Function and Regulation of the Na\(^{+}\)-Ca\(^{2+}\) Exchanger NCX3 Splice Variants in Brain and Skeletal Muscle

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Background: The Na\(^{+}\)-Ca\(^{2+}\) exchanger isoform 3 (NCX3) participates in maintaining calcium homeostasis in brain and skeletal muscle.

Results: The two splice variants of NCX3 have distinct regulation and capacity of exchange.

Conclusion: Distinct calcium uptake capacity of muscle NCX3 suggests a novel function in muscle exercise physiology.

Significance: This study brings a new understanding of the role of NCX3 variants in excitable tissues.

Isoform 3 of the Na\(^{+}\)-Ca\(^{2+}\) exchanger (NCX3) is crucial for maintaining intracellular calcium ([Ca\(^{2+}\)]\(_i\)) homeostasis in excitable tissues. In this sense NCX3 plays a key role in neuronal excitotoxicity and Ca\(^{2+}\) extrusion during skeletal muscle relaxation. Alternative splicing generates two variants (NCX3-AC and NCX3-B). Here, we demonstrated that NCX3 variants display a tissue-specific distribution in mice, with NCX3-B as mostly expressed in brain and NCX-AC as predominant in skeletal muscle. Using Fura-2-based Ca\(^{2+}\) imaging, we measured the capacity and regulation of the two variants during Ca\(^{2+}\) extrusion and uptake in different conditions. Functional studies revealed that, although both variants are activated by intracellular sodium ([Na\(^{+}\)]\(_i\)), NCX3-AC has a higher [Na\(^{+}\)] sensitivity, as Ca\(^{2+}\) influx is observed in the presence of extracellular Na\(^{+}\). This effect could be partially mimicked for NCX3-B by mutating several glutamate residues in its cytoplasmic loop. In addition, NCX3-AC displayed a higher capacity of both Ca\(^{2+}\) extrusion and uptake compared with NCX3-B, together with an increased sensitivity to intracellular Ca\(^{2+}\). Strikingly, substitution of Glu\textsuperscript{580} in NCX3-B with its NCX3-AC equivalent Lys\textsuperscript{580} recapitulated the functional properties of NCX3-AC regarding Ca\(^{2+}\) sensitivity, Lys\textsuperscript{580} presumably acting through a structure stabilization of the Ca\(^{2+}\) binding site. The higher Ca\(^{2+}\) uptake capacity of NCX3-AC compared with NCX3-B is in line with the necessity to restore Ca\(^{2+}\) levels in the sarcoplasmic reticulum during prolonged exercise. The latter result, consistent with the high expression in the slow-twitch muscle, suggests that this variant may contribute to the Ca\(^{2+}\) handling beyond that of extruding Ca\(^{2+}\).

The Na\(^{+}\)-Ca\(^{2+}\) exchanger (NCX)\(^2\) is a plasma membrane transporter that plays a major role in the maintenance of Ca\(^{2+}\) homeostasis in various cell types (1, 2). NCX catalyzes the exchange of Na\(^{+}\) and Ca\(^{2+}\) with a 3:1 stoichiometry (3). Depending on the electrochemical gradient across the plasma membrane NCX can either extrude intracellular Ca\(^{2+}\) in its forward mode or take up extracellular Ca\(^{2+}\) in its reverse mode. The NCX protein contains 10 transmembrane segments and a large central intracellular loop (4–6). The latter was shown to be responsible for the allosteric modulation of the exchanger by Na\(^{+}\) and Ca\(^{2+}\) (7). A high intracellular Na\(^{+}\) concentration ([Na\(^{+}\)]\(_i\)) or low intracellular Ca\(^{2+}\) concentration ([Ca\(^{2+}\)]\(_i\)) inactivates the reverse mode of the exchanger (7–9).

