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Identification of Novel Genetic Loci Associated with Thyroid Peroxidase Antibodies and Clinical Thyroid Disease


1 Department of Internal Medicine, Erasmus Medical Center Rotterdam, Rotterdam, The Netherlands, 2Istituto di Ricerca Genetica e Biomedica (IRGB), Consiglio Nazionale delle Ricerche, c/o Università di Monza e Brianza, Monza e Brianza, Italy, 3Division of Genetics and Cell Biology, San Raffaele Scientific Institute, Milan, Italy, 4Interfaculty Institute for Genetics and Functional Genomics, University Medicine and Ernst-Moritz-Arndt-University Greifswald, Greifswald, Germany, 5Department of Endocrinology and Diabetes, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia, 6Department of Cardiovascular Health Research Unit, Departments of Medicine, Epidemiology and Health Services, University of Washington, Seattle, Washington, United States of America, 7Institute for Genetic Epidemiology, Helmholtz Zentrum Munich, Munich/Neuherberg, Germany, 8Department of Endocrinology and Internal Medicine, University Hospital Ghent and Faculty of Medicine, Ghent University, Ghent, Belgium, 9Department of Health Evidence, Radboud University Medical Centre, Nijmegen, The Netherlands, 10Institute of Medical Epidemiology, Biostatistics, and Informatics, Martin-Luther-University Halle-Wittenberg, Halle, Germany, 11Comprehensive Cancer Center, Ohio State University, Columbus, Ohio, United States of America, 12Department of Epidemiology, Erasmus Medical Center Rotterdam, Rotterdam, The Netherlands, 13Departments of Medicine, Human Genetics, Epidemiology and Biostatistics, Lady Davis Institute, McGill University, Montreal, Canada, 14Department of Twin Research and Genetic Epidemiology, King’s College London, London, United Kingdom, 15National Institute for Health and Welfare, Helsinki, Finland, 16Hospital for Children and Adolescents, Helsinki University Central Hospital and University of Helsinki, Helsinki, Finland, 17Institute of Laboratory Medicine, Clinical Chemistry and Molecular Diagnostics, University Hospital Leipzig, Leipzig, Germany, 18Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Greifswald, Germany, 19Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, United States of America, 20Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland, 21Vaasa Health Care Centre, Diabetes Unit, Vaasa, Finland, 22Research Unit of Molecular Epidemiology Helmholtz Zentrum München - German Research Center for Environmental Health, Institute of Epidemiology II, Neuherberg, Germany, 23Department of Psychiatry and Psychotherapy, University Medicine Greifswald, HELIOS Hospital Stralsund, Greifswald, Germany, 24Pathwest Laboratory Medicine WA, Nedlands, Western Australia, Australia, 25Research Unit of Molecular Epidemiology Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany, 26School of Medicine and Pharmacology, the University of Western Australia, Crawley, Western Australia, Australia, 27School of Population Health, University of Western Australia, Nedlands, Western Australia, Australia, 28School of Pathology and Laboratory Medicine, University of Western Australia, Crawley, Western Australia, Australia, 29High Performance Computing and Network, CRIS, Parco Tecnologico della Sardegna, Pula, Italy, 30Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, United States of America, 31Department of Respiratory Medicine, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia, 32School of Medicine and Pharmacology, the University of Western Australia, Crawley, Western Australia, Australia, 33UWA Centre for Medical Research, University of Western Australian Institute for Medical Research, Perth, Western Australia, Australia, 34School of Population Health, University of Western Australia, Nedlands, Western Australia, Australia, 35MRC Lifecourse Epidemiology Unit, Southampton General Hospital, Southampton, United Kingdom, 36School of Pathology and Laboratory Medicine, University of Western Australian Institute for Medical Research, Perth, Western Australia, Australia, 37High Performance Computing and Network, CRIS, Parco Tecnologico della Sardegna, Pula, Italy, 38Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, United States of America, 39Department of Chronic Disease Prevention, National Institute for Health and Welfare, Helsinki, Finland, 40Vaasa Health Care Centre, Diabetes Unit, Vaasa, Finland, 41Department of Respiratory Medicine, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia.
Abstract

