed (Aluru and Vijayan, 2008). The UCS treatment of salmon parr. Noteworthy, in trout, an up-regulation of genes involved in cortisol production and release (mc2r and star), inactivation (11βhsd2) or cortisol receptors (gr1, gr2 and mr) after the novel stressor results in a lower responsiveness (over-) capacity of protein in cortisol production. Alternatively, transcript abundance of mc2r and 11βhsd2 in control fish could have been depleted post-stress as a result of the induction of protein production, whereas in the UCS group, transcription of these genes was already activated in order to support steroidogenesis in response to the UCS. Additional studies should clear the doubts about the time course and dynamics of stress axis-related gene expression in the head kidney.

Conclusions and perspectives

Whereas the acute stress response of fish has been extensively reviewed (Barton, 2002; Wendelaar Bonga, 1997), the effects of long-term stress on fish remain relatively unexplored. The purpose of the present study was to show the effects of UCS on Atlantic salmon parr and explore the mechanisms that drive and control the HPI axis under these conditions. We established a new UCS protocol, based on multiple stressors, applied randomly, to avoid habituation. In these conditions, the fish could experience allostatic overload or ‘wear and tear’ of the body if they cannot attenuate the mediators involved in the stress response (McEwen, 1998; McEwen and Stellar, 1993). We also examined genes relevant to all levels of the HPI axis before and after a novel acute stressor in both chronically stressed fish and controls. As predicted, gene expression data from the POA and pituitary gland demonstrate that control fish respond readily to an acute stressor (Wendelaar Bonga, 1997). In contrast, UCS fish were characterised by a dampened cortisol response that correlates with changes in stress axis-related gene expression at the level of the POA and pituitary gland. We conclude that, despite an increased crf expression in the POA, UCS treatment reduced the pituitary capacity to mount a proper stress response. Before the novel stressor, the expression of genes involved in the feedback systems as well as the initiation of interrenal cortisol production increased. After the novel stressor, mRNA levels of the stress axis regulators in the central part of the HPI axis were depleted. The interval between stress events is critical for the physiological response (Schreck, 2000). Therefore, a higher frequency of UCS episodes may reduce the time available for the fish to cope with the stressors: in such a situation, the effect of each stress episode may become cumulative and push the fish tolerance capabilities until disease and death occur.

### Table 1. Description of the stressors randomly given to Atlantic salmon parr throughout the experiment

<table>
<thead>
<tr>
<th>Stressor</th>
<th>Time (min)</th>
<th>Description</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chasing</td>
<td>5</td>
<td>Stirring in the tank with a net</td>
<td>Pavlidis et al., 2015; Tsalafouta et al., 2014</td>
</tr>
<tr>
<td>Netting</td>
<td>3</td>
<td>Net and release fish with a dip net including brief air exposure (±1 s)</td>
<td>Barton et al., 1980</td>
</tr>
<tr>
<td>Temperature shock 12°C to 4°C</td>
<td>120</td>
<td>Reduction of the water temperature from 12°C to 4°C and up to 12°C</td>
<td>Foss et al., 2012</td>
</tr>
<tr>
<td>Temperature shock 12°C to 19°C</td>
<td>120</td>
<td>Rise of the water temperature from 12°C to 19°C and down to 12°C</td>
<td>Templeman et al., 2014</td>
</tr>
<tr>
<td>Noise</td>
<td>5</td>
<td>Knocking on the tank with a metal object</td>
<td>Slabbekoorn, 2012</td>
</tr>
<tr>
<td>Darkness+flash light</td>
<td>5</td>
<td>Turn off the tank lighting and use of a white intermittent led light</td>
<td>Remen et al., 2012</td>
</tr>
<tr>
<td>Brief hypoxia</td>
<td>5</td>
<td>Closure of water inflow until the oxygen saturation of the water reaches 40%</td>
<td></td>
</tr>
<tr>
<td>Emptying the tank</td>
<td>5</td>
<td>Removal of the tank plug while leaving the water flow open with a constant 3 cm deep layer of water as a result</td>
<td>Einarsdóttir and Nilssen, 1996</td>
</tr>
</tbody>
</table>

Stressors were given three times per day, for 23 days.
The Atlantic salmon (S. salar, AquaGen strain) proved to be a fish with a high tolerance to multiple, unpredictable stressors. The procedure did not harm the fish physically and no fish died during the experiment. Accordingly, Solberg et al. (2013) showed that farmed salmon are characterised by a reduced responsiveness to stress compared with wild strains. Solberg and colleagues suggest that, during the last four decades of intensive salmon breeding programmes, the selection of individuals with increased growth rate in captivity environments is likely to have involuntarily led to the selection of fish with reduced stress responsiveness and/or higher resilience, as indeed individuals affected by stress would display impaired growth rates and therefore would not be selected as brood stock for the next generation. A comparative study on the effects of the UCS protocol on the AquaGen strain and wild salmon seems indicated.