In mammals, the NCX family consists of three separate genes: NCX1, NCX2, and NCX3 (10). NCX1 is predominantly expressed in heart, kidney, and brain (11), NCX2 is most abundantly expressed in brain (12), and NCX3 is expressed in excitable tissues such as brain and skeletal muscle (13). Numerous splice variants of NCX1 exist in various species, whereas there is only a single variant described for NCX2 (12). Finally, for NCX3, six variants have been described in human (12, 14). In mice, two variants of NCX3 have been identified, but data regarding their expression pattern are lacking. It has, therefore, been hypothesized that expression of mouse NCX3 variants follows the same tissue distribution as that observed in rat (12). This would imply that in mice NCX3-B is expressed in brain, and NCX3-AC expression is restricted to the adult skeletal muscle. These two NCX3 variants are derived from an alternative splicing of the exons A, B, and C. Given the fact that exons A and B are mutually exclusive, the alternative splicing introduces a difference of 24 residues between the NCX3-B and NCX3-AC variants. All of these residues are located within the cytoplasmic regulatory loop known to regulate the activity of the exchanger (12).

The role of NCX3 in brain has been extensively studied. In neurons, NCX3 plays an important role in pathological situations such as ischemia and excitotoxicity (15, 16). In skeletal
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muscle, NCX3 action constitutes the major Ca\(^{2+}\) extrusion mechanism during the relaxation process (17). Its absence induces muscle necrosis and aberrant Ca\(^{2+}\) homeostasis (18). However, little is known about the regulation and functional properties of NCX3-AC and NCX3-AC.

The aim of the present study was to gain insight into the properties and physiological role of NCX3-AC. To this end, the expression of both variants in various mouse tissues was assessed. Next, we investigated whether the amino acid differences in the regulatory loop of both NCX3 variants have functional consequences. Therefore, NCX3-AC and NCX3-BC were expressed at the plasma membrane in human embryonic kidney (HEK293T) cells. Subsequently, the regulation of the NCX3 variants by [Ca\(^{2+}\)], and [Na\(^{+}\)], was studied using Fura-2-based Ca\(^{2+}\) imaging. Finally, the same approach was applied to site-directed mutants to investigate the molecular mechanisms that give rise to the differences in exchange activity.

**EXPERIMENTAL PROCEDURES**

**cDNA Cloning**—Mice NCX3-B (kindly provided by Prof. Geerten Vuister, Leicester, UK) was subcloned into the XhoI and NheI sites of the bicistronic mammalian vector pcCINeo containing an internal ribosome entry site and enhanced green fluorescent protein. An HA tag was added to the N terminus of the NCX protein. Fragments corresponding to exons AC or B were obtained by amplification of cDNA from mouse hind limb skeletal muscle using a high fidelity DNA polymerase (Phusion, Thermo Fisher). The uncut fragment, containing only NCX3-AC, after digestion with XmnI was subcloned into the BlpI and NheI sites of the bicistronic mammalian vector pCINeo.

**Expression of NCX3**—Mice NCX3-B and NCX3-AC were expressed in human embryonic kidney (HEK293T) cells. Subsequently, the regulation of the NCX3 variants by [Ca\(^{2+}\)] and [Na\(^{+}\)] was studied using Fura-2-based Ca\(^{2+}\) imaging. Finally, the same approach was applied to site-directed mutants to investigate the molecular mechanisms that give rise to the differences in exchange activity.

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**Mutagenesis**—All NCX3-B mutants were generated using a QuikChange site-directed mutagenesis kit (Stratagene, La Jolla, CA) following the manufacturer’s instructions. All mutants were subsequently validated by Sanger sequencing.