Autoimmune thyroid diseases (AITD) are common, affecting 2-5% of the general population. Individuals with positive thyroid peroxidase antibodies (TPOAbs) have an increased risk of autoimmune hypothyroidism (Hashimoto’s thyroiditis), as well as autoimmune hyperthyroidism (Graves’ disease). As the possible causative genes of TPOAbs and AITD remain largely unknown, we performed GWAS meta-analyses in 18,297 individuals for TPOAb-positivity (1769 TPOAb-positives and 16,528 TPOAb-negatives) and in 12,353 individuals for TPOAb serum levels, with replication in 8,990 individuals. Significant associations (P<5x10^-8) were detected at TPO-rs11675434, ATXN2-rs653178, and BACH2-rs10944479 for TPOAb-positivity, and at TPO-rs11675434, MAGI3-rs1230666, and KALRN-rs2010099 for TPOAb levels. Individual and combined effects (genetic risk scores) of these variants on (subclinical) hypo- and hyperthyroidism, goiter and thyroid cancer were studied. Individuals with a high genetic risk score had, besides an increased risk of TPOAb-positivity (OR: 2.18, 95% CI 1.68-2.81, P = 8.1x10^-8), a higher risk of increased thyroid-stimulating hormone levels (OR: 1.51, 95% CI 1.26-1.82, P = 2.9x10^-6), as well as a decreased risk of goiter (OR: 0.77, 95% CI 0.66-0.89, P = 6.5x10^-4). The MAGI3 and BACH2 variants were associated with an increased risk of hyperthyroidism, which was replicated in an independent cohort of patients with Graves’ disease (OR: 1.37, 95% CI 1.22-1.54, P = 1.2x10^-7 and OR: 1.25, 95% CI 1.12-1.39, P = 6.2x10^-4). The MAGI3 variant was also associated with an increased risk of hypothyroidism (OR: 1.57, 95% CI 1.18-2.10, P = 1.9x10^-3). This first GWAS meta-analysis for TPOAbs identified five newly associated loci, three of which were also associated with clinical thyroid disease. With these markers we identified a large subgroup in the general population with a substantially increased risk of TPOAbs. The results provide insight into why individuals with thyroid autoimmunity do or do not eventually develop thyroid disease, and these markers may therefore predict which TPOAb-positives are particularly at risk of developing clinical thyroid dysfunction.
Environmental Health, by the German Federal Ministry of Education and Research and by the State of Bavaria. The work of KORA is supported by the German Federal Ministry of Education and Research (BMBF) in the context of the German National Genome Research Network (NGFN-2 and NGFN-plus). The present research was supported within the Munich Center of Health Sciences (MC Health) as part of LMUinnovativ. Thyroid examinations in KORA-F4 were supported by Sanofi-Aventis in the framework of the Papillon Initiative. Collection and genotyping of the NBS samples was funded in part by the European Commission (POLYGÉN - LSHC-CT-2005-018827) and a research investment grant of the Radboud University Nijmegen Medical Care. For the use of supercomputer facilities, financial support from the NWO. The Thyroid Cancer Program (P.I. Matthew Ringel) at the Ohio State University is supported by grants P30 CA16058 and P01 CA24570 from the National Cancer Institute. USA. The generation and management of GWAS genotype data for the Rotterdam Study is supported by the Netherlands Organisation of Scientific Research NWO Investments (no. 175.010.2005.011; 911-03-012). This study is funded by the Research Institute for Diseases in the Elderly (DIX XII), the Netherlands Genomics Initiative (NGI)/Netherlands Organisation for Scientific Research (NWO) project no. 050-0600-810. The Rotterdam Study is funded by Erasmus Medical Center and Erasmus University, Rotterdam, Netherlands Organization for the Advancement of Health Research and Development (ZonMw), the Research Institute for Diseases in the Elderly (RIDE2), the Ministry of Education, Culture and Science, the Ministry for Health, Welfare and Sports, and the Municipality of Rotterdam. The SardiNIA study is supported by the Intramural Research Program of the National Institute on Aging (NIA), National Institutes of Health (NIH). The Sardinia (“Sardegna”) team was supported by Contract NO1-AG-1-2109 from the US Department of Agriculture and USDA supporting in part contract 263-MA-410053 from the NIA to the University of Michigan and by research grant Hg02651. SHIP is part of the Community Medicine Research net of the University of Greifswald, Germany, which is funded by the Federal Ministry of Education and Research (grants no. 01ZZ9603, 01ZZ1003, and 01ZZ2003), the Ministry of Cultural Affairs as well as the Social Ministry of the Federal State of Mecklenburg-West Pomerania, and the network ‘Greifswald Approach to Individualized Medicine (GANI_MED)’ funded by the Federal Ministry of Education and Research (grant 031S0261A). Genome-wide data have been supported by the Federal Ministry of Education and Research (grant no. 033ZIK012) and a joint grant from Siemens Healthcare, Erlangen, Germany, and the Federal State of Mecklenburg-West Pomerania. Data analyses were further supported by the German Research Foundation (DFG Vo 955/10-2; SPP 1629: THYROID TRANS ACT WA 1328/5-1 and the Federal Ministry of Nutrition, Agriculture and Consumer’s Safety (BMELV 07 HS 003). SHIP-Trend is part of the Community Medicine Research net of the University of Greifswald, Germany, which is funded by the Federal Ministry of Education and Research (grants no. 01ZZ9603, 01ZZ1003, and 01ZZ2003), the Ministry of Cultural Affairs as well as the Social Ministry of the Federal State of Mecklenburg-West Pomerania. TwinsUK received funding from the Wellcome Trust; the Chronic Disease Research Foundation; the European Community’s Seventh Framework Program grant agreement (FP7/2007-2013; ENGAGE project grant agreement HEALTH-F4-2007-201413); the Department of Health via the National Institute for Health Research (NIHR) Comprehensive Biomedical Research Centre award to Guy’s & St Thomas’ NHS Foundation Trust in partnership with King’s College London; the Canadian Institutes of Health Research, Canadian Foundation for Innovation, Fonds de la Recherche en Santé Québec, Ministère du Développement Économique, de l’Innovation et de l’Exportation Québec and the Lady Davis Institute of the Jewish General Hospital. The Australian National Health and Medical Research Council (Project Grant GNT1000849, 1031422) and the Sir Charles Gardiner Hospital Research Fund. Val Borbera was supported by funds from Compagnia di San Paolo, Torino, Italy; Fondazione Cariplo, Italy and Ministry of Health, Ricerca Finalizzata 2008. The UK Graves’ disease cohort was funded by the Wellcome Trust grant 068181. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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* E-mail: m.medici@erasmusmc.nl

These authors contributed equally to this work.

† SS, SN and RPP also contributed equally to this work.

Introduction

Autoimmune thyroid disease (AID), including Hashimoto’s thyroiditis and Graves’ disease, is one of the most common autoimmune diseases, affecting 2–5% of the general population [1,2,3]. Thyroid dysfunction has been associated with osteoporosis, depression, atrial fibrillation, heart failure, metabolic syndrome, and mortality [4,5,6,7,8,9,10,11]. High serum antibodies against the enzyme thyroid peroxidase (TPO), which is located in the thyroid and plays a key role in thyroid hormone synthesis, are present in 90% of patients with Hashimoto’s thyroiditis [12,13], the most frequent cause of hypothyroidism and goiter. Although TPO antibodies (TPOAbs) are a useful clinical marker for the detection of early AITD, it remains controversial if these antibodies play a causative role in the pathogenesis of Hashimoto’s thyroiditis [14,15,16].

Interestingly, TPOAb-positive persons also have an increased risk of developing autoimmune hyperthyroidism (Graves’ disease) [17,18], which is caused by stimulating antibodies against the thyroid stimulating hormone (TSH) receptor [19]. Numerous studies have shown that Graves’ hyperthyroidism and Hashimoto’s thyroiditis show co-inheritance [17,20,21]. Finally, thyroid autoimmunity is the most common autoimmune disorder in women of childbearing age, and TPOAb-positive women have an increased risk of developing pregnancy complications such as miscarriage and pre-term delivery [17,18,22,23,24,25,26].

The prevalence of TPOAb-positivity in the general population ranges from 5–24%, but it is currently unknown why these people develop TPOAbs, nor is it known why not all individuals with thyroid autoimmunity develop clinical thyroid disease [27,28]. It is estimated that around 70% of the susceptibility to develop thyroid autoantibodies is due to genetic factors [29]. In this context it is remarkable to note that little is known about the genetic factors that determine TPOAb-positivity and the risk of AITD.