**MATERIALS AND METHODS**

**Fish and experimental facilities**

Atlantic salmon (S. salar, AquaGen strain) eggs obtained from a commercial farm (Aqua Gen AS, Trondheim, Norway) were hatched (March 2012) and reared at the Institute of Marine Research (IMR), Matre, Norway. Experimental fish were kept in freshwater with light and temperature according to natural winter conditions (12 h light:12 h dark, 9°C). On 11 January 2013, 740 fish (average mass 57 g) were transferred to a commercial farm (Aqua Gen AS, Trondheim, Norway) for the study on the effects of the UCS protocol on the AquaGen strain and wild salmon seems indicated.

**Experimental design**

At the beginning of the experiment (4 February 2013), the six tanks were divided into two groups of three tanks (replicates) each, receiving UCS or left undisturbed (control group). The UCS group was stressed three times per day (at 08:30 h, 13:00 h and 17:00 h) using a total of eight types of stressors given in random and unpredictable order throughout 1 week, and this protocol was then repeated over the next 2 weeks (Table 1). All stressors were chosen such that there would be no physical damage to the fish (such as scale loss or fin damage). Disturbance for the control group was reduced to a minimum and limited to routine practices of tank maintenance and sampling. Fish were fed with dry feed for the duration of 1 h (2 mm Skretting Nutra Olimpic, Stavanger, Norway) 30–60 min after each stress event (i.e. at 09:00 h–10:00 h, 13:30 h–14:30 h and 17:30 h–18:30 h). To study the fish stress response during the experiment, on days 1, 2, 5, 9, 16 and 23, the first stressor of each day was 5 min chasing stress, which was followed by sampling of 5 fish per tank (N=15) 1 h later. Undisturbed fish from the control tanks served as controls (5 fish per tank, N=15). We chose to use the same stressor before these samplings in order to avoid confounding effects of different stressor intensities.

On the last day of the experiment (day 23), 10 fish per tank (N=30) were collected from both control and stressed groups before stress (T0) to assess basal cortisol levels and gene expression, while two extra fish per tank (N=6) were collected for in situ hybridisation analysis. Immediately thereafter, another 10 fish per tank (N=30) were collected and exposed to a novel stressor which consisted of netting, air exposure for 15 s and confinement in a 10 l bucket for 5 min before being transferred to a new tank (T1) for recovery for 1 h, after which the fish were sampled for analysis (T2). All experiments were approved by the Norwegian Animal Experiment Committee (Forsøksdyruttervalget, 11.12.2012).

**Sampling**

Fish were starved for 12 h before sampling. Fish received an overdose of anaesthetics (100 mg l⁻¹ Finquel®vet) buffered with 100 mg l⁻¹ sodium bicarbonate (Finquel®vet). Fork length and body mass were recorded for each individual fish. Blood was collected using 1 ml heparinised syringes fitted with a 23 G needle and plasma was separated immediately by centrifugation at 13,000 rpm for 3 min and stored at −80°C until cortisol analysis.