**Expression Profile and Quantitative Real-time Polymerase Chain Reaction Analysis**—Three C57BL/6 mice were sacrificed. The brain, heart, and four different skeletal muscles from the hind limb were collected: gastrocnemius, soleus, extensor digitorum longus, and tibialis anterior. Tissue RNA was extracted using TRIzol total RNA isolation reagent (Life Technologies BRL, Breda, The Netherlands). After DNase treatment (Promega, Madison, WI), 1 μg of RNA was reverse-transcribed by Moloney murine leukemia virus reverse transcriptase (Invitrogen) as described previously (19). Using a CFX96 Real time PCR detection system (Bio-Rad), three calibration curves corresponding to NCX3-AC, NCX3-B, and total NCX3 were realized using serial dilutions of pCINeo-NCX3-B and pCINeo-NCX3-AC. The cDNA from tissues were used to measure the absolute copy number per μg of mRNA. The sequences were amplified using the following primers: total NCX3, 5’-ATTAGGGGAGCTGGATCTTCA-3’ (forward) and 5’-CTGGAGATTACACGGAGGC-3’ (reverse), NCX3-AC, 5’-GGGCCCCCGCATGGTGGATA-3’ (forward) and 5’-CAGCTTCTCTGTCTGACCTTCTGGA-3’ (reverse), and NCX3-B, 5’-GCATTAGGGGAGCTGGAGGTCTTCA-3’ (forward) and 5’-GTTCCACCAAGGGCAGAATTGAAG-3’ (reverse).

**Expression of Na\(^{+}/Ca\(^{2+}\) Exchange in Human Embryonic Kidney Cells**—Human embryonic kidney cells (HEK293T) were grown in Dulbecco’s modified Eagle’s medium (Bio Whittaker-Europe, Verviers, Belgium) containing 10% (v/v) fetal calf serum and 2 mM L-glutamine at 37 °C with 5% (v/v) CO\(_2\). The cells were seeded on a six-well plate and transiently transfected with the respective constructs (2 μg/well) using polyethyleneimine cationic polymer PEI (Polysciences, Inc., Warrington, PA) in accordance with the manufacturer’s instructions. The HEK293T cells were used for the experiments 48 h after transfection.

**Cell Surface Biotinylation**—Cell surface labeling with biotin was performed using the sulfo-NHS-LC-LC-biotin (0.5 mg/ml, Thermo Fisher Scientific, Rockford, IL) as described previously (20). Cells from each six-well plate were homogenized on ice in 0.5 ml of lysis buffer (150 mM NaCl, 5 mM EDTA, 0.5% (v/v) Triton X-100 (v/v), protease inhibitors, 50 mM Tris/HCl, pH 7.5) to prevent endo- and exocytosis, as described previously (21). Subsequently 5% (v/v) of the total protein amount was collected as an input sample. Next, biotinylated proteins (plasma membrane fraction) were precipitated using Neutravidin-agarose beads (Thermo Fisher). Biotinylated fractions were eluted in 2× Laemmli buffer containing 100 mM dithiothreitol and protease inhibitors (22). Finally, samples were denatured for 30 min at 37 °C. NCX3-AC and NCX3-AC expression was analyzed by immunoblot analysis for the input and the plasma membrane fraction (23).

**Immunoblot Analysis**—Lysates were subjected to SDS-PAGE 8% (w/v) and electroblotted onto PVDF membranes. Blots were incubated with 5% (w/v) nonfat dried milk in TBS-T (137 mM NaCl, 0.2% (v/v) Tween 20, and 20 mM Tris/HCl, pH 7.6). Immunoblots were incubated overnight at 4 °C with a rabbit anti-NCX3 antibody (1:3000) (kindly provided by Prof. Kenneth Phillipson, UCLA) (24) diluted in PBS-T. Immunoblots were incubated overnight at 4 °C with a rabbit anti-NCX3 antibody (1:3000) (kindly provided by Prof. Kenneth Phillipson, UCLA) (24) diluted in 1% (w/v) milk in TBS-T. PVDF membranes were incubated 1 h at room temperature with a sheep horseradish peroxidase-conjugated anti-rabbit (1:10000) (Sigma) in TBS-T. Afterward, blots were visualized using the enhanced chemiluminescence system (ECL, Thermo Fisher).

**[Ca\(^{2+}\)]\(_{i}\) and [Ca\(^{2+}\)]\(_{ER}\) Measurements**—48 h after transfection, cells were loaded with 3 μM Fura-2 acetoxymethyl ester (Fura-2/AM) and 0.01% (v/v) Pluronic F-127 for 20 min at 37 °C in Krebs solution (5.5 mM KCl, 147 mM NaCl, 1.2 mM MgCl\(_2\), 1.5 mM CaCl\(_2\), 10 mM glucose, and 10 mM HEPES/NaOH, pH 7.4). Then, the cells were washed with Krebs medium for 10 min. Finally the coverslips were placed into a perfusion chamber mounted onto the stage of an inverted microscope (Zeiss Axiovert 200 M, Carl Zeiss, Jena, Germany). Changes in medium and addition of inhibitors were facilitated using a perfusion system.