We therefore performed a genome wide association study (GWAS) meta-analysis for TPOAbs in the general population in 18,297 individuals from 11 populations. Newly identified genetic variants were studied in relation to subclinical and overt hypo- and hyperthyroidism, goiter, thyroid autoimmunity during pregnancy and thyroid cancer risk.

Results

Characteristics of the studied populations are shown in Table 1 and the Supplementary Material S1. Heritability estimates in the family-based cohorts SardiNIA, TwinsUK and Val Borbera were, respectively, 0.65, 0.66, and 0.54 for TPOAb-positivity, and 0.43, 0.66, and 0.30 for TPOAb levels.

Loci associated with TPOAb-positivity and TPOAb levels

See Table 1 and Supplementary Figure S1 for TPOAb measurements and Supplementary Table S1 for genotyping procedures. In most autoimmune diseases, both the presence and the level of autoantibodies are relevant for the disease onset [18,30,31]. Furthermore, different pathophysiological processes may be involved in the initiation and severity of the autoimmune response. We therefore performed a GWAS on TPOAb-positivity (including 1769 TPOAb-positives and 16,328 TPOAb-negatives), as well as a GWAS on continuous TPOAb levels (including 12,353 individuals) in stage 1. See Supplementary Figures S2 and S3 for QQ (quantile-quantile) and Manhattan plots.

In stage 2, we followed-up 20 stage 1 SNPs (P<5×10^{-6}, 13 TPOAb-positivity and 10 TPOAb level SNPs, with 3 SNPs overlapping) in 5 populations, including up to 8,990 individuals for TPOAb-positivity (922 TPOAb-positives and 8068 TPOAb-negatives) and 8,159 individuals for TPOAb level analyses (see Supplementary Material S1). Results of the combined stage 1 and 2
Novel Thyroid Antibody and Disease Loci

Author Summary

Individuals with thyroid peroxidase antibodies (TPOAbs) have an increased risk of autoimmune thyroid diseases (AITD), which are common in the general population and associated with increased cardiovascular, metabolic and psychiatric morbidity and mortality. As the causative genes of TPOAbs and AITD remain largely unknown, we performed a genome-wide scan for TPOAbs in 18,297 individuals, with replication in 8,990 individuals. Significant associations were detected with variants at TPO, ATXN2, BACH2, MAGI3, and KALRN. Individuals carrying multiple risk variants also had a higher risk of increased thyroid-stimulating hormone levels (including subclinical and overt hypothyroidism), and a decreased risk of goiter. The MAGI3 and BACH2 variants were associated with an increased risk of hyperthyroidism, and the MAGI3 variant was also associated with an increased risk of hypothyroidism. This first genome-wide scan for TPOAbs identified five newly associated loci, three of which were also associated with clinical thyroid disease. With these markers we identified a large subgroup in the general population with a substantially increased risk of TPOAbs. These results provide insight into why individuals with thyroid autoimmunity do or do not eventually develop thyroid disease, and these markers may therefore predict which individuals are particularly at risk of developing clinical thyroid dysfunction.

Associations with goiter

As autoimmunity significantly changes during pregnancy [25], we additionally studied these effects in an independent pregnant population. Pregnant women with a high genetic risk score had a 2.4 times increased risk of TPOAb-positivity compared to women with a low score (10.3% vs 4.8%, P = 0.03). These women did not have a higher risk of increased TSH levels. However, a borderline significant signal with a lower risk of increased TSH levels was observed at ATXN2- rs633178 (OR, 0.54 [95% CI, 0.34–0.87], P = 0.012).

Associations with thyroid disease in independent populations

a) Graves’ disease. As MAGI3- rs1230666 and BACH2- rs10944479 showed promising associations (i.e., P ≤ 0.05) with hyperthyroidism in our meta-analyses, we tested these SNPs in an independent population of 2478 patients with Graves’ disease and 2682 controls (see Supplementary Material for further details). Both were associated with an increased risk of Graves’ disease (MAGI3- rs1230666: OR, 1.37 [95% CI, 1.22–1.54]; P = 1.2 × 10^{-7}; BACH2- rs10944479: OR, 1.25 [1.12–1.39]; P = 6.2 × 10^{-7}).

b) Thyroid cancer. Supplementary Table S6 shows the associations of the 5 GWAS significant SNPs with thyroid cancer. No statistically significant associations were detected, but a borderline significant signal with an increased risk of thyroid cancer was observed at ATXN2- rs653178 (OR, 1.32 [95% CI, 1.02–1.70], P = 0.03).

Pathway analyses

Ingenuity Pathway Analyses (IPA; Ingenuity Systems, Ca, USA) and GRAIL analyses [32] were performed to identify potential pathways involved in AITD, the results of which are shown in Supplementary Tables S7 and S8, and Figure S6. The identified top pathways involved cell death, survival, movement, and OX40 signalling.

Discussion

This is the first GWAS meta-analysis investigating the genetics of TPOAbs in the normal population in up to 18,297 individuals from 11 populations with replication in up to 8,990 individuals from 5 populations. We identified 5 GWAS significant loci associated with TPOAb-positivity and/or levels.

The most significant hit for both TPOAb-positivity and TPOAb levels was located near the TPO gene itself. TPO is a membrane-bound protein located on the apical membranes of the thyroid follicular cell, catalyzing key reactions in thyroid hormone synthesis [33]. Mutations in TPO have been found in patients with congenital hypothyroidism [34,35]. Although TPOAbs are

meta-analyses, including heterogeneity analyses, are shown in Supplementary Tables S2 and S3. Regional association plots are shown in Supplementary Figures S4 and S5. The variance explained by these 5 SNPs is 10.3% vs 4.8%, P = 0.03. These women did not have a higher risk of increased TSH levels. However, a borderline significant signal with a lower risk of increased TSH levels was observed at ATXN2- rs633178 (OR, 0.54 [95% CI, 0.34–0.87], P = 0.012).

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## Table 1. Population characteristics and serum TPOAb, TSH, and FT4 level measurements specifications.