**Table 2. Primer sequences used in the real-time qPCR assays**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequences (5′→3′)</th>
<th>Amplicon (bp)</th>
<th>Accession no.</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>ef1a</td>
<td>F CCCCTCCAGAGCTTACAAA</td>
<td>57</td>
<td>AF321836</td>
<td>Olsvik et al., 2005</td>
</tr>
<tr>
<td></td>
<td>R CACAGCGCCCCACAGGTA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>20S</td>
<td>F GCAGACGCTTACCGGAGGCTA</td>
<td>85</td>
<td>BG93672</td>
<td>Olsvik et al., 2005</td>
</tr>
<tr>
<td></td>
<td>R TGGTATGGCCAGTCTCTCTCT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>crf</td>
<td>F AACCGACCCAGACCACTGTG</td>
<td>135</td>
<td>BT057824</td>
<td>L.O.E.E., unpublished sequence</td>
</tr>
<tr>
<td></td>
<td>R TCTATGGCCCTTACGCTATG</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>crf5p</td>
<td>F TGGCCCAACAGGCTATCAAG</td>
<td>81</td>
<td>BT059529</td>
<td>Leong et al., 2010</td>
</tr>
<tr>
<td></td>
<td>R TCCCTTCTACCCAGGACCAT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>crf1</td>
<td>F TGACCATCCTGGCCTCTAGCT</td>
<td>81</td>
<td>GBRB01035702</td>
<td>L.O.E.E., unpublished sequence</td>
</tr>
<tr>
<td></td>
<td>R TAAGATTTGCTGGACAGGAGCA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>pomca1</td>
<td>F CCAATCCAGCAGTGAGGAC</td>
<td>112</td>
<td>NM_001198575.1</td>
<td>Leong et al., 2014 direct submission</td>
</tr>
<tr>
<td></td>
<td>R CATTGTGGCTCAGAGGAGCA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>pomcb1</td>
<td>F CAATCCCACAGGCCCCCTGTAT</td>
<td>75</td>
<td>NM_001128604.1</td>
<td>Designed by authors</td>
</tr>
<tr>
<td>gr1</td>
<td>F ATGGCTTTTCGAGGAGGATAG</td>
<td>106</td>
<td>AF209873.1</td>
<td>Kiliarich et al., 2007</td>
</tr>
<tr>
<td>gr2</td>
<td>F TGTTGGCTCTGAGTTTCTGC</td>
<td>240</td>
<td>AY495372.1</td>
<td>Kiliarich et al., 2011</td>
</tr>
<tr>
<td>Mr</td>
<td>F TCCTTCACAGCAAAAGTTG</td>
<td>148</td>
<td>AF209873</td>
<td>Designed by authors</td>
</tr>
<tr>
<td></td>
<td>R TCTTCGCCGCACACAGGATAG</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>11thsd1</td>
<td>F GCTGGTCTACTCTGGCCA</td>
<td>74</td>
<td>G934620</td>
<td>Kiliarich et al., 2007</td>
</tr>
<tr>
<td>Star</td>
<td>F AGGATGGATGGACACCTGAG</td>
<td>163</td>
<td>DQ415678.1</td>
<td>Anukwe, 2005</td>
</tr>
<tr>
<td>mc2r</td>
<td>F AACCTCCACCTCCCCGATGTA</td>
<td>95</td>
<td>NM_001124680.1</td>
<td>Designed by authors</td>
</tr>
<tr>
<td></td>
<td>R ACCATCACAGGCTCTCCCA</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
analysis. Pituitary glands, brains and head kidneys were collected and stored in RNAlater (RNAter® RNA Stabilization Solution, Life Technology, Oslo, Norway) at 4°C for one night and subsequently stored at −80°C until RNA isolation. The sex of the fish was recorded. POAs were isolated from brains immediately before RNA purification (Bernier et al., 2008).

**Food consumption and growth rate**

Food uneaten after 15 min was collected from each tank after each feeding and left to dry in a colander for 10 min before the mass was recorded. Food consumption was calculated following the method described by Helland and colleagues (1996). The specific growth rate (SGR, % body mass gain day−1) was calculated using Eqn 1:

$$\text{SGR} = \frac{\left( \log M_2 - \log M_1 \right)}{(t_2 - t_1)} \times 100,$$

in which $M_1$ is the bulk mass at the start of the growth period ($t_1$) and $M_2$ the bulk mass at the end ($t_2$) (Houde and Schekter, 1981). Both food consumption and final SGR were calculated per tank ($N=3$).

**Blood analyses**

Plasma cortisol concentrations were quantified by radioimmunoassay (Gorissen et al., 2012) using a highly specific and sensitive commercially available antibody (xm210 cortisol antibody, Abcam, Cambridge, UK) and 11−H-cortisol (Perkin Elmer, Groningen, The Netherlands). The primary antibody shows a 100% cross-reactivity with cortisol, 0.9% with 11−deoxycorticisol, 0.6% with corticosterone and <0.01% with 11−deoxycorticosterone, progesterone, 17-hydroxyprogesterone, testosterone and oestradiol. Inter- and intra-assay variation was 12.5% and 3.5%, respectively.