**[Ca\(^{2+}\)]\(_{i}\) was monitored by exciting Fura-2 with monochromatic light of wavelength 340 and 380 nm (Polychrome IV, TILL Photonics, Gräfelfing, Germany). Fluorescence emission light was directed by a 415DCLP dichroic mirror (Omega Optical, Inc., Brattleboro, VT) through a 510WB40 emission filter (Omega Optical, Inc.) onto a CoolSNAP HQ monochrome CCD camera.
Analysis of the \([Ca^{2+}]_i\) Measurements—After initiation of the reverse mode, varying \(Ca^{2+}\) uptakes were observed in the different conditions. For each condition, the baseline value was calculated as an average of the first 60 s of recording. During investigation of the forward mode, the baseline Fura-2 ratio recorded in mock-expressing cells in \(Ca^{2+}\)-free medium was subtracted from the maximal Fura-2 ratio in the same cells after ionomycin treatment. This calculation represents the maximum amount of \(Ca^{2+}\) that is present in the ER and that could potentially be extruded via NCX3. Next, the experimental extrusion of \(Ca^{2+}\) was calculated as follow: the difference between the mock and the given condition (NCX3-AC, NCX3-B, or mock) both after addition of ionomycin. This difference was reported as a percentage of the theoretical maximal extrusion value described above. This difference is subsequently expressed as a percentage of the theoretical maximal value. During the experiments in absence of thapsigargin or ouabain, the cells responding to a removal of extracellular \(Na^+\) were quantified, by taking the Fura-2 ratio value of 0.4 as a threshold of responsiveness.
FIGURE 1. NCX3 splice variants and their tissue expression. A, alternative splicing of the mice Na\(^+\)/Ca\(^{2+}\) exchanger isoform 3 pre-mRNA and the two variants obtained after alternative splicing of the exons A, B, and C. B, alignment of amino acid sequences in alternatively spliced NCX3 exons in the mice NCX3-AC and NCX3-B using ClustalW software. Missing residues are indicated by dashes. Symbols below the sequence indicate conserved (period) highly conserved (colon) and identical residues (asterisk). C, NCX3 mRNA levels in different mice tissues. Total NCX3 (black), NCX3-B (gray), and NCX3-AC (white) transcript expression levels were quantified by real-time PCR. mRNA copy numbers were calculated from the NCX3 standard curves generated by using a diluted pCINeo-IRES-eGFP-mNCX3-AC and pCINeo-IRES-eGFP-mNCX3-B vectors. Heart has been used as a negative control to show the absence of NCX3. eGFP, enhanced GFP; IRES, internal ribosome entry site. EDL, extensor digitorum longus.

FIGURE 2. NCX3 variants exhibit functional differences in their reverse mode. A, Ca\(^{2+}\) influx in HEK293T cells expressing NCX3-AC (●), NCX3-B (▲), or an empty vector (□) and loaded with Fura-2-AM. Cells were perfused with Krebs medium (147 mM Na\(^+\)). At 100 s, internal calcium stores were depleted by applying 1 \(\mu\)M thapsigargin. At 245 s, reverse NCX mode was initiated by perfusing with a Na\(^+\)-free medium (NMDG medium). The 340/380 nm emission ratios are shown. Each point represents the mean of the data, studied in three independent experimental sessions for a number (n) of cells monitored. Mock, n = 126; NCX3-B, n = 96; NCX3-AC, n = 106. B, mean values of the maximum Fura-2 ratio after addition of thapsigargin (1 \(\mu\)M) shown in A. C, mean values of the maximum Fura-2 fluorescence shown in A after removal of extracellular Na\(^+\). *, p < 0.05 (B and C). D, quantification and representative immunoblots of total cellular NCX3 at the plasma membrane (cell surface) and in the total cell lysates (Input) in cells expressing NCX3-AC, NCX3-B, or an empty vector (Ctrl). For each condition, cells that were not treated with biotin are included for comparison (n = 4 per condition).
both variants were similarly expressed at the plasma membrane (Fig. 2d).