<table>
<thead>
<tr>
<th>Study</th>
<th>Ethnic group (origin)</th>
<th>N with TPOAb and GWAS data</th>
<th>N using thyroid medication</th>
<th>N case-control approach (cases/controls)</th>
<th>N continuous approach</th>
<th>Men (%)</th>
<th>Age (yrs) Mean (SD)</th>
<th>TPOAb-positivity (%)</th>
<th>TPOAb-positivity cut off</th>
<th>Assay (Detection range)</th>
<th>TSH Median (IQR)</th>
<th>Assay (normal range)</th>
<th>FT4 Mean (SD)</th>
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<td>47</td>
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<td>1316</td>
<td>43%</td>
<td>53.0 (17.2)</td>
<td>15.0%</td>
<td>53.0</td>
<td>Immulite 2000 chemiluminescent immunoassay (5-5000)</td>
<td>1.3 (0.9;1.9) mU/L</td>
<td>Immulite 2000 chemiluminescent immunoassay (0.4 - 4.0 mU/L)</td>
<td>16.9 (2.5) pmol/L</td>
<td>Immulite 2000 chemiluminescent immunoassay (9 – 23 pmol/L)</td>
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<td>0</td>
<td>2024 (281/1743)</td>
<td>1817</td>
<td>41%</td>
<td>74.8 (5.1)</td>
<td>13.9%</td>
<td>74.8</td>
<td>Chemiluminescent immunoassay (5-600)</td>
<td>2.3 (1.5;3.5) mU/L</td>
<td>Chemiluminescent immunoassay (0.27-4.2 mU/L)</td>
<td>1.2 (0.2) ng/dL</td>
<td>Chemiluminescent immunoassay (0.93–1.7 ng/dL)</td>
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<td>497</td>
<td>50%</td>
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<td>15.1%</td>
<td>61.0</td>
<td>Chemiluminescent immunoassay (0-1000)</td>
<td>2.0 (1.2;2.4) mU/L</td>
<td>Chemiluminescent immunoassay (0.49–4.67 mU/L)</td>
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<td>49</td>
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<td>1475</td>
<td>45%</td>
<td>60.5 (8.9)</td>
<td>5.0%</td>
<td>60.5</td>
<td>Chemiluminescent immunoassay (1–3000)</td>
<td>1.5 (0.6;2.5) mU/L</td>
<td>Chemilumineous immuneimunoassay (0.4–4.3 mU/L)</td>
<td>18.9 (2.6) pmol/L</td>
<td>Chemiluminescent immunoassay (11–25 pmol/L)</td>
</tr>
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<td>26</td>
<td>1829 (287/1542)</td>
<td>1829</td>
<td>50%</td>
<td>61.5 (10.3)</td>
<td>15.7%</td>
<td>61.5</td>
<td>Fluoro-immunometric assay (2.6–1000)</td>
<td>1.3 (0.9;2.0) mU/L</td>
<td>Immuno- luminometer assay (0.4–4.0 mU/L)</td>
<td>13.5 (2.4) pmol/L</td>
<td>Chemiluminescent immunoassay (8.0–220 pmol/L)</td>
</tr>
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<td>50</td>
<td>1577 (137/1440)</td>
<td>210</td>
<td>40%</td>
<td>70.2 (5.6)</td>
<td>8.7%</td>
<td>70.2</td>
<td>Chemiluminescent immunoassay (5-5000)</td>
<td>1.2 (0.6;2.5) mU/L</td>
<td>Chemiluminescent immunoassay (0.4–4.3 mU/L)</td>
<td>18.4 (2.4) pmol/L</td>
<td>Chemiluminescent immunoassay (11–25 pmol/L)</td>
</tr>
<tr>
<td>SardiNIA</td>
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<td>154</td>
<td>972 (108/864)</td>
<td>1257</td>
<td>49%</td>
<td>56.9 (12.5)</td>
<td>11.1%</td>
<td>56.9</td>
<td>Chemiluminescent immunoassay (5-1000)</td>
<td>1.3 (0.8;2.0) mU/L</td>
<td>Chemiluminescent immunoassay (0.4–4.0 mU/L)</td>
<td>1.3 (0.2) ng/dL</td>
<td>Chemiluminescent immunoassay (0.3–2.4 ng/dL)</td>
</tr>
<tr>
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<td>Caucasian (Germany)</td>
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<td>293</td>
<td>3803 (265/3538)</td>
<td>1818</td>
<td>52%</td>
<td>49.3 (16.3)</td>
<td>7.0%</td>
<td>49.3</td>
<td>Chemiluminescent immunoassay (1–3000)</td>
<td>0.7 (0.4;1.0) mU/L</td>
<td>Chemiluminescent immunoassay (0.3–3.0 mU/L)</td>
<td>12.8 (3.8) pmol/L</td>
<td>Chemiluminescent immunoassay (7.7–23.2 pmol/L)</td>
</tr>
<tr>
<td>SHIP-Trend</td>
<td>Caucasian (Germany)</td>
<td>986</td>
<td>99</td>
<td>887 (36/851)</td>
<td>887</td>
<td>46%</td>
<td>49.5 (13.7)</td>
<td>4.1%</td>
<td>49.5</td>
<td>Chemiluminescent immunoassay (1–3000)</td>
<td>1.2 (0.8;1.6) mU/L</td>
<td>Chemiluminescent immunoassay (0.36–3.74 mU/L)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>TwinsUK</td>
<td>Caucasian (UK)</td>
<td>2455</td>
<td>86</td>
<td>2369 (461/1893)</td>
<td>774</td>
<td>0%</td>
<td>46.9 (12.5)</td>
<td>19.5%</td>
<td>46.9</td>
<td>Chemiluminescent immunoassay (0.5–1000)</td>
<td>1.3 (0.9;1.8) mU/L</td>
<td>Chemiluminescent immunoassay (0.4-4.0 mU/L)</td>
<td>13.6 (1.9) pmol/L</td>
<td>Chemiluminescent immunoassay (9-19 pmol/L)</td>
</tr>
<tr>
<td>ValBorbera</td>
<td>Caucasian (Italy)</td>
<td>1661</td>
<td>90</td>
<td>1571 (161/1410)</td>
<td>452</td>
<td>46%</td>
<td>54.3 (18.4)</td>
<td>10.2%</td>
<td>54.3</td>
<td>Two chemiluminescent immunoassays (5.5–3000 ; 6-7500)</td>
<td>1.4 (0.9;2.0) mU/L</td>
<td>Chemiluminescent immunoassay (0.34–5.60 mU/L)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Study</td>
<td>Ethnic group (origin)</td>
<td>N with TPOAb and GWAS data</td>
<td>N using thyroid medication</td>
<td>N case-control approach (cases/controls)</td>
<td>N continuous approach</td>
<td>Age (yrs) Mean (SD)</td>
<td>TPOAb-positivity (%)</td>
<td>TPOAb-positivity cut-off</td>
<td>TPOAb specifications</td>
<td>TSH specifications</td>
<td>FT4 specifications</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>--------------------</td>
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<td>----------------------</td>
<td>---------------------</td>
<td>---------------------</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Asklepios</td>
<td>Caucasian (Belgium)</td>
<td>2418</td>
<td>109</td>
<td>2309 (245/2064)</td>
<td>2185</td>
<td>50%</td>
<td>45.9 (5.9)</td>
<td>10.6%</td>
<td>35</td>
<td>Chemiluminescent immunoassay (5–600)</td>
<td>Chemiluminescent immunoassay (0.3–4.2 mU/L)</td>
<td>1.5 (1.1; 2.1) mU/L</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CARLA</td>
<td>Caucasian (Germany)</td>
<td>1753</td>
<td>270</td>
<td>1483 (186/1297)</td>
<td>1190</td>
<td>60%</td>
<td>64.2 (10.2)</td>
<td>12.5%</td>
<td>28</td>
<td>Chemiluminescent immunoassay (5–600)</td>
<td>Chemiluminescent immunoassay (0.4–3.8 mU/L)</td>
<td>0.9 (0.6; 1.2) mU/L</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EFSOCH</td>
<td>Caucasian (UK)</td>
<td>1289</td>
<td>-</td>
<td>1289 (97/1192)</td>
<td>1233</td>
<td>64%</td>
<td>34.2 (5.9)</td>
<td>7.5%</td>
<td>34</td>
<td>Chemiluminescent immunoassay (5–600)</td>
<td>Chemiluminescent immunoassay (0.4–4.5 mU/L)</td>
<td>1.9 (1.3; 2.6) mU/L</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Health2006 Study</td>
<td>Caucasian (Danish)</td>
<td>3287</td>
<td>-</td>
<td>3287 (204/3083)</td>
<td>3285</td>
<td>45%</td>
<td>49.3 (13.0)</td>
<td>6.2%</td>
<td>100</td>
<td>Chemiluminescent immunoassay (1–3000)</td>
<td>Chemiluminescent immunoassay (0.4–3.7 mU/L)</td>
<td>1.7 (1.0; 2.0) mU/L</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SardiNIA2</td>
<td>Caucasian (Italy)</td>
<td>1387</td>
<td>30</td>
<td>765 (104/661)</td>
<td>375</td>
<td>41%</td>
<td>46.6 (17.4)</td>
<td>13.6%</td>
<td>35</td>
<td>Chemiluminescent immunoassay (5–1000)</td>
<td>Chemiluminescent immunoassay (0.4–4.0 mU/L)</td>
<td>1.6 (1.0; 2.2) mU/ml</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