**Gene expression analysis**

POA, pituitary and head kidney total RNA was isolated with Trizol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer’s instructions and then dissolved in DEPC-treated H2O. RNA concentration and purity were determined by spectrophotometry (Nanodrop, Wilmington, DE, USA). A 500 ng sample of total mRNA was incubated for 15 min at room temperature with 1 μl (1 U) DNase I (Invitrogen) in a volume of 10 μl to remove possible genomic DNA contamination. DNase was inactivated by the addition of 1 μl 15 mmol l−1 EDTA and incubation at 65°C for 10 min. To each sample, 250 ng random hexamer primers, 4 μl 5×First Strand buffer, 1 μl (10 mmol l−1) dNTP mix, 1 μl 0.1 mol l−1 DTT, 1 μl RNase OUT (10 μl μl−1) and 100 U of SuperscriptII Reverse Transcriptase (200 U μl−1; all from Invitrogen) were added and incubated for 10 min at 25°C, 50 min at 42°C and 15 min at 70°C for the synthesis of first strand cDNA. The cDNA was diluted 5 times and stored at −20°C until measurement of the relative mRNA levels by real-time quantitative PCR (RT-qPCR). Each RT-qPCR reaction consisted of 4 μl template and 10 μl SYBR Green Mastermix (BioRad, Hercules, CA, USA), 0.7 μl (10 μmol l−1) forward primers, 0.7 μl (10 μmol l−1) reverse primers and 4.6 μl DEPC-treated H2O. RT-qPCR (3 min at 95°C, 40 cycles of 15 s at 95°C and 1 min at 60°C) was carried out on a CFX96 Touch™ real-time PCR detection system (BioRad). The relative gene expression was analysed according to Vandesompele et al. (2002) and normalised to an index of two reference genes: 20S ribosomal protein (20S) and elongation factor 1a (ef1a). Sequences of primers used for the qPCR analysis are shown in Table 2.

**crf mRNA in situ hybridisation**

*In situ* hybridisation of *crf* mRNA was performed according to Ebbesson et al. (2011). In brief, three juvenile Atlantic salmon from control and stressed groups each were deeply anaesthetised with buffered Finquel® vet. and fixed by vascular perfusion with 4% paraformaldehyde in 0.1 mol l−1 Sorensens phosphate buffer (PB; 28 mmol l−1 NaH2PO4, 71 mmol l−1 Na2HPO4, pH 7.2). The brains were dissected out and post-fixed in the same fixative for 16 h at 4°C. The tissue was washed 3×20 min in PB, cryopreserved overnight in 25% sucrose in PB at 4°C, embedded in Tissue-Tek O.C.T. Compound (Sakura Fintek, Zoeterwoude, Netherlands) and stored at −80°C until sectioning. Adjacent transversal 12 μm sections were cut using a Leica CM 1850 cryostat, collected on SuperFrost Ultra Plus glasses (Menzel Glaser, Braunschweig, Germany) and dried at 60°C for 10 min. Digoxygenin labelled riboprobes were prepared using a digoxigenin (DIG)-RNA labelling mix according to the manufacturer’s instructions (Roche Diagnostics, Mannheim, Germany). The *in situ* hybridisation probe was 469 nucleotides long with a high homology to Atlantic salmon *crf I* (99%) and *crf II* (93%) transcripts, and a low homology to *irotensin I* (51%) transcripts (L.O.E.E., T.O.N., M.G. and V. Tronci, unpublished results). The probe was cloned using the following primers: forward 5′-TTTCTCGTGCAACCGTG-3′ and reverse 5′-ATCATTTTTCGTGGCAGGG-3′. The quality and quantity of the synthesised riboprobes were assessed by agarose gel electrophoresis. Prior to *in situ* hybridisation, the tissue was air dried at room temperature for 1 h and at 65°C for 10 min, rehydrated in a graded ethanol series (95–50%), washed 1 min with 2× SSC, then permeabilised with proteinase K (10 μg ml−1 in 0.1 mol l−1 Tris-HCl, pH 8.0) for 3.5 min, post-fixed in 4% paraformaldehyde in KPBS (137 mmol l−1 NaCl, 1.4 mmol l−1 K2HPO4, 2.7 mmol l−1 KCl, 4.3 mmol l−1 NaHPO4, pH 7.3) for 5 min, followed by rinsing 2 times 2 min in KPBS. Tissue was then treated with 0.1 mol l−1 tricholamine (TEA, pH 8.0, Sigma) for 3 min and then with 0.25% acetic anhydride (Sigma-Aldrich) in 0.1 mol l−1 TEA for 10 min. Finally, tissue was dehydrated in a graded ethanol series (50–100%) and air dried for 1 h. For hybridisation, 200 ng digoxigenin labelled probe in 100 μl hybridisation solution was applied to each slide. The composition of the hybridisation solution was: 10 mmol l−1 Tris-HCl, 300 mmol l−1 NaCl, 20 mmol l−1 EDTA, 0.2% Tween-20, 1% blocking solution (Roche Diagnostics), 0.1% dextran sulphate (Sigma-Aldrich), 50% deionised formamide (Sigma-Aldrich). Incubation was carried out at 65°C for 16 h, using humidity chambers and hybri-slips (Sigma-Aldrich) to prevent evaporation. The sense probe was applied as a control for non-specific staining. After hybridisation, tissue was washed 2 times 30 min in 2× SSC, 30 min in 50% deionised formamide in 2× SSC at 65°C, and 2 times 10 min in 2× SSC at 37°C. The tissue was treated for 20 min with RNase A (0.02 mg ml−1, Sigma-Aldrich) at 37°C, and washed 20 min at 65°C. The sections were incubated for 1 h with 2% blocking solution in 2× SSC with 0.05% Triton X-100 and then overnight with alkaline phosphatase-conjugated sheep anti-DIG goat antibody (1:2000, Roche Diagnostics). The tissue was washed 2 times 10 min in 1× maleate buffer and then 10 min in visualisation buffer (100 mmol l−1 Tris-HCl, 100 mmol l−1 NaCl, pH 9.5). The staining reaction with chromogen substrate [3.4 μl nitroblue tetrazolium, 3.5 μl 5-bromo-4-chloro-3-indoylphosphate (Roche Diagnostics) and 0.24 mg ml−1 levamisole in visualisation buffer] was carried out for 3 h in darkness at room temperature. The reaction was terminated with stop solution (10 mmol l−1 Tris-HCl, 1 mmol l−1 EDTA, 150 mmol l−1 NaCl, pH 8.0) and tissue was mounted in ProLong® Gold (Invitrogen). Photographs were taken using a digital camera (Leica DFC 320, Leica 350 FX) attached to a Leica DM 6000B microscope using the Leica Application Suite V 3.0.0 image acquisition and processing software.