**Forward Exchange Activity of NCX3-AC Is Most Sensitive to Intracellular Ca\(^{2+}\)**—Furthermore, to explain the reduced increase in [Ca\(^{2+}\)], upon addition of thapsigargin in NCX3-AC-expressing cells (Fig. 2b), we hypothesized that the forward mode of NCX3-AC would be more active, leading to a smaller apparent increase in [Ca\(^{2+}\)], compared with NCX3-B-expressing cells. In the absence of extracellular Na\(^+\) and Ca\(^{2+}\), conditions under which the forward mode is abolished, the Ca\(^{2+}\) level in the ER and the change in [Ca\(^{2+}\)], in response to thapsigargin, were similar in all cells (Fig. 3, a–d).

To strengthen our hypothesis, the forward mode was triggered by perfusing the cells with the Ca\(^{2+}\) ionophore ionomycin (1 \(\mu\)M) in the presence of extracellular Na\(^+\). Under these conditions, ionomycin allows for a complete release of Ca\(^{2+}\) in a fast manner from the ER to the cytoplasm, thereby inducing an activation of NCX in its forward mode of exchange. Upon addition of ionomycin, an increase in [Ca\(^{2+}\)], was recorded in the control condition (Fig. 3e). This rise in [Ca\(^{2+}\)], was greatly diminished in both variants of NCX3, confirming the extrusion of Ca\(^{2+}\) via the forward mode of the exchanger. The forward mode activity, expressed as the ability to extrude Ca\(^{2+}\), was significantly larger for NCX3-AC than for NCX3-B (78% ± 9% and 47% ± 11%, respectively, \(p < 0.05\)) (Fig. 3f).

The Activity of NCX3-AC in Reverse Exchange Is Higher Than NCX3-B—Fig. 2, a and c, show that NCX3-B and NCX3-AC can both operate in the reverse mode and that NCX3-AC induces a larger increase in [Ca\(^{2+}\)]. The capacity of the reverse mode of the NCX3 variants was then investigated by adding thapsigargin in the absence of extracellular Na\(^+\), thus preventing a contribution from NCX3 in its forward mode of exchange. The reverse mode activity, expressed as the ability to extrude Ca\(^{2+}\), was significantly larger for NCX3-AC-expressing cells than for NCX3-B (78% ± 9% and 47% ± 11%, respectively, \(p < 0.05\)) (Fig. 3f).

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**DISCUSSION**

In the present study, we show, for the first time, functional recordings of the variant NCX3-AC. These recordings indicate a higher capacity of this variant in reverse and forward mode of exchange compared with NCX3-B. A difference that can be fully attributed to the intrinsic properties of both variants considering that the plasma membrane expression in HEK293T cells did not differ between the two variants and that the Ca\textsuperscript{2+} level in the endoplasmic reticulum were similar in these cells as proven by the thapsigargin addition in absence of extracellular...
Na⁺ and Ca²⁺. Furthermore, the reduced [Ca²⁺], measured in NCX3-AC-expressing cells upon thapsigargin addition compared with NCX3-B, together with the activation of NCX3-AC reverse mode in absence of an elevation in [Ca²⁺], suggest a higher sensitivity of NCX3-AC to changes in [Ca²⁺]. Thereby, a value slightly above the [Ca²⁺], recorded during the resting state is sufficient to trigger the exchanger.