TSH Median (IQR)  TSH (normal range)  FT4 Mean (SD)  Assay (normal range)

Chemiluminescent immunoassay (0.3–1.7 ng/dl) 1.3 (0.2) ng/dL  Chemiluminescent immunoassay (0.8–2.0 ng/dL) 16.3 (2.5) pmol/L  Chemiluminescent immunoassay (12.8–20.4 pmol/L) 16.0 (2.4) pmol/L  Chemiluminescent immunoassay (11–24 pmol/L) 12.4 (1.8) pmol/L  Chemiluminescent immunoassay (9.8–18.8 pmol/L) 1.3 (0.2) ng/dL  Chemiluminescent immunoassay (0.3–2.4 ng/dl)
valid clinical biomarkers ofAITD, they are generally considered to be secondary to the thyroid damage inflicted by T-cells. The FOXE1 gene has been previously associated with hypothyroidism [36,37] and is known to regulate transcription of TPO [38]. In this context it is interesting to note that we did not find any associations of the variant near TPO with hypothyroidism. Most genes that have been associated with AITD (predominantly Graves’ disease) by candidate gene and GWAS studies so far are located in the HLA class I and II regions, or in genes involved in T-cell [i.e., CTLA-4, PTPN22] or other autoimmune responses [28,39]. Until now, the TPO gene itself had not been associated with AITD, except in one recent candidate gene analysis in a small cohort (n = 188) without replication [40]. A variant near TPO (rs11675434), which is in LD with rs11675434 (r² = 0.97 in HapMap2), has previously been associated with TSH levels by Hashimoto’s thyroiditis and Graves’ disease [55]. However, the association signal at rs2476601 was not significant when Hashimoto’s thyroiditis and Graves’ disease were studied separately. BACH2 is specifically expressed in early stages of B-cell differentiation and represses different immunoglobulin genes [56]. Interestingly, BACH2 can bind to the co-repressor SMRT (silencing mediator of retinoid and thyroid receptor), which may suggest a more direct effect on thyroid hormone secretion and action as well. Polymorphisms in ATXN2 have been associated with multiple neurodegenerative diseases, including spinocerebellar ataxia and Parkinson’s disease [57,58,59]. Different epidemiological studies have associated thyroid dysfunction with cerebellar ataxia [60,61]. Furthermore, the identified SNP in ATXN2 has been previously associated with renal function, serum urate levels and blood pressure [62,63,64]. However, this SNP is in high LD with ATXN2. This is supported by a recent study which showed that variants in LD with SH2B3, BACH2, and PTPN22 are associated with TPOAb levels in patients with type 1 diabetes [66]. Whereas the above four loci are located in genes involved in the immune response or the autoantigen, the KALRN (Kalirin) gene encodes the adaptor protein 3 (SH2B3). SH2B3 encodes the adaptor protein LNK, a key negative regulator of cytokine signaling playing a critical role in hematopoiesis. This variant is associated with susceptibility to several autoimmune diseases, including celiac disease, type 1 diabetes, vitiligo, and rheumatoid arthritis [46,51,53,65], suggesting more relevance for TPOAb levels than ATXN2. This is supported by a recent study which showed that variants in LD with SH2B3, BACH2, and PTPN22 are associated with TPOAb levels in patients with type 1 diabetes [66].

### Table 2. Newly identified loci associated with TPOAb-positivity and/or serum TPOAb levels reaching genome wide significance.