**Statistics**

Values are represented as means±s.e.m. or as a Tukey box-and-whiskers plot. Differences in SGR were assessed with Mann–Whitney U-test, while body mass data were analysed with one-way ANOVA followed by Sidak’s post hoc test. Prior to analysis, data were checked for outliers, and the replicate tanks compared by ANOVA. If there were no differences between the tanks, they were pooled for further comparison of groups. Plasma cortisol levels and gene expression were evaluated using two-way ANOVA followed by a Tukey post hoc test with Bonferroni correction for multiple comparisons applied where necessary, or Fisher’s LSD test when no correction for multiple comparisons was needed (i.e. in the case of comparison of cortisol levels between control and UCS between days). Differences were considered to be statistically different from one another when P<0.05, unless otherwise stated (i.e. in case of correction for multiple comparisons).

All statistical analyses were carried out using GraphPad Prism (version 6 for Windows, GraphPad Prism Software, La Jolla, CA, USA).
Acknowledgements

We would like to thank the staff at the IMR Research station in Matre for their help in experimental design and sampling, in particular Ivar Helge Matre, Kåre Storsæter, Marita Larsen, Ole Fredrik Skulstad, Linda Oldeide, Grethe Thorshjelm, Britt Svaeren, Karen Anita Kvestad, Tone Vågseth and Remen Mette. Further, we would like to thank F. A. Tom Spannings and Thamar N. M. Pelgrim (Radboud University) for their assistance during sampling and cortisol assays, and Jan Zethof (Radboud University) for his assistance with gene expression analyses. Finally, we want to thank Valentina Tronci (Uni Research) for her help with in situ hybridisations.

Competing interests

The authors declare no competing or financial interests.

Author contributions

A.M., R.E.O., T.S.K., L.O.E.E., G.F. and M.G. conceived and designed the experiments; A.M., R.E.O., T.S.K., L.O.E.E., G.F. and M.G. carried out the experiments and data acquisition; A.M., R.E.O., T.S.K., L.O.E.E., G.F. and M.G. analysed and interpreted the data; A.M., R.E.O. and M.G. drafted the manuscript; all authors critically revised the manuscript.

Funding

This research was funded by the European Community’s Seventh Framework Programme [FP7/2010–2014] under grant agreement no. 265957 – COPEWEEL.

References