The lower sensitivity of NCX3-B to [Ca²⁺], in both modes of exchange, could be suppressed by mutating the glutamate residue at position 580. This gain of sensitivity observed in the E580K NCX3-B mutant reveals that the activation by Ca²⁺ of the forward and reverse mode of the exchanger is regulated through the same site for which the residue 580 represents a key residue. In NCX1, the equivalent residue Lys585 is involved in the formation of a salt bridge that, in Ca²⁺-free conditions, stabilizes the Ca²⁺-binding site I and II through the preservation of the tertiary structure (29, 32). NCX1 mutation of this lysine into a glutamate causes a drop in the Ca²⁺ affinity (29, 32). In NCX3-AC, Lys⁵⁸⁰ confers to the site I of the CBD2 a high affinity for Ca²⁺ (31–33). In contrast, this variant has two Ca²⁺-binding sites in its Ca²⁺-binding domain 2, whereas isothermal titration calorimetry measurements showed that NCX3-B has three Ca²⁺-binding sites (30, 33). However, none of these Ca²⁺ sites featured a high affinity for Ca²⁺ (33). Our data concur with the literature in suggesting that during a rise in [Ca²⁺], the high-affinity site I of NCX3-AC CBD2 will bind Ca²⁺ at a lower concentration than NCX3-B. This could result in a disruption of the salt bridge at Lys⁵⁸⁰ as hypothesized for NCX1 (29) and, therefore, in an activation of the exchanger at lower [Ca²⁺].

Another important finding from the current study is that NCX3-B displayed a higher Na⁺-dependent inactivation, given that only the reverse mode of NCX3-AC could be activated, during a rise in [Na⁺] in presence of extracellular Na⁺. The partial suppression of this effect by mutating glutamate resi-
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FIGURE 5. NCX3 variants exhibit distinct sensitivities to [Na\(^+\)]. A, Fura-2 ratio (340 nm/380 nm) in HEK293T cells expressing NCX3-AC (▴), NCX3-B (▲), or an empty vector (○) after 1-h treatment with ouabain (1 µM). Each point represents the mean of the data, for a number (n) of cells monitored. Mock, n = 124; NCX3-B, n = 152; NCX3-AC, n = 176. B and C, mean values of Fura-2 ratio recorded at the baseline (6). Shown is the average of the first 60 s of the experiment shown in A, in 147 mM Na\(^+\) medium, and maximum Fura-2 ratio recorded after the switch to Na\(^+\)-free medium shown in A (C) (*, p < 0.05).

The tissue-specific distribution observed for the two variants of NCX3, with NCX3-B as mostly expressed in the brain and with NCX3-AC as the main variant detected in skeletal muscle, is consistent with the functional properties measured for these variants. In neurons, NCX3-B and its low capacity to perform Ca\(^{2+}\) uptake would have a protective effect against severe Ca\(^{2+}\) overload, a phenomenon that could have dramatic consequences on neurons. With this respect, the low sensitivity to an increase in [Na\(^+\)] would confer this preservation during the firing of neurons. In the same manner, activation of the forward mode of NCX3-B requires a substantial rise in [Ca\(^{2+}\)], This feature would have the same protective effect and yet will not interfere with the synaptic activity, highly dependent on [Ca\(^{2+}\)], (37).

In skeletal muscle, sensitivity to [Ca\(^{2+}\)], and the high capacity of NCX3-AC in the forward mode corroborates with the necessity in such fibers to activate the Ca\(^{2+}\) extrusion as soon as the contraction starts. Moreover, the high sensitivity of NCX3-AC to changes in [Na\(^+\)], and its high capacity to perform Ca\(^{2+}\) uptake in reverse mode suggest that NCX3-AC might have additional implications via its reverse mode. The operation and relevance of the reverse mode has been previously observed for several members of the NCX family in numerous tissues, in both physiological (38–40) and pathological situations (41, 42). From another angle, during prolonged exercise, extracellular Ca\(^{2+}\) has been shown to have a beneficial effect on the resistance to muscle fatigue, an effect that could only be observed in slow-twitch muscle, similar to soleus, dedicated to this type of prolonged exercise (43, 44). Together with the high expression of NCX3-AC observed in soleus, these results suggest that NCX3-AC might be implicated in Ca\(^{2+}\) influx in slow-twitch muscle, a hypothesis consistent with the findings from Danielli-Betto’s group (44). Here, the Ca\(^{2+}\)-free conditions provoke a dramatic increase of muscle fatigue in soleus, an effect also obtained by inhibition of NCX. This is also in accordance with the increased fatigability observed in NCX3\(^{-/-}\) mice (18). The underlying mechanism linking Ca\(^{2+}\) uptake and muscle fatigue is still poorly understood but presumably involves a refilling of the SR Ca\(^{2+}\). Many variants of the NCX family are already known to contribute to this latter process (45, 46), particularly...
in excitable cells (47–49). Therefore, NCX3 could in principle be involved in this process in slow-twitch muscle. Nevertheless, our study was only performed in HEK 293 cells, albeit a well used cell model to study mechanistic properties and ionic regulation of NCX isoforms (35, 50, 51). Therefore, confirmation of the latter hypothesis needs further investigations using muscular excitable tissue, in particular in soleus.