<table>
<thead>
<tr>
<th>TPOAb-positivity</th>
<th>SNP Chr. Position (Build 36)</th>
<th>Risk</th>
<th>Other</th>
<th>RAF*</th>
<th>Nearby gene</th>
<th>OR (95% CI) [b]</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs11675434</td>
<td>2</td>
<td>1386822</td>
<td>T</td>
<td>C</td>
<td>0.39</td>
<td>TPO</td>
<td>1.21 (1.15–1.28)</td>
</tr>
<tr>
<td>rs653178</td>
<td>12</td>
<td>110492139</td>
<td>C</td>
<td>T</td>
<td>0.40</td>
<td>ATXN2</td>
<td>1.14 (1.08–1.19)</td>
</tr>
<tr>
<td>rs10944479</td>
<td>6</td>
<td>90937114</td>
<td>A</td>
<td>G</td>
<td>0.16</td>
<td>BACH2</td>
<td>1.25 (1.14–1.37)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>TPOAb levels</th>
<th>SNP Chr. Position (Build 36)</th>
<th>Risk</th>
<th>Other</th>
<th>RAF*</th>
<th>Nearby gene</th>
<th>OR (95% CI) [b]</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs11675434</td>
<td>2</td>
<td>1386822</td>
<td>T</td>
<td>C</td>
<td>0.39</td>
<td>TPO</td>
<td>0.0202 (0.0046)</td>
</tr>
<tr>
<td>rs1230666</td>
<td>1</td>
<td>113974933</td>
<td>A</td>
<td>G</td>
<td>0.16</td>
<td>MAGI3</td>
<td>0.0269 (0.0064)</td>
</tr>
<tr>
<td>rs2010099</td>
<td>3</td>
<td>125782947</td>
<td>C</td>
<td>T</td>
<td>0.91</td>
<td>KALRN</td>
<td>0.0240 (0.0076)</td>
</tr>
</tbody>
</table>

Chr., chromosome
*RAF: Relative Allele Frequency
[b]Adjusted for age and gender
Expresed in sd of natural logarithm transformed serum TPOAb level, adjusted for age and gender
doi:10.1371/journal.pgen.1004123.t002
development of AITD [44,45]. Another top pathway involved was the OX40 signalling pathway, and it is of interest to note that OX40 is a T-cell activator promoting the survival of CD4+ T-cells at sites of inflammation [69].

Our results have potential clinical relevance for several reasons. Genetic risk scores based on these novel common (risk allele frequencies: 9–40%) TPOAb-associated SNPs enabled us to identify a large subgroup in the general population with a two-fold...
increased risk of TPOAb-positivity (10.4% vs 5.4%). These individuals also have a higher risk of increased TSH levels and a lower risk of goiter, suggesting an advanced stage of destruction of the thyroid due to autoimmune processes. Furthermore, pregnant women with high genetic risk scores had a 2.4 times increased risk of TPOAb-positivity during pregnancy. In this context it is interesting to note that TPOAb-positive pregnant women have an increased risk of miscarriages and preterm births independent of thyroid function [70].

Associations with thyroid disease were also found on an individual SNP level. The MAGI3 SNP was associated with a substantially increased risk of hypothyroidism, and the BACH2 SNP showed a borderline significant association ($P = 0.011$) with a higher risk of increased TSH levels, which includes subjects with subclinical and overt hypothyroidism. Furthermore, both loci were significantly associated with an increased risk of Graves’ hyperthyroidism in an independent population. To predict which patients with first or second degree relatives with documented Hashimoto’s or Graves’ disease will develop clinical thyroid disease, a clinical algorithm has been developed (i.e., the THEA score) [18]. Future studies should analyze if these genetic markers increase the sensitivity of the THEA score. Graves’ hyperthyroidism and Hashimoto’s thyroiditis co-segregate in families and subjects with TPOAbs have an increased risk of both diseases [17,18,20,21,22,26]. The current study provides insight into this phenomenon by showing that specific loci associated with TPOAbs and (subclinical) hypothyroidism, i.e. MAGI3 and BACH2, are also associated with Graves’ hyperthyroidism in an independent case-control study.

The prevalence of TPOAb-positivity in the general population is high (5–24%), but it is currently unknown why part of the individuals with thyroid autoimmunity develop clinical thyroid disease whereas others do not [27,28]. In this context it is interesting to note that the TPOAb-associated SNPs located in TPO and ATXN2 were not associated with clinical thyroid disease. This suggests that the TPOAbs in these individuals may be of less clinical relevance, providing insight into why TPOAb-positive individuals do or do not eventually develop clinical thyroid disease.

Our study has some limitations. The validity of the results is restricted to individuals from populations of European ancestry. Future GWASs in populations from non-European descent will be required to determine to which extent our results can be generalized to other ethnic groups. Secondly, we did not perform conditional analyses to further identify secondary association signals within the identified loci, nor did we perform functional studies for the identified variants. Further research is therefore needed to unravel the exact biological mechanism behind the observed associations. The fact that various TPOAbs assays were used across the participating cohorts could lead to bias. We
therefore used TPOAb-positivity cut-off values as provided by the respective assay manufacturer, instead of using one fixed cut-off value. This is also of clinical importance as in clinical practice most institutions rely on the TPOAb-positivity cut-off as provided by the assay manufacturer. Furthermore, we did not detect heterogeneity in our results, supporting the fact that results obtained with different assays can be combined across cohorts using the z-score based meta-analysis. Finally, as AITD coincides with other autoimmune diseases, our results could be driven by indirect heterogeneity in our results, supporting the fact that results obtained with different assays can be combined across cohorts using the z-score based meta-analysis. Finally, as AITD coincides with other autoimmune diseases, our results could be driven by indirect associations with other autoimmune diseases. However, AITD is the most common autoimmune disease in the general population. We furthermore show that carriage of multiple risk alleles is associated with an increased risk of thyroid dysfunction, which underlines the clinical importance of our findings.

In conclusion, this first GWAS for TPOAbs identified five newly associated loci, three of which were also associated with clinical thyroid disease. Furthermore, we show that carriage of multiple risk variants is not only associated with a substantial increased risk of TPOAb-positivity, but also with a higher risk of increased TSH levels (including subclinical and overt hypothyroidism) and a lower risk of goiter. These genetic markers not only help to identify large groups in the general population with an increased risk of TPOAb-positivity, but may also predict which TPOAb-positive persons are particularly at risk of developing clinical thyroid disease.

**Materials and Methods**

**Study cohorts**

For the TPOAb GWAS stage 1 and 2 analyses, and the hypothyroidism, hyperthyroidism and goiter analyses, individuals were recruited from 16 independent community-based and family studies. For the Graves’ disease analyses, cases were recruited from the United Kingdom Graves’ disease cohort and controls from the British 1958 Birth Cohort. Thyroid cancer cases and controls were recruited from the Nijmegen and Ohio thyroid cancer cohorts. A detailed description of the original cohorts contributing samples is provided in Table 1 and in the Supplementary Material. All participants provided written informed consent and protocols were approved by the institutional review boards or research ethics committees at the respective institutions, and conducted according to the Declaration of Helsinki.

**Phenotype definitions**

Serum TPOAb levels were determined with a range of assays. TPOAb-positives were defined as subjects with TPOAb levels above the assay-specific TPOAb-positivity cut-off, as defined by the manufacturer (Table 1). Serum TSH and free thyroxine (FT4) levels were determined using a range of assays (Table 1). Assay-specific TSH and FT4 reference ranges were used, as provided by the manufacturer (Table 1). Overt hypothyroidism was defined as a high TSH (i.e., a TSH level above the TSH reference range) and a low FT4. Increased TSH was defined as an elevated TSH, including persons with overt hyperthyroidism or subclinical hypothyroidism (i.e., high TSH with a normal FT4). Overt hyperthyroidism was defined as a low TSH and a high FT4. Decreased TSH was defined as a low TSH, including persons with subclinical or overt hyperthyroidism.

The diagnosis of goiter is described in the Supplementary Material, and the diagnosis of Graves’ disease and thyroid cancer in the respective cohorts have been described previously [41].

**Genotyping**

Samples were genotyped with a range of GWAS genotyping arrays (Supplementary Table S1). Sample and SNP quality control procedures were undertaken within each study. For each GWAS, over 2.5 million SNPs were imputed using CEU samples from Phase 2 of the International HapMap project (www.hapmap.org). Genotyping procedures in the stage 2, Graves’ disease and thyroid cancer populations are described in the Supplementary Material.

**Association analyses**

The heritabilities of TPOAb-positivity and serum TPOAb levels were estimated, as described in the Supplementary Material.

In stage 1, we performed a GWAS on TPOAb-positivity as well as a GWAS on continuous TPOAb levels. Persons taking thyroid medication were excluded. Each SNP was tested for association with TPOAb-positivity using logistic regression analyses, adjusting for age and sex. For cohorts with family structure, we approximated the probability of being affected with a linear mixed model adjusting for age and sex. The produced model was used to predict the expected proportion of “risk” (effective) alleles in cases and controls, hence giving the means to estimate odds ratios. Only unrelated individuals were considered for the SardiNIA cohort. For the GWAS of continuous TPOAb levels, samples with a TPOAb level lower than the minimum TPOAb assay detection limit (Table 1) were excluded. TPOAb levels were natural log-transformed, and sex-specific, age adjusted standardized residuals were calculated. Each SNP was tested for association with these TPOAb level residuals using linear regression analyses (additive model).

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**Table 5.** Newly identified TPOAb associated loci, genetic risk scores and the risk of goiter.

<table>
<thead>
<tr>
<th>Nearby gene</th>
<th>SNP</th>
<th>Risk allele</th>
<th>Other allele</th>
<th>OR (95% CI)*</th>
<th>P value</th>
<th>GRS Quartile</th>
<th>% Goiter (N cases/total)</th>
<th>OR (95% CI)*</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>TPO</td>
<td>rs11675434</td>
<td>T</td>
<td>C</td>
<td>0.95 (0.88–1.02)</td>
<td>0.17</td>
<td>1 (reference)</td>
<td>35.2% (588/1669)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>ATXN2</td>
<td>rs653178</td>
<td>C T</td>
<td></td>
<td>0.95 (0.88–1.03)</td>
<td>0.22</td>
<td>2</td>
<td>33.7% (570/1691)</td>
<td>0.92 (0.79–1.06)</td>
<td>0.21</td>
</tr>
<tr>
<td>BACH2</td>
<td>rs10944479</td>
<td>A G</td>
<td></td>
<td>0.94 (0.85–1.05)</td>
<td>0.28</td>
<td>3</td>
<td>31.6% (530/1675)</td>
<td>0.84 (0.72–0.98)</td>
<td>0.03</td>
</tr>
<tr>
<td>MAGI3</td>
<td>rs1230666</td>
<td>A G</td>
<td></td>
<td>0.90 (0.81–1.00)</td>
<td>0.05</td>
<td>4</td>
<td>30.4% (517/1702)</td>
<td>0.77 (0.66–0.89)</td>
<td>6.5 × 10⁻⁴</td>
</tr>
<tr>
<td>KALRN</td>
<td>rs2010099</td>
<td>C T</td>
<td></td>
<td>0.93 (0.81–1.05)</td>
<td>0.23</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Adjusted for age, gender, and body surface area.

**ATXN2-rs653178 is in high LD with SH2B3-rs3184504.**

**MAGI3-rs1230666 is in high LD with PTPN22-rs24756601.**

**do10.1371/journal.pgen.1004123.t005**
correcting for relatedness in studies with family structure. See Supplementary Table S1 for the software used for these analyses.

Before meta-analysis, SNPs with a minor allele frequency (MAF) \(< 1\%\) or a low imputation quality were excluded (Supplementary Material). Results from stage 1 and 2 were combined in a population size weighted z-score based meta-analysis using METAL [71]. Genomic control was applied to individual studies if \(k > 1.0\).

In stage 2, we followed-up stage 1 GWAS significant SNPs, as well as promising SNPs not reaching GWAS significance, in an attempt to reach GWAS significant associations by increasing sample size (Supplementary Material). Results from stage 1 and 2 were combined in a population size weighted z-score based meta-analysis using METAL [71]. A z-score based meta-analysis was used to reduce bias that might be induced by different assays. As this method does not provide betas, and we wanted to provide a rough estimate of the actual effect sizes for convenience, we calculated betas using the fixed effects (inverse variance based) meta-analysis method. Heterogeneity was tested, applying bonferroni based P-value thresholds of \(P=0.004\) for the TPOAb-positivity analyses and \(P=0.005\) for the TPOAb level analyses.