In conclusion, two variants of NCX3, which depict a tissue-specific distribution among excitable tissues, have distinct properties with regard to capacity of exchange and sensitivity to $[\text{Ca}^{2+}]_i$ and $[\text{Na}^+]_i$, with NCX3-AC functioning with a greater capacity in all situations. Our data indicated that NCX3-AC is the main variant expressed in skeletal muscle with a high expression in soleus, a muscle in which NCX have been shown to be implicated in the resistance to fatigue (44). Additionally, our results are, to the best of our knowledge, the first functional recordings of NCX3-AC and provides evidence of a high capacity to achieve $\text{Ca}^{2+}$ influx that can be triggered by a simple

**FIGURE 6. Glutamate residues are key regulators of $\text{Ca}^{2+}$ and $\text{Na}^+$ sensitivity in NCX3-AC.**

A, alignment of the amino acid sequences of the mutually exclusive exon A/B of NCX3-B to NCX3-AC. Missing residues are indicated by dashes. Specific NCX3-B single mutants were generated from corresponding amino acid of NCX3-AC (indicated by an arrow). *, $p < 0.05$ (B–E). B, mean values of the maximum Fura-2 ratio recorded during forward mode followed by the addition of ionomycin ($1 \mu M$) in $\text{Ca}^{2+}$-free conditions in cells expressing NCX3-AC, NCX3-B, or mutants of NCX3-B; NCX3-B, $n = 108$; NCX3-AC, $n = 116$; E573D NCX3-B, $n = 93$; E574K NCX3-B, $n = 110$; E575A NCX3-B, $n = 82$; E580K NCX3-B, $n = 90$; E588G NCX3-B, $n = 110$. C, mean values of the maximum Fura-2 ratio in reverse mode triggered by the switch to $\text{Na}^+$-free medium containing $1 \mu M$ thapsigargin. Values were recorded in HEK293T cells expressing NCX3 variants and mutants of NCX3-B; NCX3-B, $n = 102$; NCX3-AC, $n = 120$; E573D NCX3-B, $n = 77$; E574K NCX3-B, $n = 110$; E575A NCX3-B, $n = 121$; E580K NCX3-B, $n = 86$; E588G NCX3-B, $n = 102$. D and E, Fura-2 ratio recorded during reverse mode triggered by 1-h treatment with ouabain ($1 \mu M$) in NCX3-AC, NCX3-B, and NCX3-B mutants expressing HEK293T cells. NCX3-B, $n = 99$; NCX3-AC, $n = 88$; E573D NCX3-B, $n = 86$; E574K NCX3-B, $n = 76$; E575A NCX3-B, $n = 122$; E580K NCX3-B, $n = 123$; E588G NCX3-B, $n = 99$. D, mean values of Fura-2 ratio at the baseline and average of the first 60 s of the experiment in 147 mM $\text{Na}^+$ medium. E, mean values of the maximum Fura-2 ratio after the switch to $\text{Na}^+$-free medium during the same experiment as described in D.
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increase in [Na⁺]. This exchanger might, therefore, play a role in the resistance to muscle fatigability observed in the slow-twitch muscle. Altogether, this study provides insights into the ionic regulation of NCX3 through its alternative splicing in a tissue-specific manner. In fact, the two variants may contribute to Ca²⁺ handling beyond their role in Ca²⁺ extrusion.

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