All studies assessed and, if present, corrected for population stratification using principal-component analysis (PCA) and/or multidimensional-scaling (MDS), with the exception of SardiNIA and ValBorbera where the high isolation substantiates a lack of stratification (Table S1) [72,73]. Lambda values were all \(<1\), indicating that population stratification was overall properly accounted for (Table S1). To fully remove residual effects, we applied genomic correction to studies were lambda was \(> 1\). The final meta-analyses reported a lambda of 1.01 for both the TPOAb-positivity and the TPOAb level GWAS, thus no genomic correction was applied.

The variances explained by the GWAS significant SNPs were calculated. We subsequently studied the individual as well as the combined effects of the GWAS significant SNPs on the risk of clinical thyroid disease, as specified in the Supplementary Material. In short, to study combined effects, a genetic risk score was calculated for every person as the weighted sum of TPOAb positivity and the TPOAb level GWAS, thus no genomic correction was applied.

Various bioinformatic tools were searched for evidence for functional relevance of the GWAS significant SNPs and pathway analyses were performed on the Stage 1 lead SNPs (see Supplementary Material).

Supporting Information

Figure S1  TPOAb level distributions in persons with detectable TPOAb levels in stage 1 and 2 populations.

Figure S2  Quantile-quantile (QQ) plots for the TPOAb-positivity and TPOAb level stage 1 meta-analyses.

Figure S3  Manhattan plots for stage 1 meta-analyses for TPOAb-positivity (a) and TPOAb levels (b). SNPs are plotted on the x-axis according to their chromosomal position against TPOAb-positivity (a) or TPOAb levels (b) (shown as –log\(_{10}\) P value) on the y-axis. The horizontal grey line indicates the threshold for genome-wide statistical significance \((P<5 \times 10^{-8})\).

Genome-wide significant associations were observed near TPO (Chr 2p25; \(P=1.5 \times 10^{-12}\), at ATXN2 (Chr 12q24.1; \(P=1.6 \times 10^{-10}\)) and near HCP5 (Chr 6p21.3; \(P=4.1 \times 10^{-8}\)) for TPOAb-positivity, and near TPO (Chr 2p25; \(P=5.4 \times 10^{-15}\)) at ATXN2 (Chr 12q24.1; \(P=1.1 \times 10^{-8}\)) for TPOAbs levels.

Figure S4  Regional association plots of stage 1 lead loci for TPOAb-positivity (panels a-m). The y-axis on the left indicates the –log\(_{10}\) P value for the association with TPOAb–positivity. SNPs are plotted on the x-axis according to their chromosomal position.

The most significant stage 1 SNP is indicated in purple. The combined stage 1 and 2 result of this SNP is indicated in yellow.

The SNPs surrounding the most significant SNP are color-coded to reflect their LD with this SNP. Symbols reflect functional genomic annotation, as indicated in the legend. The blue y-axes on the right of each plot indicate the estimated recombination rates (based on HapMap Phase II); the bottom of each panel shows the respective annotated genes at the locus and their transcriptional direction. Mb, megabases.

Figure S5  Regional association plots of stage 1 lead loci for TPOAb levels (panels a-j). The y-axis on the left indicates the –log\(_{10}\) P value for the association with TPOAb levels. SNPs are plotted on the x-axis according to their chromosomal position.

The most significant stage 1 SNP is indicated in purple. The combined stage 1 and 2 result of this SNP is indicated in yellow.

The SNPs surrounding the most significant SNP are color-coded to reflect their LD with this SNP. Symbols reflect functional genomic annotation, as indicated in the legend. The blue y-axes on the right of each plot indicate the estimated recombination rates (based on HapMap Phase II); the bottom of each panel shows the respective annotated genes at the locus and their transcriptional direction. Mb, megabases.

Figure S6  GRAIL results for the stage 1 TPOAb-positivity and TPOAb level lead SNPs. GRAIL circle plot of locus connectivity where each locus is plotted in a circle, where significant connections \((P<0.05)\) based on PubMed abstracts are drawn spanning the circle. Analyses were based on the 20 stage 1 TPOAb-positivity and TPOAb level lead SNPs.

Table S1  Study sample genotyping, quality control and association analyses for stage 1 populations.

Table S2  Associations of stage 1 lead SNPs with TPOAb-positivity in stage 1 and 2.

Table S3  Associations of stage 1 lead SNPs with serum TPOAb levels in stage 1 and 2.

Table S4  Stage 1 TPOAb-positivity and TPOAb level meta-analyses results for GWAS significant SNPs reported in previous GWAS on thyroid related phenotypes.

Table S5  Genetic risk score and the risk of increased TSH levels.

Table S6  Newly identified TPOAb associated loci and the risk of thyroid cancer.
Table S7 Top IPA associated canonical pathways for the Stage 1 TPOAb-positivity and TPOAb level lead SNPs. (DOCX)

Table S8 Top IPA associated canonical pathways for the Stage 1 TPOAb-positivity and TPOAb level lead SNPs. (DOCX)

Text S1 Supplementary methods. (DOCX)

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Author Contributions

Conceived and designed the experiments: MM SJR RAJ RR AA HJG ER JIR HH LC DTB BV TDm Tj JGE Bmp AhO DS HW AdIc TMF AL LR KAK AGU Jpw KS EwAc CMe MdH TjVs TjD SgW Hv AC Dto SS SN RPP. Performed the experiments: MM EP GP AT LC SJR RAJ RR GLR TSP SHV JL MJ S llN Nh RmF BS BM GS YA AL TjVs SS SN RPP. Analyzed the data: MM EP GP AT SJR RAJ RR GLR TSP SHV JL MJ S llN Nh RmF BS BM GS LC EB HL GM LA SM NS MC MN CSp AR MH EML ER PJL SLa MV GA EwAd AP AD APb Dwpw JBp AM TF Aj JH HP EeF SFJ JIR AK DR GLS EB HH JAf BV TDm Tj JGE Pco Arh Bmp MtI AhO HW Adic RtNm SclG HmAs TMF AL Fr AGU Jpw CMe JtV TjDs SgW Hv Ac Dto RPP. Wrote the paper: MM AT LCjVs SgW Ac SS SN RPP.

References


